

Research Article

Modulation of Cell Cycle Profile by *Chlorella vulgaris* Prevents Replicative Senescence of Human Diploid Fibroblasts

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In this study, the effects of *Chlorella vulgaris* (CV) on replicative senescence of human diploid fibroblasts (HDFs) were investigated. Hot water extract of CV was used to treat HDFs at passages 6, 15, and 30 which represent young, presenescence, and senescence ages, respectively. The level of DNA damage was determined by comet assay while apoptosis and cell cycle profile were determined using FACSCalibur flow cytometer. Our results showed direct correlation between increased levels of damaged DNA and apoptosis with senescence in untreated HDFs ($P < 0.05$). Cell cycle profile showed increased population of untreated senescent cells that enter G0/G1 phase while the cell population in S phase decreased significantly ($P < 0.05$). Treatment with CV however caused a significant reduction in the level of damaged DNA and apoptosis in all age groups of HDFs ($P < 0.05$). Cell cycle analysis showed that treatment with CV increased significantly the percentage of senescent HDFs in S phase and G2/M phases but decreased the population of cells in G0/G1 phase ($P < 0.05$). In conclusion, hot water extract of *Chlorella vulgaris* effectively decreased the biomarkers of ageing, indicating its potential as an antiageing compound.

1. Introduction

Regardless of the persons' official age, senescence is associated with appearance of age-related phenotypes, decline in protein homeostasis, and accumulation of DNA damage which alters individual lifespan. The theories of ageing can be classified into two broad categories, namely, the genetic and stochastic theories [1]. The genetic theories postulated that ageing process is under the control of the same genes responsible for running the life cycle program [2]. However, the stochastic theories rely on the accumulation of cell damages and their inefficient repair as the main reason of ageing [3]. In other words, we can do nothing with ageing based on the genetic theories, but it is reversible based on the stochastic theories by improving the cellular defense system or decreasing cell damages.

Based on the free radical theory of ageing, accumulation of free radicals over times leads to cellular damages and

progressive deterioration of cells functions, tissues, and organ systems [4]. Due to their instability, the free radicals will attack the neighboring molecules to balance their unpaired electrons. Free radicals can be produced intrinsically through normal metabolic processes or extrinsically from exogenous agents [5, 6]. The mitochondria, for instance, are major contributors of reactive oxygen species (ROS) in the cells [4], and any dysfunction in this organelle can result in energy shortage and other cellular deficiency that finally give rise to age-related disorders such as muscular and neurological degeneration, heart failure, strokes, and other degenerative diseases and finally death.

Therefore, introducing natural or synthetic compounds with antioxidant properties may be beneficial to protect cells or tissues against oxidative damage which may result in age-related diseases as well as promoting longevity. Previous study showed that antioxidant activity of resveratrol in grapes and tocotrienol in palm oil extends lifespan [7–10].

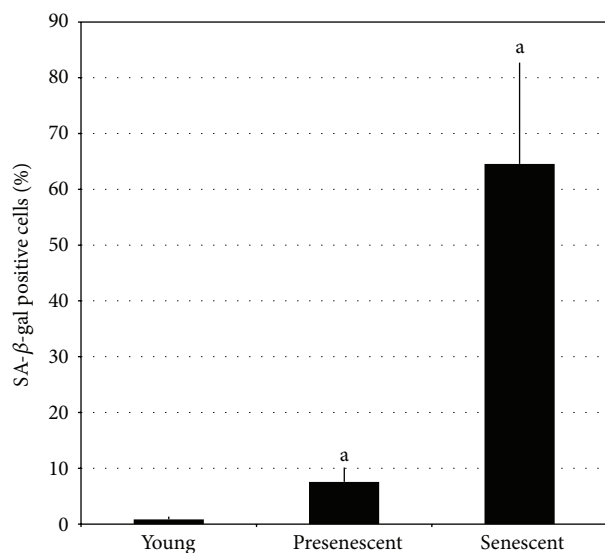


FIGURE 2: Quantitative analysis of positive SA β -gal stained cells. Data are expressed as means \pm SD, $n = 6$. ^aDenotes $P < 0.05$ compared to young HDFs.

was subsequently lyophilized to obtain *C. vulgaris* extracts (CVE) which was then added to cell culture growth medium as supplementation.

2.2. Cell Culture and Treatment Protocol. Human diploid fibroblasts (HDFs) were obtained from the foreskin of 8- to 12-year-old boys who underwent circumcision. Written informed consent was obtained from parents of all subjects. The HDFs were cultured till different passages (P6, P15 and P30), representing different age groups (young, pre-senescent, and senescent, resp.). Each age group was divided into two subgroups, treated with CV and untreated control. Young, presenescent, and senescent HDFs were treated with 400 $\mu\text{g}/\text{mL}$, 200 $\mu\text{g}/\text{mL}$ and 100 $\mu\text{g}/\text{mL}$ of *Chlorella vulgaris*, respectively for 24 h before determination of aging biomarkers which include morphological analysis, senescence-associated β -galactosidase (SA-B-gal) expression, DNA damage, apoptosis, and cell cycle profile. The different dosages used for CV treatment were obtained from viability assay. This research has been approved by the Ethics Committee of the Universiti Kebangsaan Malaysia (approval project code: FF-028-2012).

2.3. Senescence-Associated Beta-Galactosidase (SA- β -gal) Staining. Expression of SA- β -gal was determined based on the method suggested by Dimri et al. [17] with minor modification. HDFs (1×10^5) were inoculated in 6-well plates and treated with fixation buffer containing 0.2% glutaraldehyde and 2% formaldehyde. After incubation for 6-7 min at room temperature, the cells were washed with PBS (three times) and incubated for 4 h at 37°C (without CO₂) in buffer containing 1 mg/mL 3-indolyl-beta-D-galactopyranoside, 40 mM citric acid/phosphate (pH 6.0), 2 mM MgCl₂, 5 mM K₃FeCN₆, and 5 mM NaCl. Blue staining was visible after

incubation, and the percentage of blue cells observed in 100 cells under a light microscope was calculated.

