MOLECULAR AND CELLULAR BIOLOGY, Sept. 1990, p. 4935–4941 0270-7306/90/094935-07\$02.00/0
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## SPT6, an Essential Gene That Affects Transcription in Saccharomyces cerevisiae, Encodes a Nuclear Protein with an Extremely Acidic Amino Terminus

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Received 5 March 1990/Accepted 31 May 1990

SPT6 is an essential gene of Saccharomyces cerevisiae that appears to play a role in transcription. Mutations in the SPT6 (SSN20, CRE2) gene suppress  $\delta$  insertion mutations in the 5' regions of HIS4 and LYS2 and mutations in cis- and/or trans-acting elements that are required for expression of SUC2 and ADH2. We report here that SPT6 encodes a 170-kilodalton highly charged protein with an extremely acidic amino terminus. By use of an epitope-tagged SPT6 protein, we have determined by indirect immunofluorescence that the SPT6 protein is located in the nucleus.

Mutations in the Saccharomyces cerevisiae gene SPT6 (also called SSN20 [25, 26] and CRE2 [7, 8]) have been isolated as suppressors of both cis- and trans-acting mutations that alter transcription of particular genes. Mutations in SPT6 were first isolated as suppressors of an insertion mutation at the HIS4 gene (his4-9128) caused by a long terminal repeat of the transposable element Ty (32). Dominant and recessive mutations in SPT6 (called SSN20) were also isolated as extragenic suppressors of the defect in SUC2 expression caused by snf2 or snf5 mutations and were shown to act as suppressors of snf6 (26) and SUC2 mutant alleles with upstream activation sequence deletions (25). One spt6 mutation, called cre2-1, was isolated in a selection for mutations that allow ADH2 to escape glucose repression. The cre2-1 mutation also restores a moderate level of ADH2 expression in the absence of the trans-acting positive activator ADR1 (7, 8). In every case that has been examined, suppression by spt6 mutations is mediated at the level of transcription (6, 25).

The SPT6 gene was cloned by complementation of both an spt6 and an ssn20 allele, and a 4.6-kilobase (kb) mRNA was identified (6, 25). The gene was mapped to the right arm of chromosome VII (6, 25). Null alleles of SPT6 were constructed and found to be lethal in haploid cells (6, 25), consistent with the temperature-sensitive lethality conferred by many spt6 alleles (7, 26, 32). Experiments with the cloned SPT6 gene demonstrated that either increased or decreased dosage of the wild-type SPT6 gene also suppresses his4-9128 mutations (6) and SUC2 upstream regulatory region deletions (25). These results suggested that the SPT6 protein functions as part of a complex which is affected by altered stoichiometry (6, 25). Since SPT6 appears to play an essential and general role in transcription, we have characterized further the gene and its product.

Sequence of the SPT6 gene. The nucleotide sequence of the SPT6 gene was determined by the method of Sanger et al. (30). The open reading frame (Fig. 1) extended for 4353 base pairs, a size consistent with the 4.6-kb SPT6 RNA identified previously (6, 25). The nucleotide sequence predicted a

Acidic residues are characteristic of the activation domains of the yeast transcriptional activators GAL4 (21) and GCN4 (16, 17). However, the negatively charged region of SPT6 is more extensive and more acidic than those of previously characterized transcription activation domains. Acidic regions have also been found in several proteins that are thought to interact with chromatin (see reference 10 for review).

Data base searches (National Biomedical Research Foundation Protein Sequence Database release 21 and translated GenBank version 62) did not reveal proteins with significant sequence similarity to SPT6.

Identification of the SPT6 protein. To identify the SPT6 protein, we first prepared specific antibody. A trpE-SPT6 gene fusion, encoding SPT6 residues 474 to 860, was constructed by cloning a restriction fragment of SPT6 into the pATH11 vector (gift of T. J. Koerner, J. Hill, and A. Tzagoloff) to create plasmid pMS1. The TrpE-SPT6 hybrid protein was expressed in Escherichia coli HB101 (3), purifled by the method of Kleid et al. (19), and used to immunize rabbits. The SPT6-specific antibody was purified from the serum of one rabbit by chromatography on Protein A-Sepharose CL-4B (Pharmacia), followed by affinity chromatography with TrpE-SPT6 protein coupled to CNBr-activated Sepharose (Pharmacia) as the adsorbent according to instructions supplied by the manufacturer. The TrpE-SPT6 hybrid protein used to prepare the column was purified as described by Bhown and Bennett (1).

