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EXTENDED REPORT

Genetic markers of rheumatoid arthritis susceptibility in anti-citrullinated peptide antibody negative patients

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Accepted 9 April 2012

Published Online First

1 June 2012

ABSTRACT

Introduction: There are now over 30 confirmed loci predisposing to rheumatoid arthritis (RA). Studies have been largely undertaken in patients with anticyclic citrullinated peptide (anti-CCP) positive RA, and some genetic associations appear stronger in this subgroup than in anti-CCP negative disease, although few studies have had adequate power to address the question. The authors therefore investigated confirmed RA susceptibility loci in a large cohort of anti-CCP negative RA subjects.

Methods: RA patients and controls, with serological and genetic data, were available from UK Caucasian patients (n=4068 anti-CCP positive, 2040 anti-CCP negative RA) and 13,009 healthy controls. HLA-DRB1 genotypes and 36 single nucleotide polymorphisms were tested for association between controls and anti-CCP positive or negative RA.

Results: The shared epitope (SE) showed a strong association with anti-CCP positive and negative RA, although the effect size was significantly lower in the latter (effect size ratio=3.18, $p<1.0E-96$). A non-intronic marker at TNFAIP3, GIN1/C5orf30, STAT4, ANKRD55/IL6ST, BLK and PTPN22 showed association with RA susceptibility, irrespective of the serological status, the latter three markers remaining significantly associated with anti-CCP negative RA, after correction for multiple testing. No significant association with anti-CCP negative RA was detected for other markers (eg, AFF3, CD28, intronic marker at TNFAIP3), though the study power for those markers was over 80%.

Discussion: In the largest sample size studied to date, the authors have shown that the strength of association, the effect size and the number of known RA susceptibility loci associated with disease is different in the two disease serotypes, confirming the hypothesis that they might be two genetically different subsets.

INTRODUCTION

Based on the presence or absence of anticyclic citrullinated peptide (anti-CCP) antibodies, rheumatoid arthritis (RA) can be classified into anti-CCP positive and anti-CCP negative RA. Anti-CCP antibodies have been widely shown to be strong predictors of disease severity and radiological damage.^{1,2} It is currently a matter of some debate as to whether anti-CCP positive and anti-CCP negative RA are two distinct entities or represent two different subsets of one and the same disease.³⁻⁵ Linkage and association analysis revealed the shared

epitope (SE) to be associated only with anti-CCP positive RA and not with anti-CCP negative RA.⁶ A study in twin pairs has shown that the estimated heritability of anti-CCP negative RA is 66% (95% CI 21% to 82%), similar to the heritability of anti-CCP positive RA, estimated at 68% (95% CI 55% to 79%).⁷ In the same study, the SE was found to explain 18% of the genetic component of RA susceptibility in anti-CCP positive RA but only 2.4% in anti-CCP negative RA.

Several studies have investigated putative associations between different human leucocyte antigen (HLA) alleles or single nucleotide polymorphisms (SNPs) within the HLA region and predisposition to anti-CCP negative RA,^{5,6,8-11} with contradictory results. A large meta-analysis, across four European populations, found only a weakly significant association between several HLA alleles and anti-CCP negative RA, but observed marked geographical differences.¹² Lack of consistency between studies might therefore be explained by a different sample size and power, by geographical differences in allele frequencies and association patterns or by different study designs or definitions of HLA genotypes (two vs four-digit typing, different classifications for the SE). A large study, performed on Caucasians of Northern European descent, investigated several HLA-DRB1 susceptibility and protective models and SE subgroups for association with RA after stratification by autoantibody status. Significant associations between several HLA-DRB1 alleles and anti-CCP negative RA were found.¹³ Together, these findings strengthen the hypothesis that genetic factors predisposing to anti-CCP positive RA are different to those predisposing to anti-CCP negative RA.

A recent large meta-analysis brought the number of confirmed non-HLA RA susceptibility loci to 31.¹⁴ However, most studies on the identification of RA susceptibility loci published to date have been performed in largely anti-CCP positive RA cohorts, thereby biasing the search of RA susceptibility loci towards genetic variants predisposing to anti-CCP positive RA. Importantly, most studies have been underpowered to identify anti-CCP negative RA predictors.

