

Biofilm Compared to Conventional Antimicrobial Susceptibility of *Stenotrophomonas maltophilia* Isolates from Cystic Fibrosis Patients

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***Stenotrophomonas maltophilia* is a multidrug-resistant organism increasingly isolated from the lungs of cystic fibrosis (CF) patients. One hundred twenty-five *S. maltophilia* isolates from 85 CF patients underwent planktonic and biofilm susceptibility testing against 9 different antibiotics, alone and in double antibiotic combinations. When *S. maltophilia* isolates were grown as a biofilm, 4 of the 10 most effective antibiotic combinations included high-dose levofloxacin and 7 of the 10 combinations included colistin at doses achievable by aerosolization.**

Stenotrophomonas maltophilia is one of the most common multidrug-resistant pathogens infecting the airways of cystic fibrosis (CF) patients (1–3). Antibiotics to treat CF pulmonary infections are chosen based on conventional antimicrobial susceptibility testing of organisms grown planktonically (“free-floating”) in liquid. However, it is known that organisms such as *S. maltophilia* actually grow as biofilms (communities of bacteria) on airway epithelial cells, suggesting that antibiotics chosen based on biofilm susceptibility testing may be more effective in CF (4, 5).

The objectives of this study were to compare biofilm antimicrobial susceptibility to conventional, planktonic antimicrobial susceptibility (as is currently done in clinical microbiology laboratories) for *S. maltophilia*, highlight the differences in antibiotic combinations derived using the two methods, and identify potentially more effective choices for inhibiting biofilm growth of *S. maltophilia* in the CF lung.

A total of 125 CF *S. maltophilia* isolates from sputum and bronchoalveolar lavage were prospectively collected from the microbiology laboratories at the Hospital for Sick Children (74 isolates from 51 CF patients; maximum of 2 isolates per patient) and St. Michael's Hospital (51 isolates from 34 CF patients; maximum of 2 isolates per patient) in Toronto, Canada, between January 2011 and July 2012. Planktonic susceptibility testing of *S. maltophilia* isolates was performed by broth microdilution according to CLSI guidelines (6). Isolates were also grown as biofilms using a modification of the Calgary biofilm technique (7). The following antibiotics were tested alone and in double combination: ceftazidime, ticarcillin-clavulanate, tobramycin, levofloxacin, moxifloxacin, trimethoprim-sulfamethoxazole, doxycycline, colistin, and azithromycin. Tobramycin (100 mg/liter and 200 mg/liter) (8) and colistin (100 mg/liter and 200 mg/liter) (9) were tested at concentrations achievable in CF sputum by aerosolization. Levofloxacin was tested at both high concentrations (50 mg/liter and 100 mg/liter, corresponding to achievable sputum levels by aerosolization) (10, 11) and low concentrations (2 mg/liter and 4 mg/liter, corresponding to achievable serum levels).

Biofilm inocula of the 125 *S. maltophilia* isolates tested fell between 2.5×10^4 and 4.6×10^6 CFU/ml (median, 5.5×10^5 CFU/ml), requiring a range of 4.5 h to over 24 h (median, 6.5 h) for biofilm generation. When tested against individual antibiotics, significantly fewer *S. maltophilia* isolates were susceptible

to fluoroquinolones, colistin, tobramycin, doxycycline, trimethoprim-sulfamethoxazole, and β -lactams when grown as biofilms than when grown planktonically (Fig. 1). High-dose levofloxacin was the most effective antibiotic against *S. maltophilia* in both the planktonic and biofilm forms. *S. maltophilia* isolates were then tested against double combinations of antibiotics grown as a biofilm and planktonically. When *S. maltophilia* isolates were grown planktonically, 6 of the 10 most effective antibiotic combinations included high-dose (achievable by aerosolization) levofloxacin and 5 of the 10 most effective antibiotic combinations included colistin at doses achievable by aerosolization (Tables 1 and 2; see also the supplemental material for complete results). In contrast, only 4 of the 10 most effective antibiotic combinations included high-dose (achievable by aerosolization) levofloxacin and 7 of the 10 most effective antibiotic combinations included colistin at doses achievable by aerosolization when isolates were grown as a biofilm.