The SPT6 protein was identified by immunoblot analysis of total yeast proteins. The protein detected was approximately 170 kDa, the size predicted by the SPT6 DNA sequence. Moreover, the 170-kDa protein was more abun-

highly charged 1,451-amino-acid protein of 168,290 daltons (Da). In the entire protein, 19% of the residues were acidic and 15% were basic. A striking characteristic of the predicted protein was its acidic amino terminus: the first 70 residues had a net charge of -30 (50% acidic residues), and the amino-terminal third of the protein (484 residues) had a net charge of -81 (29% acidic residues). In addition, the SPT6 sequence included several consensus sites for phosphorylation by casein kinase II (20; reviewed in reference 2; G. Prelich and F. Winston, unpublished observations).

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AGCTGACTGATTCTTTATAGCAATAGGGAGGAAGGCAGTACACATATCAGCAGTGCATGGGCACAACCCGCATTACA

-321 ACGGTTAAAGCAAAAGGAGGAAGAAAGATATTGATTTGTGCTTTGAAGAATAACCCAAGCGAACAGGTCATTAATTGCCTAGAATCTTTACATAGAATTTGCCTTTT -107 1 ATG GAA GAG ACG GGA GAT TCG AAG CTG GTC CCT AGG GAC GAG GAA AAA GTA AAT GAC AAC GAT GAA ACT AAA GCG CCT Met <u>Glu Glu</u> Thr Gly <u>Asn</u> Ser Lys Leu Val Pro Arg <u>Asn Glu Glu Glu</u> Ile Val Asn <u>Asn Asn Glu</u> Thr Lys Ala Pro AGT GAG GAA GAA GAA GAA GAA GAT GTC TTT GAC TCC TCT GAG GAA GAC GAA GAT ATT GAT GAA GAC GAA GAT GAG GCA AGA Ser <u>Giu Giu Giu Giu Giy Giu Asp</u> Vai Phe <u>Asp</u> Ser Ser Gi<mark>u Giu Asp Giu Asp</mark> Ile Asp Giu Asp Giu Asp Giu Ala Arg 54 AAA GTG CAA GAA GGC TTC ATT GTC AAC GAT GAT GAT GAA AAT GAA GAC CCA GGA ACA AGT ATT TCC AAA AAA AGA AGA AAA Lys Vai Gin <u>Glu</u> Gly Phe Ile Vai Asn <u>Asp Asp Glu</u> Asn <u>Glu Asp</u> Pro Gly Thr Ser Ile Ser Lys Lys Arg Arg Lys 81 CAT AMA AGA AGA GAA AGA GAA GAA GAT GAT CGA CTA TCC GAA GAT GAT TTG GAT TTG TTA ATG GAG AAT GCT GGT GTT GAA His Lys Arg Arg <u>Glu Arg Glu Glu Asp Asp</u> Arg Leu Ser <u>Glu Asp Asp</u> Leu Asp Leu Leu Met <u>Glu</u> Asn Ala Gly Val <u>Glu</u> CGT ACC AAA GCG AGC TCC TCT TCA GGA AAA TTT AAA AGA TTA AAG AGA GTA GGA GAT GAA GGA AAC GCT GCA GAA TCA GAA ing Thr Lys Ala Sen Sen Sen Gly Lys Phe Lys Ang Leu Lys Ang Val Gly <u>Asp Glu</u> Gly Asn Ala Ala <u>Glu</u> Sen <u>Glu</u> 135 406 AGT GAC AAC GTC GCT GCA TCA AGG CAA GAT TCT ACG TCT AAA TTG GAA GAT TTC TTT TCT GAA GAT GAA GAA GAA GAA GAA Ser <u>Asp</u> Asn Val Ala Ala Ser Arg Gin <u>Asp</u> Ser Thr Ser Lys Leu <u>Giu Asp</u> Phe Phe Ser <u>Giu Asp Giu Giu Giu