

Modeling Calcium Wave Based on Anomalous Subdiffusion of Calcium Sparks in Cardiac Myocytes

Xi Chen, Jianhong Kang, Ceji Fu, Wenchang Tan*

State Key Laboratory of Turbulence and Complex Systems and Department of Mechanics and Aerospace Engineering, College of Engineering, Peking University, Beijing, People's Republic of China

Abstract

Ca^{2+} sparks and Ca^{2+} waves play important roles in calcium release and calcium propagation during the excitation-contraction (EC) coupling process in cardiac myocytes. Although the classical Fick's law is widely used to model Ca^{2+} sparks and Ca^{2+} waves in cardiac myocytes, it fails to reasonably explain the full-width at half maximum (FWHM) paradox. However, the anomalous subdiffusion model successfully reproduces Ca^{2+} sparks of experimental results. In this paper, in the light of anomalous subdiffusion of Ca^{2+} sparks, we develop a mathematical model of calcium wave in cardiac myocytes by using stochastic Ca^{2+} release of Ca^{2+} release units (CRUs). Our model successfully reproduces calcium waves with physiological parameters. The results reveal how Ca^{2+} concentration waves propagate from an initial firing of one CRU at a corner or in the middle of considered region, answer how large in magnitude of an anomalous Ca^{2+} spark can induce a Ca^{2+} wave. With physiological Ca^{2+} currents (2pA) through CRUs, it is shown that an initial firing of four adjacent CRUs can form a Ca^{2+} wave. Furthermore, the phenomenon of calcium waves collision is also investigated.

Citation: Chen X, Kang J, Fu C, Tan W (2013) Modeling Calcium Wave Based on Anomalous Subdiffusion of Calcium Sparks in Cardiac Myocytes. PLoS ONE 8(3): e57093. doi:10.1371/journal.pone.0057093

Editor: Yulia Komarova, University of Illinois at Chicago, United States of America

Received: October 28, 2012; **Accepted:** January 17, 2013; **Published:** March 6, 2013

Copyright: © 2013 Chen et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by the National Natural Science Foundation of China (Grant No. 11272014 and No. 10825208), and National Key Basic Research Program of China (Grant No. 2013CB531200). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: tanwch@mech.pku.edu.cn

Introduction

Nomenclature

x, y	spatial coordinates, μm
t	time, ms
β	fractional order of the spatial derivative.
D_{C_x}, D_{C_y}	Ca^{2+} diffusion coefficients along x -axis and y -axis, $\mu\text{m}^2 \cdot \text{ms}^{-1}$
$[Ca^{2+}]$	free Ca^{2+} concentration, μM
$[Ca^{2+}]_{\infty}$	resting Ca^{2+} concentration, μM
$[CaF]$	Ca-bound fluo-3 concentration, μM
$[F]_T$	total fluo-3 concentration, μM
$[CaB_n]$	Ca-bound endogenous buffer concentration, μM
$[B_n]_T$	total endogenous buffer concentration, μM
k_F^+, k_n^+	forward rate constants for dye and endogenous buffer reactions, $\mu\text{M}^{-1} \cdot \text{s}^{-1}$
k_F^-, k_n^-	reverse rate constants for dye and endogenous buffer reactions, s^{-1}
l_x, l_y	spatial separation of CRUs along x -axis and y -axis, μm
I_{CRU}	current through the CRU, pA
F	Faraday's constant, $\text{C} \cdot \text{mol}^{-1}$
K_{pump}	SR pump Michaelis constant, μM
V_{pump}^{max}	maximum SR pump rate, $\text{mM} \cdot \text{s}^{-1}$
m	SR pump Hill coefficient
n	CRU Hill coefficient

σ	molar flux of a clustered RyR channel, $\text{mM} \cdot \text{s}^{-1}$
T_{open}	open time of CRU, ms
S	stochastic switching function equaling either 0 or 1
K	Ca^{2+} sensitivity parameter, μM
P	probability of Ca^{2+} spark occurrence, / calcium release unit/ms
P_{max}	maximum probability of Ca^{2+} spark occurrence, / calcium release unit/ms
v_x	wave velocity along x -axis, mm/s
v_y	wave velocity along y -axis, mm/s

In the endoplasmic or sarcoplasmic reticulum (SR) of cardiac cells, there stores plenty of Ca^{2+} , the concentration of which is 2–3 orders of magnitude greater than that in the cytosol. During the excitation-contraction (EC) coupling process, triggered by L-type Ca^{2+} channels, Ca^{2+} is released from SR through ryanodine receptors (RyRs) on the z-lines [1–4], where RyR is one kind of Ca^{2+} release units (CRUs). This event is called “ Ca^{2+} spark”. Ca^{2+} -induced Ca^{2+} release (CICR) makes RyRs fire in succession such that Ca^{2+} concentration rises [1,5], the process of which is called calcium transient. Physiologically, calcium homeostasis is important for the contraction and relaxation of the heart muscle. However, in some pathological conditions, spontaneous propagating wave of Ca^{2+} may occur, which is called “calcium wave”. The occurrence of calcium wave can affect the heart's normal

function, and may induce some disease, such as ventricular arrhythmias [6].

The model of Ca^{2+} spark using Fick's Law failed to reproduce the full-width at half maximum (FWHM) of experimental results for Ca^{2+} sparks. Simulated results for Ca^{2+} spark based on Fick's Law presented a lower FWHM ($\sim 1.0 \mu\text{m}$), which was only half the width of experimental result ($\sim 2.0 \mu\text{m}$). Izu et al. [7] tried to increase the current through RyR to get larger FWHM, however, the spark amplitude also increased (~ 10 times), which is far beyond experimental results and physiological conditions. In contrast, the results obtained with the anomalous subdiffusion model of Ca^{2+} spark were found to be in close agreement with the experimental ones so that the "FWHM Paradox" was successfully explained [8–10]. Therefore, it is confirmed that diffusion of Ca^{2+} in cytoplasm obeys no longer Fick's Law, but the anomalous subdiffusion.

A Ca^{2+} wave is formed from propagation of Ca^{2+} sparks. According to the results for Ca^{2+} sparks, Ca^{2+} wave should also obey the anomalous subdiffusion. However, all previous work on Ca^{2+} waves were based on Fick's Law [11–14]. Anisotropic Ca^{2+} diffusion was studied by Girard et al. [11]. Keizer and Smith [12] investigated Ca^{2+} waves under stochastic firing of CRUs. Izu et al. [13] combined large CRU currents [7], stochastic firing of CRUs, asymmetric distribution of CRUs and anisotropic Ca^{2+} diffusion to investigate the propagation of Ca^{2+} waves. Lu et al. [14] studied the effect of rogue RyRs on Ca^{2+} waves in ventricular myocytes with heart failure.

