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INTERLEUKIN-4- AND INTERLEUKIN-13-MEDIATED ALTERNATIVELY ACTIVATED MACROPHAGES: ROLES IN HOMEOSTASIS AND DISEASE

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Abstract

The macrophage, a versatile cell type prominently involved in host defense and immunity, assumes a distinct state of alternative activation in the context of polarized type 2 immune responses such as allergic inflammation and helminth infection. This alternatively activated phenotype is induced by the canonical type 2 cytokines interleukin (IL)-4 and IL-13, which mediate expression of several characteristic markers along with a dramatic shift in macrophage metabolic pathways that influence surrounding cells and tissues. We discuss recent advances in the understanding of IL-4- and IL-13-mediated alternatively activated macrophages and type 2 immune responses; such advances have led to an expanded appreciation for functions of these cells beyond immunity, including maintenance of physiologic homeostasis and tissue repair.

Keywords

Th2 cell; type 2 immunity; helminth; allergy; metabolism

INTRODUCTION

An essential element of metazoan survival is the capacity to detect and respond to foreign material encountered at barrier surfaces. This ability constitutes immunity at its most basic level, serving to rapidly and efficiently defend the host from intrusion by pathogenic invaders, although similar mechanisms can also select and retain agents that provide some host benefit (1). On a cellular level, the task of distinguishing foreign material is primarily carried out by the macrophage, an incredibly versatile cell type that has been widely conserved throughout phylogeny; macrophage-like sentinel cells that exhibit hallmark phagocytic and microbicidal functions arise even among colonies of the social amoeba *Dictyostelium discoideum* (2). Macrophages not only possess a broad array of cell surface receptors and intracellular mediators that enable recognition, engulfment, and destruction of pathogens, but they are also equipped with essential secretory molecules that serve to regulate the activities of neighboring cells. And although a century has passed since Metchnikoff first described the critical immune function of macrophages and phagocytosis in the detection and elimination of harmful microbes—observations that have since formed a fundamental tenet of modern immunology—new and diverse roles for macrophages during immune responses and various physiologic processes continue to be discovered.

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Metchnikoff's initial concept now incorporates a host of molecular detail about how macrophages recognize, phagocytose, and kill microbes. In this respect, the characterization of several germ line–encoded pattern-recognition receptors (PRRs) and subsequent intracellular signaling events induced upon PRR ligation has been instrumental in providing molecular links between the initial sensing of foreign material and the distinct cellular changes that occur in response to different classes of microbial patterns (3, 4). This paradigm is preserved throughout many phyla, constituting most invertebrate innate immunity, but also underlies the highly evolved adaptive immune system in mammals and other vertebrates. A hallmark example occurs within mammalian macrophages upon exposure to Toll-like receptor (TLR) ligands and the cytokine interferon- γ (IFN- γ), which induce a phenotype referred to as the classically activated macrophage (CAM), marked by a metabolic shift to anaerobic glycolysis, increased production of proinflammatory cytokines, and new synthesis of reactive oxygen and nitrogen species, enabling efficient killing of engulfed microbes. Elements of this system are rooted in similar macrophage-like phagocytic cells found among invertebrates, notably *Drosophila*, that exhibit striking homology with mammalian innate immune function on both genetic and cellular levels (5).

Mammals also engender a unique tissue response, encompassed by the term type 2 immunity, that diverges significantly from that associated with the CAM and is prominently associated with parasite infection and allergic disorders; however, it is also increasingly recognized to be coupled with broader mechanisms that maintain homeostasis in larger organisms (6, 7). Type 2 immunity consists of a complex orchestration of cells and cytokines involving an initial response, typically generated at mucosal barriers, which is amplified by activation and recruitment of innate and adaptive effector cells and followed by a resolution phase involving tissue repair mechanisms. Although many cell types and molecules are involved, most of these aspects are coordinated by the activities of the canonical type 2 cytokines interleukin (IL)-4 and IL-13, two closely related cytokines that play both overlapping and distinct roles in type 2 immunity. This cytokine-mediated program is required for the generation of high-affinity IgE antibodies, mucus overproduction, and smooth muscle alterations that produce the “weep and sweep” response in mammals, which mediates expulsion of large extracellular parasites but is also engaged in the context of asthma and allergic disorders for reasons that remain unclear (6--8). Chronic infection with some parasites that subvert the expulsive response leads to walled-off granulomas containing mixed macrophage- and eosinophil-rich inflammatory infiltrates, suggesting that type 2 immunity also functions as a means to suppress unrestricted inflammation, an important property in the maintenance of tissue homeostasis and host survival.

In this respect, type 2 immune responses direct phenotypic and functional changes within macrophages that are distinct from, and often oppose, the CAM phenotype. Particular alterations in endocytic and enzymatic activities shift cellular resources away from the generation of molecules associated with proinflammatory cytokine and nitric oxide (NO) production and toward the synthesis of anti-inflammatory and tissue repair factors. This distinct state, defined by exposure to IL-4 or IL-13, was first described *in vitro* as the alternatively activated macrophage (AAM) by Gordon and colleagues (9), who contrasted the effects of IFN- γ on macrophages with those of IL-4; these findings were subsequently recapitulated using IL-13 (10, 11). CAM and AAM have been assigned the additional terms M1 and M2, respectively, corresponding to the classic Th1 and Th2 designations, which have provided an essential framework for CD4 T cell research for decades (12). Much remains unclear, though, about how the multiple macrophage phenotypes induced by cytokines and other stimuli relate to specific transcriptional programs and/or epigenetic changes and how these factors might correspond to the heterogeneity and plasticity within the macrophage lineage, both in the steady state and during inflammatory processes

(13--15). In light of these considerations, we focus this review on the central role of the canonical type 2 cytokines IL-4 and IL-13 in mediating the alternatively activated phenotype in macrophages, emphasizing recent insights about how regulation of these cytokines can lead to differential signaling events and a profound metabolic shift that is reflected in specific tissue responses—not only in the context of type 2 immunity but also in the maintenance of homeostasis during complex physiologic processes (Figure 1).

MACROPHAGE DEVELOPMENT AND ACTIVATION

In mammals, macrophages develop from precursors that arise during multiple waves of embryonic hematopoiesis, occurring initially within extraembryonic yolk sac tissue and then subsequently in the aortic endothelium of the aorta-gonad-mesonephros region and in the placenta, fetal liver, and bone marrow (16, 17). Most adult macrophage, monocyte, and dendritic cell (DC) populations derive from bone marrow–derived myeloid precursors, which critically depend on cytokine signaling via colony-stimulating factor 1 receptor (CSF1-R) and transcription factors such as the Ets family member PU.1 (18, 19). Many tissue macrophages, however, are refractory to irradiation protocols and are poorly replaced after bone marrow transplantation, suggesting that not all adult macrophage populations derive from the bone marrow–derived hematopoietic compartment. Interestingly, recent fate-mapping studies have demonstrated that primitive yolk sac–derived myeloid precursors contribute substantially to microglial cell populations in the adult central nervous system and also, to varying degrees, to several tissue-resident macrophage populations found in kidney, lung, liver, pancreas, spleen, and skin, some of which exhibit self-renewal capability in the absence of Myb-dependent, bone marrow–derived stem cells and PU.1 (20--23). In addition, although most bone marrow–derived populations depend primarily on colony-stimulating factor 1 (CSF-1) signaling via CSF1-R for their development and survival, some self-renewing tissue macrophages, in particular microglia, require tissue-restricted signals derived exclusively from the alternate ligand of CSF1-R, IL-34 (24).

The issue of macrophage development and turnover is relevant, as in situ self-renewal and proliferation may define a unique property among AAM precursor populations during type 2 inflammation. This concept is supported by evidence that tissue-resident macrophages proliferate locally during helminth infection or in response to IL-4 treatment (22). The capacity for in situ self-renewal, however, does not appear to preclude overlaid contributions of blood-derived monocyte populations that assume the AAM phenotype during inflammatory responses, although monocyte subset characteristics in this regard appear to vary by experimental system (22, 25, 26). Further to this point, microglial cells can assume an IL-4-dependent AAM phenotype in vitro and during experimental autoimmune encephalomyelitis (27); however, during fungal invasion of the central nervous system only recruited monocyte-derived macrophages, not microglia, exhibit AAM markers (28). Among differentiated macrophages, self-renewal depends on suppression of the transcription factors MafB and c-Maf along with concomitant upregulation of the stem cell-inducing factors Krüppel-like factor 4 (KLF4) and c-Myc, offering potential mechanistic insights into how in situ proliferation may occur during AAM polarization (29). Notable in this regard is the finding that KLF4 is induced by IL-4 in a signal transducer and activator of transcription 6 (STAT6)-dependent manner and can directly regulate the AAM phenotype in vitro and in vivo (30). Aspects of this type of regulation may also apply to recruited monocytic AAM precursors, as bone marrow–derived macrophages deficient in *Jmjd3*, which is implicated in the epigenetic regulation of AAM polarization (discussed further below), show impaired cell cycle progression and decreased expression of cell cycle regulatory genes, including c-Myc and c-Myb, in response to CSF-1 signals (31). In human monocytes IL-4 treatment also induces c-Myc, which can regulate expression of a subset of AAM genes, including STAT6 (32).

PHENOTYPIC CHARACTERISTICS OF AAMS

Macrophages are inherently difficult to classify by marker analysis, due to phenotypic heterogeneity *in vitro* and *in vivo* as these cells develop, migrate, take up residence in diverse tissues, and become activated by various stimuli (13, 14). Nevertheless, AAMs have been characterized by a distinct set of cell surface receptors and intracellular factors that are induced upon exposure to IL-4/IL-13, and their *in vivo* functional relevance is beginning to be explored using models of genetic deficiency. Many of these factors have become variably employed as definitive markers of AAMs, but important caveats to such approaches exist: expression that is not restricted to macrophages, induction by stimuli other than IL-4/IL-13, and differences in expression between species (e.g., mouse versus human macrophages). A notable example in this regard is the signature mouse AAM marker arginase 1 (Arg1), which does not mark human AAMs (33), possibly reflecting compensation by functional isozymes or expression by other cell types. Consequently, although gene profiling studies have yielded additional markers to verify (34--37), these caveats caution against the exclusive use of any single marker in establishing the AAM phenotype; rather, a combinatorial approach should be employed in parallel with an assessment of contributions by IL-4 or IL-13 (14, 15). Some of the most prominent characteristic AAM markers are discussed below.

Arginase 1

Arg1 is a manganese metalloenzyme that catalyzes the conversion of L-arginine into L-ornithine and urea, a critical final step in the urea cycle that occurs primarily in the liver. Arg1 is also induced by IL-4/IL-13 in macrophages and functions to counterbalance inflammatory signals that generate inducible nitric oxide synthase (iNOS), which also utilizes L-arginine to produce nitric oxide and L-citrulline (38). As such, L-arginine metabolism in macrophages is a key determinant in defense against infection by both intracellular and extracellular pathogens and has emerged as a defining feature of alternative versus classical macrophage activation. Several studies support a model in which IL-4/IL-13-induced Arg1 and IFN- γ -induced iNOS compete for limiting L-arginine substrate, such that expression of either enzyme shifts the subsequent generation of metabolites, thereby suppressing the opposing pathway (39--43). Although general aspects of this paradigm are well supported, the *in vivo* effects of manipulating various stages in this metabolic pathway have revealed a more complex regulatory network.

A key step is Arg1-mediated production of L-ornithine, which can subsequently be decarboxylated by L-ornithine aminodecarboxylase (ODC) to produce polyamines or converted to proline by L-ornithine aminotransferase (OAT), implicating Arg1 in regulation of cell growth and collagen production, respectively. Indeed, when ODC activity was inhibited during *Schistosoma mansoni* egg-induced liver inflammation, granuloma size and fibrosis increased, presumably via increased substrate availability to OAT for proline production and collagen synthesis (43). Mice with myeloid cell-specific Arg1 deficiency, however, died at an accelerated rate in response to chronic *S. mansoni* infection, which was associated with increased type 2 cytokine production, granulomatous inflammation, and liver fibrosis (44). This is consistent with findings from acutely *S. mansoni*-infected Arg1-deficient bone marrow chimeric mice (45), suggesting that Arg1 in myeloid cells is not required for fibrotic responses in this system but instead plays a critical role in suppressing unrestrained pathologic inflammation. Further to this point, although Arg1-deficient macrophages induced expression of other AAM markers [mannose receptor (MR) and the chi-lectin Ym1] after type 2 cytokine exposure, they failed to inhibit *in vitro* T cell proliferation in a manner that could be restored by addition of exogenous L-arginine. This indicated that macrophage Arg1 activity depleted an essential proliferative factor, thereby providing a mechanistic basis for the exacerbated Th2 pathology observed *in vivo* (44). In support of these findings, high arginase activity in nonhealing *Leishmania major* lesions

impaired lesional CD4 T cell proliferation and IFN- γ production, whereas inhibition of arginase or administration of exogenous L-arginine restored T cell function and improved control of parasite replication (46), echoing earlier studies that described the T cell-suppressive effect of parasite-elicited macrophages (47).