2.4. Determination of DNA Damage. DNA damage was determined by single cell gel electrophoresis (SCGE) assay also known as the comet assay according to the method previously described by Singh et al. [18] with slight modifications. Briefly, 1×10^4 cells in 5 μL medium were suspended in 70 μL warm 0.6% low-melting point (LMA) agarose (Boehringer Mannheim, Germany) (DNase-free, RNase-free) and was quickly pipetted onto a prepared frosted slide coated with 80 μL of 0.6% normal-melting (NMA) agarose. Slides with cell suspension were covered with glass coverslips and subsequently placed in a Coplin jar containing chilled lysis solution (10% DMSO, 1% Triton X in alkaline lysis buffer: 2.5 M NaCl, 10 mM Tris, 100 mM Na₂EDTA, pH 10) for 1 h to completely digest cellular proteins. Next, the slides were removed from lysis solution and placed in a horizontal gel electrophoresis unit filled with cold electrophoresis buffer (10 mM NaOH and 200 mM Na₂EDTA at pH 13.2). Slides were allowed to sit in the alkaline buffer for 20 min to allow unwinding of DNA strands and expression of alkali-labile damage. Electrophoresis was performed for 20 min at 300 mA and 25 V. Following dropwise neutralization (Tris-HCl, pH 7.5) for 5 min, cells were stained by applying 30 μL 1X ethidium bromide. The slides were examined and the tail length was measured with a fluorescence microscope (Carl Zeiss, Germany). The DNA migration of 100 randomly selected cells was examined for each sample. A total damage score was determined by multiplying the number of cells assigned to each grade of damage by the numeric value of the grade according to methods described by Heaton et al. [19]. Total DNA damage score was calculated as follows.

Total DNA damage = $[(0 \times n_0) + (1 \times n_1) + (2 \times n_2) + (3 \times n_3) + (4 \times n_4)]$ where n_0 = cells with Score 0, n_1 = cells with Score 1, n_2 = cells with Score 2, n_3 = cells with Score 3, and n_4 = cells with Score 4.

2.5. Apoptosis Determination by Annexin V-FITC. Externalization of phosphatidylserine (PS) at the cell surface of HDFs as an early apoptotic event was assessed by Annexin V-FITC Apoptosis Detection Kit II (BD Pharmingen, San Diego, CA, USA) according to the manufacturer's instructions. Cells were washed twice with cold PBS and resuspended in 1X binding buffer to prepare 1×10^6 cells/mL concentration. Then 5 μL of Annexin V-FITC and 5 μL of PI staining solution were added into a 5 mL culture tube containing 100 μL of cells suspension, followed by incubation for 15 min in the dark at room temperature (25°C). Finally, cells were suspended in 400 μL 1X binding buffer and analyzed within 1 h by FACSCalibur flow cytometer (Becton Dickinson, San Jose, CA, USA).

2.6. Determination of Cell Cycle Profile. HDFs were harvested at desired time points after trypsinization and were rinsed 3 times with buffer solution with adjusted concentration 1×10^6 cells/mL and prepared using CycleTEST PLUS DNA Reagent Kit (Becton Dickinson, USA) according to

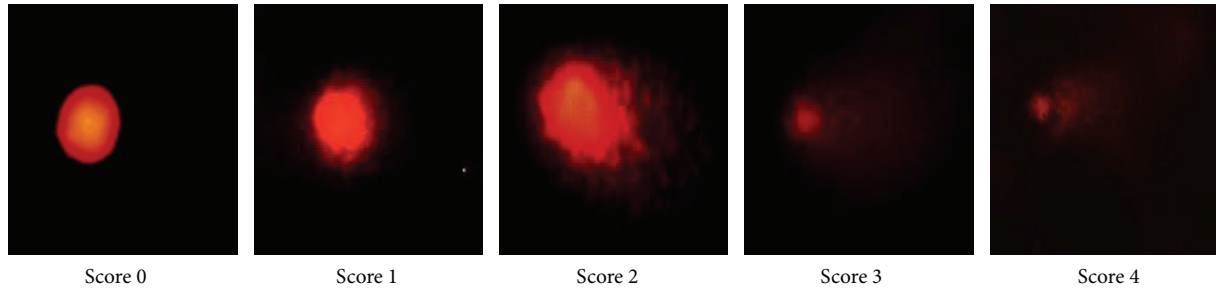


FIGURE 3: DNA damage. The cell nuclei were classified into five categories: (Score 0), undamaged (nuclei without comet tail); (Score 1), low damaged (nuclei with comet tails up to two- fold longer than nucleus diameter); (Score 2), damaged (nuclei with comet tail two- to threefold longer than nucleus diameter); (Score 3), highly damaged (nuclei with comet tails threefold longer than nucleus diameter); and (Score 4), severely damaged (cell nuclei were almost not visible with long and dispersed comet tails).

the manufacturer's instruction. Cell cycle status was analyzed by flow cytometer using propidium iodide (PI) as a specific fluorescent dye probe. The PI fluorescence intensity of 10,000 cells was measured for each sample using a Becton-Dickinson FACSCalibur flow cytometer.

2.7. Statistical Analysis. Each experiment was carried out in duplicates from three independent cultures. Data were reported as means \pm SD. Comparison between groups was made by Student's *t*-test (two-tailed). $P < 0.05$ was considered statistically significant

3. Results

3.1. Senescence-Associated Beta-Galactosidase (SA- β -gal) Expression. Young HDFs display spindle-shaped morphology while senescent fibroblasts were larger and flattened in shape containing cytoplasmic inclusions (Figure 1). Senescent HDFs at passage 30 contained numerous blue stain granules compared to HDFs at lower passages (young and pre-senescent HDFs). Quantitative analysis showed a significant increase in SA- β -gal in pre-senescent and senescent HDFs as compared to young cells ($P < 0.05$) (Figure 2).

3.2. Determination of DNA Damage. The cell nuclei were classified into five categories: (0), undamaged (nuclei without Comet tail); (1), low damaged (nuclei with Comet tails up to two fold longer than nucleus diameter); (2), damaged (nuclei with Comet tail two- to threefold longer than nucleus diameter); (3), highly damaged (nuclei with Comet tails threefold longer than nucleus diameter); and (4), severely damaged (cell nuclei were almost not visible with long and dispersed Comet tails) (Figure 3). Senescent cells showed a significant increase in damaged DNA as compared to young and pre-senescent cells ($P < 0.05$). Treatment with CV, however, significantly decreased the damaged DNA in all age groups of HDFs ($P < 0.05$) (Figure 4).