Giu Giu</u> Giu 162 TCT GGT TTA CGC AAC GGT AGG AAC AAT GAA TAT GGG CGT GAT GAA GAA GAC CAT GAA AAT AGG AAC AGA ACA GCT GAC AAA Ser Gly Leu Arg Asn Gly Arg Asn Asn <u>Glu</u> Tyr Gly Arg <u>Asp Glu Glu Asp</u> His <u>Glu</u> Asn Arg Asn Arg Thr Ala <u>Asp</u> Lys 189 GGT GGA ATC TTG GAC GAA TTG GAT GAT TTC ATT GAA GAT GAT GAA TTT TCT GAT GAG GAC GAA ACC AGA CAA AGG AGG Gly Gly Ile Leu Asp Glu Leu Asp Asp Phe Ile Glu Asp Asp Glu Phe Ser Asp Glu Asp Asp Glu Thr Arg Gln Arg Arg 216 ATC CAA GAA AAG AAG CTT TTA AGA GAA CAA TCC ATC AAA CAA CCT ACA CAG ATT ACT GGT CTA TCG TCG GAT AAG ATT GAC Ile Gin Giu Lys Lys Leu Leu Arg Giu Gin Ser Ile Lys Gin Pro Thr Gin Ile Thr Giy Leu Ser Ser Asp Lys Ile Asp GAG ATG TAT GAC ATT TTT GGT GAT GGT CAT GAC TAC GAT TGG GCT TTA GAA ATT GAA AAT GAA GAA CTA GAA AAT GGT AAC <u>Glu</u> Met Tyr <u>Asp</u> Ile Phe Gly <u>Asp</u> Gly His <u>Asp</u> Tyr <u>Asp</u> Trp Ala Leu <u>Glu</u> Ile <u>Glu</u> Asn <u>Glu Glu</u> Leu <u>Glu</u> Asn Gly Asn 270 GAC AAC AAT GAA GCT GAA GAA GAA GAA GAA GAA GAA GAA ACT GGT GCT ATA AAA AGC ACC AAG AAA AAG ATA TCT CTA CAA Aso Aso Aso Qiu Ala Qiu Qiu Qiu Qiu Ile Aso Qiu Qiu Thr Giy Ala Ile Lys Ser Thr Lys Lys Lys Ile Ser Leu Gin 297 892 GAC ATT TAT GAT TTA GAG GAT TTG AAA AAA AAC TTG ATG ACT GAA GGA GAC ATG AAA ATT AGA AAG ACA GAT ATT CCA GAA Asp Ile Tyr Asp Leu Giu Asp Leu Lys Lys Asn Leu Met Thr Giu Gly Asp Met Lys Ile Arg Lys Thr Asp Ile Pro Giu 324 AGA TAT CAA GAA TTA AGA GCA GGT ATT ACT GAC TAC GGA AAT ATG TCA TCG GAG GAT CAA GAA TTA GAA AGA AAC TGG ATA Arg Tyr Gin <u>Gin</u> Leu Arg Ala Gly Ile Thr <u>Asp</u> Tyr Gly Asn Met Ser Ser <u>Gin Asp</u> Gin <u>Gin</u> Leu <u>Gin</u> Arg Asn Trp Ile 351 1054 GCA GAA AAA ATT TCT GTG GAT AAG AAC TTC GAT GCC AAT TAT GAT CTC ACT GAA TTT AAA GAA GCA ATT GGG AAC GCA ATC Als <u>Giu</u> Lys Ile Ser Val <u>Asp</u> Lys Asn Phe <u>Asp</u> Als Asn Tyr <u>Asp</u> Leu Thr <u>Giu</u> Phe Lys <u>Giu</u> Als Ile Gly Asn Als Ile 1135 AMA TIT ATC ACC AMA GAM AMC TTG GAM GTC CCT TIT ATA TAT GCT TAC CGT CGT AMC TAT ATT TCC TCA AGA GAM AMA GAT Lys Phe Ile Thr Lys <u>Qiu</u> Asn Leu <u>Qiu</u> Val Pro Phe Ile Tyr Ala Tyr Arg Arg Asn Tyr Ile Ser Ser Arg <u>Qiu</u> Lys <u>Asp</u> 405 1216 GGG TIT CTT TTG ACT GAA GAT GAC CTT TGG GAT ATA GTT AGC CTT GAC ATC GAA TTT CAC AGT CTT GTG AAC AAA AAG GAT Gly Phe Leu Leu Thr Glu Asp Asp Leu Trp Asp Ile Val Ser Leu Asp Ile Glu Phe His Ser Leu Val Asn Lys Asp TAT GTC CAG AGA TTT TAT GCA GAA TTA CAT ATC GAT GAT CCT ATT GTC ACT GAA TAC