This study is the first to examine the antimicrobial susceptibility of a large collection of predominantly CF *S. maltophilia* isolates grown both planktonically and in a biofilm. In a biofilm environment, traditional antibiotics used to treat CF patients, β -lactams and aminoglycosides, are not very effective, as β -lactams target rapidly dividing bacteria and aminoglycosides act on aerobically growing organisms (12, 13). Our study confirmed that *S. maltophilia* growing as a biofilm is very rarely susceptible to β -lactams and aminoglycosides (to which it is intrinsically resistant) (14), with fewer than 10% of isolates being susceptible to ceftazidime and ticarcillin-clavulanate and only 20% of isolates being susceptible to high-dose tobramycin which correlates with levels achiev-

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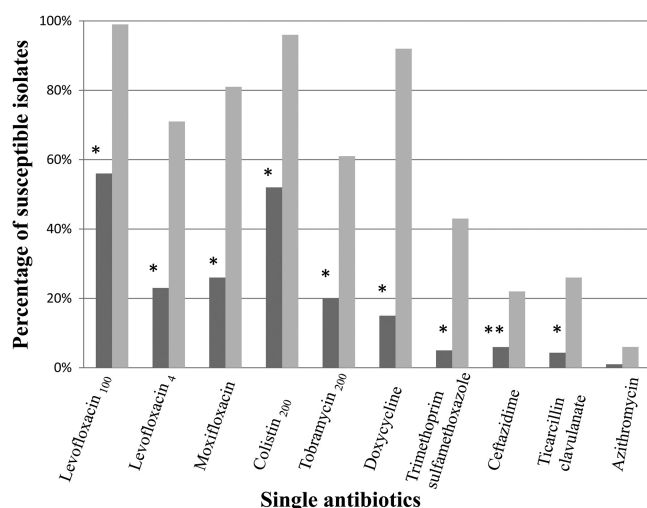


FIG 1 Percentage of *S. maltophilia* isolates susceptible to single antibiotics when grown as a biofilm (dark gray) compared to planktonic (light gray) (*, $P < 0.0001$; **, $P < 0.05$, by Fisher's exact test). Levofloxacin₁₀₀, levofloxacin tested at a maximum concentration of 100 mg/liter achievable in sputum after aerosolization; levofloxacin₄, levofloxacin tested at a maximum concentration of 4 mg/liter achievable in serum; colistin₂₀₀, colistin tested at a maximum concentration of 200 mg/liter achievable in sputum after aerosolization; and tobramycin₂₀₀, tobramycin tested at a maximum concentration of 200 mg/liter achievable in sputum after aerosolization.

able by aerosolization. Trimethoprim-sulfamethoxazole is often considered the drug of choice in the treatment of *S. maltophilia* infections; however, *S. maltophilia* resistance to trimethoprim-sulfamethoxazole has been increasingly described (15). In our assays, only half of *S. maltophilia* isolates were susceptible to trimethoprim-sulfamethoxazole alone using planktonic susceptibility testing; fewer still (less than 10%) were susceptible when grown as a biofilm.

In our study, colistin was included in many of the most effective double antibiotic combinations, and the majority of *S. maltophilia* isolates were susceptible to colistin when grown planktonically or as a biofilm. It is important to note, however, that very high concentrations of colistin (to approximately the levels achievable by aerosolization) were used in this assay based on previous *in vitro* susceptibility reports (9) and high lung concentrations achieved in animal models (16–18). However, the pulmonary concentration of colistin that can be achieved through inha-

TABLE 1 Most effective antibiotic combinations against planktonically grown *S. maltophilia* isolates