L-arginine availability is additionally regulated by expression of cationic amino acid transporter 2 (CAT2), which is induced in concert with Arg1 and also acts to dampen T cell responses (38, 48). Absence of substrate in this setting also affects the iNOS-mediated pathway of NO production, as CAT2-deficient mice are more susceptible to infection with *Toxoplasma gondii*, in part due to impaired NO production, and CAT2-deficient mice infected with *S. mansoni* die at an accelerated rate, exhibiting exacerbated granuloma formation and fibrosis associated with increased arginase activity in fibroblasts and AAMs (49). Notably, mice deficient in CAT2 exhibit spontaneous inflammation in the lung marked by increased numbers of eosinophils, neutrophils, memory T cells, activated DCs, and alveolar macrophages that produce lower NO in response to IFN- γ and lipopolysaccharide (LPS) stimulation (50). It remains unclear, however, how steady-state lung inflammation in these mice might relate to the in vivo phenotypes induced by pathogen infection. Nevertheless, other potential regulators of L-arginine metabolism include polyamines themselves, as a recent report shows that IL-4 can induce polyamine production via Arg1-independent pathways, further modulating AAM marker levels (51). Other studies indicate that suppression of T cell proliferation by AAMs relies on cell-cell contact (52), perhaps due to a receptor-mediated process, such as ligation of the inhibitory PD-1 receptor on T cells by AAM-expressed ligands during helminth infection (53, 54).

Importantly, although IL-4-mediated Arg1 induction in AAMs relies on STAT6 (42), Arg1 can also be induced by STAT3-dependent activation in the absence of other AAM markers after TLR stimulation or *Mycobacterium tuberculosis* infection, via signaling through the TLR adapter myeloid differentiation marker 88 (MyD88) and the transcription factor CCAAT/enhancer binding protein β (C/EBP β) (55, 56). These studies not only reinforce the need to examine multiple markers in combination with IL-4/IL-13/STAT6 analysis in definitively establishing the AAM phenotype but also reveal a complex integration of host- and pathogen-derived signals within macrophages that converges on a critical effector pathway. Expression of Arg1 in this system can dramatically influence subsequent biological outcomes, as illustrated in an apparent immune evasion strategy employed by the intracellular pathogens *T. gondii* and *M. tuberculosis*, which act to inhibit protective NO production by engaging the STAT6-independent Arg1 pathway, effectively enhancing pathogen survival; the absence of macrophage Arg1 promotes host survival and reduced pathogen load (55, 56). Furthermore, specific strains of *T. gondii* encode rhopty kinase ROP16, which can directly activate STAT6 to induce host Arg1 expression (57, 58); however, ROP16-deficient *T. gondii* also exhibit increased replication and dissemination in vivo, possibly resulting from increased arginine availability (59), suggesting particular strain-specific effects. Nevertheless, other pathogens, such as *Francisella tularensis*, also induce a switch from the CAM to the AAM phenotype that results in blunted pathogen killing (60), and *Helicobacter pylori* (61) and *Leishmania* (62) encode endogenous arginases that suppress host macrophage NO production in establishing an immune-suppressed niche. Notably, concurrent STAT3 activation observed in some systems may represent a point of intersection with additional cytokines, such as IL-6, GM-CSF, and IL-10, as well as suppressor of cytokine signaling (SOCS) proteins, particularly SOCS1 and SOCS3, which have also been implicated in the regulation of AAM polarization (56, 63). Exactly how these factors interconnect remains to be established, but their participation in generation of AAM markers may be recapitulated in other settings where IL-4/IL-13 is lacking.

Mannose Receptor

MR (CD206; Mrc1) is a C-type lectin receptor (CLR) expressed widely among tissue macrophages, DCs, and endothelial cells. MR can be cell surface bound or shed in soluble form and contains multiple carbohydrate recognition domains that enable binding and clearance of several serum glycoproteins (64) in addition to mediating endocytosis and potentially playing a role in antigen presentation (65). Increased expression of MR on isolated macrophages in response to treatment with IL-4/IL-13 also corresponds to increased endocytosis of mannosylated ligands by AAMs, an observation that formed a key initial determinant in the description of the alternatively activated state (9, 11). MR has been subsequently employed in many studies as a marker for the AAM phenotype, although expression tends to be broad among tissue macrophage populations and, like that of other AAM markers, can be modulated by signals other than IL-4/IL-13, such as IL-10, in the context of helminth infection (66). In addition to binding endogenous glycoproteins, MR also recognizes carbohydrate moieties on several antigenic glycoproteins associated with type 2 immune responses and thus may participate with other CLRs during innate sensitization to stimuli such as fungal particles that are rich in mannoproteins (67). In this respect, although MR-dependent binding of fungal spores can be observed *in vitro* (68), MR-deficient mice display normal responses to infection with *Candida albicans* (69) and *Pneumocystis carinii*, albeit with some alterations in lung pathology (70), but are more susceptible to pulmonary challenge with *Cryptococcus neoformans* (71). Notably, in MR-deficient mice, the steady-state elevation of several serum glycoproteins, particularly lysosomal hydrolases (64), could impact the *in vivo* phenotypes observed during fungal exposure independently from MR-mediated fungal recognition. Nevertheless, these findings and others highlight redundancy between MR and other fungal PRRs, such as dectin-1, that appear to act sequentially and exhibit fungal stage-specific binding corresponding to exposure of fungal wall carbohydrate structures (72--75). Similar findings have emerged from studies of MR in the context of parasite infection; for example, MR-deficient mice mount normal immune responses to *Leishmania* (76), and although *Trichuris muris* excretory/secretory products bind MR and induce IL-6 production by bone marrow-derived macrophages, MR-deficient mice expel *T. muris* parasites with normal kinetics (77).

Dectin-1

Dectin-1 (Clec7a) is an AAM-associated CLR that is expressed on macrophages, neutrophils, and DCs, among other cells, and is increasingly recognized as the principal PRR in mediating fungal recognition in mice (78, 79) and humans (80). In contrast to MR, dectin-1 exhibits specificity for β -glucans and contains a hemITAM in its cytoplasmic domain that transduces signals via Src and Syk family kinases to induce respiratory burst and cell type-specific transcription factor activity, mediating protection in several models of fungal infection (81). Thus, although dectin-1 expression is strongly induced by IL-4/IL-13 (82), subsequent engagement of dectin-1 results in a different signaling response than that mediated by IL-4/IL-13. Particularities in downstream activation, however, are observed in a context- and cell subtype-dependent manner, hinting that signaling events induced by fungal recognition may represent an important point of divergence from the classical and alternative paradigm. This is borne out in the pervasive induction of mixed CAM/AAM and Th1/Th2/Th17 responses in models of experimental fungal infection, in which protection has been associated with productive dectin-1-mediated signal activation and Th1/Th17-mediated responses, whereas increased fungal burdens and poorer outcomes have been associated with Th2 and AAM responses (74, 81, 83). Related to these findings perhaps, is the broader issue of plasticity within macrophage populations, which *in vitro* can switch expression levels of hallmark CAM or AAM markers after initial polarization, resulting in mixed marker patterns (84, 85); whether dectin-1-mediated recognition among AAMs is

pertinent to initiating such changes during the course of in vivo fungal models has not been fully explored.

Chitinases and Chi-lectins

Chitin, a biopolymer of β -1,4-*N*-acetylglucosamine and a component of many parasites and allergenic organisms such as fungi and dust mites, induces a localized innate cell infiltration characterized by eosinophils, basophils, and AAMs (25, 86). Mammals do not synthesize chitin but as a result of gene duplication events express several members of the glycosyl hydrolase family 18, including active chitinases that bind and degrade chitin as well as several homologous chi-lectins that exhibit some carbohydrate specificity but lack enzymatic activity (87, 88). Intriguingly, a close linkage observed between the human chitinase/chi-lectin gene family and major histocompatibility complex paralogon genes on chromosome 1 has led to the suggestion that recent expansion of this family is related to the acquisition of adaptive immunity (88). Both chitinases and chi-lectins, particularly AMCCase (25, 89) and Ym1 (Chi313)/Ym2 (Chi314) (34, 35, 90, 91), are induced in an IL-4/IL-13- and STAT6-dependent manner in vitro and during type 2 immune responses. Copious production of the chi-lectin Ym1 leads to formation of large eosinophilic crystals in lung tissue from SHP-1-deficient viable motheaten (*mev*) and *p47^{phox}*-deficient mice; similar crystals are also observed in SHIP-deficient mice, CD40L-deficient mice spontaneously infected with *P. carinii*, and mice expressing a lung-specific soluble TNFRII transgene (92--94). Ym2 has been associated with hyaline gastric and lung lesions in aging 129S4/SvJae and B6.129 strains (95). Yet, although both Ym2 and AMCCase are highly upregulated in a STAT6-dependent manner during helminth infection, lung-specific transgenic expression of either AMCCase or Ym2 in Balb/c mice did not result in spontaneous lung inflammation (25; T. Reese and R. Locksley, unpublished observations), indicating that these proteins do not mediate lung inflammation per se in the absence of accompanying stimuli. Another chi-lectin, breast regression protein 39 (BRP-39; Chi311; YKL-40 in humans), has been associated with several inflammatory disorders, including severe asthma (96), and mice lacking BRP-39 display diminished Th2 responses and AAM generation following allergen challenge (97). Thus, the recognition and degradation of chitin by mammalian chi-lectins and chitinases associated with AAMs during type 2 immune responses, when considered in the context of widespread environmental chitin derived from insects, parasites, and fungi, suggest a conserved role for this family in mediating mammalian host defense functions, perhaps representing convergence with similar chitin-recognition systems employed in plants (98).

Relma

Resistin-like molecule α [Relm α ; also known as Retnla, found in inflammatory zone (FIZZ1), or hypoxia-induced mitogenic factor (HIMF)] is a member of the Relm/FIZZ family of cysteine-rich secretory proteins, initially identified in bronchoalveolar lavage fluid from allergen-challenged mice (99). Relm α is highly induced during helminth- and allergen-induced type 2 immune responses and is primarily expressed by epithelial cells, eosinophils, and AAMs (35, 91, 99--101). Analysis of helminth-infected Relm α -deficient mice by two separate groups (102, 103) has revealed an important regulatory role during type 2 inflammation, as Relm α -deficient mice displayed increased inflammation associated with increased granuloma size, liver fibrosis, and exacerbated type 2 cytokine production after *S. mansoni* infection—intriguingly echoing results from myeloid-specific Arg1-deficient mice—along with increased resistance to *Nippostrongylus brasiliensis*. The exact mechanism of regulation by Relm α in the setting of helminth infection, as well as the identity of the relevant Relm α -expressing cell type responsible for mediating the effects in vivo, remains unclear. Notably, however, additional in vitro experiments indicated that Relm α -deficient AAMs exerted direct effects on CD4 T cells, possibly through modulation of Bruton's

tyrosine kinase signaling (102). Contrasting roles for Relm α , however, have been described in other systems in which recombinant Relm α directly mediated eosinophilia, epithelial thickening, and fibrotic responses (101), whereas Relm α -deficient mice exhibited normal type 2 cytokine production, mucus production, and inflammatory responses during allergy-induced lung inflammation (100).

IL-4 AND IL-13: CANONICAL TYPE 2 CYTOKINES

AAMs arise within the context of tissues responding to the actions of the type 2 cytokines IL-4 and IL-13, which are released by various cell types and mediate effects through receptor complexes that share subunits and several downstream signaling components. The specific roles of IL-4, IL-13, and their associated receptor subunits are now becoming clearer through studies employing various mouse strains that lack specific cells, cytokines, or critical signaling components. Reporter mice, too, have allowed unprecedented spatiotemporal tracking of signature cytokine- and enzyme-expressing cells *in vivo*, unveiling complexities in how the appearance of these cells is tightly correlated with induction of AAMs during type 2 immune responses and associated physiologic processes.

IL-4/IL-13 Production

IL-4 and IL-13 are closely related members of the short-chain four-helix bundle cytokine family that bind shared receptor subunits but exhibit only 25% identity at the amino acid level, accounting for significant structural differences that influence receptor binding (104--109). And, although IL-4 and IL-13 can mediate similar physiologic effects, the two cytokines appear to be differentially regulated and play distinct roles during *in vivo* type 2 immune responses, due to additional exclusive receptor subunit binding and segregation of expression among different cellular and tissue sources. In line with these differences, recent studies employing reporter mice have indicated a partitioning of *in vivo* IL-4 and IL-13 production between distinct tissue sites, a concept reinforced by mice deficient in cytokines or cytokine-producing cells or lacking specific receptor subunits. These studies have ascribed nonredundant roles to IL-4, which mediates most events involved in the generation of high-affinity IgE antibodies, and IL-13, which mediates localized tissue effects such as chemokine secretion, goblet cell hyperplasia, mucus production, and smooth muscle alterations (8, 110, 111). Important species-specific differences exist, however, as human B cells induce proliferation and Ig switching in response to IL-13, whereas mouse B cells do not, owing to differential expression of IL-13-specific receptor subunits (112). During helminth infection, primary sources of IL-4 in the affected lung tissue include basophils and conventional Th2 cells, whereas T follicular helper (Tfh) cells are the main IL-4 producers in the tissue-draining lymph nodes (110, 111). Other cells, including mast cells, eosinophils, natural killer T cells, and innate lymphoid type 2 cells (ILC2; a.k.a. nuocytes, Ih2 cells, or natural helper cells), assume a constitutively poised state expressing IL-4 mRNA but do not secrete IL-4 cytokine in this context, as assessed by protein reporter mice. In contrast, tissue Th2 cells and ILC2, but not Tfh cells, are major producers of IL-13 (111, 113--115). Whether any of these cellular sources contributes IL-4/IL-13 to induce AAM development in various *in vivo* settings remains to be determined, but Arg1-reporter mice have revealed functional relationships between closely associated AAMs and eosinophils during innate responses to chitin administration and during maintenance of metabolic homeostasis (25, 116). Accordingly, macrophage depletion or deficiency during type 2 inflammation induced by IL-33, chitin, or helminth infection results in reduced recruitment of eosinophils (25, 117, 118). This regulation may exert reciprocal effects, as genetic deficiency of eosinophils also impairs accumulation of AAMs in white adipose tissue (WAT) (116).

