

Corrections

BIOCHEMISTRY

Correction for “Confinement of caspase-12 proteolytic activity to autoprocessing,” by Sophie Roy, Jeffrey R. Sharom, Caroline Houde, Thomas P. Loisel, John P. Vaillancourt, Wei Shao, Maya Saleh, and Donald W. Nicholson, which appeared in issue 11, March 18, 2008, of *Proc Natl Acad Sci USA* (105:4133–4138; first published March 10, 2008; 10.1073/pnas.0706658105).

The authors note that Fig. 6 appeared incorrectly. There was an error in the alignment of the molecular mass markers, and minor adjustments have been made to the assignment of caspase-12 bands. The authors also note that the source of the anti-caspase-12 antibody for Fig. 6 was Sigma (clone 14F7). This error does not affect the conclusions of the article. The corrected figure and its corresponding legend appear below.

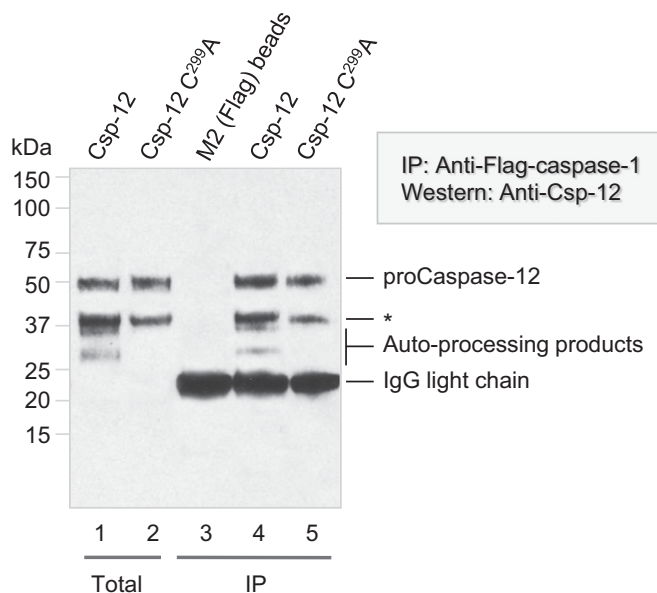


Fig. 6. Procasase-12 forms a complex with caspase-1 and is partially autoprocessed in the complex. HEK293T cells were cotransfected with expression vectors harboring Flag-tagged procaspase-1 (all lanes) plus either procaspase-12 (lanes 1 and 4) or the catalytically incapacitated C²⁹⁹A mutant (lanes 2 and 5). After 24 h, cells were harvested and lysed. One tenth of the lysate was directly applied to SDS/PAGE (lanes 1 and 2), and the remainder was immunoharvested with antibodies directed against the caspase-1 Flag epitope tag (lanes 4 and 5; lane 3 was processed in the same way, except that only lysis buffer was used). Immunoblotting for the large subunit (p20) of caspase-12 revealed that procaspase-12 and the resulting autocleavage product were both immunoharvested with caspase-1. The asterisk indicates a band of unknown identity that is detected by prebleed control serum.

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CHEMISTRY

Correction for “Direct asymmetric vinylogous Michael addition of cyclic enones to nitroalkenes via dienamine catalysis,” by Giorgio Bencivenni, Patrizia Galzerano, Andrea Mazzanti, Giuseppe Bartoli, and Paolo Melchiorre, which appeared in issue 48, November 30, 2010, of *Proc Natl Acad Sci USA* (107:20642–20647; first published June 21, 2010; 10.1073/pnas.1001150107).

The authors note that they incorrectly assigned the structure of the reaction product reported in Scheme 4. The published structure represents the γ -aminated adduct **8**, when it should instead be the α -analogue arising from an α -site selective pathway. As a result of this, Scheme 4 and its related comments should be removed from the article.

On page 20642, left column, within the Abstract, lines 16–18, “Finally, we describe the extension of the dienamine catalysis-induced vinylogous nucleophilicity to the asymmetric γ -amination of cyclohexene carbaldehyde” should be removed from the article.

On page 20645, right column, third full paragraph, lines 1–8, to page 20646, left column, first paragraph, lines 1–2, “Finally, to explore the potential of the chiral primary amine-induced vinylogous nucleophilicity, we wondered whether this unique reactivity concept may be translated to an aldehyde derivative adorned with a six-membered ring scaffold, reminiscent of the β -substituted cyclohexanone framework. Although the vinylogous Michael addition of 1-cyclohexene-1-carboxaldehyde **7** to nitrostyrene **2a** did not proceed at all, the combination with *tert*-butylazodicarboxylate under the catalysis of **A** furnished the γ -amination product **8** with perfect regio- and enantioselectivity (Scheme 4)” should be removed from the article.