Very few studies have systematically compared the genetic basis of anti-citrullinated protein autoantibody (ACPA)-positive to ACPA-negative RA outside the HLA region. In a genome-wide association study (GWAS) recently performed in 774 anti-CCP negative RA patients, 1147 anti-

CCP positive RA patients and 1079 common controls,¹⁵ no SNP achieved genome-wide significance in the comparison between anti-CCP negative RA and controls, while the *PTPN22* gene was associated with anti-CCP positive RA, together with hundreds of SNPs located within the HLA locus on chromosome 6.

Candidate gene association studies and a subsequent meta-analysis have confirmed an association of *STAT4* polymorphisms with anti-CCP positive and negative RA.^{16 17} *IRF5* polymorphisms have been shown in independent studies to be more strongly associated with anti-CCP negative RA than with anti-CCP positive RA in Caucasians,^{18 19} while this differential association is controversial in Asians.^{20 21} No other SNPs have been convincingly associated with anti-CCP negative RA. Interestingly, the association of *PTPN22* polymorphisms with anti-CCP negative RA is controversial, with some investigators reporting association in anti-CCP negative patients.^{22 23} For example, a recent study investigating the usefulness of data derived from electronic health records tested multiple non-HLA RA susceptibility markers in 871 anti-CCP positive RA patients, 378 anti-CCP negative RA patients and 1212 common healthy controls.²⁴ Only *PTPN22* showed an association with anti-CCP negative RA with a *p* value < 0.05. Due to the small sample size of the anti-CCP negative subgroup, conclusions could be made only for SNPs in aggregate rather than for individual SNPs. The authors conclude that there is a partial overlap between the genetic basis of anti-CCP positive and anti-CCP negative RA.

We hypothesised that currently known RA susceptibility SNPs would show a differential association pattern in anti-CCP negative RA compared with anti-CCP positive RA. Therefore, we tested the 31 RA confirmed susceptibility loci for association with RA in a dataset comprising between 1935 and 3827 anti-CCP positive RA patients, between 808 and 1918 anti-CCP negative RA patients and between 11468 and 12392 healthy controls per genetic marker, the largest number of anti-CCP negative patients studied so far in this setting.

METHODS

Data collection, SNP selection and genotyping

Confirmed RA susceptibility SNPs were selected from the large meta-analysis by Stahl *et al.*¹⁴ Most of the SNPs had been already genotyped in RA cases and controls as part of several other projects conducted at our laboratory. Cohorts and patient characteristics are presented in the online supplementary table S1; genotyping and quality control procedures have been described elsewhere.^{22 25–27} All patients and controls were Caucasians originating from the UK satisfying the 1987 American College of Rheumatology classification criteria for RA. Only RA cases with an available anti-CCP status, as determined with the second generation CCP (CCP2) assay, were included in the analysis. Several studies/cohorts did not contain any controls. Therefore, anti-CCP positive RA cases or anti-CCP negative RA cases from different studies were pooled together and compared with controls. If a SNP reported in the study of Stahl was not available in a cohort, a SNP proxy was selected to fulfil the following requirements: linkage disequilibrium (r^2) ≥ 0.90 with the original SNP and a maximum of three different SNPs for one specific locus across all datasets. If these requirements could not be fulfilled for one dataset, it was excluded from the analysis for that particular locus. Therefore, different numbers of cases and controls were available for analysis for different loci. The following HLA-DRB1 genotypes were considered as SE alleles: '0101' '0102' '0104' '0105' '0107' '0108' '0110' '0111' '0401' '0404' '0405' '0408' '0409' '0410' '0413' '0416' '0419' '0421'

'0423' '0426' '0428' '0429' '0430' '0433' '0434' '0435' '0438' '0440' '0442' '0443' '0445' '1001' '1113' '1126' '1134' '1402' '1409' '1413' '1417' '1419' '1420' '1421' '1429' '1430' '1431' '1432' '1434' '1441' '1446' '1447' '1448'.