Antibiotic combination	% susceptible isolates (no.)
Levofloxacin ₁₀₀ -azithromycin	99 (124)
Levofloxacin ₁₀₀ -trimethoprim-sulfamethoxazole	99 (124)
Levofloxacin ₁₀₀ -ticarcillin-clavulanate	99 (124)
Levofloxacin ₁₀₀ -colistin ₂₀₀	99 (124)
Doxycycline-colistin ₂₀₀	98 (123)
Levofloxacin ₁₀₀ -ceftazidime	98 (123)
Colistin ₂₀₀ -trimethoprim-sulfamethoxazole	98 (123)
Tobramycin-levofloxacin ₁₀₀	98 (123)
Levofloxacin ₄ -colistin ₂₀₀	98 (122)
Moxifloxacin-colistin ₂₀₀	98 (122)

TABLE 2 Most effective antibiotic combinations against biofilm-grown *S. maltophilia* isolates

Antibiotic combination	% susceptible isolates (no.)
Ceftazidime-colistin ₂₀₀	65 (81)
Levofloxacin ₁₀₀ -ticarcillin-clavulanate	62 (78)
Colistin ₂₀₀ -trimethoprim-sulfamethoxazole	62 (78)
Moxifloxacin-colistin ₂₀₀	61 (76)
Doxycycline-colistin ₂₀₀	60 (75)
Levofloxacin ₁₀₀ -ceftazidime	59 (74)
Levofloxacin ₁₀₀ -azithromycin	58 (73)
Levofloxacin ₁₀₀ -colistin ₂₀₀	58 (72)
Levofloxacin ₄ -colistin ₂₀₀	58 (72)
Ticarcillin-clavulanate-colistin ₂₀₀	58 (72)

lation is limited by several factors, including significant bronchospasm and hypersensitivity pneumonitis (19–21). Colistin may thus be less effective *in vivo* with lower achievable pulmonary concentrations (22, 23) than has been demonstrated *in vitro* against *S. maltophilia*.

The most effective antibiotic tested alone against planktonic and biofilm-grown *S. maltophilia* isolates in our study was high-dose levofloxacin. Previous *in vitro* studies have demonstrated that fluoroquinolones, such as levofloxacin, can disrupt *S. maltophilia* biofilms and significantly reduce *S. maltophilia* biofilm mass (24, 25). In addition, high lung concentrations of aerosolized levofloxacin can be achieved in mouse models of lung infection (10) and in CF patients (11, 26). Inhaled levofloxacin may thus represent a potentially effective suppressive antimicrobial therapy for patients chronically infected with *S. maltophilia*, although antimicrobial resistance may develop with long-term use.

This study has several limitations. Based on current clinical practice, double, not triple, antibiotic combinations known to have *in vitro* activity against *S. maltophilia* were tested (9, 27, 28). However, current practices are losing efficacy, and different solutions may be required. Results may also be biased toward patients with repeated samples, although the majority of susceptibility results of isolates from the same patient were different in our study.

In conclusion, both colistin and levofloxacin, at levels achievable by inhalation, were effective at inhibiting the growth of CF *S. maltophilia* isolates under biofilm conditions. Further prospective studies are needed to determine whether aerosolized levofloxacin treatment can significantly decrease the pulmonary burden of *S. maltophilia* and improve clinical outcomes in CF patients.

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We have no conflicts of interest to declare.

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REFERENCES

- Gibson RL, Burns JL, Ramsey BW. 2003. Pathophysiology and management of pulmonary infections in cystic fibrosis. *Am. J. Respir. Crit. Care Med.* 168:918–951.
- Ballesteros S, Virseda I, Escobar H, Suarez L, Baquero F. 1995.

