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# High-Field <sup>13</sup>C DNP with a Radical Mixture

Vladimir K. Michaelis<sup>1,2,‡</sup>, Albert A. Smith<sup>1,2,‡</sup>, Björn Corzilius<sup>1,2</sup>, Oleysa Haze<sup>2</sup>, Timothy M. Swager<sup>2</sup>, and Robert G. Griffin<sup>1,2,\*</sup>

<sup>1</sup>Francis Bitter Magnet Laboratory, Massachusetts Institute of Technology, Cambridge, Massachusetts, 02139 USA

<sup>2</sup>Department of Chemistry, Massachusetts Institute of Technology, Cambridge, Massachusetts, 02139 USA

## Abstract

We report direct <sup>13</sup>C dynamic nuclear polarization at 5 T under magic angle spinning (MAS) at 82 K using a mixture of monoradicals with narrow EPR linewidths. We show the importance of optimizing both EPR linewidth and electron relaxation times by studying direct DNP of <sup>13</sup>C using SA-BDPA and trityl radical, and achieve <sup>13</sup>C enhancements above 600. This new approach may be better suited for dissolution DNP and for studies of <sup>1</sup>H depleted biological and other non-protonated solids.

Over the past few decades, many techniques have been developed for studying chemical structure. NMR spectroscopy in particular has achieved widespread use due to its ability to characterize biological molecules ranging from chemical structure, dynamics, and medium-range (4–6 Å) intra- and intermolecular structure. Solid-state NMR in particular has been especially important in structurally characterizing disordered biological solids, which are inaccessible to traditional diffraction based methods. However, the success of these experiments is limited due to the low Boltzmann polarization of nuclear spins, leading to long acquisition times. To address this issue, solution NMR and magnetic resonance imaging focuses primarily on high- $\gamma$ , abundant nuclei such as <sup>1</sup>H, <sup>19</sup>F and <sup>31</sup>P, while solid-state NMR methods utilizes magic angle spinning (MAS), cross-polarization (CP) and high magnetic fields to obtain modest gains in sensitivity and resolution.

More recently high-field dynamic nuclear polarization (DNP) has been a valuable approach for studying structure, function, and reaction pathways because it allows significant reduction in acquisition times. In a DNP experiment, the large thermal electron spin polarization of a paramagnetic compound is transferred to surrounding nuclei, a process that is driven by irradiating the sample with microwaves.<sup>1,2</sup> Immense gains in sensitivity have been reported for various low- nuclei (e.g., <sup>13</sup>C, <sup>15</sup>N, <sup>17</sup>O, <sup>27</sup>Al and <sup>29</sup>Si) using indirect DNP polarization.<sup>3–6</sup> Typically, a nitroxide-based biradical (e.g., TOTAPOL) is used as the electron polarization source and polarizes <sup>1</sup>H (theoretically reaching ( $\gamma_e/\gamma_H$ )  $\mathbf{e} \approx 660$ ).<sup>7,8</sup> This polarization is then transferred to lower- $\gamma$  nuclei by a CP step, reaching enhancement

#### Author Contributions

<sup>\*</sup>Corresponding Author: Robert G. Griffin, rgg@mit.edu. \*These authors contributed equally.

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Supporting Information: Experimental methods and sample preparation. This material is available free of charge via the Internet at http://pubs.acs.org.

factors (*e*) as high as 248.<sup>9</sup> This approach has been used successfully for structural biology and more recently surface science studies.<sup>4,10–18</sup>

Indirect polarization is extremely attractive as many systems contain <sup>1</sup>H's that are easily polarized. Strong <sup>1</sup>H–<sup>1</sup>H and e'–<sup>1</sup>H couplings allow for efficient DNP and dispersion of polarization via spin-diffusion. Additionally, the wide availability of nitroxide-based biradicals, with broad EPR lines, allows high- $\gamma$  nuclei such as <sup>1</sup>H to be efficiently polarized by the cross-effect (CE). Unfortunately, many chemical systems do not fall into this category, because they are severely lacking <sup>1</sup>H's which limits the ability to efficiently cross-polarize. An attractive approach in these circumstances is to perform direct polarization of low- $\gamma$  nuclei such as <sup>13</sup>C without the CP step from <sup>1</sup>H.