TTC AAA AAT CAG AAT ACT GCA TCT Tyr Val Gin Arg Phe Tyr Ala <u>Giu</u> Leu His Ile <u>Asp Asp</u> Pro Ile Val Thr <u>Giu</u> Tyr Phe Lys Asn Gin Asn Thr Ala Ser 459 EcoRI ATT GCG GAA CTG AAT TCT CTA CAG GAT ATT TAT GAC TAC CTA GAA TTC AAA TAT GCC AAC GAA ATC AAT GAA ATG TIT ATA Ile Als Glu Leu Asn Ser Leu Gin Asp Ile Tyr Asp Tyr Leu Glu Phe Lys Tyr Als Asn Glu Ile Asn Glu Met Phe Ile AAC CAC ACT GGA AAG ACT GGT AAG AAA CAT TTG AAA AAT TCC AGT TAT GAA AAA TTT AAA GCT AGT CCT CTT TAT CAA GCG 513 Asn His Thr Gly Lys Thr Gly Lys Lys His Leu Lys Asn Ser Ser Tyr QLu Lys Phe Lys Ala Ser Pro Leu Tyr Gln Ala GTT AGT GAT ATT GGT ATA TCA GCT GAG GAT GTT GGT GAA AAT ATC AGT TCC CAG CAT CAA ATC CAC CCT CCC GTA GAT CAT Val Ser <u>Asp</u> Ile Gly Ile Ser Ala <u>Glu Asp</u> Val Gly <u>Glu</u> Asn Ile Ser Ser Gln His Gln Ile His Pro Pro Val <u>Asp</u> His 540 CCA AGT TCC AAA CCA GTA GAA GTG ATA GAA TCT ATA TTG AAT GCA AAC AGC GGT GAT TTG CAA GTC TTT ACG TCC AAT ACT 567 Pro Ser Ser Lys Pro Val Qiu Val Ile Qiu Ser Ile Leu Asn Ala Asn Ser Qiy Asn Leu Qin Val Phe Thr Ser Asn Thr ANG CTG GCA ATT GAT ACG GTC CAA AMA TAC TAC TCT TTG GAA TTG TCT AMA AAT ACA AMA ATA AGG GAA AMA GTT AGA TCC Lys Leu Ala Ile <u>Asp</u> Thr Val Gin Lys Tyr Tyr Ser Leu <u>Glu</u> Leu Ser Lys Asn Thr Lys Ile Arg <u>Glu</u> Lys Vel Arg Ser 594 1783 GAT TIT TCC AAA TAT TAT CTG GCT GAC GTT GTG TTA ACT GCT AAA GGT AAA GAA ATT CAA AAG GGA TCT CTG TAT GAG 621 Amp Phe Ser Lys Tyr Tyr Leu Ala Amp Val Val Leu Thr Ala Lys Gly Lys Lys Glu Ile Gln Lys Gly Ser Leu Tyr Glu GAC ATA AAA TAT GCC ATC AAT AGA ACT CCA ATG CAC TTC CGT AGG GAT CCA GAC GTT TTT TTG AAA ATG GTC GAG GCT GAG <u>Asp</u> Ile Lys Tyr Ala Ile Asn Arg Thr Pro Met His Phe Arg Arg <u>Asp</u> Pro <u>Asp</u> Val Phe Leu Lys Met Val <u>Glu</u> Ala <u>Glu</u> 648 1945 TCT TTG AAC CTG CTC AGT GTT AAG TTA CAC ATG TCG TCA CAA GCC CAA TAT ATA GAG CAT TTA TTC CAA ATT GCA CTT GAA Ser Leu Asn Leu Leu Ser Val Lys Leu His Met Ser Ser Gln Ala Gln Tyr Ile Glu His Leu Phe Gln Ile Ala Leu Glu 675 ACT ACC AAT ACC TCG GAC ATC GCA ATA GAA TGG AAT AAC TTC CGT AAA CTG GCA TTC AAC CAA GCG ATG GAC AAG ATT TTC 702 Thr Thr Asn Thr Ser Asp Ile Ala Ile <u>Qiu</u> Trp Asn Asn Phe Arg Lys Leu Ala Phe Asn Qin Ala Met <u>Asp</u> Lys Ile Phe 2107 CAG GAT ATA TCT CAA GAA GTC AAA GAC AAT TTA ACA AAA AAT TGT CAA AAA TTG GTA GCC AAG ACT GTT CGC CAT AAG TTT Gin <u>Asp</u> Ile Ser Gin <u>Giu</u> Val Lys <u>Asp</u> Asn Leu Thr Lys Asn Cys Gin Lys Leu Val Ala Lys Thr Val Arg His Lys Phe