In this work, we develop a mathematical 2D model based on anomalous subdiffusion of Ca^{2+} sparks. The anomalous subdiffusion model is used to study Ca^{2+} waves propagation from an initial firing of one CRU at a corner or in the middle of the considered region. We reproduce wave velocities of experimental results using a small current through CRUs which is close to the physiological conditions. The phenomenon of calcium waves collision is also investigated. With physiological Ca^{2+} currents (2 pA) through CRUs, an initial firing of four adjacent CRUs is shown to form a Ca^{2+} wave. Furthermore, study on how the system becomes unstable is also performed by changing the transverse distance of CRUs.

Methods

0.1 Anomalous Diffusion Model for Calcium Waves

Figure 1 shows a 2-dimensional schematic of a cardiac myocyte (establishing line resources along z -axis [13]) which contains plenty of CRUs. The regular intervals of CRUs are l_x along x -axis and l_y along y -axis. The governing equation for Ca^{2+} waves based on the anomalous subdiffusion model can be expressed as

$$\frac{\partial[\text{Ca}^{2+}]}{\partial t} = D_x \frac{\partial^\beta[\text{Ca}^{2+}]}{\partial x^\beta} + D_y \frac{\partial^\beta[\text{Ca}^{2+}]}{\partial y^\beta} + J_{dye} + J_{buffer} + J_{pump} + J_{leak} + J_{CRU}, \quad (1)$$

where $[\text{Ca}^{2+}]$ is the free Ca^{2+} concentration; D_x and D_y are diffusion coefficients for anisotropic diffusion with $D_x = 0.30 \text{mm}^2 \text{ms}^{-1}$ and $D_y = 0.15 \text{mm}^2 \text{ms}^{-1}$ [15]; J_{dye} and J_{buffer} are fluxes due to Ca^{2+} fluorescent indicator dye and endogenous stationary buffers; J_{pump} is pumping rate of SR Ca^{2+} -ATPase, and J_{leak} is a SR leak that is to balance J_{pump} ; The

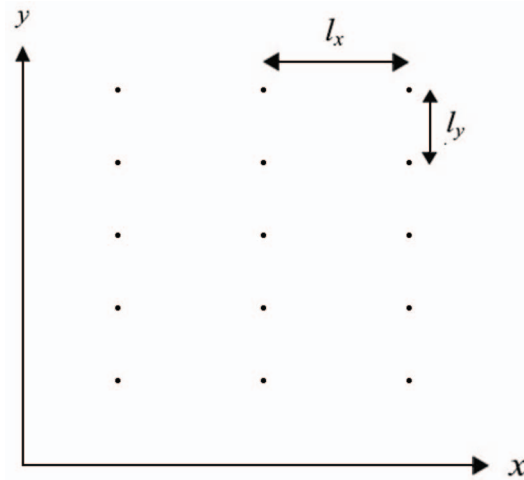


Figure 1. The 2D model of a cardiac myocyte. The black dots represent CRUs which distribute regularly spaced l_x along x -axis and l_y along y -axis.

doi:10.1371/journal.pone.0057093.g001

expressions of J_{dye} , J_{buffer} , J_{pump} , and J_{leak} are

$$J_{dye} = -k_F^+ [\text{Ca}^{2+}] ([F]_T - [\text{CaF}]) + k_F^- [\text{CaF}], \quad (2)$$

$$J_{buffer} = \sum_n J_n, \quad (3)$$

$$J_n = -\frac{\partial[\text{CaB}_n]}{\partial t}, \quad (4)$$

$$\frac{\partial[\text{CaB}_n]}{\partial t} = k_n^+ [\text{Ca}^{2+}] ([B_n]_T - [\text{CaB}_n]) - k_n^- [\text{CaB}_n], \quad (5)$$

$$J_{pump} = -\frac{V_{pump}^{max} ([\text{Ca}^{2+}])^m}{(K_{pump})^m + ([\text{Ca}^{2+}])^m}, \quad (6)$$

$$J_{leak} = -J_{pump}([\text{Ca}^{2+}]_\infty) = \frac{V_{pump}^{max} ([\text{Ca}^{2+}]_\infty)^m}{(K_{pump})^m + ([\text{Ca}^{2+}]_\infty)^m}, \quad (7)$$

where n identifies each of buffer species. $[F]_T$ and $[B_n]_T$ represent total concentration of the indicator and buffers, respectively. $[\text{CaF}]$ and $[\text{CaB}_n]$ are concentration of the Ca^{2+} -bound complexes. k_F^+ , k_F^- , k_n^+ and k_n^- are reaction kinetics. K_{pump} is the affinity constant for SR pumps, m the Hill constant, and V_{pump}^{max} the maximum rate. SR leak is used to balance J_{pump} in resting state.

J_{CRU} is the flux of Ca^{2+} release from CRU, the expression of which is the same as that of Izu et al. [13],

$$J_{CRU} = \sigma \sum_{ij} \delta(x - x_i) \delta(y - y_j) S(x_i, y_j, t; T_{open}), \quad (8)$$

where $\sigma = 0.64 I_{CRU} / 2F$ is a molar flux of a clustered RyR

channel(I_{CRU} is current through the CRU and F is Faraday's constant), and δ the Dirac delta function, S a stochastic function which controls the firing of the CRU, and T_{open} the firing time. Within a time interval Δt , the probability that the CRU fires is $P(C(x,y,t),K,n)\Delta t$, where $P(C(x,y,t),K,n) = P_{max}[Ca^{2+}]^n / (K^n + [Ca^{2+}]^n)$ with $P_{max} = 0.3/CRU/ms$ the maximum probability of Ca^{2+} spark occurrence, $K = 15mM$ Ca^{2+} the sensitivity parameter and $n = 1.6$ the Hill coefficient.

The anomalous space diffusion is model used in Eq.(1), where $\frac{\partial^\beta}{\partial x^\beta}$ is the Riemann-Liouville operators which is defined as

$$\frac{\partial^\beta [Ca^{2+}]}{\partial x^\beta} = \frac{1}{\Gamma(k-\beta)} \frac{\partial^k}{\partial x^k} \int_0^x \frac{[Ca^{2+}]}{(x-\tau)^{1+\beta-k}} d\tau, \quad (9)$$

$$\frac{\partial^\beta [Ca^{2+}]}{\partial y^\beta} = \frac{1}{\Gamma(k-\beta)} \frac{\partial^k}{\partial y^k} \int_0^y \frac{[Ca^{2+}]}{(y-\tau)^{1+\beta-k}} d\tau, \quad (10)$$

where k is an integer with $\beta < k < \beta + 1$ ($1 < \beta < 3$). In Eq.(1), when $1 < \beta < 2$, anomalous space superdiffusion occurs, while anomalous space subdiffusion occurs when $2 < \beta < 3$. Particularly, our model reduces to Fick's Law when $\beta = 2$. According to Li et al. [10], calcium sparks follow the anomalous space subdiffusion of $\beta = 2.25$, so we only consider the space subdiffusion in the following sections.

0.2 Numerical Methods

Our simulation is performed on a rectangular region with size of $20\mu m \times 20\mu m$, which is meshed with a uniform grid size of $0.1\mu m \times 0.1\mu m$. The time-step size is 0.005ms.