In this paper, we utilize a mixture of two narrow-linewidth radicals that have EPR resonance frequencies that are approximately separated by the <sup>13</sup>C nuclear Larmor frequency as the polarizing agent. Concurrently, since that exhibit different relaxation rates, we can optimizing both the CE matching condition and the DNP kinetics. With this mixture, we obtain record <sup>13</sup>C DNP enhancements > 600 (nearly 25 % of the theoretical enhancement ( $\gamma_e/\gamma_C$ )  $\mathbf{e} \approx 2620$ ).

The development of high-field DNP has focused on the CE mechanism, since typical solid effect (SE) enhancements had been considerably lower than those for CE.<sup>19</sup> However, recent results have shown that SE may be useful for polarization using transition-metal based polarizing agents<sup>20</sup> and can also give enhancements ~100;<sup>21,22</sup> with sufficient microwave field strength, sensitivity gains may match those of CE.<sup>23</sup> The dominant polarization transfer process depends on the nucleus being polarized and the EPR characteristics of the polarizing agent. In particular, the relative magnitudes of the electron homogeneous ( $\delta$ ) and inhomogeneous ( $\Delta$ ) linewidths, and the nuclear Larmor frequency ( $\omega_{01}$ ) determine the dominant polarization mechanism.

The SE mechanism is a two-spin process which is dominant when  $\omega_{0I} > \delta$ ,  $\Delta$  and microwave irradiation is applied at the electron-nuclear zero- or double-quantum frequency.<sup>24–26</sup> This matching condition is given by

$$\omega_{\rm mw} = \omega_{\rm os} \pm \omega_{\rm ol}$$
 (1)

where  $\omega_{0S}$  is the electron Larmor frequency and  $\omega_{mw}$  is the micro- wave frequency.

The CE mechanism may be described as a three-spin flipflop-flip process between two electrons and a nucleus, which is dominant when  $\Delta > \omega_{0I} > \delta$ . In order to achieve maximum efficiency the difference between the two electron Larmor frequencies must be near the nuclear Larmor frequency.<sup>27–31</sup>

$$\omega_{0I} = \omega_{0S_2} - \omega_{0S_1}$$
 (2)

Optimizing the polarizing agents used for SE and CE is a non-trivial task, as discussed recently by Hu.<sup>32</sup> Both the EPR line-shape and the electron spin-lattice relaxation time ( $T_{1,S}$ ) must be considered. For SE, one applies microwave irradiation at the matching condition given in (eq, 1); ideally, one uses a polarizing agent with a narrow EPR spectrum, thus allowing the matching condition to be met for the majority of unpaired electrons in the system. Also, a short electron  $T_{1,S}$  allows quick 'recycling' of SE, since the electron must quickly recover its polarization in order to polarize many nuclei. However, there is an

optimum  $T_{1S}$  since a short  $T_{1S}$  leads to paramagnetic relaxation of nearby nuclei, thus destroying the polarization already transferred to nuclei.

For CE, one destroys the thermal polarization of one electron with microwave irradiation. This electron then recovers its polarization via a flip-flop-flip process with a second electron and a nucleus. This process is efficient when eq. 2 is satisfied. Therefore, the ideal polarizing agent includes two different radicals, each with narrow EPR resonances, which are separated by the nuclear Larmor frequency. Note that the recovery of polarization of the first electron occurs via two competing processes. The first process is the CE mechanism as just described. However, the second process is the usual electron  $T_{1,S}$  relaxation. Therefore, if the  $T_{1,S}$  of the first electron is long, the CE mechanism dominates and polarization transfer is more efficient, as recently demonstrated by Zagdoun et al.<sup>8</sup> This is not a complete picture, however. The second electron must provide polarization, as does the electron in SE. Therefore, quick 'recycling' of the CE mechanism relies on a sufficiently short  $T_{1,S}$  of the second electron. If the two electrons have the same  $T_{1,S}$ , as is the case for most nitroxide biradicals, then one must compromise on the  $T_{1,S}$ . However, if two different radicals are used, then one can select polarizing agents such that the first electron has a long  $T_{1,S}$  and the second electron has a shorter  $T_{1,S}$ .

By using a mixture of SA-BDPA and trityl, we demonstrate efficient cross effect using two radicals with relatively narrow EPR linewidths, being 28 and 50 MHz, respectively. The centers of the EPR spectra are separated by roughly the <sup>13</sup>C Larmor frequency (34 MHz separation vs. 53 MHz Larmor frequency at 5 T; trityl is broad enough to make up the difference). Additionally, SA-BDPA has a long  $T_{1,S}$ , whereas the  $T_{1,S}$  of trityl is shorter<sup>33</sup>, giving improved CE performance when irradiating near the SA-BDPA resonance.