3.3. Apoptotic Changes Detected by Annexin V-FITC. Figure 5 shows the contour diagram of FITC-Annexin V/PI double staining by flow cytometry analysis. The three quadrants

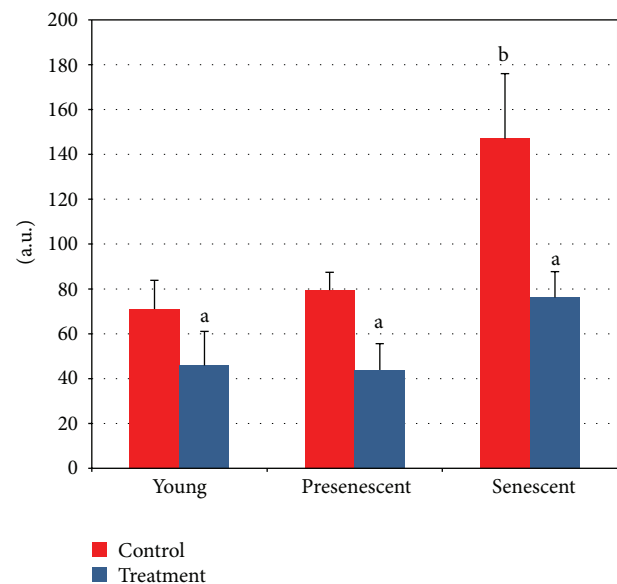
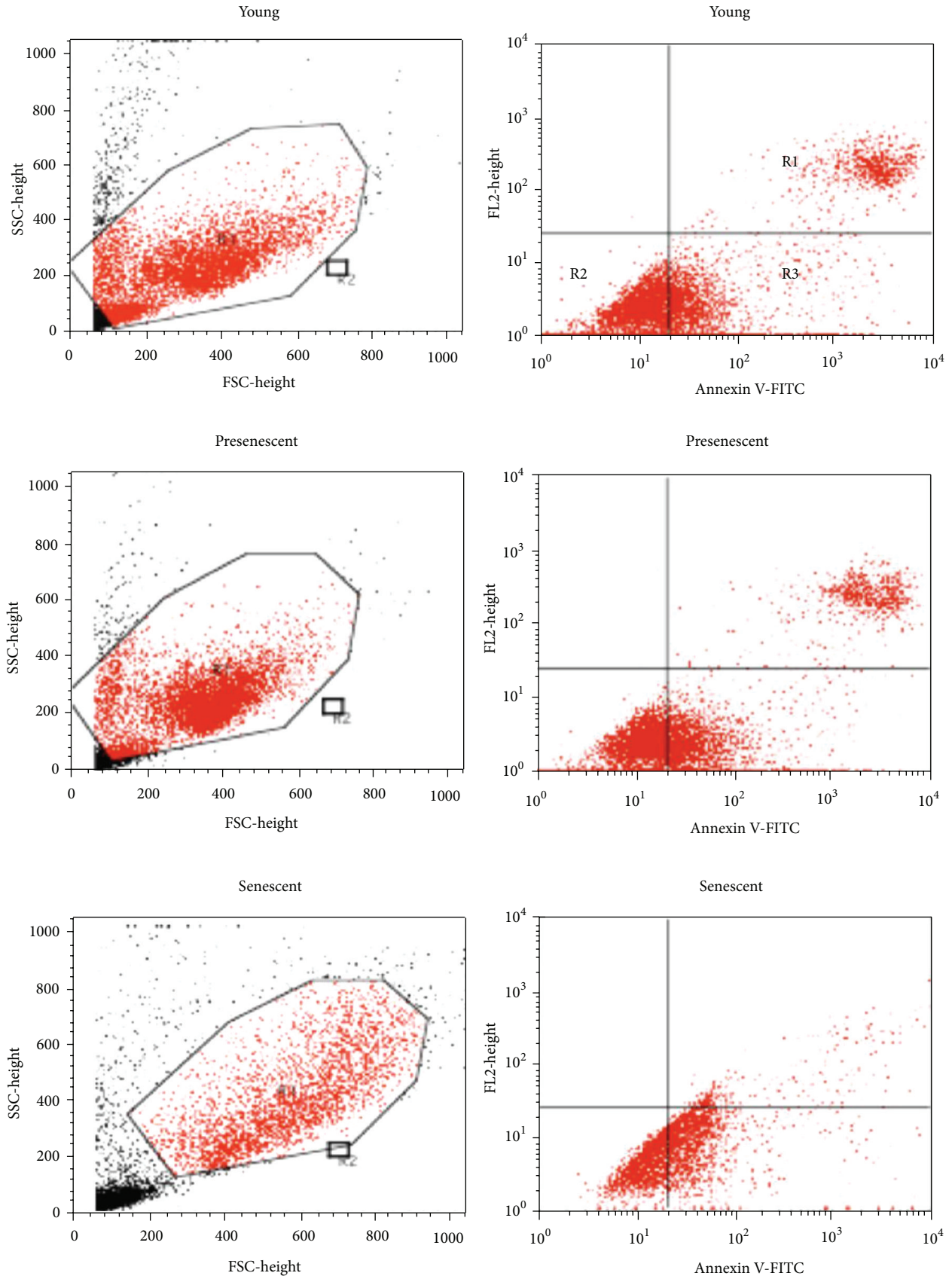


FIGURE 4: Protective effects of CV against DNA damage in young, presenescent, and senescent HDFs. ^aDenotes $P < 0.05$ compared to untreated HDFs, ^b $P < 0.05$ compared to untreated young HDFs. Data are expressed as means \pm SD, $n = 6$.