- Stenotrophomonas maltophilia* in cystic fibrosis patients. Eur. J. Clin. Microbiol. Infect. Dis. 14:728–729.
3. Steinkamp G, Wiedemann B, Rietschel E, Krahl A, Gielen J, Barmeier H, Ratjen F. 2005. Prospective evaluation of emerging bacteria in cystic fibrosis. J. Cyst. Fibros. 4:41–48.
 4. Pompilio A, Crocetta V, Confalone P, Nicoletti M, Petrucca A, Guarnieri S, Fiscarelli E, Savini V, Piccolomini R, Di Bonaventura G. 2010. Adhesion to and biofilm formation on IB3-1 bronchial cells by *Stenotrophomonas maltophilia* isolates from cystic fibrosis patients. BMC Microbiol. 10:102.
 5. Keays T, Ferris W, Vandemheen KL, Chan F, Yeung SW, Mah TF, Ramotar K, Saginur R, Aaron SD. 2009. A retrospective analysis of biofilm antibiotic susceptibility testing: a better predictor of clinical response in cystic fibrosis exacerbations. J. Cyst. Fibros. 8:122–127.
 6. Clinical and Laboratory Standards Institute. 2012. Performance standards for antimicrobial susceptibility testing; 22nd informational supplement M100-S22. Clinical and Laboratory Standards Institute, Wayne, PA.
 7. Ceri H, Olson ME, Stremick C, Read RR, Morck D, Buret A. 1999. The Calgary Biofilm Device: new technology for rapid determination of antibiotic susceptibilities of bacterial biofilms. J. Clin. Microbiol. 37:1771–1776.
 8. Dales L, Ferris W, Vandemheen K, Aaron SD. 2009. Combination antibiotic susceptibility of biofilm-grown *Burkholderia cepacia* and *Pseudomonas aeruginosa* isolated from patients with pulmonary exacerbations of cystic fibrosis. Eur. J. Clin. Microbiol. Infect. Dis. 28:1275–1279.
 9. San Gabriel P, Zhou J, Tabibi S, Chen Y, Trauzzi M, Saiman L. 2004. Antimicrobial susceptibility and synergy studies of *Stenotrophomonas maltophilia* isolates from patients with cystic fibrosis. Antimicrob. Agents Chemother. 48:168–171.
 10. Sabet M, Miller CE, Nolan TG, Senekeo-Effenberger K, Dudley MN, Griffith DC. 2009. Efficacy of aerosol MP-376, a levofloxacin inhalation solution, in models of mouse lung infection due to *Pseudomonas aeruginosa*. Antimicrob. Agents Chemother. 53:3923–3928.
 11. Geller DE, Flume PA, Griffith DC, Morgan E, White D, Loutit JS, Dudley MN. 2011. Pharmacokinetics and safety of MP-376 (levofloxacin inhalation solution) in cystic fibrosis subjects. Antimicrob. Agents Chemother. 55:2636–2640.
 12. Hassett DJ, Cuppoletti J, Trapnell B, Lyman SV, Rowe JJ, Yoon SS, Hilliard GM, Parvatiyar K, Kamani MC, Wozniak DJ, Hwang SH, McDermott TR, Ochsner UA. 2002. Anaerobic metabolism and quorum sensing by *Pseudomonas aeruginosa* biofilms in chronically infected cystic fibrosis airways: rethinking antibiotic treatment strategies and drug targets. Adv. Drug Deliv. Rev. 54:1425–1443.
 13. Worlitzsch D, Tarran R, Ulrich M, Schwab U, Cekici A, Meyer KC, Birrer P, Bellon G, Berger J, Weiss T, Botzenhart K, Yankaskas JR, Randell S, Boucher RC, Doring G. 2002. Effects of reduced mucus oxygen concentration in airway *Pseudomonas* infections of cystic fibrosis patients. J. Clin. Invest. 109:317–325.
 14. Leclercq R, Canton R, Brown DF, Giske CG, Heisig P, Macgowan AP, Mouton JW, Nordmann P, Rodloff AC, Rossolini GM, Soussy CJ, Steinbakk M, Winstanley TG, Kahlmeter G. 2011. EUCAST expert rules in antimicrobial susceptibility testing. Clin. Microbiol. Infect. [Epub ahead of print.] doi:10.1111/j.1469-0691.2011.03703.x.