Statistical analysis

Multinomial logistic regression was applied to compute OR, 95% CI and *p* values for association between the minor allele at every locus and either anti-CCP-positive ($OR_{\text{ccp-positive}}$) or anti-CCP-negative RA ($OR_{\text{ccp-negative}}$), assuming additivity on the log-odds scale (ie, every locus was coded as 0,1 or 2 corresponding to the copy number of the minor allele). The minor allele was defined according to the allele frequency in the total population, including cases and controls. To test for differences between $OR_{\text{ccp-positive}}$ and $OR_{\text{ccp-negative}}$, the linear combination $\beta_+ - \beta_-$, where β_+ is $\log(OR_{\text{ccp-positive}})$ and β_- is $\log(OR_{\text{ccp-negative}})$ was calculated, along with its standard error. This enables a *p* value for the difference in association to be calculated.

Due to a high pretest probability for association in anti-CCP positive RA, *p* values were not corrected for multiple testing in this subset. Based on previously published data, 32 of the 35 non-HLA SNPs tested have been confirmed to represent independent effects. Therefore, without assuming any prior probability for association in the anti-CCP negative RA subset, which is a stringent assumption, the Bonferroni-corrected significance threshold would be 1.6E-03. Statistical analysis was performed with Stata V.10.1 (Stata Statistical Software: Release 10; Stata Corp., College Station, Texas, USA).

RESULTS

Thirty-one confirmed RA susceptibility loci,¹⁴ some of which contain independent effects, were considered for analysis. Together, this represents a total of 36 markers, plus the SE. Actual SNPs and proxies are presented for each locus in table 1. The combination of datasets available led to a total number of 4068 anti-CCP-positive, 2040 anti-CCP-negative RA and 13009 healthy UK controls. The actual number of cases and controls varies for every locus from 1935 to 3827 for anti-CCP positive RA, from 808 to 1918 for anti-CCP negative RA and from 11468 to 12392 for healthy controls. Basic cohort characteristics are presented in the online supplementary table S1. The results of the association analysis are presented in table 2. As expected, every locus showed an association with anti-CCP positive RA with a *p* value < 0.05. The SE and the corresponding tag SNP were highly associated with anti-CCP positive RA with an OR of 4.08 and 2.68, respectively. Three other markers at *AFF3*, *CD28*, *PTPN22* and two at the *TNFAIP3* locus reached genome-wide significance (<5E-08) in anti-CCP positive RA with OR of 1.17, 1.18, 1.91, 1.29 and 1.45, respectively. By contrast, only six non-HLA loci in total reach a *p* value below 0.05 in anti-CCP negative RA: *TNFAIP3*, *GIN1/C5orf30*, *STAT4*, *ANKRD55/IL6ST*, *BLK* and *PTPN22*. The three last loci remained significant at the Bonferroni corrected threshold of 1.6E-03. The SE, and its tag SNP, show an association with anti-CCP negative RA with an OR of 1.28 and 1.15, respectively. The online supplementary table S2 shows association results stratified by rheumatoid factor and anti-CCP positivity. The probability of obtaining, by chance, at least 7/36 associated loci at a significance level of 0.05 is 1.8E-03. Therefore, under a prior hypothesis of no association, there is still a significant accumulation of RA susceptibility loci associated with anti-CCP negative RA.

Due to the different number of cases in anti-CCP positive and negative RA, simply comparing *p* values would be misleading.

Table 1 Single nucleotide polymorphism (SNP) markers and their proxys for the independent rheumatoid arthritis susceptibility loci considered for analysis. The shared epitope is not defined by SNP markers, but by a list of 4-digit HLA-DRB1 alleles described in the methods section

Chromosome	Locus name	Single nucleotide polymorphism markers		
1	CD2/CD58	rs11586238		
1	FCGR2A	rs12746613	rs10494360	
1	MMEL1/TNFRSF14	rs10910099	rs3890745	
1	PTPN22	rs2476601	rs6679677	
1	PTPRC	rs10919563	rs1932435	
2	AFF3 locus 1	rs1160542	rs11676922	rs9653442
2	AFF3 locus 2	rs10865035		
2	CD28	rs1980422		
2	CTLA4	rs3087243	rs231804	
2	REL	rs13031237		
2	SPRED2	rs934734	rs17534670	
2	STAT4	rs7574865		
3	DNASE1L3/PXK	rs13315591	rs9813011	
4	IL2/IL21	rs6822844	rs13151961	
4	RBPJ	rs874040	rs10517086	
5	ANKRD55/IL6ST	rs6859219		
5	GIN1/C5orf30	rs26232	rs35797	
6	CCR6	rs3093023	rs6907666	rs3093024
6	HLA-DRB1 0401 tag	rs6910071		
6	PRDM1	rs548234		
6	TAGAP	rs394581	rs169858	
6	TNFAIP3 locus 1	rs6920220	rs2327832	
6	TNFAIP3 locus 2	rs13207033	rs10499194	
6	TNFAIP3 locus 3	rs5029937	rs5029939	
7	IRF5	rs10488631	rs12531711	
8	BLK	rs2736340		
9	CCL21 locus 1	rs951005		
9	CCL21 locus 2	rs2812378	rs10814138	
9	TRAF1/C5	rs3761847		
10	IL2RA locus 1	rs706778	rs10795791	rs7072793
10	IL2RA locus 2	rs2104286		
10	PRKCQ	rs4750316	rs10796045	
11	RAG1/TRAF6	rs540386	rs5030437	rs1046864
12	KIF5A/PIP4K2C	rs1678542	rs11172254	
20	CD40	rs4810485	rs1569723	
22	IL2RB	rs3218253	rs3218258	