Suppression of aberrant IL-4 and IL-13 production appears to be under steady-state regulation at peripheral tissue sites. Rudensky and colleagues (119) have demonstrated that

this suppression is primarily imposed by induced regulatory T (iTreg) cells, which act to restrain type 2 immune activation in the steady-state, particularly at mucosal barriers. In this model, mutation of the Foxp3 enhancer CNS1 in mice results in a lack of the normal complement of iTreg cells; these mice concurrently exhibit spontaneous airway hyperresponsiveness and type 2 immune inflammation in the lung, characterized by the increased presence of acidophilic macrophages, Arg1 and Ym1 expression, and Ym1 crystal formation, all of which correlate with the increased presence of IL-4-, IL-5-, and IL-13-producing T cells in mucosal tissues (119). The exact mechanism by which iTregs repress steady-state activation of type 2 immunity remains unclear, but this process represents an important insight into how the maintenance of immune homeostasis might be subject to multiple layers of regulation. Furthermore, several factors aside from IL-4/IL-13, particularly IL-10, IL-21R, galectin-3, and TGF β , play relevant synergistic or inhibitory roles during the induction of AAM responses; exactly how these factors impinge upon IL-4/IL-13 and STAT6-mediated aspects also remains to be elucidated but may involve similar regulation at the cellular level or engagement of several interconnected complementary or inhibitory signaling pathway elements (93, 120--122).

IL-4/IL-13 Signaling

IL-4 or IL-13 signaling is initiated via two types of heterodimeric transmembrane receptor complexes: the type I receptor, which exclusively binds IL-4 and is comprised of IL-4R α and γ_c subunits (the latter of which also functions as a subunit in IL-2, IL-7, IL-9, IL-15, and IL-21 receptor complexes), and the type II receptor, which binds both IL-4 and IL-13 and is comprised of IL-4R α and IL-13R α 1 subunits. Thus, exclusive expression of type I receptor complexes on any particular cell type limits responsiveness to IL-4, as is the case for most mouse B and T cells, whereas expression of the type II receptor permits signaling in response to both IL-4 and IL-13, as occurs in many nonhematopoietic cell types. Macrophages express both type I and type II receptors and thus may initiate signaling and induction of the AAM phenotype under all three circumstances: engagement of the type I receptor by IL-4 or engagement of the type II receptor by either IL-4 or IL-13. Recent work detailing the ternary structure of each ligand-receptor binding state has revealed critical differences in affinity that influence subunit assembly as IL-4R α is paired with γ_c versus IL-13R α 1 in the presence of ligand and also, unexpectedly, that the type II receptor varies in signaling potencies and kinetics when bound by IL-4 versus IL-13. This latter observation reflects the differences in sequential assembly of the shared subunits with consequences for the strength and duration of downstream signaling events (109). The implication of these findings is that both cytokine and receptor subunit availability influence subsequent signaling; studies demonstrating that macrophages exhibit differing signaling sensitivities depending on the presence or absence of γ_c and the exposure to either IL-4 or IL-13 provide further support (123). Also noteworthy in this respect is the potential contribution in vivo of the decoy receptor IL-13R α 2, which contains additional IL-13 binding sites that confer an affinity for IL-13 that is four orders of magnitude greater than that of IL-13R α 1 (124, 125). The in vivo relevance of this inhibitory receptor is borne out in IL-13 α 2-deficient mice, which display enhanced IL-13 tissue expression and suppression of NO macrophage responses (126).

Differential signaling events occurring downstream of type I versus type II receptor engagement may also influence the induction of gene programs in AAMs. Similarly to other cytokine receptor complexes, both type I and type II receptors initiate signaling via Jak/STAT-mediated phosphorylation events but differ in particular subunit-specific adapters: namely, Jak3 associates with γ_c , whereas Jak2 or Tyk2 associates with IL-13R α 1 (127). The shared IL-4R α chain associates with Jak1 and contains multiple tyrosine phosphorylation sites that serve as binding sites for proteins containing Src homology 2

(SH2) domains (e.g., STAT6) and phosphotyrosine-binding (PTB) domains [e.g., insulin receptor substrate (IRS) proteins]; various binding events within specific regions of the IL-4R α chain correspond to different cellular transcriptional and growth functions (128). After initial receptor dimerization and tyrosine phosphorylation, STAT6 associates with IL-4R α and is phosphorylated, dissociates, homodimerizes, and then translocates into the nucleus and binds to promoter elements within IL-4/IL-13-responsive genes. In macrophages, IL-4 induces γ_c -dependent tyrosine phosphorylation of IRS-2 and association with the p85 subunit of phosphatidylinositol 3-kinase (PI3K) and growth factor receptor-bound protein 2 (Grb2); these responses correlate with heightened expression of the AAM markers Arg1, Relm α , and Ym1 compared with IL-13, even in conditions of similar STAT6 phosphorylation, indicating that type I receptor engagement by IL-4 may more sensitively induce AAM polarization (129). Consistent with these findings, other groups demonstrated that IL-13R α 1 was dispensable for induction of some AAM markers but not others during allergen challenge and helminth infection (130, 131). Hence, induction of the AAM phenotype can be subject to several types of regulation solely at the level of IL-4/IL-13: availability of IL-4 and IL-13 cytokines, expression levels of both types of receptor complexes, presence or absence of IL-13R α 2, and differential engagement of signaling components downstream of each subunit.

IL-4/IL-13-induced signaling events can be further modulated by an immunoreceptor tyrosine-based inhibitory motif (ITIM) within the cytoplasmic tail of IL-4R α , a site recognized by Src family kinases and inhibitory phosphatases that are also implicated in the regulation of the AAM phenotype. Evidence for steady-state regulation of AAM polarization emerged from mice containing mutations in SH2 domain-containing inositol-5'-phosphatase (SHIP) and SH2 domain-containing protein tyrosine phosphatase 1 (SHP-1), both negative regulators of the phosphoinositide 3-kinase (PI3K)-mediated signaling pathway in hematopoietic cells. SHIP^{-/-} mice have a short life span, abnormal hematopoiesis, and marked lung pathology (132), mirroring phenotypes observed in motheaten (me) or viable motheaten (mev) SHP-1-deficient states and reflecting the broad immunosuppressive activities associated with this family of signaling molecules (133--135). Peritoneal and alveolar macrophages from SHIP^{-/-} mice display constitutively increased levels of Arg1 and phosphatidylinositol (3,4,5)-trisphosphate (PIP₃) as a result of increased PI3K activity, accompanied by lung pathology characterized by spontaneous type 2 inflammation, fibrosis, and the conspicuous presence of Ym1 eosinophilic crystals (93, 136). Although initially interpreted as an indication that SHIP was acting in a macrophage-intrinsic manner during development to repress a separate, IL-4/IL-13-independent pathway of AAM skewing (93), more recent evidence indicates that the *in vivo* AAM phenotype could arise from aberrant IL-4 production, perhaps by hypersensitive basophils or mast cells (137).

A similar picture has emerged in the case of SHP-1, as the severity of the spontaneous type 2 lung inflammation observed in mev mice was significantly ameliorated by concomitant loss of IL-13 or STAT6 (138). As with SHIP deficiency, the AAM phenotype in SHP-1-deficient mice may arise indirectly as a result of aberrantly activated cells that produce IL-4/IL-13 upstream of macrophage activation. In line with this, SHP-1 has also been described to play a critical role in restraining hyperresponsiveness and cytokine production in a variety of cell types, such as mast cells (139). This model gains further support from mice with combined deficiency of Src family kinases Lyn and Hck, which exhibit similarly complex hematopoietic defects along with lung AAM infiltration. This phenotype could be attenuated by expression of a membrane-bound form of SHIP in hematopoietic stem cells, and further mechanistic insights also suggest a role for increased IL-4 production by basophils (140). In contrast to these studies, however, other work indicates that IL-4-induced STAT6 transcription and expression of Ym1 and Arg1 are dependent on cell-intrinsic SHIP

degradation and the PI3K pathway in macrophages during AAM polarization (141). In addition, bone marrow–derived macrophages from mice in which the IL-4R α ITIM, a region previously demonstrated to interact with SHP-1, SHP-2 and SHIP (142), was inactivated by mutagenesis of the tyrosine (Y709) residue into phenylalanine (F709) exhibited enhanced STAT6 phosphorylation and AAM marker expression in response to IL-13 and, to a lesser degree, IL-4 (143). Although the mechanistic details remain cloudy, these systems, along with observations made in the Foxp3 CNS1 mutant mouse, indicate a clear role for steady-state suppression of aberrant IL-4 and IL-13 signaling and/or production among various potential cellular sources, without which deleterious type 2-mediated pathology arises and proceeds unabated.

Other Factors

Several additional intracellular factors act in concert with the IL-4/IL-13–STAT6 axis to regulate the AAM phenotype. In particular, IL-4/IL-13- and STAT6-mediated signals induce the lipid-activated transcription factors peroxisome proliferator–activated receptor γ (PPAR γ) and δ (PPAR δ), along with PPAR γ coactivator 1 β (PGC1 β), which mediate several aspects of AAM polarization particularly important in the context of glucose and lipid metabolism, as discussed further below (144--147). In the case of PPAR γ , an interaction between STAT6 and PPAR γ on promoters of PPAR γ target genes appears to act as a transcriptional switch to promote DNA binding and increased gene expression in AAMs (148). These factors may act in parallel with other STAT6-inducible transcription factors such as c-Myc and KLF4, which appear to amplify STAT6-mediated signals in AAMs while, in the case of KLF4, simultaneously suppressing CAM-inducing NF- κ B activation signals (30, 32). This type of signal integration is further apparent upon exposure to CAM-polarizing stimuli, as PPAR γ expression in macrophages is suppressed upon LPS exposure (149); however, interestingly, macrophages induced to become LPS tolerant exhibit an AAM phenotype by a mechanism regulated by the p50 subunit of NF- κ B, an observation supported by the finding that p50-deficient mice show reduced AAM responses during allergy and helminth challenge models (150).

Epigenetic changes may also impinge upon the IL-4/IL-13-mediated AAM program. For instance, IL-4 signals induce STAT6 binding to the promoter of the chromatin-remodeling factor Jmjd3, which was shown to demethylate histone H3 at lysine 27 and thereby regulate expression of AAM marker genes such as Arg1, Ym1, and Relm α (151). Because Jmjd3 deficiency in mice results in perinatal lethality, Akira and colleagues (31) generated chimeric mice using Jmjd3-deficient fetal liver cells; these mice displayed normal responses to CAM-inducing stimuli, but the induction of AAMs was impaired in response to intraperitoneal chitin administration, M-CSF exposure, and helminth infection. Curiously, however, IL-4-induced AAM marker expression among bone marrow–derived macrophages from these mice was preserved, in contrast to the prior study (151), which demonstrated using siRNA knockdown of Jmjd3 that IL-4 induction of AAM genes acted via a Jmjd3-dependent mechanism. Further, impaired IL-4 and IL-13 production and lymph node T cell defects were observed during *N. brasiliensis* infection in Jmjd3-deficient chimeric mice, which, when coupled with the observation that Jmjd3-deficient macrophages showed impaired proliferation in response to M-CSF (31), suggest broader and more fundamental effects of Jmjd3 deficiency apart from AAM marker induction. Indeed, Jmjd3 is also induced in response to LPS and inflammatory cytokines to bind and regulate polycomb group target gene activity in macrophages (152). Nevertheless, the additional demonstration that the gene encoding the transcription factor interferon regulatory factor 4 (*Irf4*) is the Jmjd3 target gene responsible for controlling AAM marker expression (31) is of potential relevance in light of other recent studies showing that, in contrast to AAM polarization, CAM polarization can be directed by IRF5 (153) or IRF8 via the Notch1–RBP-J axis (154).

PHYSIOLOGIC FUNCTIONS OF AAMS

Although IL-4 and IL-13 production and AAM development have generally been studied in the context of type 2 immunity to helminthes or allergic airways disease, a crucial role for AAMs in metabolic homeostasis is also emerging, in concert with the increasing appreciation of a role for hematopoietic cells in this and other physiologic processes; these roles may ultimately have roots in basic mechanisms of tissue maintenance and repair (Figure 2). In vivo functional studies are limited, however, and tools to selectively deplete AAMs or AAM products are hampered by reagents that lack complete specificity for macrophages or display incomplete penetrance among macrophage populations.