For the fractional differential term, we used the right-shifted Grünwald formula to make a finite difference approximation [16].

$$\frac{\partial^\beta f(x,y,t)}{\partial x^\beta} = \frac{1}{\Gamma(-\beta)} \lim_{N_x \rightarrow \infty} \frac{1}{h_x^\beta} \sum_{k=0}^{N_x} \frac{\Gamma(k-\beta)}{\Gamma(k+1)} f(x-(k-1)h), \quad (11)$$

$$\frac{\partial^\beta f(x,y,t)}{\partial y^\beta} = \frac{1}{\Gamma(-\beta)} \lim_{N_y \rightarrow \infty} \frac{1}{h_y^\beta} \sum_{k=0}^{N_y} \frac{\Gamma(k-\beta)}{\Gamma(k+1)} f(y-(k-1)h), \quad (12)$$

where N_x and N_y are positive integers, and $h_x = x/N_x$, $h_y = y/N_y$. Γ denotes the gamma function. The shifted Grünwald approximation for fractional order derivative has been shown to be unconditionally stable [16].

Considering simple impermeability of the cell boundary to the diffusing ions, reflecting boundary conditions $\partial C / \partial n|_{boundary} = 0$ are taken on all edges [14]. The scale of our computation time is 200–500ms so that a CRU would not reopen after firing and closing.

Standard values of parameters used in the current study are listed in Table 1 and Table 2. β , I_{CRU} and l_x are changeable parameters whose effects on the results will be investigated.

Results and Discussion

Modeling a Ca^{2+} wave from a Single Ca^{2+} Spark

Ca^{2+} waves have been shown to be initiated and sustained by Ca^{2+} sparks [17]. Under pathological conditions, Ca^{2+} sparks fire spontaneously and stochastically, so whether a single spark can

Table 1. Standard parameter values.

Parameter	Value
D_{C_x}	0.30
D_{C_y}	0.15
l_y	0.8
$[Ca^{2+}]_\infty$	0.1
F	96500
V_{pump}^{max}	208
K_{pump}	0.184
K	15
m	3.9
n	1.6
T_{open}	10
P_{max}	0.3

doi:10.1371/journal.pone.0057093.t001

trigger a Ca^{2+} wave is important to the stability of cardiac myocytes [4]. Simulations based on Fick's Law reveal that large currents through CRUs and high calcium concentrations are needed to trigger a Ca^{2+} wave [13]. In this work, based on the anomalous subdiffusion model, we find the current which can trigger a normal Ca^{2+} wave initiating from a single Ca^{2+} spark at the corner of considered region.

According to Li et al. [10], calcium sparks follow the anomalous space subdiffusion of $\beta = 2.25$, so this subdiffusion order is also taken in our simulation. In our model, initial source is a 10ms opening of one CRU, the longitudinal intervals are $l_x = 2.0\mu m$. When we take $I_{CRU} = 5pA$, the longitudinal wave velocity ($v_x = 96\mu m/s$ initiating from the corner) is in good agreement with the experimental result ($\approx 100\mu m/s$ [17]).

Figure 2 shows Ca^{2+} waves propagating on a discrete rectangle lattice initiating from a 10ms opening of the CRU at point (2,0.8). The snapshots are the Ca^{2+} concentration distribution at 10, 30, 50, 70, 90, 110, 130, 150, 170, 190 and 200ms (left to right, top to bottom). From image to image, we can see CRUs fire stochastically by turns while Ca^{2+} concentration wave propagates to the points of CRUs. At the beginning, CRUs fire one by one, and the amplitude (maximum of Ca^{2+} concentration in a region) is not very large; but with the increase of time, some of CRUs fire simultaneously in a short time so that Ca^{2+} sparks influence each other, and the amplitudes of Ca^{2+} sparks become larger and larger. For example, the CRU at (2,1.6) fires at $t = 16ms$, the CRU at (2,2.4) fires at $t = 31ms$; at $t = 67ms$, the CRUs at (4,0.8) and (4,2.4) fires simultaneously, in a short time interval, at $t = 68ms$ the CRU at (2,4.8) fires. In image 10, sparks occur at (18,4.0), (18,4.8), (18,5.6), (18,6.4), (18,7.2), (18,8.0), (18,8.8) and (18,9.6) in rapid succession, which may trigger a calcium transient. Image 11 shows that the boundary limits the propagation of Ca^{2+} wave, but in an actual cardiac myocyte with size $100\mu m \times 20\mu m$, calcium transient will be observed. In addition, though the space intervals of CRUs along y-axis are more compact than along x-axis, transverse wave velocity $v_y = 82\mu m/s$ is smaller than that along x-axis v_x (because the diffusion coefficient along y-axis $D_{C_y} = 0.15\mu m^2 \cdot ms^{-1}$ is smaller than that along x-axis $D_{C_x} = 0.30\mu m^2 \cdot ms^{-1}$).

Our numerical results show that a single Ca^{2+} spark can trigger a normal Ca^{2+} wave under the pathological condition of

Table 2. Standard parameter values for dye and endogenous buffers.

buffers	k^+	k^-	$[B]_T$
dye	80	90	50
Calmodulin	100	38	24
Troponin	39	20	70
SR	115	100	47
SL	115	1000	1124

doi:10.1371/journal.pone.0057093.t002

$I_{CRU} = 5pA$ which is consistent with the experimental results (longitudinal wave velocity $v_x \approx 100\mu m/s$). Physiological current through CRUs is about 2pA. However, the current may be increased (but not so large as 20pA [7,13]) by external or internal factors, such as some disease or some electroneurographic signals. Our model present that a spontaneous Ca^{2+} spark can form a Ca^{2+} wave. The physical reason is, subdiffusion of Ca^{2+} is slower than that for Fick's diffusion. When an event of Ca^{2+} spark occurs, high value of Ca^{2+} concentration may stay in a larger region around the firing CRU (for one Ca^{2+} spark, FWHM is $2.0\mu m$ for subdiffusion and $1.0\mu m$ for Fick's diffusion), the firing probability of adjacent CRUs becomes higher. Then the fire-diffuse-fire process can be initiated and sustained, a Ca^{2+} wave can propagate. So a smaller current and fewer sparks are needed to form a Ca^{2+} wave with our model than that using Fick's Law.

Because of the large FWHM for one spark due to anomalous subdiffusion, one firing CRU will trigger a Ca^{2+} wave. Then we prohibit the event of another spontaneous spark so that it will not affect the initial Ca^{2+} wave. In other words, when local Ca^{2+} concentration is larger than resting Ca^{2+} concentration, Ca^{2+} sparks may occur. So the "wall" of high Ca^{2+} concentration spreads from the left corner to the top of cardiac myocyte. Image 12 shows the sequence of CRU firing along $y = 2.4\mu m$ (the longitudinal linescan). The horizontal axis denotes time t (from left to right, 200ms), and the vertical axis denotes spatial coordinate x (20mm). Except for initial two sparks, CRUs fire at nearly regular intervals, and the "wall" of high Ca^{2+} concentration is nearly a straight line. Here, from the initial spark to the second spark, it takes more time than those of the subsequent sparks, which is different from the results by Izu et al. [13] (in their simulation, the transverse linescan is adopted, but the qualitative profile must be the same). It is because the subsequent sparks are triggered by two or more adjacent sparks, and the longitudinal wave velocity approaches to a constant value, but it takes more time to trigger the next CRUs from the initial signal spark.