Therefore, the significance of the observed difference in association between anti-CCP positive and negative RA was addressed by computing a comparison OR (or effect size OR) and its corresponding p value (column 'comparison' in table 2). This allows the classification of the loci into three distinct categories (table 3 and figure 1): the *HLA-DRB1 SE*, *PTPN22* and one marker at *TNFAIP3* are associated with anti-CCP positive and negative RA, but show a clear differential association with an effect size significantly higher in anti-CCP positive RA (category 1). Other loci, like *C5orf30* or *STAT4* are associated with RA irrespective of the serological status, with the effect size not differing significantly between subsets (category 2: anti-CCP independent associations). A third category comprises anti-CCP positive specific loci with no significant association detected in anti-CCP negative RA; however, the p value for the effect size ratio is below 0.05. The remaining loci could not be classified into one of these three categories, because although they are associated with anti-CCP positive RA with a p value < 0.05, the effect in anti-CCP-negative RA is not significantly different from that in anti-CCP-positive RA, nor significantly different from the null. This last situation can only be explained by a lack of power. Indeed, for the vast majority of markers in category 3, the study has, in the anti-CCP

negative subgroup, a power between 60% and 90% at the 0.05 significance level to detect an association of the same effect size as observed in the anti-CCP positive subgroup, while power drops to between 15% and 50% for most of the unclassifiable markers. As an example in category 3, an association of an effect size of 1.17 would be detected for *AFF3 locus 1* with a power of 89.3% at the 0.05 significance level for a minor allele frequency (MAF) of 45.5% in controls. An association of a larger effect size of 1.45 would be detected for the intronic *TNFAIP3* marker with a power of 81.6% at the 0.05 significance level for a MAF of 3.5% in controls. The detection power drops for smaller effects around 1.13, but is still 64.4% *CCL21 locus 2* (MAF in controls 34.2%). However, although *CD2/CD58* has been genotyped in 1918 anti-CCP negative cases, it is not possible to classify it as being associated with anti-CCP positive, negative or both because the power to detect an effect of 1.11 in anti-CCP negative with a MAF of 24% in controls is only 46.1%.

Of note, is the fact, that the effect size of all but two loci, *ANKRD55* and *BLK* (figure 1), is larger in anti-CCP positive RA than in anti-CCP negative RA. However, due to relatively wide CIs, the effect size ratio is not significant, so these two loci are both classified in category 2.

Table 2 OR with 95% CI and p value for association of the minor allele at every locus with anticyclic citrullinated peptide (anti-CCP) positive or anti-CCP negative rheumatoid arthritis (RA). The minor allele is defined according to the frequency in the total population, including cases and controls.