2188 ATG ACA AMA TTA GAC CAG GCT CCA TTC ATT CCT AAT GTC AGG GAT CCA AMA ATT CCA AMA ATC TTA TCT TTA ACC TGT GGA Met Thr Lys Leu <u>Asp</u> Gin Ala Pro Phe Ile Pro Asn Val Arg <u>Asp</u> Pro Lys Ile Pro Lys Ile Leu Ser Leu Thr Cys Gly 2289 CAG GGT AGA TTC GGA GCC GAC GCT ATA ATT GCT GTC TAC GTC AAC AGA AAG GGT GAT TTT ATA AGA GAT TAC AAG ATT GTC Gin Giy Arg Phe Giy Ala Asp Ala Ile Ile Ala Val Tyr Val Asn Arg Lys Giy Asp Phe Ile Arg Asp Tyr Lys Ile Val 783 2350 GAC AAT CCA TIT GAT AAG ACG AAT CCT GAA AAA TIT GAA GAC ACC TIG GAT AAT ATC ATT CAA AGC TGT CAA CCG AAT GCC Asn Asn Pro Phe Asn Lys Thr Asn Pro Giu Lys Phe Giu Asp Thr Leu Asp Asn Ile Ile Gin Ser Cys Gin Pro Asn Ala 810 2431 ATC GGA ATC AAT GGC CCT AAC CCA AAG ACT CAA AAA TIT TAC AAA AGA TTA CAA GAA GTT CTA CAT AAG AAG CAA ATC GTC Ile Gly Ile Asn Gly Pro Asn Pro Lys Thr Gln Lys Phe Tyr Lys Arg Leu Gln Glu Val Leu His Lys Lys Gln Ile Val 837 EcoRI 2512 GAC AGT AGA GGA CAT ACT ATT CCA ATC ATT TAC GTT GAG GAC GAA GTC GCT ATC CGT TAT CAG AAT TCC GAA AGA GCT GCT Asp Ser Arg Gly His Thr Ile Pro Ile Ile Tyr Val Glu Asp Glu Val Ala Ile Arg Tyr Gln Asn Ser Glu Arg Ala Ala 864 CAA GAA TTC CCT AAT AAA CCT CCT CTA GTT AAA TAC TGT ATC GCC TTG GCG CGC TAT ATG CAT TCC CCA TTG TTG GAA TAT Gin <u>Giu</u> Phe Pro Asn Lys Pro Pro Leu Val Lys Tyr Cys Ile Ala Leu Ala Arg Tyr Met His Ser Pro Leu Leu <u>Giu</u> Tyr 891 GCT AAT TTA ACA AGT GAA GAA GTG AGA TCA TTG TCA ATT CAT CCA CAC CAA AAT CTG TTA TCC TCA GAA CAA TTG AGT TGG Ala Asn Leu Thr Ser <u>Glu Glu</u> Val Arg Ser Leu Ser Ile His Pro His Gln Asn Leu Leu Ser Ser <u>Glu</u> Gln Leu Ser Tro 918 2755 GCT CTT GAA ACC GCT TTC GTT GAT ATT GTC AAC CTG GTA AGT GTT GAA GTT AAC AAA GCC ACA GAT AAT AAT TAC TAC GCT Ala Leu Giu Thr Ala Phe Val Asn Ile Val Asn Leu Val Ser Val Giu Val Asn Lys Ala Thr Asn Asn Asn Tyr Tyr Ala 2836 AGT GCG CTG AMA TAC ATC TCT GGC TTT GGA AMA CGT AMA GCT ATT GAT TTC TTA CAG TCC CTT CAA AGG CTA AMT GAA CCA Ser Ala Leu Lys Tyr Ile Ser Gly Phe Gly Lys Arg Lys Ala Ile <u>Asp</u> Phe Leu Gln Ser Leu Gln Arg Leu Asn <u>Glu</u> Pro 972 2917 TTA