Helminth Infection

Several helminths elicit highly polarized type 2 immune responses, marked by induction of IL-4 and IL-13, which direct IgE antibody and mucus overproduction, eosinophilia, and smooth muscle and fibroblast alterations along with AAM development. AAMs have been described to play important roles in modulating various pathologic features of helminth infection, including suppression of T cell responses, modulation of fibrosis, and formation of multinucleated giant cells found within parasite-induced granulomas. Brombacher and colleagues (155) described a vital function for AAMs after acute *S. mansoni* infection, in which myeloid-restricted IL-4R α deficiency resulted in early mortality, phenocopying results from mice with global IL-4R α deficiency. In contrast to global IL-4R α deficiency, however, myeloid-restricted IL-4R α -deficient mice retained robust eosinophilic granuloma formation and fibrosis, along with antigen-specific Th2 responses (155). Similar effects of myeloid cell-restricted Arg1 deficiency during *S. mansoni* infection indicate that AAM-derived Arg1 is a key mediator in this process, acting to restrain both unrestricted Th2-mediated fibrotic pathology and intestinal damage associated with increased Th1/Th17 cytokines, NOS-2 levels, and endotoxemia (44, 45). These effects appear to vary among helminths, however, as myeloid-specific IL-4R α -deficient mice exhibited normal cytokine and antibody production, parasite clearance, and mucous goblet cell numbers in response to *N. brasiliensis* infection (155), and Arg1 blockade or deficiency in hematopoietic and endothelial cell lineages had little effect on responses to acute and chronic infection with *T. muris* (156). Macrophage depletion, however, impaired eosinophil recruitment and tissue repair processes during *N. brasiliensis* infection (117, 157) and abrogated memory responses to the gastrointestinal helminth *Heligmosomoides*; further, chemical inhibition of Arg1 in this system attenuated expulsion of *Heligmosomoides* (158). Thus, roles for AAMs during parasite infection depend on the infection model, along with the particulars of AAM depletion, perhaps resulting from inadequate drivers of deletion in these mouse models or compensatory factors aside from IL-4/IL-13—particularly IL-10, which can also mediate important AAM functions. For example, IL-10-induced expression of selenoprotein P contributes to lowered pathogenicity and associated susceptibility in a model of African trypanosomiasis (121).

Asthma and Allergic Inflammation

Type 2 immune responses are also prominently associated with asthma and allergic inflammation, in which many features of helminth infection, such as elevated IgE, eosinophilia, mucus production, and smooth muscle hypercontractility, are recapitulated in a manner dependent on IL-4/IL-13 and STAT6. Animal models of allergic airways disease have yielded a consistent view of distinct yet overlapping roles for IL-4 and IL-13 in mediating most aspects of associated pathology (8). For instance, hallmarks of human asthma such as airway hyperreactivity (AHR; increased sensitivity to bronchoconstricting agents), eosinophilia, and mucus overproduction can be reproduced in rodents by transgenic overexpression or exogenous administration of IL-13, whereas these effects are inhibited by

IL-13 neutralization or IL-13R α 1 deficiency, albeit somewhat variably, in the context of allergen-induced airway inflammation (130, 131, 159--161). Although IL-4 mediates similar effects, IL-13 in this context appears to act directly on smooth muscle cells and epithelial cells expressing IL-4R α and STAT6, rather than on hematopoietic cells. For instance, transgenic expression of hSTAT6 in CC10-expressing epithelial cells restores IL-13-induced mucus overproduction and AHR in STAT6-deficient mice (162). In contrast, AHR develops normally in allergen-challenged mice lacking IL-4R α on epithelial cells expressing Clara cell secretory protein (163), indicating the involvement of other IL-13 targets such as airway smooth muscle cells. Accordingly, mice engineered to express IL-4R α exclusively on smooth muscle cells show normal induction of IL-13-mediated AHR, but this restricted expression is sufficient but not necessary to induce AHR in response to cytokine or allergen inhalation (164). These findings extend to human studies, in which treatment with an IL-13-neutralizing antibody led to improved lung function in asthmatic adult populations, particularly in patients exhibiting a Th2-high phenotype marked by elevated blood eosinophils, serum IgE, and periostin (165).

Prominent expression of IL-4/IL-13 also leads to AAM development in several allergen-driven models, much like in helminth infection, although a functional role for AAMs in mediating pathology in these settings is less clear. For example, myeloid deficiency of IL-4R α in mice reduced Ym1, Arg1, and Relm α AAM marker expression during ovalbumin (Ova)- or house dust mite extract-induced allergic inflammation, but this reduction resulted in minor alterations to other parameters, such as AHR, antibody and cytokine production, mucus hypersecretion, eosinophil levels, and collagen deposition (166). These findings were mirrored by a similarly limited impact of Relm α deficiency in the context of allergy-induced lung inflammation (100). A separate study, however, showed that AAMs contribute to eosinophil recruitment in an Ova-driven allergy model, as demonstrated by the ability in mice of adoptively transferred bone marrow-derived macrophages to acquire AAM markers and enhance lung eosinophil recruitment, in contrast to IL-4R α -deficient macrophages (167). In support of this finding, IL-33 administration, which induces AAM development via an IL-13-dependent mechanism in the lung, also mediated eosinophil recruitment in a macrophage-dependent manner (118). Thus, it seems likely that the involvement of AAMs in allergy-driven models may depend critically on the nature of the stimulus and exposure regimen, as extracellular parasite infection is often accompanied by a significant tissue injury component that may be lacking in most allergen-driven model systems.

Tissue Damage and Repair

Parasite migration through host tissues causes extensive tissue destruction that simultaneously invokes repair processes, which are characterized by deposition of collagen and other extracellular matrix proteins and by increased mucus production and smooth muscle effects directed at generating the mechanical force needed to expel large foreign entities. Engagement of these processes suggests that type 2 immunity may have evolved from a more generalized innate response to tissue injury (7). Tissue repair and regeneration processes, such as those triggered during virus-induced lung inflammation, invoke close collaboration among the stem cell compartment, innate lymphoid cells, and AAMs (168). The kinetics and magnitude of AAM induction during innate and adaptive immune responses indicate that early innate sources of IL-4/IL-13 stimulate an immediate basal AAM polarization that is reinforced and potentiated by subsequent Th2 cell-derived IL-4/IL-13. For example, recent work confirming earlier observations (169, 170) indicates that normal wound healing responses are critically dependent on macrophages present during the first several days following sterile tissue damage such as surgical trauma. These early time points are concurrent with the IL-4R α 1-dependent innate induction of AAM markers among macrophages, a response that wanes over subsequent days unless accompanied by an

adaptive Th2 response induced by antigenic stimulation (170--172). In the case of parasite infection, this later engagement by the adaptive immune system serves to amplify early innate responses and is required for normal expulsion of many parasites, such as *N. brasiliensis*. As such, worsening of hemorrhage and inflammation were seen when macrophages were depleted during the early stages of *N. brasiliensis* migration through the lung (157), corroborating results from the surgical injury model. Myeloid KLF4 deficiency, which led to impaired AAM polarization, also resulted in significantly delayed wound healing in a mouse skin injury model (30). Repair processes after injury critically involve additional factors, such as IL-10 and TGF- β 1 signaling, but, as in other type 2-mediated processes, how they interact with IL-4/IL-13 remains undefined.

The induction of multinucleated giant cells in response to granulomatous diseases or foreign bodies, such as surgical implants, that cannot be otherwise phagocytosed may involve AAMs in a manner resembling osteoclast function in bone degradation, although in vivo data are lacking. In vitro, IL-4 or IL-13 can trigger formation of multinucleated cells and induction of fusogenic molecules, depending on the macrophage source, adherence to particular substrates, and membrane and cytoskeletal rearrangements. Several molecules have been implicated in regulating the fusogenic state, including surface receptors E-cadherin and DC-STAMP (173). Intriguingly, recent evidence implicates microRNAs as exerting transcriptional control over macrophage fusion and DC-STAMP induction by IL-4, particularly through miR-7a-1, which negatively regulates fusion by directly targeting DC-STAMP mRNA, providing a mechanistic basis for feedback suppression that may be involved in the differential induction of this phenotype within various settings (174). Additionally, microRNAs may control other aspects of AAMs, as miR-155 targets IL-13R α 1 in human macrophages to modulate AAM gene expression (175).

Metabolism and Homeostasis

Initial appreciation that a significant inflammatory component present in obese WAT may contribute to the pathogenesis of insulin resistance, diabetes, and metabolic syndrome gave way to more recent findings that, upon transition from a lean to an obese state in mice, WAT macrophages switch phenotypes from AAM to CAM (145, 176). Subsequently, the maintenance of AAMs in WAT was found to be dependent on macrophage-intrinsic PPAR γ and IL-4/IL-13-STAT6 signals, likely derived from WAT-associated eosinophils; reduction of either cell type in vivo resulted in diet-induced metabolic dysregulation characterized by insulin resistance and glucose intolerance (116, 145). In addition, impaired AAM polarization due to myeloid KLF4 deficiency promoted insulin resistance (30). Other PPARs contribute to metabolic homeostasis, as similar metabolic effects and AAM marker impairment were also observed in the absence of PPAR δ (146, 147); however, metabolic dysregulation was not observed in chimeric mice reconstituted with PPAR γ -, PPAR δ -, or doubly deficient bone marrow (177), perhaps reflecting different genetic backgrounds of the mice used in these studies. Notable, too, is a recent report indicating that synthetic PPAR γ agonists may target cells other than macrophages, such as adipose tissue Tregs, which play a role in thiazolidinedione-mediated restoration of insulin sensitivity in obese mice (178). Nevertheless, IL-4 mediates dramatic changes in macrophages that are relevant to metabolism on both a cellular and a tissue level, including activation and maintenance of PPAR γ , PPAR δ , and PGC1 β . These factors antagonize the program of anaerobic glycolysis induced within CAMs by hypoxia-inducible transcription factor (HIF)-1 α and instead promote a program of aerobic metabolism characterized by β -oxidation of fatty acids. This program enables long-term alternative activation and sustainment of macrophages, which may be relevant not only to the homeostatic maintenance of metabolic parameters but also during prolonged type 2 immune responses typified by chronic helminth infection (179).

The endogenous regulators of this immunometabolic unit remain undefined, but points of integration with several other physiologic systems suggest a default state that can be perturbed by various stressors. For example, exposure to cold temperatures can induce an IL-4-mediated AAM response in adipose tissue macrophages, which directly produce norepinephrine in mediating an adaptive thermogenic program in both WAT and brown adipose tissue, indicating that type 2 immune cells operate in parallel with the nervous system to integrate physiologic stress signals (180). Also, the appearance of AAMs or macrophages exhibiting AAM markers in several tissues such as bone (181), skeletal muscle (182), mammary gland (183), and testes (184) suggests potential involvement of type 2 immune cells in the regulation of several other physiologic processes, but these possibilities also remain largely unexplored. The notably mixed CAM/AAM population and angiogenic response to hypoxic environments in the context of tumors, for example, may reflect metabolic energy flux that occurs within macrophages and that can also contribute to general tissue homeostasis. Glucose metabolism, which is controlled by several kinases acting via the pentose phosphate pathway of glycolysis, may thus be a critical branch point during induction of AAM/CAM phenotypes. A recent study implicated the carbohydrate kinase-like protein CARKL as a key rheostat in macrophage metabolism, acting to maintain oxygen consumption in AAMs, whose metabolic profile resembled nonpolarized macrophages; CAMs, in contrast, downregulated CARKL to increase glycolysis and decrease oxygen consumption rate. CARKL also downregulated SOCS3 expression in CAMs, resulting in enhanced STAT3 phosphorylation (185) and thereby representing an intriguing link between metabolic flux and modulation of cytokine signaling within macrophages.

SUMMARY AND NEW DIRECTIONS

The critical role of macrophages in host defense has been reinforced by a century of research since Metchnikoff's initial description. New appreciation for the spectrum of phenotypes that these cells can assume within a wide range of cellular milieus invoked by homeostatic and inflammatory physiologic conditions has extended their functions into previously unexplored realms. Thus, an important area for future work will be the careful characterization of how these various phenotypic states are influenced and how they function in vivo, taking into consideration recent findings detailing the development and differentiation of distinct macrophage populations. Differential exposure to IL-4 versus IL-13 and receptor type expression may contribute substantially to heterogeneous responses even among AAMs, but the in vivo relevance of this remains to be established. Also, the induction and modulation of AAM markers by stimuli or cytokines other than IL-4/IL-13, such as IL-10 and TGF- β , suggest that specific aspects of AAMs are preserved in a redundant fashion. How these states of activation relate to or modify AAMs, in the context of IL-4/IL-13 or otherwise, is also unclear but may be important in the context of tissue responses that exhibit a complement of macrophages with mixed CAM/AAM phenotypic markers, such as tumor-associated macrophages or myeloid-derived suppressor cells.

As such, the definition of the AAM phenotype as a signature of IL-4 or IL-13 activation remains useful to guide interrogation of relevant cell types or particular microenvironmental cues that thus induce AAM markers in contexts other than highly polarized type 2 immune responses such as helminth infection or allergic inflammation. The complex roles and regulation of IL-4 and IL-13 are now becoming clearer as new tools allowing in vivo functional analysis have revealed an element of innate activation, originally inferred from the tissue repair and immunosuppressive processes induced by parasites, that appears to be relevant in maintaining tissue homeostasis. New appreciation of the dramatic metabolic changes occurring upon macrophage activation that shift amino acid, oxygen, and glucose processing on a cellular level has led to a greater understanding of the connections between immune cells and broader physiologic tissue responses. In addition, as the critical producers

of IL-4/IL-13 are parsed in various experimental systems, attention has focused on the initiation of type 2 immunity upstream of canonical IL-4/IL-13 production, where newly identified innate cells and epithelial cytokines have been implicated. In particular, IL-25, IL-33, and TSLP appear to play critical roles in priming type 2 immunity and activating ILC2, a rare population of innate effector lymphoid cells found in multiple tissues that is poised to rapidly produce IL-5 and IL-13 in response to these cytokines (6). Whether these cells contribute to the development of AAMs remains to be established, but their involvement in the innate production of type 2 cytokines and the recruitment of other cells competent for cytokine production is intriguing and raises broader questions about how they fit into the initiation of polarized type 2 immunity and other physiologic processes that invoke similar innate responses. Finally, although AAMs have been described in several physiologic settings, their functions have remained elusive due to a lack of adequate in vivo tools. Novel mouse strains exhibiting genetic deficiencies of various cytokines and cytokine receptor subunits, along with deleter and reporter mice, have been greatly informative, but caveats exist, including deletion efficiency, lineage specificity, marker analysis, and particulars of the mouse model systems employed. Thus, a systems approach using careful in vivo evaluation must guide future research aimed at elucidating AAM function within the larger context of type 2 immune biology.