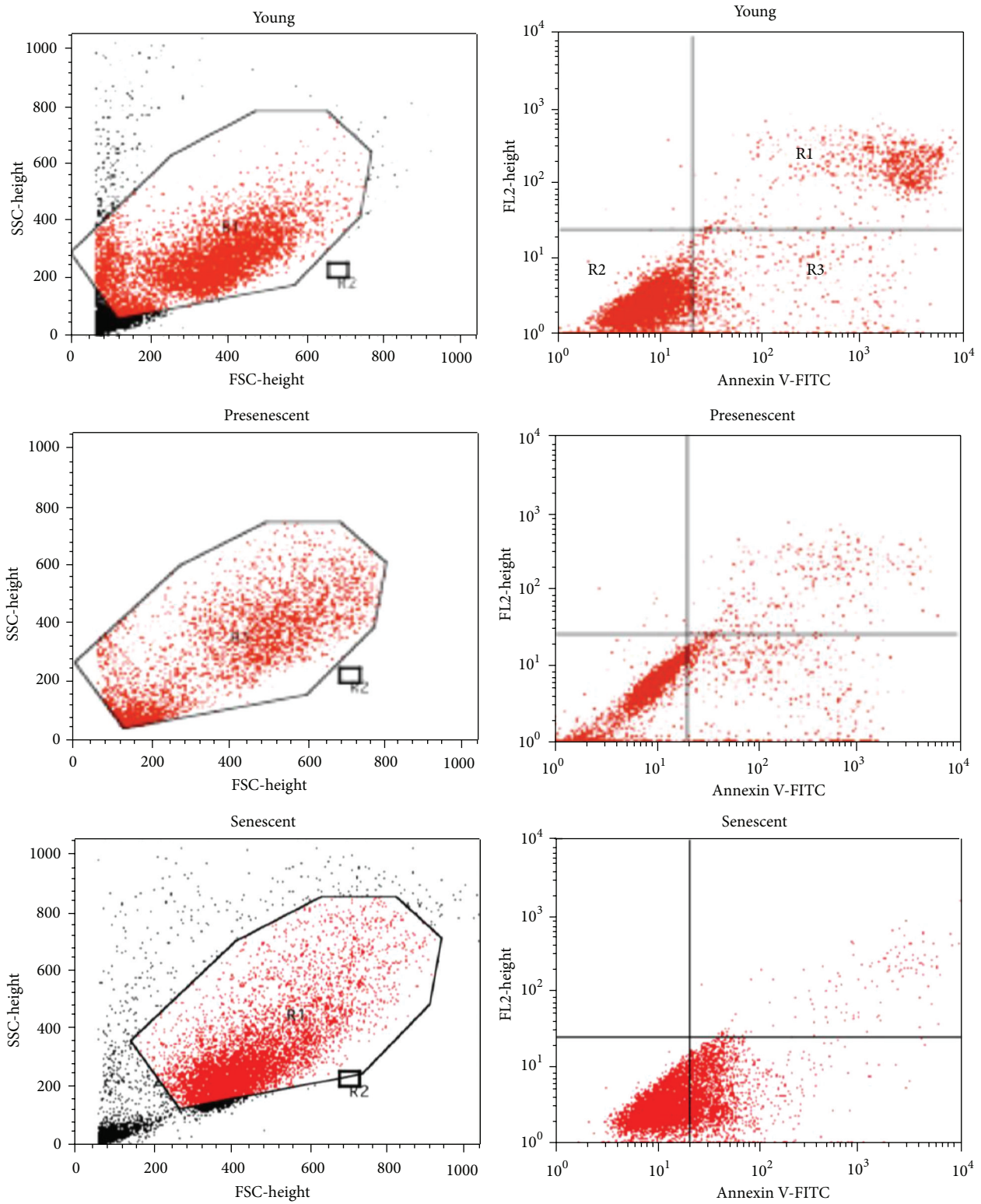
represent different cells conditions; the upper right quadrant (R1) indicates nonviable, late apoptotic, and necrotic cells (FITC⁺/PI⁺), lower left quadrant (R2) indicates viable cells (FITC⁻/PI⁻) and lower right quadrant (R3) indicates early apoptotic cells (FITC⁺/PI⁻), which is demonstrated by Annexin V binding and cytoplasmic membrane integrity. The percentage of cells at early apoptotic stage in all age groups was significantly decreased in HDFs treated with CV as compared to untreated control ($P < 0.05$) (Figure 6).

3.4. Cell Cycle Profile Analysis. Analysis on cell cycle profile showed that HDFs population in G0/G1 phase was significantly increased while S phase and G2/M phase cells decreased in senescent cells as compared to young HDFs (Figures 7 and 8) ($P < 0.05$). Treatment with CV significantly increased the number of cells in S phase and G2/M phase



(a)

FIGURE 5: Continued.



(b)

FIGURE 5: Contour diagram of FITC-Annexin V/PI double staining by flow cytometry. The three quadrants represent different cells conditions the upper right quadrant (R1), nonviable, late apoptotic, and necrotic cells (FITC⁺/PI⁺); lower left quadrant (R2), viable cells (FITC⁻/PI⁻) and lower right quadrant (R3), early apoptotic cells (FITC⁺/PI⁻). (a) Untreated control HDFs. (b) CV-treated HDFs.

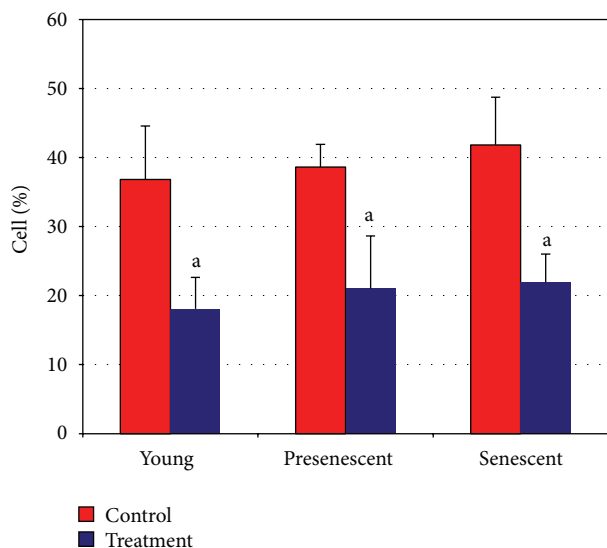


FIGURE 6: Percentage of cells at early apoptotic stage demonstrated by FITC⁺/PI⁻. ^aDenotes $P < 0.05$ compared to untreated HDFs. Data are expressed as means \pm SD, $n = 6$.

($P < 0.05$) in all age groups of HDFs. In contrast, population of cells in G0/G1 phase decreased significantly ($P < 0.05$) after CV treatment in all age groups (Figure 8).

4. Discussion

For the past few decades, researchers have tried to find a way to postpone ageing and extend lifespan. Findings from ageing research provide promising data to prolong maximum lifespan of human being [20]. Normal metabolic processes which involved respiratory reactions in electron transport chain in the mitochondria produce reactive oxygen species (ROS) such as O_2^- , H_2O_2 , ROO^- , OH^\bullet , and O_3^- . These free radicals are hazardous to cells and can initiate senescence pathway [21, 22]. Although eukaryotic cells are able to remove the free radicals, their ability is limited as they cannot cope with the deleterious effects of accumulated free radicals during life [23, 24]. Accumulation of free radicals causes damage to DNA due to increase in mutation rate and cells' inability to repair the damaged DNA [25]. Damaged DNA will further activate tumor suppressor genes such as p53 and Rb and halt cell proliferation, leading to senescence [23, 25, 26]. Therefore, reduction of oxidative stress by an antiageing compound is considered as an effective antiageing intervention.