CTG GCT CGT CAA CAA TTA ATT ACT CAT AAC ATT CTT CAC AAG ACT ATT TTT ATG AAT TCC GCG GGA TTC CTC TAT ATC Leu Leu Ala Arg Gin Gin Leu Ile Thr His Asn Ile Leu His Lys Thr Ile Phe Met Asn Ser Ala Gly Phe Leu Tyr Ile 999 TCA TGG AAT GAA AAA AGA CAA AAA TAC GAA GAT TTG GAA CAT GAT CAA CTA GAT AGC ACT AGA ATT CAT CCA GAA GAC TAC Ser Trp Asn Glu Lys Arg Gln Lys Tyr Glu Asp Leu Glu His Asp Gln Leu Asp Ser Thr Arg Ile His Pro Glu Asp Tyr 1026 CAT TTG GCC ACC AAG GTT GCC GCT GAT GCT TTA GAA TAC GAT CCT GAT ACT ATT GCC GAA AAA GAA GAA CAG GGG ACT ATG His Leu Ala Thr Lys Val Ala Ala <u>Asp</u> Ala Leu <u>Glu</u> Tyr <u>Asp</u> Pro <u>Asp</u> Thr Ile Ala <u>Glu</u> Lys <u>Glu Glu</u> Gln Gly Thr Met 1053 AGT GAA TTC ATT GAA CTG TTG AGA GAA GAT CCT GAC CGT AGA GCT AAA CTA GAA TCA CTA AAT CTA GAA TCA TAC GCA GAA 1080 Ser <u>Giu</u> Phe Ile <u>Giu</u> Leu Leu Arg <u>Giu Asn</u> Pro <u>Asn</u> Arg Arg Ala Lys Leu <u>Giu</u> Ser Leu Asn Leu <u>Giu</u> Ser Tyr Ala <u>Giu</u> 3241 GAA CTT GAG AAG AAT ACC GGA TTA AGA AAA CTT AAT ATA CTA AAT ACA ATT GTC CTT GAA TTG TTG GAT GGA TTT GAA GAA <u>Giu</u> Leu <u>Giu</u> Lys Asn Thr Giy Leu Arg Lys Leu Asn Asn Leu Asn Thr Ile Val Leu <u>Giu</u> Leu <u>Asp</u> Giy Phe <u>Giu Giu</u> 3322 TTG AGA AAT GAC TTT CAT CCT TTG CAA GGT GAT GAA ATT TTC CAA AGT TTG ACT GGT GAG TCT GAA AAG ACG TTT TTC AAG Leu Arg Asn <u>Asp</u> Phe His Pro Leu Gin Gly <u>Asp Giu</u> Ile Phe Gin Ser Leu Thr Gly <u>Giu</u> Ser <u>Giu</u> Lys Thr Phe Phe Lys 1134 3403 GGT AGT ATT ATT CCA GTC AGA GTA GAA AGA TTC TGG CAC AAC GAT ATA ATT TGC ACT ACA AAC TCT GAA GTT GAA TGT GTA 1161 Gly Ser Ile Ile Pro Val Arg Val Glu Arg Phe Trp His Asn Asn Ile Ile Cys Thr Thr Asn Ser Glu Val Glu Cys Val GTA AAT GCT CAA CGT CAC GCA GGT GCA CAA TTA AGA AGA CCT GCA AAT GAA ATA TAC GAA ATT GGT AAA ACA TAT CCA GCA Val Asn Ala Gin Arg His Ala Giy Ala Gin Leu Arg Arg Pro Ala Asn <u>Giu</u> Ile Tyr <u>Giu</u> Ile Giy Lys Thr Tyr Pro Ala 1188 AAG GTG ATA TAT ATT GAC TAT GCT AAT ATT ACT GCA GAA GTT TCC TTA TTA GAT CAT GTC AAA CAG CAA TAT GTT CCA Lys Val Ile Tyr Ile Asp Tyr Ala