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LITERATURE CITED

1. Nussbaum JC, Locksley RM. Infectious (non)tolerance—frustrated commensalism gone awry? *Cold Spring Harb. Perspect. Biol.* 2012; 4:a007328. [PubMed: 22456498]
2. Chen G, Zhuchenko O, Kuspa A. Immune-like phagocyte activity in the social amoeba. *Science.* 2007; 317:678–81. [PubMed: 17673666]
3. Medzhitov R. Recognition of microorganisms and activation of the immune response. *Nature.* 2007; 449:819–26. [PubMed: 17943118]
4. Takeuchi O, Akira S. Pattern recognition receptors and inflammation. *Cell.* 140:805–20. [PubMed: 20303872]
5. Hoffmann JA. The immune response of *Drosophila*. *Nature.* 2003; 426:33–38. [PubMed: 14603309]
6. Locksley RM. Asthma and allergic inflammation. *Cell.* 2010; 140:777–83. [PubMed: 20303868]
7. Allen JE, Maizels RM. Diversity and dialogue in immunity to helminths. *Nat. Rev. Immunol.* 2011; 11:375–88. [PubMed: 21610741]
8. Finkelman FD, Hogan SP, Hershey GK, Rothenberg ME, Wills-Karp M. Importance of cytokines in murine allergic airway disease and human asthma. *J. Immunol.* 2010; 184:1663–74. [PubMed: 20130218]
9. Stein M, Keshav S, Harris N, Gordon S. Interleukin 4 potently enhances murine macrophage mannose receptor activity: a marker of alternative immunologic macrophage activation. *J. Exp. Med.* 1992; 176:287–92. [PubMed: 1613462]
10. Doherty TM, Kastelein R, Menon S, Andrade S, Coffman RL. Modulation of murine macrophage function by IL-13. *J. Immunol.* 1993; 151:7151–60. [PubMed: 7903102]
11. Doyle AG, Herbein G, Montaner LJ, Minty AJ, Caput D, et al. Interleukin-13 alters the activation state of murine macrophages in vitro: comparison with interleukin-4 and interferon- γ . *Eur. J. Immunol.* 1994; 24:1441–45. [PubMed: 7911424]
12. Mosmann TR, Cherwinski H, Bond MW, Giedlin MA, Coffman RL. Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. *J. Immunol.* 1986; 136:2348–57. [PubMed: 2419430]

13. Gordon S, Taylor PR. Monocyte and macrophage heterogeneity. *Nat. Rev. Immunol.* 2005; 5:953–64. [PubMed: 16322748]
14. Martinez FO, Helming L, Gordon S. Alternative activation of macrophages: an immunologic functional perspective. *Annu. Rev. Immunol.* 2009; 27:451–83. [PubMed: 19105661]
15. Murray PJ, Wynn TA. Protective and pathogenic functions of macrophage subsets. *Nat. Rev. Immunol.* 2011; 11:723–37. [PubMed: 21997792]
16. Lichanska AM, Hume DA. Origins and functions of phagocytes in the embryo. *Exp. Hematol.* 2000; 28:601–11. [PubMed: 10880746]
17. Orkin SH, Zon LI. Hematopoiesis: an evolving paradigm for stem cell biology. *Cell.* 2008; 132:631–44. [PubMed: 18295580]
18. Auffray C, Sieweke MH, Geissmann F. Blood monocytes: development, heterogeneity, and relationship with dendritic cells. *Annu. Rev. Immunol.* 2009; 27:669–92. [PubMed: 19132917]
19. Geissmann F, Manz MG, Jung S, Sieweke MH, Merad M, Ley K. Development of monocytes, macrophages, and dendritic cells. *Science.* 2010; 327:656–61. [PubMed: 20133564]
20. Ajami B, Bennett JL, Krieger C, Tetzlaff W, Rossi FM. Local self-renewal can sustain CNS microglia maintenance and function throughout adult life. *Nat. Neurosci.* 2007; 10:1538–43. [PubMed: 18026097]
21. Ginhoux F, Greter M, Leboeuf M, Nandi S, See P, et al. Fate mapping analysis reveals that adult microglia derive from primitive macrophages. *Science.* 2010; 330:841–45. [PubMed: 20966214]
22. Jenkins SJ, Ruckerl D, Cook PC, Jones LH, Finkelman FD, et al. Local macrophage proliferation, rather than recruitment from the blood, is a signature of TH2 inflammation. *Science.* 2011; 332:1284–88. [PubMed: 21566158]
23. Schulz C, Gomez Perdiguero E, Chorro L, Szabo-Rogers H, Cagnard N, et al. A lineage of myeloid cells independent of Myb and hematopoietic stem cells. *Science.* 2012; 336:86–90. [PubMed: 22442384]
24. Wang Y, Szretter KJ, Vermi W, Gilfillan S, Rossini C, et al. IL-34 is a tissue-restricted ligand of CSF1R required for the development of Langerhans cells and microglia. *Nat. Immunol.* 2012; 13:753–60. [PubMed: 22729249]
25. Reese TA, Liang HE, Tager AM, Luster AD, Van Rooijen N, et al. Chitin induces accumulation in tissue of innate immune cells associated with allergy. *Nature.* 2007; 447:92–96. [PubMed: 17450126]
26. Auffray C, Fogg D, Garfa M, Elain G, Join-Lambert O, et al. Monitoring of blood vessels and tissues by a population of monocytes with patrolling behavior. *Science.* 2007; 317:666–70. [PubMed: 17673663]
27. Ponomarev ED, Maresz K, Tan Y, Dittel BN. CNS-derived interleukin-4 is essential for the regulation of autoimmune inflammation and induces a state of alternative activation in microglial cells. *J. Neurosci.* 2007; 27:10714–21. [PubMed: 17913905]
28. Stenzel W, Muller U, Kohler G, Heppner FL, Blessing M, et al. IL-4/IL-13-dependent alternative activation of macrophages but not microglial cells is associated with uncontrolled cerebral cryptococcosis. *Am. J. Pathol.* 2009; 174:486–96. [PubMed: 19147811]
29. Aziz A, Soucie E, Sarrazin S, Sieweke MH. MafB/c-Maf deficiency enables self-renewal of differentiated functional macrophages. *Science.* 2009; 326:867–71. [PubMed: 19892988]
30. Liao X, Sharma N, Kapadia F, Zhou G, Lu Y, et al. Krüppel-like factor 4 regulates macrophage polarization. *J. Clin. Investig.* 2011; 121:2736–49. [PubMed: 21670502]
31. Satoh T, Takeuchi O, Vandenbon A, Yasuda K, Tanaka Y, et al. The Jmjd3-Irf4 axis regulates M2 macrophage polarization and host responses against helminth infection. *Nat. Immunol.* 2010; 11:936–44. [PubMed: 20729857]
32. Pello OM, De Pizzol M, Mirolo M, Soucek L, Zammataro L, et al. Role of c-MYC in alternative activation of human macrophages and tumor-associated macrophage biology. *Blood.* 2012; 119:411–21. [PubMed: 22067385]
33. Raes G, Van den Bergh R, De Baetselier P, Hassanzadeh Ghassabeh G. Arginase-1 and Ym1 are markers for murine, but not human, alternatively activated myeloid cells. *J. Immunol.* 2005; 174:6561–62. [PubMed: 15905489]

34. Loke P, Nair MG, Parkinson J, Guiliano D, Blaxter M, Allen JE. IL-4 dependent alternatively-activated macrophages have a distinctive in vivo gene expression phenotype. *BMC Immunol.* 2002; 3:7. [PubMed: 12098359]
35. Raes G, De Baetselier P, Noel W, Beschin A, Brombacher F, Hassanzadeh Ghassabeh G. Differential expression of FIZZ1 and Ym1 in alternatively versus classically activated macrophages. *J. Leukoc. Biol.* 2002; 71:597–602. [PubMed: 11927645]
36. Scotton CJ, Martinez FO, Smelt MJ, Sironi M, Locati M, et al. Transcriptional profiling reveals complex regulation of the monocyte IL-1 β system by IL-13. *J. Immunol.* 2005; 174:834–45. [PubMed: 15634905]
37. Martinez FO, Gordon S, Locati M, Mantovani A. Transcriptional profiling of the human monocyte-to-macrophage differentiation and polarization: new molecules and patterns of gene expression. *J. Immunol.* 2006; 177:7303–11. [PubMed: 17082649]
38. Bronte V, Zanovello P. Regulation of immune responses by l-arginine metabolism. *Nat. Rev. Immunol.* 2005; 5:641–54. [PubMed: 16056256]
39. Modolell M, Corraliza IM, Link F, Soler G, Eichmann K. Reciprocal regulation of the nitric oxide synthase/arginase balance in mouse bone marrow-derived macrophages by TH1 and TH2 cytokines. *Eur. J. Immunol.* 1995; 25:1101–4. [PubMed: 7537672]
40. Munder M, Eichmann K, Modolell M. Alternative metabolic states in murine macrophages reflected by the nitric oxide synthase/arginase balance: competitive regulation by CD4⁺ T cells correlates with Th1/Th2 phenotype. *J. Immunol.* 1998; 160:5347–54. [PubMed: 9605134]
41. Munder M, Eichmann K, Moran JM, Centeno F, Soler G, Modolell M. Th1/Th2-regulated expression of arginase isoforms in murine macrophages and dendritic cells. *J. Immunol.* 1999; 163:3771–77. [PubMed: 10490974]
42. Rutschman R, Lang R, Hesse M, Ihle JN, Wynn TA, Murray PJ. Cutting edge: Stat6-dependent substrate depletion regulates nitric oxide production. *J. Immunol.* 2001; 166:2173–77. [PubMed: 11160269]
43. Hesse M, Modolell M, La Flamme AC, Schito M, Fuentes JM, et al. Differential regulation of nitric oxide synthase-2 and arginase-1 by type 1/type 2 cytokines in vivo: Granulomatous pathology is shaped by the pattern of l-arginine metabolism. *J. Immunol.* 2001; 167:6533–44. [PubMed: 11714822]
44. Pesce JT, Ramalingam TR, Mentink-Kane MM, Wilson MS, El Kasmi KC, et al. Arginase-1-expressing macrophages suppress Th2 cytokine-driven inflammation and fibrosis. *PLoS Pathog.* 2009; 5:e1000371. [PubMed: 19360123]
45. Herbert DR, Orekov T, Roloson A, Ilies M, Perkins C, et al. Arginase I suppresses IL-12/IL-23p40-driven intestinal inflammation during acute schistosomiasis. *J. Immunol.* 2010; 184:6438–46. [PubMed: 20483789]
46. Modolell M, Choi BS, Ryan RO, Hancock M, Titus RG, et al. Local suppression of T cell responses by arginase-induced l-arginine depletion in nonhealing leishmaniasis. *PLoS Negl. Trop. Dis.* 2009; 3:e480. [PubMed: 19597544]
47. Allen JE, Lawrence RA, Maizels RM. APC from mice harbouring the filarial nematode, *Brugia malayi*, prevent cellular proliferation but not cytokine production. *Int. Immunol.* 1996; 8:143–51. [PubMed: 8671598]
48. Rodriguez PC, Zea AH, DeSalvo J, Culotta KS, Zabaleta J, et al. l-arginine consumption by macrophages modulates the expression of CD3 ζ chain in T lymphocytes. *J. Immunol.* 2003; 171:1232–39. [PubMed: 12874210]
49. Thompson RW, Pesce JT, Ramalingam T, Wilson MS, White S, et al. Cationic amino acid transporter-2 regulates immunity by modulating arginase activity. *PLoS Pathog.* 2008; 4:e1000023. [PubMed: 18369473]
50. Rothenberg ME, Doepker MP, Lewkowich IP, Chiamonte MG, Stringer KF, et al. Cationic amino acid transporter 2 regulates inflammatory homeostasis in the lung. *Proc. Natl. Acad. Sci. USA.* 2006; 103:14895–900. [PubMed: 17003120]
51. Van den Bossche J, Lamers WH, Koehler ES, Geuns JM, Alhonen L, et al. Pivotal advance: Arginase-1-independent polyamine production stimulates the expression of IL-4-induced