<sup>13</sup>C direct polarization magic-angle spinning NMR experiments were performed using two organic water-soluble polarizing agents, SA-BDPA<sup>33</sup> and trityl OX063,<sup>34</sup> which are depicted in Figure 1. Both SE and CE must be considered in this study. To evaluate the dominating DNP mechanism, the <sup>13</sup>C DNP enhancement field-profiles were measured via direct detection, where the magnetic field was adjusted between 4977 and 4990 mT (Figure 2) and 8 W of microwave output power was chosen for long term stability (>6 hours).

In Figure 2A, we show the EPR spectra of SA-BDPA and trityl, acquired at 140 GHz (the field axis is adjusted to align with the DNP experiments at 139.66 GHz). We also mark the center of each spectrum in black, and mark the field positions that are predicted to be optimal for SE DNP for both SA-BDPA (blue) and trityl (red). In Figure 2B–D, <sup>13</sup>C-DNP field profiles are shown for SA-BDPA, trityl, and a 1:1 mixture, each with a total radical concentration of 40 mM.

Figure 2B shows the field profile using only SA-BDPA. From the EPR spectrum, we see that the line of SA-BDPA is narrower than the <sup>13</sup>C Larmor frequency. Therefore, we expect that SE is strongly dominating the DNP transfer. This is confirmed by the fact that there is a plateau in the enhancement at the center of the DNP field profile, which is characteristic of a well-resolved SE. Also the position of maximum enhancement is in good agreement with the position predicted for SE.

The trityl EPR spectrum is considerably broader than that of SA-BDPA, and with a linewidth of 50 MHz it is possible for both SE and CE to contribute to the DNP enhancements. The trityl DNP profile is given in Figure 2C. The asymmetry of enhancements (–380 vs. 480), and the lack of a plateau in the center of the field profile suggest that CE is making some contribution to the DNP enhancement. However, the extrema of the DNP profile are in good agreement with those predicted for SE DNP, and so one sees that SE also likely is making a major contribution.

In Figure 2D, the <sup>13</sup>C DNP field profile is shown for a mixture of SA-BDPA and trityl. In this case, there are many contributing DNP processes: SE resulting from SA-BDPA, SE and CE resulting from trityl, and finally CE resulting from the interaction between SA-BDPA and trityl. Careful examination reveals that the major features seen in the field profile are at nearly the same positions as seen in the pure SA-BPDA and trityl profiles. We first examine the features seen at 4980.7 and 4984.4 mT, which correspond to SE from SA-BDPA. The intensity of these features is remarkable, giving enhancements at these positions of  $\pm 535$  and 575, respectively. Although other processes contribute to these large enhancements, it appears that SE from SA-BDPA is at least as effective in this sample as it was in the pure SA-BDPA sample, despite the total concentration of SA-BDPA of the mixed sample being half of that in the pure sample. This is further confirmed in Figure 3, where the SA-BDPA DNP field profile is subtracted from the mixture field profile, using a coefficient of 1.1. This coefficient was chosen in order to fully remove the features resulting from SE using SA-BDPA, without introducing other features. This implies that SE resulting from SA-BDPA is somehow more effective in this sample than it is in the pure sample. In fact, this is a result of a reduced electron  $T_{1S}$  for SA-BDPA in the mixed sample. Paramagnetic relaxation from trityl actually shortens the SA-BDPA  $T_{1S}$ , allowing SA-BDPA to become more efficient at polarizing the <sup>13</sup>C nuclei. This is seen in Table 1, where  $T_{1S}$  of SA-BDPA changes from 28.9 ms in the pure SA-BDPA sample to 3.6 ms in the mixture. We also note that any change in the trityl  $T_{1S}$  is negligible.