These errors do not affect the conclusions of the article of the vinylogous Michael addition of cyclic enones to nitroalkenes. The ability of primary amine catalysis to address the synthetic issue connected with the enantioselective carbon–carbon bond formation gamma to a carbonyl group, promoting vinylogous nucleophilicity upon selective activation of unmodified cyclic unsaturated ketones, is fully supported by the separated results presented in Tables 1, 2, and 3, and Schemes 2 and 3.

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MICROBIOLOGY

Correction for “Commensal bacteria play a role in mating preference of *Drosophila melanogaster*,” by Gil Sharon, Daniel Segal, John M. Ringo, Abraham Hefetz, Ilana Zilber-Rosenberg, and Eugene Rosenberg, which appeared in issue 46, November 16, 2010, of *Proc Natl Acad Sci USA* (107:20051–20056; first published November 1, 2010; 10.1073/pnas.1009906107).

The authors note the following: “The mating frequencies reported in Table 1 of this paper did not follow a multinomial distribution, making the statistical analysis inapplicable. This problem was obviated by considering only the first matings in each experimental unit and computing odds ratios. After submitting the paper, we continued to perform experiments

identical in design to those we reported. In the table below, we combined the results of those additional replicate experiments with those already reported. From the new analysis, we now find that experiment 4, in which flies were infected with a mixture of *Lactobacillus* spp., assortative mating was not restored. Otherwise, the conclusions of the article were not changed by our reanalysis. We acknowledge the statistical advice of Dan Yekutieli and thank Tal Lahav for calculating the odds ratios and their 95% confidence intervals, and for performing the chi-squared tests presented in the corrected Table 1.”

The corrected Table 1 appears below.

Table 1. The role of bacteria in diet-induced mating preference of *D. melanogaster*

Experiment	Fly treatment*	N [†]	OR [‡]	95% CI	P value [‡]
1	Starch-grown × CMY-grown	18	3.21	2.14–4.81	1.8 × 10 ⁻⁸
2	Experiment 1 after antibiotics	11	1.04	0.63–1.71	0.9888
3	Experiment 2 after infection of starch-grown flies with homologous bacteria [§]	6	2.68	1.40–5.11	0.0477
4	Experiment 3 with <i>Lactobacillus</i> spp. replacing homologous bacteria	4	1.76	0.74–4.19	0.2912
5	Experiment 3 with <i>Lactobacillus plantarum</i> replacing homologous bacteria	7	2.14	1.35–3.39	0.0019
6	Infection control (no added bacteria)	4	1.26	0.53–3.00	0.7712

*After all treatments, the flies were grown for one generation in CMY medium before performing the mating preference test.

[†]N is the number of replicate experiments.

[‡]Cochran-Mantel-Haenszel Odds Ratio and P value are from the Cochran-Mantel-Haenszel Chi-squared test (34, 35).

[§]Antibiotic-treated starch- and CMY-grown flies were infected with bacteria isolated from their respective growth medium (before antibiotic treatment).

34. Mantel N, Haenszel W (1959) Statistical aspects of the analysis of data from retrospective studies of disease. *J Natl Cancer Inst* 22(4):719–748.

35. Cochran WD (1954) Some methods for strengthening the common χ^2 tests. *Biometrics* 10:417–451.

www.pnas.org/cgi/doi/10.1073/pnas.1302326110

MEDICAL SCIENCES

Correction for “Interaction of intracellular β amyloid peptide with chaperone proteins,” by Virginia Fonte, Vadim Kapulkin, Andrew Taft, Amy Fluet, David Friedman, and Christopher D. Link, which appeared in issue 14, July 9, 2002, of *Proc Natl Acad Sci USA* (99:9439–9444; first published June 27, 2002; 10.1073/pnas.152313999).

The authors note that the author name Vadim Kapulkin should instead appear as Wadim Jan Kapulkin. The corrected author line appears below. The online version has been corrected.

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SYSTEMS BIOLOGY

Correction for “Expanded methyl-sensitive cut counting reveals hypomethylation as an epigenetic state that highlights functional sequences of the genome,” by Alejandro Colaneri, Nickolas Staffa, David C. Fargo, Yuan Gao, Tianyuan Wang, Shyamal D. Peddada, and Lutz Birnbaumer, which appeared in issue 23, June 7, 2011, of *Proc Natl Acad Sci USA* (108:9715–9720; first published May 20, 2011; 10.1073/pnas.1105713108).

The authors note that, within the corresponding author footnote on page 9715, the email address “colaneria@niehs.nih.gov” should instead appear as “acolaneri_2000@yahoo.com.ar”.

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