Several antiageing compounds and natural products have been identified which reduced the oxidative damage of mitochondrial DNA and decreased mitochondrial reactive oxygen species production [27, 28]. For instance, vitamin E has been proven to have antiageing properties. Tocotrienol, a type of vitamin E, was reported to prevent telomere shortening [29] and decreased oxidative damage in ageing model [27]. Another natural product with antiageing properties is garlic which acts as an efficient anti-inflammatory factor and an antioxidant agent that can remove reactive oxygen species and peroxy radicals [30, 31]. Similar effects have

been shown by Curcumin derived from turmeric [32, 33]. Therefore introducing antioxidant compounds will improve physiological functions of cells and delay the onset of age-related phenomenon in the cells [34].

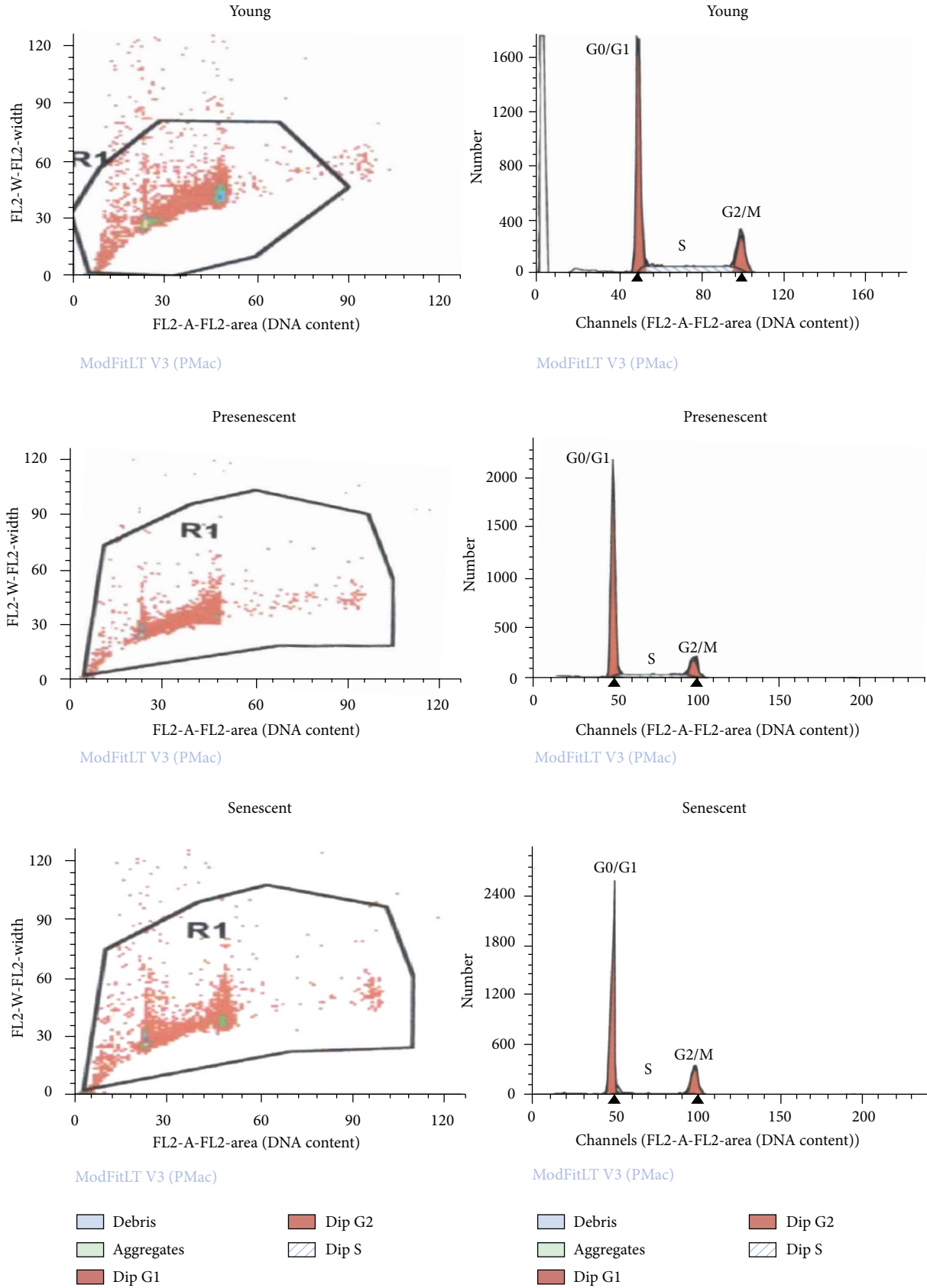
Chlorella vulgaris (CV) is a spherical (2–10 μm) and nonflagellated green algae belonging to the phylum Chlorophyta. It contains proteins, fibers, minerals, vitamins, and chlorophylls and is considered as a rich source of antioxidant such as α - and β -carotene, lutein, ascorbic acid, and vitamin E [35–37]. CV is well recognized as a biological modifier that can modulate immune responses against viruses, bacteria, and tumors by increasing CD4⁺CD8⁺ T cells, granulocytes, and macrophage [11, 38, 39]. Because of its nutritional values, CV has been used as a supplement in many countries such as Japan, Taiwan, and the USA [11, 38, 40].

Therefore CV with its antioxidant properties may contribute in postponing ageing by decreasing oxidative stress in cells. In this study, the effects of CV on a few ageing biomarkers including DNA damage, apoptosis, and cell cycle arrest were evaluated. Primary human diploid fibroblast cells derived from foreskin were cultured until passage 30 which represents cellular senescence. Senescent cells were confirmed by the presence of SA- β -gal which forms aggregates in autophagic lysosomal vacuoles when cells reached senescence [41–43]. The findings of this study showed that 64.6% of the cells in passage 30 were stained for SA- β -gal indicating senescent cells. It has been reported that increased autophagic vacuoles and β -galactosidase activity during cell ageing are associated with increase in lysosomal mass and accumulation of lipofuscin [43, 44].

Since DNA damage is one of the critical factors that trigger ageing, we also investigated the effects of CV on DNA damage. Our results showed increased damaged DNA with cellular ageing and CV could significantly reduce DNA damage in all age groups of HDFs. This could be due to its antioxidant properties which quenched the reactive ROS and free radicals from attacking the DNA. Thus CV conserves the DNA from destruction and can effectively delay cellular ageing.