Effect of the Anomalous Subdiffusion Order

The anomalous diffusion order β , which determines the diffusion mode of Ca^{2+} waves, was shown to affect the wave velocity considerably in last subsection (comparing with Fick's Law). When $\beta > 2$, a large value of β means a wild spread of initial concentration, but with the increase of time, the remanent concentration at initial point will be smaller due to the wild spread of calcium concentration. So the anomalous diffusion order β affects not only the wave velocity, but also the amplitude of each CRU, and further the average amplitude of a Ca^{2+} wave. Here, β is taken to be 2.00, 2.05, 2.15 and 2.25 in order to figure out whether the amplitudes of Ca^{2+} waves will change obviously with

the variation of wave velocities. The initial condition is still a 10ms opening of the CRU at the point (2,0.8).

Table 3 presents the effect of anomalous fractional order β on the longitudinal wave velocity v_x and the average amplitude, respectively. Comparing with the results based on Fick's law, velocities of Ca^{2+} waves increase considerably when anomalous subdiffusion order β becomes bigger. For $\beta = 2.25$, the wave velocity ($96\mu m/s$) is almost twice as big as that based on Fick's law ($\beta = 2.00$, $57\mu m/s$). It is because FWHM along x -axis for $\beta = 2.25$ ($\approx 2.2\mu m$) is almost twice as large as that for $\beta = 2.00$ ($\approx 1.1\mu m$). Here, in Ca^{2+} waves, FWHM for one CRU is affected by adjacent sparks, so it is a little bigger than FWHM of a single Ca^{2+} spark ($1.97\mu m$ [10]). In contrast to wave velocity, the variation of amplitudes is not very considerable. For $\beta = 2.25$, amplitude is 81% as that for $\beta = 2.00$. The physical reason is that although for $\beta = 2.25$, FWHM along x -axis is twice as big as that for $\beta = 2.00$, the full duration at half maximum (FDHM) along x -axis for one spark still has a obvious decrease. So when the total release of Ca^{2+} concentration is almost the same, under the expansion of spatial affection and the decrease of temporal continuity, amplitude of Ca^{2+} waves does not decrease obviously. In addition, wave velocities and amplitudes do not vary linearly with β . When β is larger, the effect of subdiffusion on Ca^{2+} waves is greater. It is because when the variation of β is small, the other parameters, such as the speed of diffusion D_{Ca} and the release strength of sparks I_{CRU} , play important roles in Ca^{2+} waves. When β becomes bigger, replacing the primary position of the diffusion speed and release strength, diffusion mode affects Ca^{2+} waves significantly (wave velocities).

Effect of Initial Location

Propagation of a Ca^{2+} wave from a corner of the cardiac myocyte has been studied. It is found that the reflecting boundaries increase the amplitude of the initial spark, then further promote the propagation of the Ca^{2+} wave. In order to figure out the boundaries determine the propagation of the Ca^{2+} wave or just affect the wave velocity, we change the location of the initial Ca^{2+} spark and study how the reflecting boundaries affect Ca^{2+} waves. In general, the process in which more than two sparks firing together, then several Ca^{2+} waves propagating, meeting and dissipating is very common in cardiac myocytes. This event is called Ca^{2+} waves collision, and it was observed in experiments [17]. We will discuss in the following the interaction of several Ca^{2+} waves.

As shown in Fig. 3, it takes only 120ms for a Ca^{2+} wave initiating from a 10ms opening of the CRU at a middle point (10,9.6) to propagates to the left and the bottom boundaries. Triggering from the middle of the region, the "walking distance" of a Ca^{2+} wave becomes shorter, and it will spread more quickly to the boundary. Due to the shorter "walking distance", less sparks will occur simultaneously, and it will not make the Ca^{2+} wave develop sufficiently; but trigger from the corner, while the Ca^{2+} wave spreads wildly, large amount of sparks will fire together in a small region (Figure 2, right corner of Image 10). Comparing with the Ca^{2+} wave initiating from a corner without the effect of the boundaries, the initial concentration of the region will be smaller, then the probability of CRUs firing will be lower, so the events of Ca^{2+} sparks are more stochastic and irregular. Affected by the absence of the reflecting boundaries and the shorter "walking distance", longitudinal wave velocity v_x reduces to $84\mu m/s$, and v_y reduces to $69\mu m/s$. So Ca^{2+} waves are easier to occur at the boundaries of cardiac myocytes, which can be compared with the