Chr.	Locus Name	UK anti-CCP positive RA						UK anti-CCP negative RA						Comparison	
		OR	95% CI	p Value	N cases	N cont	OR	95% CI	p Value	N cases	N cont	ORpos/ ORneg	p Value		
1	CD2/CD58	1.11	1.05 to 1.18	3.68E-04	3827	11468	1.05	0.97 to 1.13	0.242	1918	11468	1.06	0.183		
1	FCGR2A	1.16	1.07 to 1.25	2.28E-04	3467	11468	1.08	0.96 to 1.20	0.195	1563	11468	1.08	0.241		
1	MIMEL1/TNFRSF14	0.88	0.77 to 0.93	2.12E-05	3485	12394	0.97	0.89 to 1.05	0.426	1574	12394	0.91	0.045		
1	PTPN22	1.91	1.73 to 2.05	3.91E-65	3813	12398	1.20	1.07 to 1.33	1.34E-03	1894	12398	1.60	7.11E-15		
1	PTPRC	0.91	0.84 to 0.99	0.022	3600	11427	0.96	0.86 to 1.08	0.504	1580	11427	0.94	0.384		
2	AFF3 locus 1	1.17	1.11 to 1.23	5.36E-09	3777	11832	1.05	0.98 to 1.13	0.165	1881	11832	1.11	0.008		
2	AFF3 locus 2	1.17	1.09 to 1.25	2.64E-06	2378	8425	1.06	0.97 to 1.17	0.187	991	8425	1.10	0.086		
2	CD28	1.18	1.12 to 1.26	3.15E-08	3826	11469	1.08	1.00 to 1.17	0.055	1912	11469	1.09	0.049		
2	CTLA4	0.91	0.87 to 0.96	9.32E-04	3626	11975	0.94	0.88 to 1.02	0.121	1592	11975	0.97	0.475		
2	REL	1.13	1.07 to 1.20	1.58E-05	3196	10878	1.02	0.95 to 1.11	0.554	1437	10878	1.11	0.028		
2	SPRED2	0.91	0.86 to 0.97	3.69E-03	2706	9032	0.97	0.89 to 1.06	0.565	1132	9032	0.94	0.198		
2	STAT4	1.13	1.06 to 1.21	1.34E-04	3401	11822	1.11	1.02 to 1.20	0.015	1824	11822	1.02	0.656		
3	DNASE1L3/PXK	1.13	1.02 to 1.24	0.014	3445	11486	0.93	0.80 to 1.08	0.338	1432	11486	1.21	0.020		
4	IL2/IL21	0.88	0.82 to 0.95	1.03E-03	3402	11767	0.95	0.86 to 1.04	0.264	1814	11767	0.93	0.211		
4	RBPJ	1.14	1.07 to 1.21	1.98E-05	3195	10879	1.02	0.94 to 1.11	0.681	1434	10879	1.12	0.021		
5	ANKRD55/IL6ST	0.85	0.78 to 0.92	9.62E-05	2377	8428	0.80	0.70 to 0.90	2.42E-04	991	8428	1.07	0.357		
5	GIN1/C5orf30	0.88	0.83 to 0.94	4.86E-05	3372	11260	0.88	0.80 to 0.95	2.32E-03	1410	11260	1.01	0.835		
6	CCR6	1.13	1.07 to 1.19	1.48E-05	3423	11313	0.99	0.92 to 1.07	0.851	1508	11313	1.14	0.004		
6	HLA-DRB1 0401 tag	2.68	2.50 to 2.88	9.97E-169	2378	8428	1.15	1.03 to 1.28	9.55E-03	991	8428	2.33	6.30E-47		
6	HLA-DRB1 SE	4.08	3.64 to 4.56	1.18E-131	2366	1352	1.28	1.14 to 1.45	5.18E-05	1315	1352	3.18	2.56E-97		
6	PRDM1	1.06	1.00 to 1.12	0.041	3827	11469	1.04	0.97 to 1.12	0.275	1916	11469	1.02	0.692		
6	TAGAP	0.90	0.85 to 0.95	1.57E-04	3824	11466	0.97	0.90 to 1.04	0.416	1914	11466	0.92	0.068		
6	TNFAIP3 locus 1	1.29	1.21 to 1.37	3.70E-16	3659	12392	1.09	1.00 to 1.18	0.048	1862	12392	1.18	3.52E-04		
6	TNFAIP3 locus 2	0.89	0.84 to 0.95	2.72E-04	3425	11923	0.93	0.85 to 1.02	0.109	1497	11923	0.96	0.383		
6	TNFAIP3 locus 3	1.45	1.28 to 1.65	1.33E-08	3414	12021	0.91	0.73 to 1.13	0.374	1460	12021	1.60	7.36E-05		
7	IRF5	1.12	1.02 to 1.23	0.016	2739	9043	1.09	0.95 to 1.24	0.213	1175	9043	1.03	0.696		
8	BLK	1.09	1.03 to 1.16	3.77E-03	3490	11481	1.15	1.05 to 1.25	1.53E-03	1525	11481	0.95	0.332		
9	CCL21 locus 1	0.92	0.85 to 1.00	0.048	3199	10878	1.01	0.91 to 1.13	0.833	1439	10878	0.91	0.145		
9	CCL21 locus 2	1.13	1.07 to 1.20	7.68E-06	3642	11936	1.04	0.97 to 1.12	0.274	1855	11936	1.09	0.046		
9	TRAF1/C5	1.10	1.04 to 1.16	7.54E-04	3135	11544	1.02	0.94 to 1.10	0.698	1447	11544	1.08	0.073		
10	IL2RA locus 1	1.14	1.06 to 1.23	3.21E-04	1935	5477	1.07	0.96 to 1.19	0.224	808	5477	1.07	0.240		
10	IL2RA locus 2	0.88	0.83 to 0.93	2.71E-05	3806	12396	0.97	0.90 to 1.05	0.450	1896	12396	0.91	0.033		
10	PRKCC	0.87	0.82 to 0.94	1.18E-04	3657	12427	0.95	0.86 to 1.04	0.281	1624	12427	0.92	0.132		
11	RAG1/TRAF6	0.92	0.85 to 0.99	0.032	3440	11379	0.98	0.88 to 1.09	0.723	1556	11379	0.94	0.285		
12	KIF5A/PIP4K2C	0.90	0.85 to 0.95	1.81E-04	3688	12452	0.95	0.88 to 1.02	0.135	1881	12452	0.95	0.237		
20	CD40	0.89	0.84 to 0.95	4.36E-04	3640	11971	1.02	0.94 to 1.11	0.594	1858	11971	0.87	0.005		
22	IL2RB	1.15	1.08 to 1.21	3.15E-06	3811	12392	1.08	1.00 to 1.16	0.057	1900	12392	1.06	0.164		