- alternatively activated macrophage markers while inhibiting LPS-induced expression of inflammatory genes. *J. Leukoc. Biol.* 2012; 91:685–99. [PubMed: 22416259]
52. Loke P, MacDonald AS, Robb A, Maizels RM, Allen JE. Alternatively activated macrophages induced by nematode infection inhibit proliferation via cell-to-cell contact. *Eur. J. Immunol.* 2000; 30:2669–78. [PubMed: 11009101]
 53. Terrazas LI, Montero D, Terrazas CA, Reyes JL, Rodriguez-Sosa M. Role of the programmed Death-1 pathway in the suppressive activity of alternatively activated macrophages in experimental cysticercosis. *Int. J. Parasitol.* 2005; 35:1349–58. [PubMed: 16126211]
 54. Huber S, Hoffmann R, Muskens F, Voehringer D. Alternatively activated macrophages inhibit T-cell proliferation by Stat6-dependent expression of PD-L2. *Blood.* 2010; 116:3311–20. [PubMed: 20625006]
 55. El Kasmi KC, Qualls JE, Pesce JT, Smith AM, Thompson RW, et al. Toll-like receptor-induced arginase 1 in macrophages thwarts effective immunity against intracellular pathogens. *Nat. Immunol.* 2008; 9:1399–406. [PubMed: 18978793]
 56. Qualls JE, Neale G, Smith AM, Koo MS, DeFreitas AA, et al. Arginine usage in mycobacteria-infected macrophages depends on autocrine-paracrine cytokine signaling. *Sci. Signal.* 2010; 3:ra62. [PubMed: 20716764]
 57. Saeij JP, Collier S, Boyle JP, Jerome ME, White MW, Boothroyd JC. *Toxoplasma* co-opts host gene expression by injection of a polymorphic kinase homologue. *Nature.* 2007; 445:324–27. [PubMed: 17183270]
 58. Jensen KD, Wang Y, Wojno ED, Shastri AJ, Hu K, et al. *Toxoplasma* polymorphic effectors determine macrophage polarization and intestinal inflammation. *Cell Host Microbe.* 2011; 9:472–83. [PubMed: 21669396]
 59. Butcher BA, Fox BA, Rommereim LM, Kim SG, Maurer KJ, et al. *Toxoplasma gondii* rhoptry kinase ROP16 activates STAT3 and STAT6 resulting in cytokine inhibition and arginase-1-dependent growth control. *PLoS Pathog.* 2011; 7:e1002236. [PubMed: 21931552]
 60. Shirey KA, Cole LE, Keegan AD, Vogel SN. *Francisella tularensis* live vaccine strain induces macrophage alternative activation as a survival mechanism. *J. Immunol.* 2008; 181:4159–67. [PubMed: 18768873]
 61. Gobert AP, McGee DJ, Akhtar M, Mendz GL, Newton JC, et al. *Helicobacter pylori* arginase inhibits nitric oxide production by eukaryotic cells: a strategy for bacterial survival. *Proc. Natl. Acad. Sci. USA.* 2001; 98:13844–49. [PubMed: 11717441]
 62. Gaur U, Roberts SC, Dalvi RP, Corraliza I, Ullman B, Wilson ME. An effect of parasite-encoded arginase on the outcome of murine cutaneous leishmaniasis. *J. Immunol.* 2007; 179:8446–53. [PubMed: 18056391]
 63. Whyte CS, Bishop ET, Ruckerl D, Gaspar-Pereira S, Barker RN, et al. Suppressor of cytokine signaling (SOCS)1 is a key determinant of differential macrophage activation and function. *J. Leukoc. Biol.* 2011; 90:845–54. [PubMed: 21628332]
 64. Lee SJ, Evers S, Roeder D, Parlow AF, Risteli J, et al. Mannose receptor-mediated regulation of serum glycoprotein homeostasis. *Science.* 2002; 295:1898–901. [PubMed: 11884756]
 65. Gazi U, Martinez-Pomares L. Influence of the mannose receptor in host immune responses. *Immunobiology.* 2009; 214:554–61. [PubMed: 19162368]
 66. Dewals BG, Marillier RG, Hoving JC, Leeto M, Schwegmann A, Brombacher F. IL-4R α -independent expression of mannose receptor and Ym1 by macrophages depends on their IL-10 responsiveness. *PLoS Negl. Trop. Dis.* 2010; 4:e689. [PubMed: 20502521]
 67. Wills-Karp M. Allergen-specific pattern recognition receptor pathways. *Curr. Opin. Immunol.* 2010; 22:777–82. [PubMed: 21093238]
 68. Ezekowitz RA, Williams DJ, Koziel H, Armstrong MY, Warner A, et al. Uptake of *Pneumocystis carinii* mediated by the macrophage mannose receptor. *Nature.* 1991; 351:155–58. [PubMed: 1903183]
 69. Lee SJ, Zheng NY, Clavijo M, Nussenzweig MC. Normal host defense during systemic candidiasis in mannose receptor-deficient mice. *Infect. Immun.* 2003; 71:437–45. [PubMed: 12496194]

70. Swain SD, Lee SJ, Nussenzweig MC, Harmsen AG. Absence of the macrophage mannose receptor in mice does not increase susceptibility to *Pneumocystis carinii* infection in vivo. *Infect. Immun.* 2003; 71:6213–21. [PubMed: 14573639]
71. Dan JM, Kelly RM, Lee CK, Levitz SM. Role of the mannose receptor in a murine model of *Cryptococcus neoformans* infection. *Infect. Immun.* 2008; 76:2362–67. [PubMed: 18391001]
72. Hohl TM, Van Epps HL, Rivera A, Morgan LA, Chen PL, et al. *Aspergillus fumigatus* triggers inflammatory responses by stage-specific β -glucan display. *PLoS Pathog.* 2005; 1:e30. [PubMed: 16304610]
73. Steele C, Rapaka RR, Metz A, Pop SM, Williams DL, et al. The β -glucan receptor dectin-1 recognizes specific morphologies of *Aspergillus fumigatus*. *PLoS Pathog.* 2005; 1:e42. [PubMed: 16344862]
74. Gersuk GM, Underhill DM, Zhu L, Marr KA. Dectin-1 and TLRs permit macrophages to distinguish between different *Aspergillus fumigatus* cellular states. *J. Immunol.* 2006; 176:3717–24. [PubMed: 16517740]
75. Heinsbroek SE, Taylor PR, Martinez FO, Martinez-Pomares L, Brown GD, Gordon S. Stage-specific sampling by pattern recognition receptors during *Candida albicans* phagocytosis. *PLoS Pathog.* 2008; 4:e1000218. [PubMed: 19043561]
76. Akilov OE, Kasuboski RE, Carter CR, McDowell MA. The role of mannose receptor during experimental leishmaniasis. *J. Leukoc. Biol.* 2007; 81:1188–96. [PubMed: 17261547]
77. deSchoolmeester ML, Martinez-Pomares L, Gordon S, Else KJ. The mannose receptor binds *Trichuris muris* excretory/secretory proteins but is not essential for protective immunity. *Immunology.* 2009; 126:246–55. [PubMed: 18624733]
78. Taylor PR, Tsoni SV, Willment JA, Dennehy KM, Rosas M, et al. Dectin-1 is required for β -glucan recognition and control of fungal infection. *Nat. Immunol.* 2007; 8:31–38. [PubMed: 17159984]
79. Iliiev ID, Funari VA, Taylor KD, Nguyen Q, Reyes CN, et al. Interactions between commensal fungi and the C-type lectin receptor Dectin-1 influence colitis. *Science.* 2012; 336:1314–17. [PubMed: 22674328]
80. Ferwerda B, Ferwerda G, Plantinga TS, Willment JA, van Sriel AB, et al. Human dectin-1 deficiency and mucocutaneous fungal infections. *N. Engl. J. Med.* 2009; 361:1760–67. [PubMed: 19864674]
81. Goodridge HS, Wolf AJ, Underhill DM. β -glucan recognition by the innate immune system. *Immunol. Rev.* 2009; 230:38–50. [PubMed: 19594628]
82. Willment JA, Lin HH, Reid DM, Taylor PR, Williams DL, et al. Dectin-1 expression and function are enhanced on alternatively activated and GM-CSF-treated macrophages and are negatively regulated by IL-10, dexamethasone, and lipopolysaccharide. *J. Immunol.* 2003; 171:4569–73. [PubMed: 14568930]
83. Romani L. Immunity to fungal infections. *Nat. Rev. Immunol.* 2011; 11:275–88. [PubMed: 21394104]
84. Stout RD, Jiang C, Matta B, Tietzel I, Watkins SK, Suttles J. Macrophages sequentially change their functional phenotype in response to changes in microenvironmental influences. *J. Immunol.* 2005; 175:342–49. [PubMed: 15972667]
85. Mylonas KJ, Nair MG, Prieto-Lafuente L, Paape D, Allen JE. Alternatively activated macrophages elicited by helminth infection can be reprogrammed to enable microbial killing. *J. Immunol.* 2009; 182:3084–94. [PubMed: 19234205]
86. Van Dyken SJ, Garcia D, Porter P, Huang X, Quinlan PJ, et al. Fungal chitin from asthma-associated home environments induces eosinophilic lung infiltration. *J. Immunol.* 2011; 187:2261–67. [PubMed: 21824866]
87. Bussink AP, Speijer D, Aerts JM, Boot RG. Evolution of mammalian chitinase(-like) members of family 18 glycosyl hydrolases. *Genetics.* 2007; 177:959–70. [PubMed: 17720922]
88. Funkhouser JD, Aronson NN Jr. Chitinase family GH18: evolutionary insights from the genomic history of a diverse protein family. *BMC Evol. Biol.* 2007; 7:96. [PubMed: 17594485]

89. Zhu Z, Zheng T, Homer RJ, Kim YK, Chen NY, et al. Acidic mammalian chitinase in asthmatic Th2 inflammation and IL-13 pathway activation. *Science*. 2004; 304:1678–82. [PubMed: 15192232]
90. Welch JS, Escoubet-Lozach L, Sykes DB, Liddiard K, Greaves DR, Glass CK. TH2 cytokines and allergic challenge induce Ym1 expression in macrophages by a STAT6-dependent mechanism. *J. Biol. Chem.* 2002; 277:42821–29. [PubMed: 12215441]
91. Nair MG, Cochrane DW, Allen JE. Macrophages in chronic type 2 inflammation have a novel phenotype characterized by the abundant expression of Ym1 and Fizz1 that can be partly replicated in vitro. *Immunol. Lett.* 2003; 85:173–80. [PubMed: 12527225]
92. Guo L, Johnson RS, Schuh JC. Biochemical characterization of endogenously formed eosinophilic crystals in the lungs of mice. *J. Biol. Chem.* 2000; 275:8032–37. [PubMed: 10713123]
93. Rauh MJ, Ho V, Pereira C, Sham A, Sly LM, et al. SHIP represses the generation of alternatively activated macrophages. *Immunity*. 2005; 23:361–74. [PubMed: 16226502]
94. Harbord M, Novelli M, Canas B, Power D, Davis C, et al. Ym1 is a neutrophil granule protein that crystallizes in p47^{Phox}-deficient mice. *J. Biol. Chem.* 2002; 277:5468–75. [PubMed: 11733538]
95. Ward JM, Yoon M, Anver MR, Haines DC, Kudo G, et al. Hyalinosis and Ym1/Ym2 gene expression in the stomach and respiratory tract of 129S4/SvJae and wild-type and CYP1A2-null B6,129 mice. *Am. J. Pathol.* 2001; 158:323–32. [PubMed: 11141507]
96. Chupp GL, Lee CG, Jarjour N, Shim YM, Holm CT, et al. A chitinase-like protein in the lung and circulation of patients with severe asthma. *N. Engl. J. Med.* 2007; 357:2016–27. [PubMed: 18003958]
97. Lee CG, Hartl D, Lee GR, Koller B, Matsuura H, et al. Role of breast regression protein 39 (BRP-39)/chitinase 3-like-1 in Th2 and IL-13-induced tissue responses and apoptosis. *J. Exp. Med.* 2009; 206:1149–66. [PubMed: 19414556]
98. Gust AA, Willmann R, Desaki Y, Grabherr HM, Nurnberger T. Plant LysM proteins: modules mediating symbiosis and immunity. *Trends Plant Sci.* 2012; 17:495–502. [PubMed: 22578284]
99. Holcomb IN, Kabakoff RC, Chan B, Baker TW, Gurney A, et al. FIZZ1, a novel cysteine-rich secreted protein associated with pulmonary inflammation, defines a new gene family. *EMBO J.* 2000; 19:4046–55. [PubMed: 10921885]
100. Munitz A, Cole ET, Karo-Atar D, Finkelman FD, Rothenberg ME. Resistin-like molecule- α regulates IL-13-induced chemokine production but not allergen-induced airway responses. *Am. J. Respir. Cell Mol. Biol.* 2012; 46:703–13. [PubMed: 22246861]
101. Doherty TA, Khorram N, Sugimoto K, Sheppard D, Rosenthal P, et al. *Alternaria* induces STAT6-dependent acute airway eosinophilia and epithelial FIZZ1 expression that promotes airway fibrosis and epithelial thickness. *J. Immunol.* 2012; 188:2622–29. [PubMed: 22327070]
102. Nair MG, Du Y, Perrigoue JG, Zaph C, Taylor JJ, et al. Alternatively activated macrophage-derived RELM- α is a negative regulator of type 2 inflammation in the lung. *J. Exp. Med.* 2009; 206:937–52. [PubMed: 19349464]
103. Pesce JT, Ramalingam TR, Wilson MS, Mentink-Kane MM, Thompson RW, et al. Relna (Relm α /Fizz1) suppresses helminth-induced Th2-type immunity. *PLoS Pathog.* 2009; 5:e1000393. [PubMed: 19381262]
104. Rozwarski DA, Gronenborn AM, Clore GM, Bazan JF, Bohm A, et al. Structural comparisons among the short-chain helical cytokines. *Structure*. 1994; 2:159–73. [PubMed: 8069631]
105. Walter MR, Cook WJ, Zhao BG, Cameron RP Jr, Ealick SE, et al. Crystal structure of recombinant human interleukin-4. *J. Biol. Chem.* 1992; 267:20371–76. [PubMed: 1400355]
106. Wlodawer A, Pavlovsky A, Gustchina A. Crystal structure of human recombinant interleukin-4 at 2.25 Å resolution. *FEBS Lett.* 1992; 309:59–64. [PubMed: 1511746]
107. Moy FJ, Diblasio E, Wilhelm J, Powers R. Solution structure of human IL-13 and implication for receptor binding. *J. Mol. Biol.* 2001; 310:219–30. [PubMed: 11419948]
108. Eisenmesser EZ, Horita DA, Altieri AS, Byrd RA. Solution structure of interleukin-13 and insights into receptor engagement. *J. Mol. Biol.* 2001; 310:231–41. [PubMed: 11419949]
109. LaPorte SL, Juo ZS, Vaclavikova J, Colf LA, Qi X, et al. Molecular and structural basis of cytokine receptor pleiotropy in the interleukin-4/13 system. *Cell*. 2008; 132:259–72. [PubMed: 18243101]