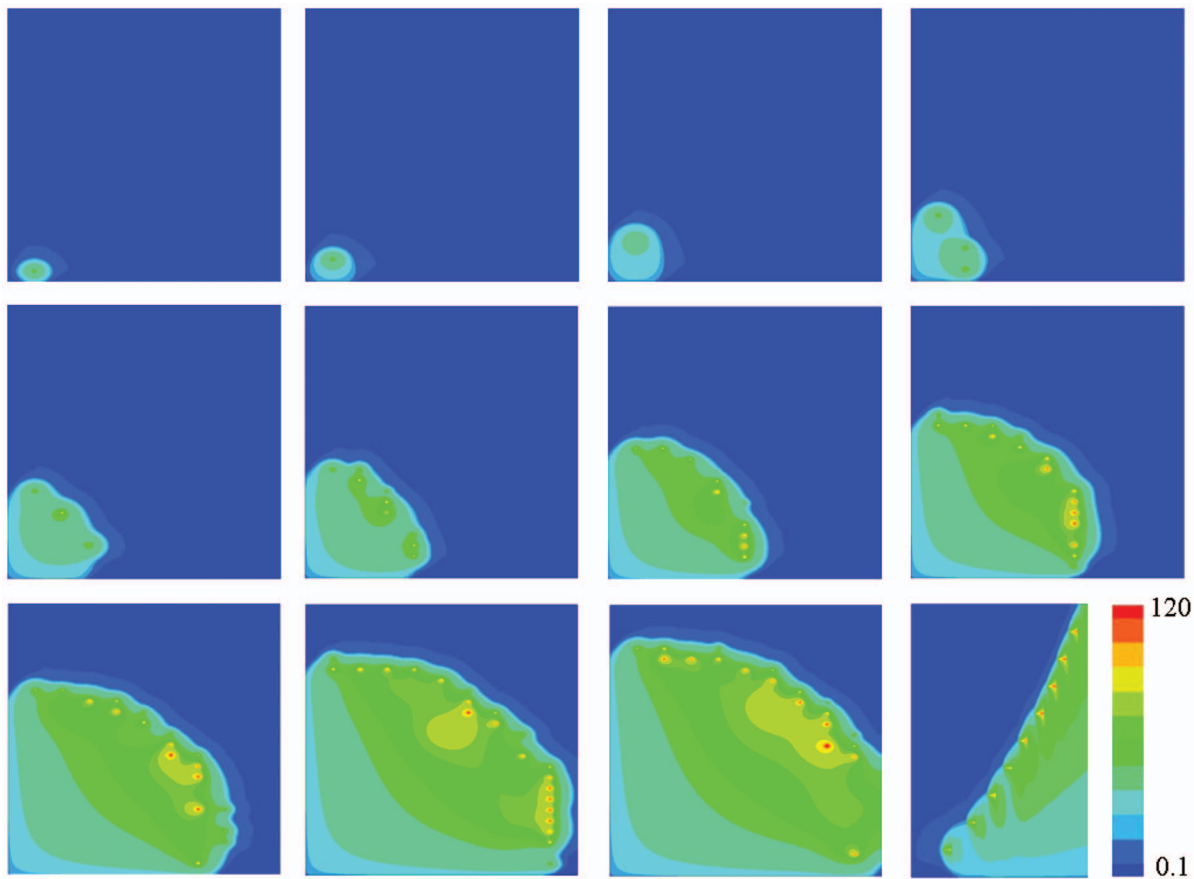


Figure 2. Snapshots of Ca^{2+} waves initiating from a 10ms opening of the CRU at the point (2,0.8). Snapshots are taken at 10, 30, 50, 70, 90, 110, 130, 150, 170, 190 and 200ms (left to right, top to bottom). Image 12 is the longitudinal linescan images along $y=2.4\mu\text{m}$. The value of parameters are $\beta=2.25$, $I_{CRU}=5\text{pA}$ and $l_x=2.0\mu\text{m}$. The concentration is from 0.1mM to 120mM . doi:10.1371/journal.pone.0057093.g002

experimental results [17], [18], [19]. Initiation of Free Ca^{2+} waves [18] and spontaneous Ca^{2+} waves [19] is kinetically favored near the boundaries, and the waves initialing from the boundaries are also easier to propagate. In ref. [17], though the results are obtained by line-scan, initiation of the waves is always near the endpoints of the line, and the waves are always triggered near the boundaries of cardiac myocytes. However, the amplitude is smaller than that of the wave from the corner though the change is not obvious. It is because the initial condition is an opening of only one spark, the reflecting effects of the boundaries are not sufficiently obvious. So the reflecting boundaries can increase the propagating probability of Ca^{2+} waves, though they are not the crucial factor for the propagation of Ca^{2+} waves. In contrast, the anomalous subdiffusion mode of Ca^{2+} concentration is the decisive factor for whether the Ca^{2+} wave can be formed by a

single Ca^{2+} spark.

Figure 4 presents the event of two Ca^{2+} waves collision(3D images). The initial condition is 10ms opening of CRUs at points (2,9.6) and (18,9.6), and they will form two Ca^{2+} waves. Two Ca^{2+} waves will meet at the middle as shown in Image 4, at $t=120\text{ms}$. Several sparks fire at the same time, and local Ca^{2+} concentration reaches a peak value. With increasing time, Ca^{2+} concentration will return to a lower value under the effect of buffers and pump, and no sparks will occur because of the CRUs' "refractory period". The 2D linescan image shows the process of two Ca^{2+} waves collision and vanishment(Fig. 8 in [17]), but it cannot show how the propagating direction changes. When Ca^{2+} concentration reaches a peak value in the middle, CRUs along x -axis($y=9.6\mu\text{m}$) are closed, but CRUs along y -axis($x=10\mu\text{m}$) have never been opened before. Therefore, Ca^{2+} waves can propagate along the line of $x=10\mu\text{m}$. Finally, all CRUs are closed and will not reopen.

Table 3. The effect of anomalous fractional order β to longitudinal wave velocity v_x and amplitude.

β	2.00	2.05	2.15	2.25
v_x	57	62	75	96
amplitude	135	128	118	110

doi:10.1371/journal.pone.0057093.t003

Modeling Ca^{2+} Waves Under a Physiological Current

To reproduce the feature of calcium waves found in experiments(primary result is wave velocity), a large current through CRU has been used in the former subsection. However, physiological value of I_{CRU} is about 2pA . So in the following discussion, $I_{CRU}=2\text{pA}$ is adopted to study how many adjacent normal Ca^{2+} sparks can trigger a Ca^{2+} wave and find out the longitudinal interval of CRUs which could make a single Ca^{2+}