Once the contribution of SE from SA-BDPA is removed, as is shown in Figure 3, we see a highly asymmetric DNP field profile remaining. This profile contains contributions from pure trityl SE and CE, but also CE resulting from SA-BDPA and trityl. We already know that the pure trityl field profile is not nearly as asymmetric as this profile, as seen in Figure 2C, and so this asymmetry is likely a result of CE from SA-BDPA and trityl. In fact, this is exactly what we expect. If we irradiate near to the center of the SA-BDPA spectrum, the SA-BDPA radical saturates easily, since it has a longer  $T_{1S}$ . Therefore, the polarization recovery of the SA-BDPA electron occurs primarily via the CE mechanism. Furthermore, the fast  $T_{1S}$  of trityl allows for quick recycling of the CE mechanism, giving an overall more efficient CE mechanism. However, when irradiating closer to the center of the trityl spectrum, saturation is more difficult because the  $T_{1,S}$  of trityl is shorter, and the source of polarization, SA-BDPA, recovers more slowly. This leads to an overall less efficient CE mechanism. We note that the peaks do not lie exactly on the SA-BDPA or trityl centers. This is both because SA-BDPA and trityl do not have exactly the correct separation for the <sup>13</sup>C cross effect matching, and also because contributions to the DNP enhancement remain from the pure trityl. One should note that previous experiments by Hu et al. showed that mixtures of TEMPO and trityl were efficient in polarizing <sup>1</sup>H.<sup>19</sup> In particular, DNP was efficient when irradiating the narrow, trityl radical. However, it was not clear whether improvements were primarily due to a longer electron T<sub>1</sub> for trityl, or due to the considerably narrower lineshape of trityl compared to TEMPO.

Both nuclear spin-lattice relaxation  $(T_{1I})$  and polarization build-up  $(T_B)$  times for <sup>13</sup>C were measured at 82 K. Both  $T_B$  and  $T_{1I}$  values were within error of one another for each radical composition, and so only  $T_B$  is listed in Table 1. The polarization build-up times varied between the radicals, however, with trityl being the shortest and SA-BDPA being the longest. Radical concentrations and sample conditions were identical, enabling direct comparison between the effect of the nuclear and electron relaxation characteristics. The observed reduction in polarization times for each must be attributed to the inherent electron  $T_{1S}$  of the radical, and the type of DNP mechanisms that are active.

Direct <sup>13</sup>C polarization enhancements were measured at the maximum positive field positions using optimized recycle delays  $(1.26 \times T_B)$  for each radical composition. Off-

signals (0 W micro-wave power) were acquired under identical conditions as the on-signals (10 W microwave power) for SA-BDPA (4984.5 mT), trityl (4983.2 mT), and mixture (4983.1 mT), respectively. SA-BDPA and trityl provided enhancements of 300 and 480, respectively, at 82 K. SA-BDPA is comparable to previous studies on <sup>13</sup>C detection using TOTAPOL obtaining enhancements of 305 under similar sample conditions.<sup>35</sup> Trityl proves slightly more efficient in polarizing as the breadth better satisfies the cross-effect mechanism, and provides ~60% larger enhancement. The mixed radical sample satisfies the CE matching condition most efficiently, and the beneficial differences in electron relaxation times (*T*<sub>1,5</sub>) lead to a very effective cross-effect enhancement of 620. The mixture outperforms trityl (> 130 %) and SA-BDPA (> 200 %), and also outperforms previous experiments which utilize TOTAPOL for <sup>13</sup>C DNP (> 200 %). <sup>35</sup>

These narrow line radicals may be ideally suited for directly polarizing low- $\gamma$  nuclei that are either poorly cross-polarized by high- $\gamma$  nuclei (e.g., <sup>1</sup>H or <sup>19</sup>F) or are found within environments in which high- $\gamma$  nuclei are absent. Introducing a mixture approach whereby better satisfying the CE matching condition and having different  $T_{1,S}$  relaxation times could have drastic implications providing a more efficient CE mechanism and may help compensate the decrease in enhancements as DNP experiments move to higher magnetic fields. These findings may be beneficial for dissolution DNP methods that commonly use trityl radical, since one may combine SA-BDPA and trityl. Then, polarizing at liquid He temperatures could lead to significant polarization gains on labeled dissolution experiments without modification of any hardware. Finally, if trityl and SA-BDPA can be tethered together to create a biradical, it should be possible to reduce radical concentrations, increase the contribution of CE to the DNP enhancements, and potentially increase DNP gains in the future.

#### Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

#### Acknowledgments

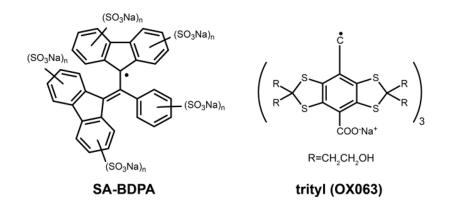
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### References

- 1. Carver TR, Slichter CP. Physical Review. 1953; 92:212.
- 2. Overhauser AW. Physical Review. 1953; 92:411.
- Becerra L, Gerfen G, Temkin R, Singel D, Griffin R. Physical Review Letters. 1993; 71:3561. [PubMed: 10055008]
- Lesage A, Lelli M, Gajan D, Caporini MA, Vitzthum V, Mieville P, Alauzun J, Roussey A, Thieuleux C, Mehdi A, Bodenhausen G, Coperet C, Emsley L. Journal of the American Chemical Society. 2010; 132:15459. [PubMed: 20831165]
- Michaelis VK, Markhasin E, Daviso E, Herzfeld J, Griffin RG. The Journal of Physical Chemistry Letters. 2012; 3:2030. [PubMed: 23024834]
- Vitzthum V, Mieville P, Carnevale D, Caporini MA, Gajan D, Coperet C, Lelli M, Zagdoun A, Rossini AJ, Lesage A. Chemical Communications. 2012; 48:1988. [PubMed: 22237253]
- 7. Song C, Hu KN, Joo CG, Swager TM, Griffin RG. J Am Chem Soc. 2006; 128:11385. [PubMed: 16939261]

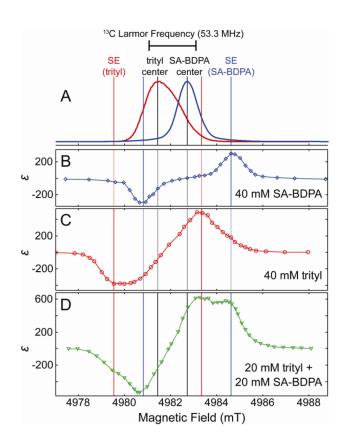
- Zagdoun A, Casano G, Ouari O, Lapadula G, Rossini AJ, Lelli M, Baffert M, Gajan D, Veyre L, Maas WE, Rosay MM, Weber RT, Thieuleux C, Coperet C, Lesage A, Tordo P, Emsley L. J Am Chem Soc. 2012; 134:2284. [PubMed: 22191415]
- 9. Kiesewetter MK, Corzilius B, Smith AA, Griffin RG, Swager TM. J Am Chem Soc. 2012; 134:4537. [PubMed: 22372769]
- Lelli M, Gajan D, Lesage A, Caporini MA, Vitzthum V, Mieville P, Heroguel F, Rascon F, Roussey A, Thieuleux C, Boualleg M, Veyre L, Bodenhausen G, Coperet C, Emsley L. J Am Chem Soc. 2011; 133:2104. [PubMed: 21280606]
- Debelouchina GT, Bayro MJ, van dWPCA, Caporini MA, Barnes AB, Rosay M, Maas WE, Griffin RG. Phys Chem Chem Phys. 2010; 12:5911. [PubMed: 20454733]
- Bajaj VS, Mak-Jurkauskas ML, Belenky M, Herzfeld J, Griffin RG. Proc Natl Acad Sci U S A, Early Ed. 2009; 106:9244.
- Hall DA, Maus DC, Gerfen GJ, Inati SJ, Becerra LR, Dahlquist FW, Griffin RG. Science. 1997; 276:930. [PubMed: 9139651]
- 14. Van, dWPCA.; Hu, K-N.; Lewandowski, J.; Griffin, RG. J Am Chem Soc. 2006; 128:10840. [PubMed: 16910679]
- Bayro MJ, Debelouchina GT, Eddy MT, Birkett NR, MacPhee CE, Rosay M, Maas WE, Dobson CM, Griffin RG. J Am Chem Soc. 2011; 133:13967. [PubMed: 21774549]
- Sergeyev IV, Day LA, Goldbourt A, McDermott AE. Journal of the American Chemical Society. 2011; 133:20208. [PubMed: 21854063]
- Akbey Ü, Franks WT, Linden A, Lange S, Griffin RG, van Rossum BJ, Oschkinat H. Angewandte Chemie International Edition. 2010; 49:7803.
- Renault M, Pawsey S, Bos MP, Koers EJ, Nand D, Tommassenvan Boxtel R, Rosay M, Tommassen J, Maas WE, Baldus M. Angewandte Chemie International Edition. 2012; 51:2998.
- 19. Hu K, Bajaj V, Rosay M, Griffin R. J Chem Phys. 2007; 126:044512. [PubMed: 17286492]
- Corzilius B, Smith AA, Barnes AB, Luchinat C, Bertini I, Griffin RG. Journal of the American Chemical Society. 2011; 133:5648. [PubMed: 21446700]
- Smith AA, Corzilius B, Barnes AB, Maly T, Griffin RG. Journal of Chemical Physics. 2012; 136:015101. [PubMed: 22239801]
- 22. Corzilius B, Smith AA, Griffin RG. J Chem Phys. 2012; 137:054201. [PubMed: 22894339]
- Smith AA, Corzilius B, Bryant JA, DeRocher R, Woskov PP, Temkin RJ, Griffin RG. J Magn Reson. 2012; 223:170. [PubMed: 22975246]
- 24. Abragam A, Proctor WGCR. Hebd Seances Acad Sci. 1958; 246:2253.
- 25. Jeffries CD. Physical Review Phys Rev PR. 1960; 117:1056.
- 26. Hovav Y, Feintuch A, Vega S. Journal of Magnetic Resonance. 2010; 207:176. [PubMed: 21084205]
- Kessenikh AV, Lushchikov VI, Manenkov AA, Taran YV. Soviet Physics Solid State. 1963; 5:321.
- 28. Hwang CF, Hill DA. Physical Review Letters. 1967; 18:110.
- 29. Hu KN, Debelouchina GT, Smith AA, Griffin RG. Journal of Chemical Physics. 2011; 134:125105. [PubMed: 21456705]
- Hovav Y, Feintuch A, Vega S. Journal of Magnetic Resonance. 2012; 214:29. [PubMed: 22119645]
- 31. Thurber KR, Tycko R. Journal of Chemical Physics. 2012; 137:084508. [PubMed: 22938251]
- 32. Hu KN. Solid State Nuclear Magnetic Resonance. 2011; 40:31. [PubMed: 21855299]
- 33. Haze O, Corzilius B, Smith AA, Griffin RG, Swager TM. J Am Chem Soc. 2012; 134:14287. [PubMed: 22917088]
- Ardenkjaer-Larsen J, Laursen I, Leunbach I, Ehnholm G, Wistrand L, Petersson J, Golman K. J Magn Reson. 1998; 133:1. [PubMed: 9654463]
- 35. Maly T, Miller AF, Griffin RG. ChemPhysChem. 2010; 11:999. [PubMed: 20169604]