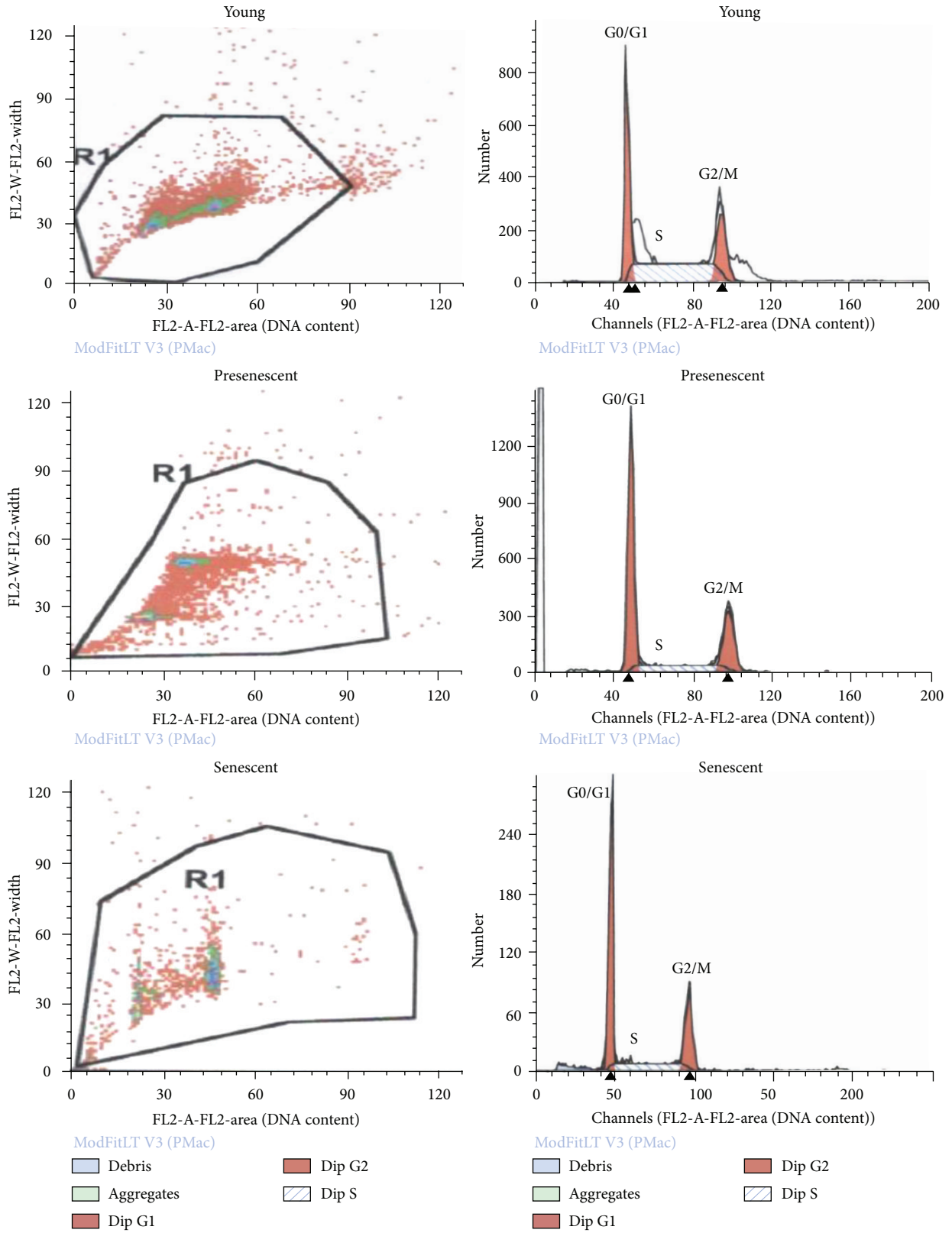
Our results showed an extensive decline in the percentage of senescent HDFs that enter into S phase while the percentage of cells in G0/G1 phase was increased with senescence. CV treatment to HDFs increased the percentage of cells that enter into S and G2/M phases and at the same time decreased the percentage of cells that enter into G1 phase. The direct relationship between decreased DNA damage and increased cell replication suggests improvement in cell division. This could be due to a more efficient repair of the DNA or prevention of DNA damage in cells treated with CV.

In addition, since programmed cell death or apoptosis is the outcome of DNA damage and normally occurs in more than 10% of all living cells [21], the effect of CV on this parameter was also investigated. Our findings showed that CV could effectively decrease apoptosis in all stages of ageing (young, pre-senescence, and senescence). Therefore in senescence, the level of damaged DNA increases, which leads to apoptosis. Subsequently the cells can be directed to two different fates: either they undergo senescence or



(a)

FIGURE 7: Continued.



(b)

FIGURE 7: Flow cytometry analysis of cell cycle progression in young, presenescent, and senescent HDFs. (a) Untreated HDFs. (b) CV-treated HDFs.

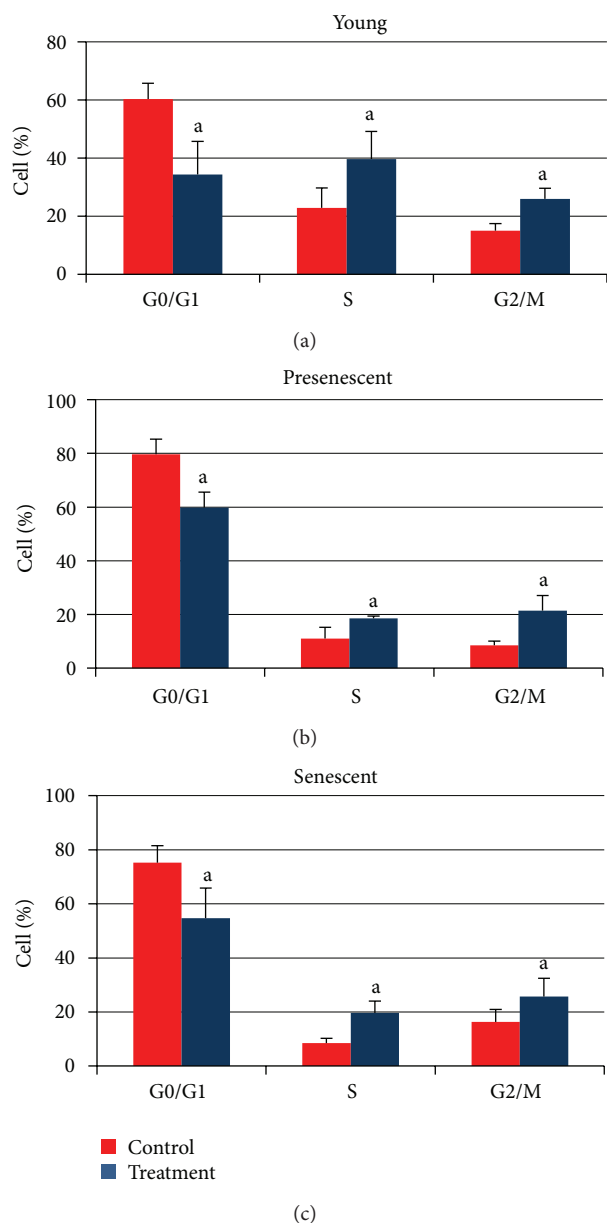


FIGURE 8: Quantitative analysis of cell cycle progression in young, presenescent, and senescent HDFs with and without CV treatment. ^aDenotes $P < 0.05$ compared to untreated HDFs. Comparison was done between HDFs in the same phase of cell cycle. Data are expressed as means \pm SD, $n = 6$.