N cases, cont: number of cases or controls with available genotype. Comparison ORpos/ORneg: effect size ratio between the OR in CCP-positive and CCP-negative RA, with the p value for its significance. The CI of the effect size OR is not shown to avoid overloading the table

Interestingly, the three known independent effects at *TNFAIP3* are classified in at least two different categories (table 3). At this locus, the most profound discordance between anti-CCP positive and negative RA is seen for the intronic marker (figure 1), which displays a genome-wide significant association of a ‘large’ effect size in anti-CCP positive RA with an OR above one (OR 1.45, 95% CI 1.28 to 1.65, $p=1.33 \times 10^{-8}$), representing a risk factor for this disease. However, no significant association is detected in anti-CCP negative RA (OR 0.91, 95% CI 0.73 to 1.13, $p=0.37$). The effect size ratio is highly significant (OR 1.60, 95% CI 1.27 to 2.02, $p=7.36 \times 10^{-5}$).

DISCUSSION

With a number of anti-CCP-negative RA patients ranging from 808 to 1918 per locus, our study represents the largest genetic study on anti-CCP negative RA to date. The 31 confirmed RA susceptibility loci identified so far have been primarily established in anti-CCP positive RA, and the meta-analysis by Stahl *et al*¹⁴ included only seropositive RA. The effect size and strength of association of the loci identified in the meta-analysis by Stahl *et al* and in the anti-CCP positive RA subset of this study are very consistent. Slight differences might be related to the difference of power for certain loci and to the use, in some instances, of several different proxies for one locus. It should be noted that we defined the minor allele frequency according to the frequency in the total population, including cases and controls. Since the minor allele frequency is close to 50% for *SPRED2* in the meta-analysis by Stahl, where the minor allele frequency is based on controls only, the G allele is the minor allele, while it is the major allele in this study. This results in an inversion of the OR for association with anti-CCP positive RA. All loci tested here are associated with anti-CCP positive RA with $p < 0.05$, and five loci reach genome-wide significance ($< 5E-08$).