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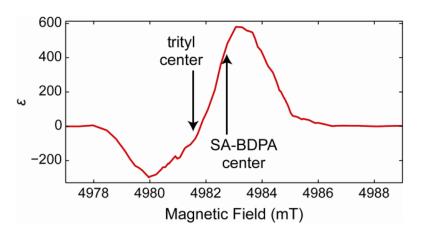
Narrowline monoradical chemical structures of SA-BDPA and trityl (OX063).



#### Figure 2.

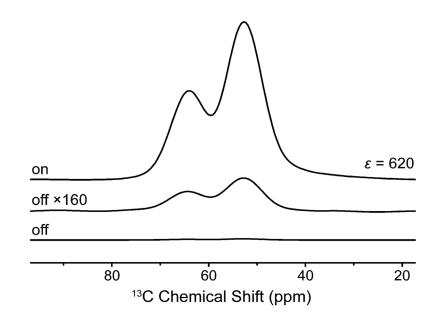
Field-dependent <sup>13</sup>C DNP enhancement profiles of SA-BDPA (B), trityl (C), and a mixture (D), with EPR spectra of SA-BDPA and trityl (A). Field profiles were recorded at 82 K with a microwave frequency of 139.66 GHz, 8 W of microwave power and a MAS frequency of 4.8 kHz.





#### Figure 3.

Subtracted DNP field profile, obtained by subtracting the SA-BDPA DNP field profile (Figure 2B) multiplied by 1.1 from the SA-BDPA and trityl mixture DNP field profile.



#### Figure 4.

<sup>13</sup>C direct polarization using the mixed radical SA-BDPA (20 mM) and trityl (20 mM) on U-<sup>13</sup>C-glycerol-d8 (60%) in D<sub>2</sub>O (30%) and H<sub>2</sub>O (10%) at 82 K and 10 W (B<sub>o</sub> = 4983.1 mT).

#### Table 1

<sup>13</sup>C direct detection enhancement e, electron spin relaxation time  $T_{1S}$  and DNP build-up time  $T_{B}$ .

Radical	е	<i>T</i> <sub>15</sub> (ms)	$T_{\rm B}\left({ m s} ight)$
SA-BDPA (40 mM)	300	28.9	287 (16)
trityl (40 mM)	480	1.4	167 (7)
Mixture (20 mM of each)	620	3.6 (SA-BPDA) 1.4 (trityl)	216 (3)