 15. Toleman MA, Bennett PM, Bennett DM, Jones RN, Walsh TR. 2007. Global emergence of trimethoprim/sulfamethoxazole resistance in *Stenotrophomonas maltophilia* mediated by acquisition of sul genes. Emerg. Infect. Dis. 13:559–565.
 16. Lu Q, Girardi C, Zhang M, Bouhemad B, Louchahi K, Petitjean O, Wallet F, Becquemini MH, Le Naour G, Marquette CH, Rouby JJ. 2010. Nebulized and intravenous colistin in experimental pneumonia caused by *Pseudomonas aeruginosa*. Intensive Care Med. 36:1147–1155.
 17. Aoki N, Tateda K, Kikuchi Y, Kimura S, Miyazaki C, Ishii Y, Tanabe Y, Gejyo F, Yamaguchi K. 2009. Efficacy of colistin combination therapy in a mouse model of pneumonia caused by multidrug-resistant *Pseudomonas aeruginosa*. J. Antimicrob. Chemother. 63:534–542.
 18. Marchand S, Gobin P, Brillault J, Baptista S, Adier C, Olivier JC, Mimoz O, Couet W. 2010. Aerosol therapy with colistin methanesulfonate: a biopharmaceutical issue illustrated in rats. Antimicrob. Agents Chemother. 54:3702–3707.
 19. Beringer P. 2001. The clinical use of colistin in patients with cystic fibrosis. Curr. Opin. Pulm. Med. 7:434–440.
 20. Cunningham S, Prasad A, Collyer L, Carr S, Lynn IB, Wallis C. 2001. Bronchoconstriction following nebulised colistin in cystic fibrosis. Arch. Dis. Child. 84:432–433.
 21. Leong KW, Ong S, Chee HL, Lee W, Kwa AL. 2010. Hypersensitivity pneumonitis due to high-dose colistin aerosol therapy. Int. J. Infect. Dis. 14:e1018–e1019.
 22. Reed MD, Stern RC, O’Riordan MA, Blumer JL. 2001. The pharmacokinetics of colistin in patients with cystic fibrosis. J. Clin. Pharmacol. 41:645–654.
 23. Athanassa ZE, Markantonis SL, Fousteri MZ, Myrianthefs PM, Boutzouka EG, Tsakris A, Baltopoulos GJ. 2012. Pharmacokinetics of inhaled colistimethate sodium (CMS) in mechanically ventilated critically ill patients. Intensive Care Med. [Epub ahead of print.] doi:10.1007/s00134-012-2628-7.
 24. Passerini de Rossi B, Garcia C, Calenda M, Vay C, Franco M. 2009. Activity of levofloxacin and ciprofloxacin on biofilms and planktonic cells of *Stenotrophomonas maltophilia* isolates from patients with device-associated infections. Int. J. Antimicrob. Agents. 34:260–264.
 25. Di Bonaventura G, Spedicato I, D’Antonio D, Robuffo I, Piccolomini R. 2004. Biofilm formation by *Stenotrophomonas maltophilia*: modulation by quinolones, trimethoprim-sulfamethoxazole, and ceftazidime. Antimicrob. Agents Chemother. 48:151–160.
 26. Geller DE, Flume PA, Staab D, Fischer R, Loutit JS, Conrad DJ. 2011. Levofloxacin inhalation solution (MP-376) in patients with cystic fibrosis with *Pseudomonas aeruginosa*. Am. J. Respir. Crit. Care Med. 183:1510–1516.
 27. Waters V, Ratjen F. 2006. Multidrug-resistant organisms in cystic fibrosis: management and infection-control issues. Expert Rev. Anti-Infect. Ther. 4:807–819.
 28. Weiss K, Restieri C, De Carolis E, Laverdiere M, Guay H. 2000. Comparative activity of new quinolones against 326 clinical isolates of *Stenotrophomonas maltophilia*. J. Antimicrob. Chemother. 45:363–365.