may return to a replicative life cycle. Our data on cell cycle profile suggested that CV returns the HDF cells back to the replicative state and thus prevents cellular ageing.

5. Conclusion

Hot water extract of *Chlorella vulgaris* (CV) is able to delay cellular ageing by decreasing the level of DNA damage and apoptosis as well as promote cell cycle progression. Therefore, *Chlorella vulgaris* extracts are a potent antiageing

compound; however, further studies are needed before it can be introduced as an effective antiageing supplement.

Conflict of Interests

The authors declare no conflict of interests.

Acknowledgments

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References

- [1] B. R. Troen, "The biology of aging," *The Mount Sinai Journal of Medicine*, vol. 70, no. 1, pp. 3–22, 2003.
- [2] C. E. Finch and R. E. Tanzi, "Genetics of aging," *Science*, vol. 278, no. 5337, pp. 407–411, 1997.
- [3] S. I. S. Rattan, "Increased molecular damage and heterogeneity as the basis of aging," *Biological Chemistry*, vol. 389, no. 3, pp. 267–272, 2008.
- [4] D. Harman, "Aging: a theory based on free radical and radiation chemistry," *Journal of Gerontology*, vol. 11, no. 3, pp. 298–300, 1956.
- [5] I. Fridovich, "Superoxide dismutases. An adaptation to a paramagnetic gas," *Journal of Biological Chemistry*, vol. 264, no. 14, pp. 7761–7764, 1989.
- [6] R. S. Sohal and R. Weindruch, "Oxidative stress, caloric restriction, and aging," *Science*, vol. 273, no. 5271, pp. 59–63, 1996.
- [7] C. K. Sen, S. Khanna, and S. Roy, "Tocotrienols: Vitamin E beyond tocopherols," *Life Sciences*, vol. 78, no. 18, pp. 2088–2098, 2006.
- [8] T. Li, G. X. Fan, W. Wang, T. Li, and Y. K. Yuan, "Resveratrol induces apoptosis, influences IL-6 and exerts immunomodulatory effect on mouse lymphocytic leukemia both in vitro and in vivo," *International Immunopharmacology*, vol. 7, no. 9, pp. 1221–1231, 2007.
- [9] B. B. Aggarwal, C. Sundaram, S. Prasad, and R. Kannappan, "Tocotrienols, the vitamin E of the 21st century: its potential against cancer and other chronic diseases," *Biochemical Pharmacology*, vol. 80, no. 11, pp. 1613–1631, 2010.
- [10] S. Schaffer, W. E. Müller, and G. P. Eckert, "Tocotrienols: constitutional effects in aging and disease," *Journal of Nutrition*, vol. 135, no. 2, pp. 151–154, 2005.
- [11] T. Hasegawa, K. Noda, S. Kumamoto, Y. Ando, A. Yamada, and Y. Yoshikai, "*Chlorella vulgaris* culture supernatant (CVS) reduces psychological stress-induced apoptosis in thymocytes of mice," *International Journal of Immunopharmacology*, vol. 22, no. 11, pp. 877–885, 2000.
- [12] P. Y. Morven, A. S. Lengume, H. Gendrot, and R. Vallee, "Non isotopic and specific detection of collagen synthesised by human skin fibroblasts," in *Poster IFSCC*, Berlin, Germany, 2000.
- [13] S. Makpol, N. Yaacob, A. Zainuddin, Y. A. M. Yusof, and W. Z. W. Ngah, "*Chlorella vulgaris* modulates hydrogen peroxide-induced DNA damage and telomere shortening of human fibroblasts derived from different aged individuals,"