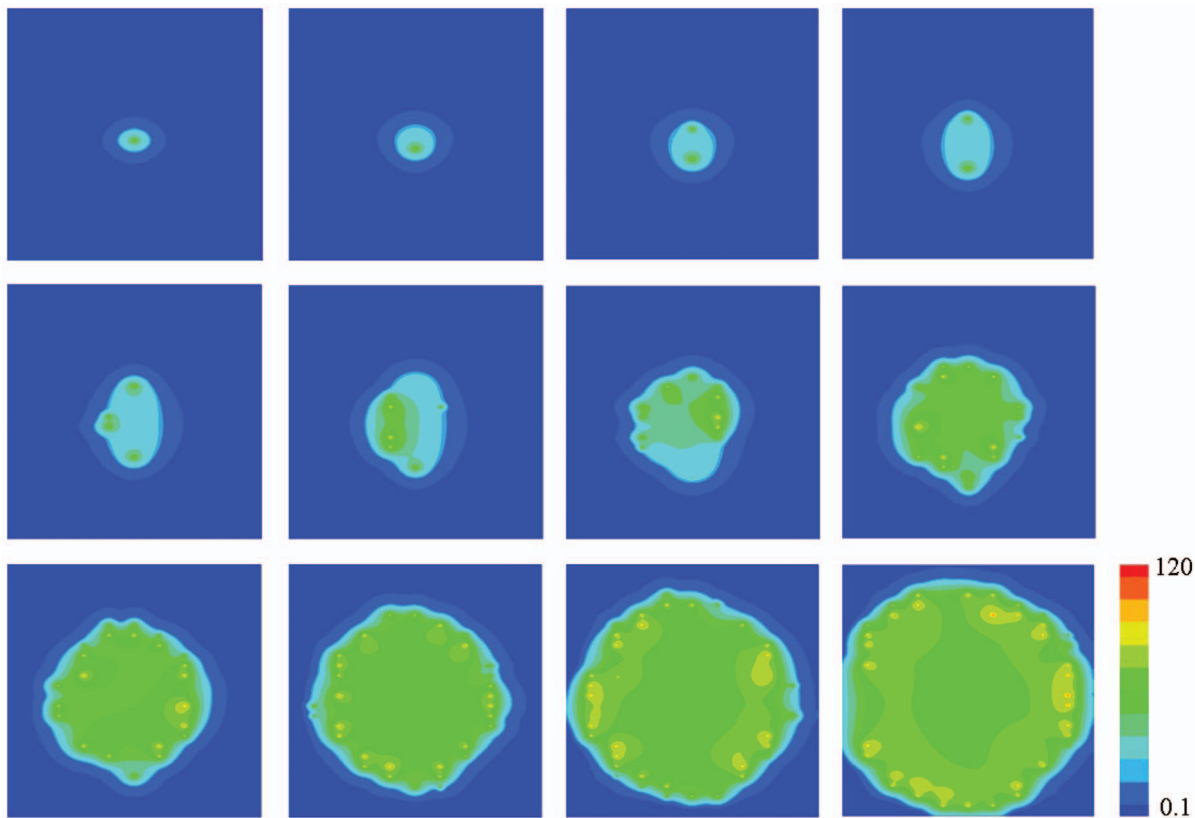


Figure 3. Snapshots of Ca^{2+} waves initiating from a 10ms opening of the CRU at the point (10,9.6). Snapshots are taken at 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110 and 120ms (left to right, top to bottom). The value of parameters are $\beta=2.25$, $I_{CRU}=5\text{pA}$ and $l_x=2.0\mu\text{m}$. The concentration is from $0.1\mu\text{M}$ to $120\mu\text{M}$.
doi:10.1371/journal.pone.0057093.g003

spark trigger a normal Ca^{2+} wave. Because of the small value of I_{CRU} , wave velocity and amplitude will be smaller. In order to make Ca^{2+} waves spread all over the region, the computation time is prolonged to 500ms. To diminish the effect of the reflecting boundaries, the initial location is chosen at the middle of the region.

Figure 5a shows whether a Ca^{2+} wave can be triggered by one spark at the middle of the region for $l_x=2.0\mu\text{m}$. When $t=490\text{ms}$, the wave only spreads through half the region, and the events of Ca^{2+} sparks are almost isolate. Under physiological conditions, even considering anomalous subdiffusion, high value of Ca^{2+} concentration may stay in a larger region around the firing CRU. For a small current, the amplitude of one spark is still small, the Ca^{2+} wave cannot propagate to the whole region, so the cardiac myocyte is stable when a normal spontaneous spark occurs.

The initial number of firing sparks is changed to study how many adjacent normal Ca^{2+} sparks can trigger a Ca^{2+} wave. The result is shown in Fig. 5b. It can be seen that four CRUs firing simultaneously at the middle will form a “weak” Ca^{2+} wave in the region. At $t=490\text{ms}$, the wave reaches the top, bottom and right boundaries, and several sparks can be found at the same time. However, three adjacent normal Ca^{2+} sparks can only form a local Ca^{2+} wave. So with the computation time of 500ms, for $\beta=2.25$, $I_{CRU}=2.0\text{pA}$ and $l_x=2.0\mu\text{m}$, four adjacent CRUs firing together is the critical initial condition to trigger a Ca^{2+} wave. However, wave velocity and amplitude here is very small ($v_x \approx 20\mu\text{m/s}$), and the Ca^{2+} concentration of the whole region is much smaller than that in Figs. 2, 3, 4.

In Figure 6a, the longitudinal interval is changed. For the case of $l_x=1.0\mu\text{m}$, it takes only 110ms for the wave to reach the left boundary. With the simultaneous firing of several CRUs, an obvious Ca^{2+} concentration wave is observed. Although the amplitude is smaller, longitudinal wave velocity ($v_x=94\mu\text{m/s}$) is comparative with the case of $I_{CRU}=5\text{pA}$, $l_x=2.0\mu\text{m}$. The physical reason for such a significant change which happens by changing $l_x=2.0\mu\text{m}$ to $l_x=1.0\mu\text{m}$ is that FWHM for $I_{CRU}=2.0\text{pA}$ is about $2.0\mu\text{m}$, and if the interval between two CRUs reduces to $1.0\mu\text{m}$, the half maximum value of a spark can “reach” adjacent CRUs easily. In addition, less interval makes more CRUs fire together, and the wave will be easier to propagate.

Our results have revealed that two factors (l_x and number of firing CRUs) can both make a Ca^{2+} wave propagate. But which the effect is more significant? Figure 6 shows when $l_x=1.0\mu\text{m}$, three Ca^{2+} waves trigger from one, four, and nine initial adjacent Ca^{2+} sparks, respectively. From 6a to 6c, both longitudinal wave velocity and amplitude become larger ($v_x=94,100,113\mu\text{m/s}$, amplitudes are $51,65,88\mu\text{M}$), but the difference is not obvious as that between Figs. 5a and 6a. So the longitudinal interval of CRUs affects Ca^{2+} waves more significantly than the number of firing CRUs. If the longitudinal interval of CRUs becomes smaller due to some reasons, such as cardiac myocytes deformation, Ca^{2+} waves will easily occur, then cardiac myocytes will be unstable.

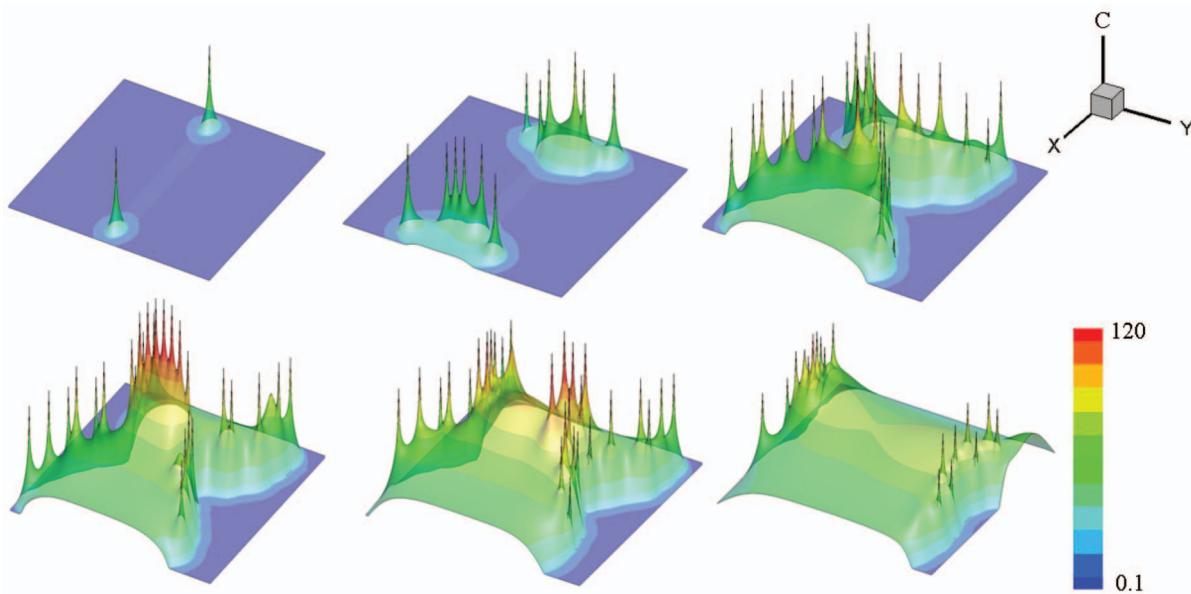


Figure 4. Snapshots of Ca^{2+} waves collision. Snapshots are taken at 10, 70, 110, 120, 130 and 150ms (left to right, top to bottom). The value of parameters are $\beta=2.25$, $I_{CRU}=5\text{pA}$ and $l_x=2.0\mu\text{m}$. The concentration is from $0.1\mu\text{M}$ to $120\mu\text{M}$.
doi:10.1371/journal.pone.0057093.g004