110. Sullivan BM, Liang HE, Bando JK, Wu D, Cheng LE, et al. Genetic analysis of basophil function in vivo. *Nat. Immunol.* 2011; 12:527–35. [PubMed: 21552267]
111. Liang H-E, Reinhardt RL, Bando JK, Sullivan BM, Ho IC, Locksley RM. Divergent expression patterns of IL-4 and IL-13 define unique functions in allergic immunity. *Nat. Immunol.* 2011; 13:58–66. [PubMed: 22138715]
112. Zurawski G, de Vries JE. Interleukin 13, an interleukin 4-like cytokine that acts on monocytes and B cells, but not on T cells. *Immunol. Today.* 1994; 15:19–26. [PubMed: 7907877]
113. Moro K, Yamada T, Tanabe M, Takeuchi T, Ikawa T, et al. Innate production of T_H2 cytokines by adipose tissue-associated c-Kit⁺Sca-1⁺ lymphoid cells. *Nature.* 2010; 463:540–44. [PubMed: 20023630]
114. Neill DR, Wong SH, Bellosi A, Flynn RJ, Daly M, et al. Nuocytes represent a new innate effector leukocyte that mediates type-2 immunity. *Nature.* 2010; 464:1367–70. [PubMed: 20200518]
115. Price AE, Liang HE, Sullivan BM, Reinhardt RL, Eisley CJ, et al. Systemically dispersed innate IL-13-expressing cells in type 2 immunity. *Proc. Natl. Acad. Sci. USA.* 2010; 107:11489–94. [PubMed: 20534524]
116. Wu D, Molofsky AB, Liang HE, Ricardo-Gonzalez RR, Jouihan HA, et al. Eosinophils sustain adipose alternatively activated macrophages associated with glucose homeostasis. *Science.* 2011; 332:243–47. [PubMed: 21436399]
117. Voehringer D, van Rooijen N, Locksley RM. Eosinophils develop in distinct stages and are recruited to peripheral sites by alternatively activated macrophages. *J. Leukoc. Biol.* 2007; 81:1434–44. [PubMed: 17339609]
118. Kurowska-Stolarska M, Stolarski B, Kewin P, Murphy G, Corrigan CJ, et al. IL-33 amplifies the polarization of alternatively activated macrophages that contribute to airway inflammation. *J. Immunol.* 2009; 183:6469–77. [PubMed: 19841166]
119. Josefowicz SZ, Niec RE, Kim HY, Treuting P, Chinen T, et al. Extrathymically generated regulatory T cells control mucosal TH2 inflammation. *Nature.* 2012; 482:395–99. [PubMed: 22318520]
120. Pesce J, Kaviratne M, Ramalingam TR, Thompson RW, Urban JF Jr, et al. The IL-21 receptor augments Th2 effector function and alternative macrophage activation. *J. Clin. Investig.* 2006; 116:2044–55. [PubMed: 16778988]
121. Bosschaerts T, Guillemins M, Noel W, Herin M, Burk RF, et al. Alternatively activated myeloid cells limit pathogenicity associated with African trypanosomiasis through the IL-10 inducible gene selenoprotein P. *J. Immunol.* 2008; 180:6168–75. [PubMed: 18424738]
122. MacKinnon AC, Farnworth SL, Hodgkinson PS, Henderson NC, Atkinson KM, et al. Regulation of alternative macrophage activation by galectin-3. *J. Immunol.* 2008; 180:2650–58. [PubMed: 18250477]
123. Junttila IS, Mizukami K, Dickensheets H, Meier-Schellersheim M, Yamane H, et al. Tuning sensitivity to IL-4 and IL-13: differential expression of IL-4R α , IL-13R α 1, and γ c regulates relative cytokine sensitivity. *J. Exp. Med.* 2008; 205:2595–608. [PubMed: 18852293]
124. Lupardus PJ, Birnbaum ME, Garcia KC. Molecular basis for shared cytokine recognition revealed in the structure of an unusually high affinity complex between IL-13 and IL-13R α 2. *Structure.* 2010; 18:332–42. [PubMed: 20223216]
125. Madala SK, Dolan MA, Sharma D, Ramalingam TR, Wilson MS, et al. Mapping mouse IL-13 binding regions using structure modeling, molecular docking, and high-density peptide microarray analysis. *Proteins.* 2011; 79:282–93. [PubMed: 21064130]
126. Wood N, Whitters MJ, Jacobson BA, Witek J, Sypek JP, et al. Enhanced interleukin (IL)-13 responses in mice lacking IL-13 receptor α 2. *J. Exp. Med.* 2003; 197:703–9. [PubMed: 12642602]
127. Kelly-Welch AE, Hanson EM, Boothby MR, Keegan AD. Interleukin-4 and interleukin-13 signaling connections maps. *Science.* 2003; 300:1527–28. [PubMed: 12791978]
128. Nelms K, Keegan AD, Zamorano J, Ryan JJ, Paul WE. The IL-4 receptor: signaling mechanisms and biologic functions. *Annu. Rev. Immunol.* 1999; 17:701–38. [PubMed: 10358772]

129. Heller NM, Qi X, Junttila IS, Shirey KA, Vogel SN, et al. Type I IL-4Rs selectively activate IRS-2 to induce target gene expression in macrophages. *Sci. Signal.* 2008; 1:ra17. [PubMed: 19109239]
130. Ramalingam TR, Pesce JT, Sheikh F, Cheever AW, Mentink-Kane MM, et al. Unique functions of the type II interleukin 4 receptor identified in mice lacking the interleukin 13 receptor α 1 chain. *Nat. Immunol.* 2008; 9:25–33. [PubMed: 18066066]
131. Munitz A, Brandt EB, Mingler M, Finkelman FD, Rothenberg ME. Distinct roles for IL-13 and IL-4 via IL-13 receptor α 1 and the type II IL-4 receptor in asthma pathogenesis. *Proc. Natl. Acad. Sci. USA.* 2008; 105:7240–45. [PubMed: 18480254]
132. Helgason CD, Damen JE, Rosten P, Grewal R, Sorensen P, et al. Targeted disruption of SHIP leads to hemopoietic perturbations, lung pathology, and a shortened life span. *Genes Dev.* 1998; 12:1610–20. [PubMed: 9620849]
133. Ward JM. Pulmonary pathology of the motheaten mouse. *Vet. Pathol.* 1978; 15:170–78. [PubMed: 664185]
134. Shultz LD, Coman DR, Bailey CL, Beamer WG, Sidman CL. “Viable motheaten,” a new allele at the motheaten locus. I. Pathology. *Am. J. Pathol.* 1984; 116:179–92. [PubMed: 6380298]
135. Tsui FW, Martin A, Wang J, Tsui HW. Investigations into the regulation and function of the SH2 domain-containing protein-tyrosine phosphatase, SHP-1. *Immunol. Res.* 2006; 35:127–36. [PubMed: 17003515]
136. Oh SY, Zheng T, Bailey ML, Barber DL, Schroeder JT, et al. Src homology 2 domain-containing inositol 5-phosphatase 1 deficiency leads to a spontaneous allergic inflammation in the murine lung. *J. Allergy Clin. Immunol.* 2007; 119:123–31. [PubMed: 17208593]
137. Kuroda E, Antignano F, Ho VW, Hughes MR, Ruschmann J, et al. SHIP represses Th2 skewing by inhibiting IL-4 production from basophils. *J. Immunol.* 2011; 186:323–32. [PubMed: 21131429]
138. Oh SY, Zheng T, Kim YK, Cohn L, Homer RJ, et al. A critical role of SHP-1 in regulation of type 2 inflammation in the lung. *Am. J. Respir. Cell Mol. Biol.* 2009; 40:568–74. [PubMed: 18952567]
139. Zhang L, Oh SY, Wu X, Oh MH, Wu F, et al. SHP-1 deficient mast cells are hyperresponsive to stimulation and critical in initiating allergic inflammation in the lung. *J. Immunol.* 2010; 184:1180–90. [PubMed: 20042576]
140. Xiao W, Hong H, Kawakami Y, Lowell CA, Kawakami T. Regulation of myeloproliferation and M2 macrophage programming in mice by Lyn/Hck, SHIP, and Stat5. *J. Clin. Investig.* 2008; 118:924–34. [PubMed: 18246197]
141. Weisser SB, McLarren KW, Voglmaier N, van Netten-Thomas CJ, Antov A, et al. Alternative activation of macrophages by IL-4 requires SHIP degradation. *Eur. J. Immunol.* 2011; 41:1742–53. [PubMed: 21469115]
142. Kashiwada M, Giallourakis CC, Pan PY, Rothman PB. Immunoreceptor tyrosine-based inhibitory motif of the IL-4 receptor associates with SH2-containing phosphatases and regulates IL-4-induced proliferation. *J. Immunol.* 2001; 167:6382–87. [PubMed: 11714803]
143. Tachdjian R, Al Khatib S, Schwinglshackl A, Kim HS, Chen A, et al. In vivo regulation of the allergic response by the IL-4 receptor α chain immunoreceptor tyrosine-based inhibitory motif. *J. Allergy Clin. Immunol.* 2010; 125:1128–36e8. [PubMed: 20392476]
144. Vats D, Mukundan L, Odegaard JI, Zhang L, Smith KL, et al. Oxidative metabolism and PGC-1 β attenuate macrophage-mediated inflammation. *Cell Metab.* 2006; 4:13–24. [PubMed: 16814729]
145. Odegaard JI, Ricardo-Gonzalez RR, Goforth MH, Morel CR, Subramanian V, et al. Macrophage-specific PPAR γ controls alternative activation and improves insulin resistance. *Nature.* 2007; 447:1116–20. [PubMed: 17515919]
146. Odegaard JI, Ricardo-Gonzalez RR, Red Eagle A, Vats D, Morel CR, et al. Alternative M2 activation of Kupffer cells by PPAR δ ameliorates obesity-induced insulin resistance. *Cell Metab.* 2008; 7:496–507. [PubMed: 18522831]
147. Kang K, Reilly SM, Karabacak V, Gangl MR, Fitzgerald K, et al. Adipocyte-derived Th2 cytokines and myeloid PPAR δ regulate macrophage polarization and insulin sensitivity. *Cell Metab.* 2008; 7:485–95. [PubMed: 18522830]