- African Journal of Traditional, Complementary and Alternative Medicines*, vol. 6, no. 4, pp. 560–572, 2009.
- [14] H. Jiang, W. Chen, L. Qu et al., “ELISA for aging biomarkers induced by telomere dysfunction in human plasma,” *Journal of Biomedicine and Biotechnology*, vol. 2010, Article ID 121947, 4 pages, 2010.
- [15] H. M. Wang, J. L. Pan, C. Y. Chen et al., “Identification of anti-lung cancer extract from *Chlorella vulgaris* C-C by antioxidant property using supercritical carbon dioxide extraction,” *Process Biochemistry*, vol. 45, no. 12, pp. 1865–1872, 2010.
- [16] T. Hasegawa, K. Ito, S. Ueno et al., “Oral administration of hot water extracts of *Chlorella vulgaris* reduces IgE production against milk casein in mice,” *International Journal of Immunopharmacology*, vol. 21, no. 5, pp. 311–323, 1999.
- [17] G. P. Dimri, X. Lee, G. Basile et al., “A biomarker that identifies senescent human cells in culture and in aging skin in vivo,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 92, no. 20, pp. 9363–9367, 1995.
- [18] N. P. Singh, R. E. Stephens, and E. L. Schneider, “Modifications of alkaline microgel electrophoresis for sensitive detection of DNA damage,” *International Journal of Radiation Biology*, vol. 66, no. 1, pp. 23–28, 1994.
- [19] P. R. Heaton, R. Ransley, C. J. Charlton et al., “Application of single-cell gel electrophoresis (comet) assay for assessing levels of DNA damage in canine and feline leukocytes,” *Journal of Nutrition*, vol. 132, no. 6, 2002.
- [20] K. Merker, A. Stolzing, and T. Grune, “Proteolysis, caloric restriction and aging,” *Mechanisms of Ageing and Development*, vol. 122, no. 7, pp. 595–615, 2001.
- [21] P. Laun, G. Heeren, M. Rinnerthaler et al., “Senescence and apoptosis in yeast mother cell-specific aging and in higher cells: a short review,” *Biochimica et Biophysica Acta*, vol. 1783, no. 7, pp. 1328–1334, 2008.
- [22] J. L. Teo and M. Kahn, “The Wnt signaling pathway in cellular proliferation and differentiation: a tale of two coactivators,” *Advanced Drug Delivery Reviews*, vol. 62, no. 12, pp. 1149–1155, 2010.
- [23] E. G. Nette, Y. P. Xi, and Y. K. Sun, “A correlation between aging and DNA repair in human epidermal cells,” *Mechanisms of Ageing and Development*, vol. 24, no. 3, pp. 283–292, 1984.
- [24] L. K. Heilbronn and E. Ravussin, “Calorie restriction and aging: review of the literature and implications for studies in humans,” *American Journal of Clinical Nutrition*, vol. 78, no. 3, pp. 361–369, 2003.
- [25] M. H. Goyns, “Genes, telomeres and mammalian ageing,” *Mechanisms of Ageing and Development*, vol. 123, no. 7, pp. 791–799, 2002.
- [26] D. Kappei and J. A. Londoño-Vallejo, “Telomere length inheritance and aging,” *Mechanisms of Ageing and Development*, vol. 129, no. 1-2, pp. 17–26, 2008.
- [27] L. Lavie, A. Z. Reznick, and D. Gershon, “Decreased protein and puromycinyl-peptide degradation in livers of senescent mice,” *Biochemical Journal*, vol. 202, no. 1, pp. 47–51, 1982.
- [28] M. D. Brand, C. Affourtit, T. C. Esteves et al., “Mitochondrial superoxide: production, biological effects, and activation of uncoupling proteins,” *Free Radical Biology and Medicine*, vol. 37, no. 6, pp. 755–767, 2004.
- [29] S. Makpol, A. Z. Abidin, K. Sairin, M. Mazlan, G. M. Top, and W. Z. W. Ngah, “ γ -tocotrienol prevents oxidative stress-induced telomere shortening in human fibroblasts derived from different aged individuals,” *Oxidative Medicine and Cellular Longevity*, vol. 3, no. 1, pp. 35–43, 2010.
- [30] M. De La Fuente and V. M. Victor, “Anti-oxidants as modulators of immune function,” *Immunology & Cell Biology*, vol. 78, no. 1, pp. 49–54, 2000.
- [31] L. Svendsen, S. I. S. Rattan, and B. F. C. Clark, “Testing garlic for possible anti-ageing effects on long-term growth characteristics, morphology and macromolecular synthesis of human fibroblasts in culture,” *Journal of Ethnopharmacology*, vol. 43, no. 2, pp. 125–133, 1994.
- [32] S. Salvioli, E. Sikora, E. L. Cooper, and C. Franceschi, “Curcumin in cell death processes: a challenge for CAM of age-related pathologies,” *Evidence-Based Complementary and Alternative Medicine*, vol. 4, no. 2, pp. 181–190, 2007.
- [33] R. A. Sharma, A. J. Gescher, and W. P. Steward, “Curcumin: the story so far,” *European Journal of Cancer*, vol. 41, no. 13, pp. 1955–1968, 2005.
- [34] M. Ichioka and H. Endo, “Effect of light on cellular carotenoids formation in *Chlorella regularis* S-50 grown on glucose,” *Annual Report of the Yakult Institute of Microbiology Research*, vol. 4, pp. 91–99, 1973.
- [35] M. L. S. Queiroz, C. O. Torello, S. M. C. Perhs et al., “*Chlorella vulgaris* up-modulation of myelossuppression induced by lead: the role of stromal cells,” *Food and Chemical Toxicology*, vol. 46, no. 9, pp. 3147–3154, 2008.
- [36] H. Endo, S. Fukuchi, and I. Nakano, “Effect of culture condition on tocopherol formation in *Chlorella Regularis*,” *Annual Report of the Yakult Institute of Microbiology Research*, vol. 5, pp. 91–98, 1974.
- [37] S. Fukuchi and H. Endo, “Effect of culture conditions on ubiquinone formation in *Chlorella Regularis*,” *Annual Report of the Yakult Institute of Microbiology Research*, vol. 6, pp. 1–8, 1975.
- [38] D. C. M. Dantas and M. L. S. Queiroz, “Effects of *Chlorella vulgaris* on bone marrow progenitor cells of mice infected with *Listeria monocytogenes*,” *International Journal of Immunopharmacology*, vol. 21, no. 8, pp. 499–508, 1999.
- [39] F. Konishi, K. Tanaka, S. Kumamoto et al., “Enhanced resistance against *Escherichia coli* infection by subcutaneous administration of the hot-water extract of *Chlorella vulgaris* in cyclophosphamide-treated mice,” *Cancer Immunology Immunotherapy*, vol. 32, no. 1, pp. 1–7, 1990.
- [40] Y. A. Mohd-Yusof, J. M. Hassan-Basari, N. A. Mukti et al., “Fatty acids composition of microalgae *Chlorella vulgaris* can be modulated by varying carbon dioxide concentration in outdoor culture,” *African Journal of Biotechnology*, vol. 10, no. 62, pp. 13536–13542, 2011.
- [41] Y. Y. Park, S. Lee, M. Karbowski, A. Neutzner, R. J. Youle, and H. Cho, “Loss of MARCH5 mitochondrial E3 ubiquitin ligase induces cellular senescence through dynamin-related protein 1 and mitofusin 1,” *Journal of Cell Science*, vol. 123, no. 4, pp. 619–626, 2010.
- [42] F. Debacq-Chainiaux, J. D. Erusalimsky, J. Campisi, and O. Toussaint, “Protocols to detect senescence-associated β -galactosidase (SA- β gal) activity, a biomarker of senescent cells in culture and in vivo,” *Nature Protocols*, vol. 4, no. 12, pp. 1798–1806, 2009.
- [43] D. J. Kurz, S. Decary, Y. Hong, and J. D. Erusalimsky, “Senescence-associated β -galactosidase reflects an increase in lysosomal mass during replicative ageing of human endothelial cells,” *Journal of Cell Science*, vol. 113, no. 20, pp. 3613–3622, 2000.

- [44] L. M. Gerland, S. Peyrol, C. Lallemand, R. Branche, J. P. Magaud, and M. Ffrench, "Association of increased autophagic inclusions labeled for β -galactosidase with fibroblastic aging," *Experimental Gerontology*, vol. 38, no. 8, pp. 887–895, 2003.