Conclusion

In this work, we present a mathematical model based on anomalous subdiffusion of Ca^{2+} concentration in the process of Ca^{2+} wave triggered by Ca^{2+} sparks. Ca^{2+} waves propagating from an initial firing of one single CRU at a corner or in the middle of a 2D rectangular region is numerically simulated. Our results can reproduce wave velocities of experimental results using

a small current. We show that Ca^{2+} waves can be triggered by one single Ca^{2+} spark under a small CRU current (5pA). When anomalous subdiffusion order β becomes bigger, velocities of Ca^{2+} waves increase obviously, but the variation of amplitude is not very considerable. The phenomenon of calcium waves collision is also simulated. Under physiological Ca^{2+} currents (2pA) through CRUs, an initial firing of four adjacent CRUs is

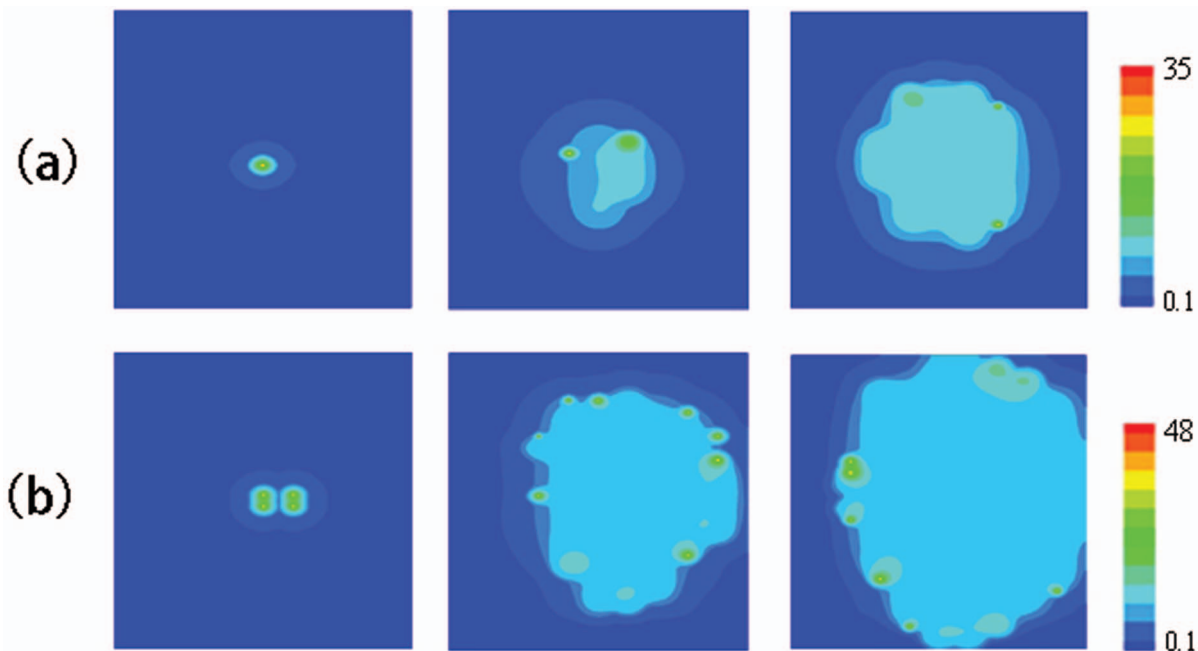


Figure 5. Illustration of Ca^{2+} waves induced by physiological Ca^{2+} sparks. (a) Snapshots of Ca^{2+} waves initiating from a 10ms opening of the CRU at the point (10,9.6), snapshots are taken at 10, 330 and 490ms. The value of parameters are $\beta=2.25$, $I_{CRU}=2\text{pA}$ and $l_x=2.0\mu\text{m}$. (b) Snapshots of Ca^{2+} waves initiating from 10ms opening of the CRUs at the point (10,9.6), (12,9.6), (10,10.4) and (12,10.4), snapshots are taken at 10, 330 and 490ms. The value of parameters are $\beta=2.25$, $I_{CRU}=2\text{pA}$ and $l_x=2.0\mu\text{m}$.
doi:10.1371/journal.pone.0057093.g005