148. Szanto A, Balint BL, Nagy ZS, Barta E, Dezso B, et al. STAT6 transcription factor is a facilitator of the nuclear receptor PPAR γ -regulated gene expression in macrophages and dendritic cells. *Immunity*. 2010; 33:699–712. [PubMed: 21093321]
149. Barish GD, Downes M, Alaynick WA, Yu RT, Ocampo CB, et al. A nuclear receptor atlas: macrophage activation. *Mol. Endocrinol.* 2005; 19:2466–77. [PubMed: 16051664]
150. Porta C, Rimoldi M, Raes G, Brys L, Ghezzi P, et al. Tolerance and M2 (alternative) macrophage polarization are related processes orchestrated by p50 nuclear factor κ B. *Proc. Natl. Acad. Sci. USA*. 2009; 106:14978–83. [PubMed: 19706447]
151. Ishii M, Wen H, Corsa CA, Liu T, Coelho AL, et al. Epigenetic regulation of the alternatively activated macrophage phenotype. *Blood*. 2009; 114:3244–54. [PubMed: 19567879]
152. De Santa F, Totaro MG, Prosperini E, Notarbartolo S, Testa G, Natoli G. The histone H3 lysine-27 demethylase Jmjd3 links inflammation to inhibition of polycomb-mediated gene silencing. *Cell*. 2007; 130:1083–94. [PubMed: 17825402]
153. Krausgruber T, Blazek K, Smallie T, Alzabin S, Lockstone H, et al. IRF5 promotes inflammatory macrophage polarization and TH1-TH17 responses. *Nat. Immunol.* 2011; 12:231–38. [PubMed: 21240265]
154. Xu H, Zhu J, Smith S, Foldi J, Zhao B, et al. Notch-RBP-J signaling regulates the transcription factor IRF8 to promote inflammatory macrophage polarization. *Nat. Immunol.* 2012; 13:642–50. [PubMed: 22610140]
155. Herbert DR, Holscher C, Mohrs M, Arendse B, Schwegmann A, et al. Alternative macrophage activation is essential for survival during schistosomiasis and downmodulates T helper 1 responses and immunopathology. *Immunity*. 2004; 20:623–35. [PubMed: 15142530]
156. Bowcutt R, Bell LV, Little M, Wilson J, Booth C, et al. Arginase-1-expressing macrophages are dispensable for resistance to infection with the gastrointestinal helminth *Trichuris muris*. *Parasite Immunol.* 2011; 33:411–20. [PubMed: 21585399]
157. Chen F, Liu Z, Wu W, Rozo C, Bowdridge S, et al. An essential role for TH2-type responses in limiting acute tissue damage during experimental helminth infection. *Nat. Med.* 2012; 18:260–66. [PubMed: 22245779]
158. Anthony RM, Urban JF Jr, Alem F, Hamed HA, Rozo CT, et al. Memory T_H2 cells induce alternatively activated macrophages to mediate protection against nematode parasites. *Nat. Med.* 2006; 12:955–60. [PubMed: 16892038]
159. Grunig G, Warnock M, Wakil AE, Venkayya R, Brombacher F, et al. Requirement for IL-13 independently of IL-4 in experimental asthma. *Science*. 1998; 282:2261–63. [PubMed: 9856950]
160. Wills-Karp M, Luyimbazi J, Xu X, Schofield B, Neben TY, et al. Interleukin-13: central mediator of allergic asthma. *Science*. 1998; 282:2258–61. [PubMed: 9856949]
161. Zhu Z, Homer RJ, Wang Z, Chen Q, Geba GP, et al. Pulmonary expression of interleukin-13 causes inflammation, mucus hypersecretion, subepithelial fibrosis, physiologic abnormalities, and eotaxin production. *J. Clin. Investig.* 1999; 103:779–88. [PubMed: 10079098]
162. Kuperman DA, Huang X, Koth LL, Chang GH, Dolganov GM, et al. Direct effects of interleukin-13 on epithelial cells cause airway hyperreactivity and mucus overproduction in asthma. *Nat. Med.* 2002; 8:885–89. [PubMed: 12091879]
163. Kuperman DA, Huang X, Nguyenvu L, Holscher C, Brombacher F, Erle DJ. IL-4 receptor signaling in Clara cells is required for allergen-induced mucus production. *J. Immunol.* 2005; 175:3746–52. [PubMed: 16148120]
164. Perkins C, Yanase N, Smulian G, Gildea L, Orekov T, et al. Selective stimulation of IL-4 receptor on smooth muscle induces airway hyperresponsiveness in mice. *J. Exp. Med.* 2011; 208:853–67. [PubMed: 21464224]
165. Corren J, Lemanske RF, Hanania NA, Korenblat PE, Parsey MV, et al. Lebrikizumab treatment in adults with asthma. *N. Engl. J. Med.* 2011; 365:1088–98. [PubMed: 21812663]
166. Nieuwenhuizen NE, Kirstein F, Jayakumar J, Emedi B, Hurdal R, et al. Allergic airway disease is unaffected by the absence of IL-4R α -dependent alternatively activated macrophages. *J. Allergy Clin. Immunol.* 2012; 130:743–50. [PubMed: 22552110]

167. Ford AQ, Dasgupta P, Mikhailenko I, Smith EM, Noben-Trauth N, Keegan AD. Adoptive transfer of IL-4R α ⁺ macrophages is sufficient to enhance eosinophilic inflammation in a mouse model of allergic lung inflammation. *BMC Immunol.* 2012; 13:6. [PubMed: 22292924]
168. Gorski SA, Hufford MM, Braciale TJ. Recent insights into pulmonary repair following virus-induced inflammation of the respiratory tract. *Curr. Opin. Virol.* 2012; 2:233–41. [PubMed: 22608464]
169. Leibovich SJ, Ross R. The role of the macrophage in wound repair. A study with hydrocortisone and antimacrophage serum. *Am. J. Pathol.* 1975; 78:71–100. [PubMed: 1109560]
170. Lucas T, Waisman A, Ranjan R, Roes J, Krieg T, et al. Differential roles of macrophages in diverse phases of skin repair. *J. Immunol.* 2010; 184:3964–77. [PubMed: 20176743]
171. Loke P, Gallagher I, Nair MG, Zang X, Brombacher F, et al. Alternative activation is an innate response to injury that requires CD4⁺ T cells to be sustained during chronic infection. *J. Immunol.* 2007; 179:3926–36. [PubMed: 17785830]
172. Daley JM, Brancato SK, Thomay AA, Reichner JS, Albina JE. The phenotype of murine wound macrophages. *J. Leukoc. Biol.* 2010; 87:59–67. [PubMed: 20052800]
173. Helming L, Gordon S. Molecular mediators of macrophage fusion. *Trends Cell Biol.* 2009; 19:514–22. [PubMed: 19733078]
174. Sissons JR, Peschon JJ, Schmitz F, Suen R, Gilchrist M, Aderem A. Cutting edge: microRNA regulation of macrophage fusion into multinucleated giant cells. *J. Immunol.* 2012; 189:23–27. [PubMed: 22661094]
175. Martinez-Nunez RT, Louafi F, Sanchez-Elsner T. The interleukin 13 (IL-13) pathway in human macrophages is modulated by microRNA-155 via direct targeting of interleukin 13 receptor α 1 (IL13R α 1). *J. Biol. Chem.* 2010; 286:1786–94. [PubMed: 21097505]
176. Lumeng CN, Bodzin JL, Saltiel AR. Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *J. Clin. Investig.* 2007; 117:175–84. [PubMed: 17200717]
177. Marathe C, Bradley MN, Hong C, Chao L, Wilpitz D, et al. Preserved glucose tolerance in high-fat-fed C57BL/6 mice transplanted with PPAR γ ^{-/-}, PPAR δ ^{-/-}, PPAR γ δ ^{-/-}, or LXR α β ^{-/-} bone marrow. *J. Lipid Res.* 2009; 50:214–24. [PubMed: 18772483]
178. Cipolletta D, Feuerer M, Li A, Kamei N, Lee J, et al. PPAR- γ is a major driver of the accumulation and phenotype of adipose tissue Treg cells. *Nature.* 2012; 486:549–53. [PubMed: 22722857]
179. Odegaard JI, Chawla A. Alternative macrophage activation and metabolism. *Annu. Rev. Pathol. Mech. Dis.* 2011; 6:275–97.
180. Nguyen KD, Qiu Y, Cui X, Goh YPS, Mwangi J, et al. Alternatively activated macrophages produce catecholamines to sustain adaptive thermogenesis. *Nature.* 2011; 480:104–8. [PubMed: 22101429]
181. Zaiss MM, Kurowska-Stolarska M, Bohm C, Gary R, Scholtysek C, et al. IL-33 shifts the balance from osteoclast to alternatively activated macrophage differentiation and protects from TNF- α -mediated bone loss. *J. Immunol.* 2011; 186:6097–105. [PubMed: 21515798]
182. Prokop S, Heppner FL, Goebel HH, Stenzel W. M2 polarized macrophages and giant cells contribute to myofibrosis in neuromuscular sarcoidosis. *Am. J. Pathol.* 2011; 178:1279–86. [PubMed: 21356378]
183. O'Brien J, Lyons T, Monks J, Lucia MS, Wilson RS, et al. Alternatively activated macrophages and collagen remodeling characterize the postpartum involuting mammary gland across species. *Am. J. Pathol.* 2010; 176:1241–55. [PubMed: 20110414]
184. Maresz K, Ponomarev ED, Barteneva N, Tan Y, Mann MK, Dittel BN. IL-13 induces the expression of the alternative activation marker Ym1 in a subset of testicular macrophages. *J. Reprod. Immunol.* 2008; 78:140–48. [PubMed: 18329106]
185. Haschemi A, Kosma P, Gille L, Evans CR, Burant CF, et al. The sedoheptulose kinase CARKL directs macrophage polarization through control of glucose metabolism. *Cell Metab.* 2012; 15:813–26. [PubMed: 22682222]

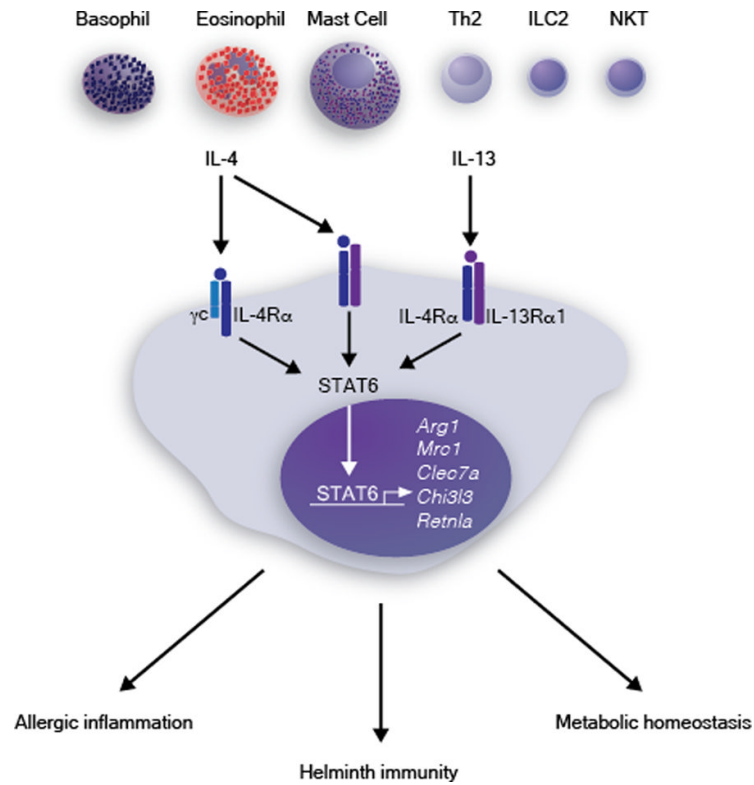


FIGURE 1. Alternatively activated macrophages in disease and homeostasis

Various cell types involved in type 2 immune responses or maintenance of metabolic homeostasis are capable of producing the canonical type 2 cytokines IL-4 and IL-13. These cytokines induce the alternatively activated phenotype in macrophages, which is characterized by the expression of a distinct set of genes (examples shown for mouse macrophages). Alternatively activated macrophages are induced in tissues associated with type 2 immune responses such as those that occur with allergic inflammation and helminth infection as well as in adipose tissue during maintenance of metabolic homeostasis. Abbreviations: ILC2, innate lymphoid type 2 cell; NKT, natural killer T cell; STAT6, signal transducer and activator of transcription 6; Th2, type 2 helper T cell.

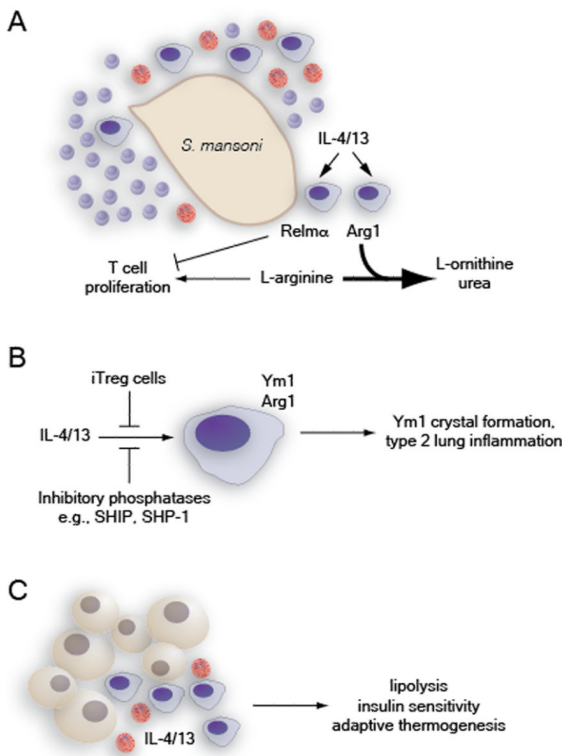


FIGURE 2. Functional aspects of alternatively activated macrophages (AAMs) during immunity and homeostasis

(a) IL-4 and/or IL-13 produced in the context of *Schistosoma mansoni*-induced granuloma formation in the liver induces AAMs. In mouse tissues, arginase 1 (Arg1) is produced by AAMs and mediates T cell suppression by catabolizing available L-arginine into L-ornithine and urea, thereby depleting an essential T cell proliferative factor. Relm α is also produced by AAMs and mediates T cell suppressive effects. (b) IL-4/IL-13 production and/or receptor signaling is suppressed in the steady state by induced T regulatory (iTreg) cells and inhibitory phosphatases such as SHIP and SHP-1. In their absence, the AAM phenotype is spontaneously induced, resulting in excessive expression in vivo of Arg1 and Ym1, which leads to Ym1 crystal formation and type 2 inflammation in the lungs. (c) Adipose tissue in lean animals contains eosinophils and AAMs, which mediate maintenance of metabolic homeostasis via lipolysis, insulin sensitivity, and stress-induced thermogenesis; in obese animals, AAMs are replaced by classically activated macrophages and these effects are diminished.