Interestingly, we show a strong and highly significant association of the SE with anti-CCP negative RA; the association remains present after stratification for rheumatoid factor (online supplementary table S2), which is consistent with the findings of a large study performed in patients from the same genetic background.¹³ However, several smaller studies, performed in populations of different origins, did not detect significant association, likely due to a lack of power or ethnic differences. A large European meta-analysis, taking only 2-digit typing into account across several different European populations, concluded that there were weak, but no robust associations between any HLA-DRB1 alleles and anti-CCP negative RA.¹² The number of anti-CCP negative RA patients tested in our study is much higher

Table 3 Schematic classification of RA susceptibility loci into three categories depending on their association pattern in anti-CCP positive and negative RA

Category	Associations	Locus Name
1	Both CCP positive and negative RA, stronger in CCP positive RA	HLA-DRB1 SE, PTPN22, TNFAIP3 locus 1
2	Both CCP positive and negative RA, equally strong in both	ANKRD55, BLK, C5orf30, STAT4
3	CCP positive RA only, significant difference between CCP positive and negative RA	AFF3 locus 1, CCR6, CCL21 locus 2, IL2RA locus 2, CD28, CD40, PXX, REL, RBPJ, TNFRSF14, TNFAIP3 locus 3
Not classifiable	CCP positive RA only, but no significant difference between CCP positive and negative RA	All others

CCP, cyclic citrullinated peptide; RA, rheumatoid arthritis.

than that in the meta-analysis or any other study, and confirms that the SE is associated with anti-CCP negative RA in UK Caucasians.

Compared with the results in anti-CCP positive RA, where all loci show evidence for association, only seven loci achieve a p value below 0.05 in the anti-CCP negative subgroup. However, this number is higher than reported to date, and represents a statistically significant overlap ($p=0.0018$). Among the loci tested here, 32 non-HLA loci have been convincingly shown to be independently associated in anti-CCP positive RA. Therefore, without assuming any prior probability for association, the Bonferroni-corrected significance threshold would be $1.6E-03$. Three non-HLA loci would remain significantly associated with anti-CCP negative RA after the Bonferroni correction: *ANKRD55/IL6ST*, *BLK* and *PTPN22*.

Interestingly, both *BLK* and *STAT4* are confirmed systemic lupus erythematosus (SLE) susceptibility loci and it would be interesting to investigate the overlap of other SLE loci with anti-CCP negative disease. Our findings confirm those of the previous reports showing association in both anti-CCP positive and negative RA subgroups at the *STAT4* locus. *IRF5*, which has been found to be associated with anti-CCP negative RA by previous investigators, is not associated with anti-CCP negative RA in this study. However, it should be noted that the current study only had 19.6% power to detect an association of an effect size of 1.12 at the 0.05 significance level. In addition, a differential association of this marker with RA and its different serotypes in different ethnic groups, or gene-environment interactions, might further account for the lack of association seen here in anti-CCP negative patients.

The effect size of *BLK* and *ANKRD55/IL6ST* is larger in anti-CCP negative RA than in anti-CCP positive RA, although this difference does not reach statistical significance. All other loci show a higher effect size in anti-CCP positive RA. This can be explained by the fact that loci tested here have been primarily identified in cohorts containing mainly or exclusively anti-CCP positive patients.

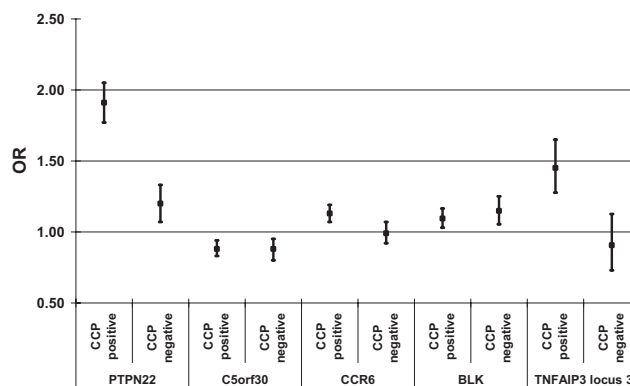


Figure 1 OR and 95% CI for different single nucleotide polymorphism association patterns. *PTPN22*: significant association in both serotypes, significantly different. *C5orf30*: significant association in both serotypes with the same effect size. *CCR6*: only associated in anticyclic citrullinated peptide (anti-CCP) positive rheumatoid arthritis (RA). *BLK*: associated in both serotypes, effect size slightly, but not significantly, larger in anti-CCP negative RA. *TNFAIP3 locus 3*: only associated in anti-CCP positive RA, highly significant difference. *PTPN22*, *C5orf30*, *CCR6* are prototypic examples illustrating the three categories presented in table 3.