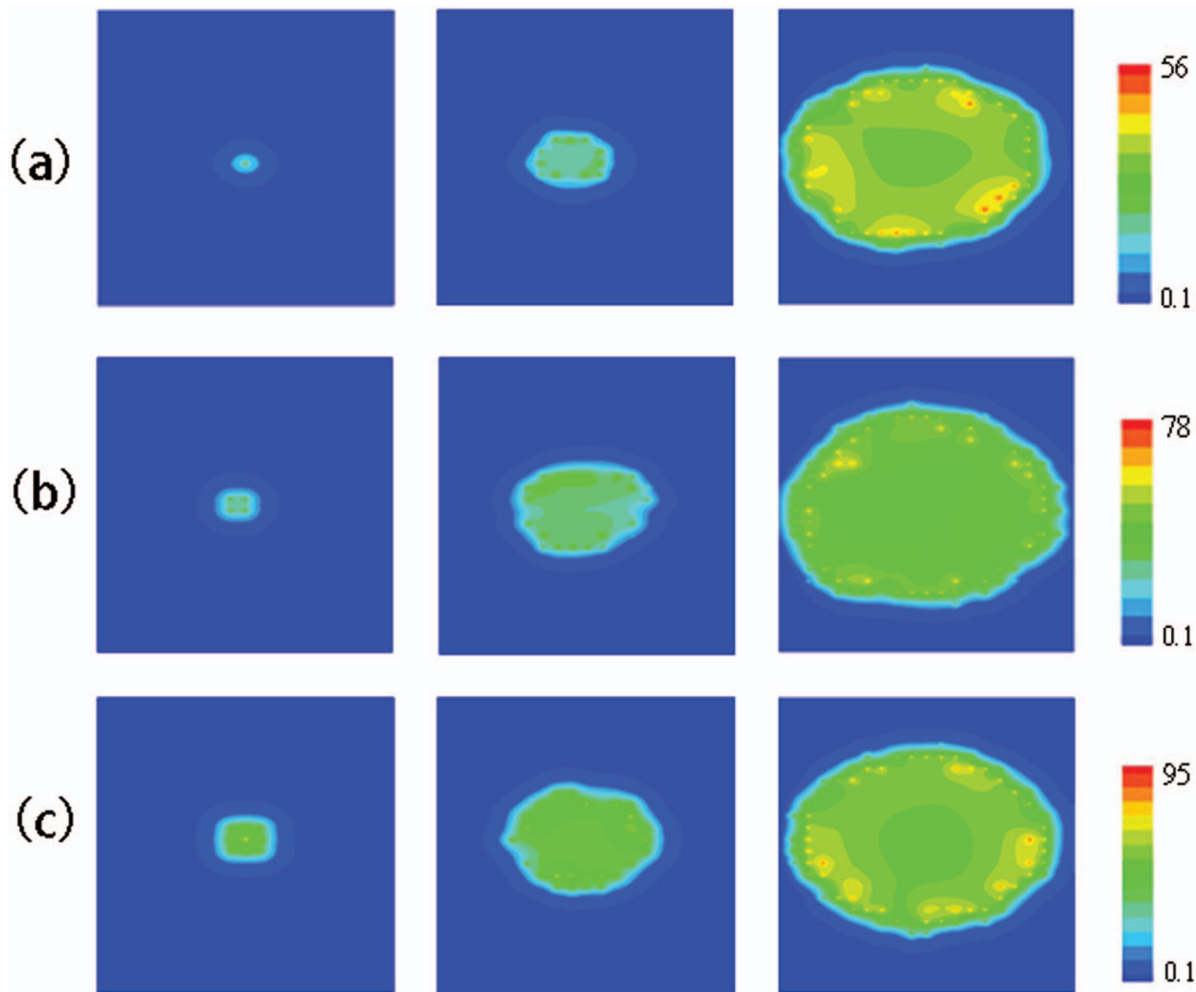


Figure 6. With smaller longitudinal intervals, the effect of initial Ca^{2+} sparks numbers. (a) Snapshots of Ca^{2+} waves initiating from a 10ms opening of the CRU at the point (10,9.6), snapshots were taken at 10, 60 and 110ms. (b) Snapshots of Ca^{2+} waves initiating from a 10ms opening of the CRUs at the point (9,9.6), (10,9.6), (9,10.4) and (10,10.4), snapshots are taken at 10, 50 and 90ms. (c) Snapshots of Ca^{2+} waves initiating from 10ms opening of the CRUs at the point (9,9.6), (10,9.6), (11,9.6), (9,10.4), (10,10.4), (11,10.4), (9,11.2), (10,11.2), (11,11.2), snapshots are taken at 10, 40 and 70ms. The value of parameters for a, b and c are $\beta = 2.25$, $I_{CRU} = 2\text{pA}$ and $l_x = 1.0\mu\text{m}$. doi:10.1371/journal.pone.0057093.g006

shown to form a Ca^{2+} wave. When $l_x = 2.0\text{mm}$, an isolated spark cannot trigger a Ca^{2+} wave, so the system is stable under physiological condition. Then the longitudinal interval of CRUs is changed to study how the system becomes unstable and how an obvious Ca^{2+} wave is formed. Our work is based on a more realistic diffusion model of Ca^{2+} sparks with the parameters close

to physiological values. The simulation results may be useful in further studies about Ca^{2+} waves.

Author Contributions

Designed and performed the numerical calculation: XC CF WT. Analyzed the data: XC JK CF WT. Wrote the paper: XC JK WT.

References

- Cheng H, Lederer W, Cannell M (1993) Calcium sparks: elementary events underlying excitation-contraction coupling in heart muscle. *Science* 262: 740–744.
- Cheng H, Lederer M, Xiao R, Gomez A, Zhou Y, et al. (1996a) Excitation-contraction coupling in heart: new insights from Ca^{2+} sparks. *Cell Calcium* 20: 129–140.
- Cannell M, Cheng H, Lederer W (1995) The control of calcium release in heart muscle. *Cell Calcium* 268: 1045–1049.
- Lopez-Lopez J, Shacklock P, Balke C, Wier W (1995) Local calcium transients triggered by single l-type calcium channel currents in cardiac cells. *Science* 268: 1042–1045.
- Fabiato A (1985) Time and calcium dependence of activation and inactivation of calcium-induced release of calcium from the sarcoplasmic reticulum of a skinned canine cardiac purkinje cell. *J Gen Physiol* 85: 247–289.
- Lakatta E, Guarnieri T (1993) Spontaneous myocardial calcium oscillations: Are they linked to ventricular fibrillation. *J Cardiovasc Electrophysiol* 44: 73–89.
- Izu L, Mauban J, Balke C, Wier W (2001a) Large currents generate cardiac Ca^{2+} sparks. *Biophys J* 80: 88–102.
- Tan W, Fu C, Xie W, Cheng H (2007) An anomalous subdiffusion model for calcium spark in cardiac myocytes. *Appl Phys Lett* 91: 183901.
- Cheng H, Lederer W (2008) Calcium sparks. *Physiol Rev* 88: 1491–1545.
- Li K, Fu C, Cheng H, Tan W (2011) Anomalous subdiffusion of calcium spark in cardiac myocytes. *Cell Mol Bioeng* 4: 457–465.
- Girard S, Luckhoff A, Lechleiter J, Sneyd J, Clapham D (1992) Two-dimensional model of calcium waves reproduces the patterns observed in xenopus oocytes. *Biophys J* 61: 509–517.
- Keizer J, Smith G (1998) Spark-to-wave transition: saltatory transmission of calcium waves in cardiac myocytes. *Biophys Chem* 72: 87–100.

13. Izu L, Gil W, William B (2001b) Evolution of cardiac calcium waves from stochastic calcium sparks. *Biophys J* 80: 103–120.
14. Lu L, Xia L, Ye X, Cheng H (2010) Simulation of the effect of rogue ryanodine receptors on a calcium wave in ventricular myocytes with heart failure. *Phys Biol* 7: 026005.
15. Baylor S, Hollingworth S (1998) Model of sarcomeric ca^{2+} movements, including atp ca^{2+} binding and diffusion, during activation of frog skeletal muscle. *J Gen Physiol* 112: 297–316.
16. Charles T, Mark M, Hans-Peter S (2006) A second-order accurate numerical approximation for the fractional diffusion equation. *J Comput Phys* 213: 205–213.
17. Cheng H, Lederer M, Lederer W, Cannell M (1996b) Calcium sparks and $[ca^{2+}]_i$ waves in cardiac myocytes. *Am J Physiol* 39: C148–C159.
18. John C, Lionel F, Ellis B, George T (1978) A free calcium wave tranverses the activating egg of the medaka, *oryzias latipes*. *J Cell Biology* 76: 448–466.
19. Takamatsu K, Wier W (1990) Calcium waves in mammalian heart: quantification of origin, magnitude, waveform, and velocity. *J Cell Biology* 4: 1519–1525.