The genetic difference between the two RA serotypes can be particularly illustrated by the association pattern of the intronic locus at *TNFAIP3*: it is strongly associated with anti-CCP positive RA, but not with anti-CCP negative RA, despite a detection power of over 80%; the difference in OR is statistically significant. The intronic locus at *TNFAIP3* therefore represents a risk factor for anti-CCP positive RA, but not for anti-CCP negative RA in this analysis.

The present study shows a genetic contrast between anti-CCP positive and anti-CCP negative RA and allows the classification of known RA susceptibility SNPs in different categories. The first category comprises markers associated with both subsets, but their effect size is significantly larger in anti-CCP positive RA. Although anti-CCP negative RA could not be clearly divided into distinct clinical subphenotypes in a recent study in the Netherlands,²⁸ it might still comprise several genetically and serologically different subsets, based for example on the presence of ACPA, other than anti-CCP antibodies.²⁹ The second category contains SNPs, similarly associated in both anti-CCP positive and negative RA, with the same effect size. The third category comprises SNPs associated with anti-CCP positive RA, but not with anti-CCP negative RA and the effect size ratio is statistically significant. A lack of association of some markers, while others are associated with disease irrespective of the serological status, suggests that RA susceptibility markers might cluster to different molecular pathways, some associated with autoantibody production, others not.

In summary, among 33 independent genetic loci tested in this study, 18 could be classified into three different categories, according to their association pattern, while 15 could not, mainly due to lack of power. Seven markers show an association with anti-CCP negative RA, while 11 others are unlikely to be associated. The use of a multinomial logistic regression analysis leading to three p values, as described here (anti-CCP positive RA, anti-CCP negative RA, effect size ratio), represents a straightforward method to classify markers into three meaningful categories or to exclude them from classification, if only one p value out of three is significant. This latter situation occurs mainly when the power is low. The accuracy of classification depends on the definition of the significance threshold. If a Bonferroni corrected p value is used for classification, markers with p values between 0.05 and 1.6E-03 would change category. Markers with highly significant associations and effect size ratio like *PTPN22* can be considered as accurately categorised. Future studies might identify more categories; for example, for markers associated exclusively with anti-CCP negative RA, or displaying a larger effect size in anti-CCP negative RA than in anti-CCP positive RA.

Despite this being the largest sample of anti-CCP negative cases studied to date, the main limitation remains lack of power for many markers. This is particularly pertinent to the loci, which could not be classified into one of the three categories presented here. Larger sample sizes will be required to explore these loci more fully. Six thousand five hundred anti-CCP negative patients and 11 000 controls would be required to detect an effect size of 1.10 with a power of 80% for a marker present at a MAF of 28% in controls (average MAF of RA susceptibility loci in controls reported in the study by Stahl *et al*¹⁴).

The low number of anti-CCP positive RA susceptibility loci associated with anti-CCP negative RA highlights the need for a well-powered GWAS for the discovery of yet unknown anti-CCP negative RA specific loci. The current study presents genetic differences and similarities between anti-CCP positive and anti-CCP negative RA. Although the two disease serotypes

show significant differences in disease course and severity, anti-CCP antibodies are currently not used to guide treatment decisions in clinical practice. However, the results presented here highlight the need for genetic analyses of susceptibility, severity and treatment response to consider the two serotypes both separately and together for future investigations.

Contributors Data analysis: SV, DP and JB. Statistical analysis: SV and ML. Manuscript preparation: SV, SE, AB and JW.

Acknowledgements SV was initially supported by a grant from the Swiss National Science Foundation, grant no. PBGEP3-129009; he is currently supported by a research grant from the Swiss Foundation for Medical-Biological Scholarships (SSMBS), managed by the Swiss National Science Foundation (grant reference number PASMP3_134380). This grant is financed by a donation of Novartis to the SSMBS. The authors thank Arthritis Research UK (grant ref 17552) and the NIHR Manchester Biomedical Research Centre for support.

Funding Swiss National Science Foundation/Swiss Foundation for Medical-Biological Scholarships/Arthritis Research UK

Competing interests None.

Patient Consent Obtained.

Ethics approval The University of Manchester Ethics Committee approved this study.

Provenance and peer review Not commissioned; externally peer reviewed.

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