Asn Ile Thr Ala Glu Val Ser Leu Leu Asp His Asp Val Lys Gin Gin Tyr Val Pro 1215 ATA AGC TAC AGT AAA GAT CCT TCC ATT TGG GAC TTG AAA CAA GAA CTG GAA GAT GCC GAA GAG GAG AGG AAA TTG ATG ATG Ile Ser Tyr Ser Lys Asp Pro Ser Ile Trp Asp Leu Lys Gin Giu Leu Giu Asp Ale Giu Giu Giu Arg Lys Leu Met Met 1242 GCA GAA GCC CGT GCA AAG AGA ACA CAT CGT GTT ATC AAT CAT CCT TAC TAT TTC CCT TTC AAC GGC AGA CAG GCT GAG GAT 1269 Ala Glu Ala Arg Ala Lys Arg Thr His Arg Val Ile Asn His Pro Tyr Tyr Phe Pro Phe Asn Gly Arg Gln Ala Glu Aso. TAC TTA AGG AGT AAA GAA CGT GGT GAA TTC GTG ATC AGA CAG TCT AGC CGA GGT GAT GAC CAC TTG GTT ATC ACC TGG AAA Tyr Leu Arg Ser Lys Giu Arg Gly Giu Phe Val Ile Arg Gin Ser Ser Arg Gly Asp Asp His Leu Val Ile Thr Trp Lys 1296 TTG GAT ANG GAT TTG TTT CAA CAT ATT GAT ATC CAA GAA TTA GAA AAA GAA AAT CCT TTG GCT TTA GGT AAA GTC TTG ATT 3889 Leu Asp Lys Asp Leu Phe Gin His Ile Asp Ile Gin Giu Leu Giu Lys Giu Asn Pro Leu Als Leu Giy Lys Val Leu Ile 1323 GTC GAC AAT CAG AAA TAC AAT GAT TTA GAC CAG ATC ATT GTA GAA TAT CTT CAA AAC AAG GTA AGG CTC TTG AAT GAA ATG 1350 Val <u>Asp</u> Asn Gin Lys Tyr Asn <u>Asp</u> Leu <u>Asp</u> Gin Ile Ile Val <u>Giu</u> Tyr Leu Gin Asn Lys Val Arg Leu Leu Asn <u>Giu</u> Met 4051 ACA TCT AGT GAA AAA TTC AAA AGC GGT ACT AAG AAA GAT GTG GTC AAG TTT ATT GAA GAC TAC TCT AGA GTG AAT CCA AAT 1377 Thr Ser Ser <u>Giu</u> Lys Phe Lys Ser Gly Thr Lys Lys <u>Asp</u> Val Val Lys Phe Ile <u>Giu Asp</u> Tyr Ser Arg Val Asn Pro Asn 4132 AAG TCT GTG TAC TAT TTC AGT TTG AAC CAC GAT AAC CCT GGT TGG TTT TAC TTG ATG TTC AAG ATT AAC GCA AAT AGC AAA 1404 Lys Ser Val Tyr Tyr Phe Ser Leu Asn His Asn Asn Pro Gly Trp Phe Tyr Leu Met Phe Lys Ile Asn Ala Asn Ser Lys 4213 TTA TAC ACA TGG MAT GTG AMA TTA ACG AMC ACT GGT TAT TTC CTG GTA AMC TAC AMT TAT CCA AGT GTT ATC CAG CTT TGT Leu Tyr Thr Trp Asn Val Lys Leu Thr Asn Thr Gly Tyr Phe Leu Val Asn Tyr Asn Tyr Pro Ser Val Ile Gin Leu Cys 1431 AAT GGT TIT AAG ACG CIT CTA AAA TCT AAC AGT AGT AAG AAT AGA ATG AAC TAC CGT TAG ATGCGTATGTAGTGTCCATTGTT 1451 Asn Gly Phe Lys Thr Leu Leu Lys Ser Asn Ser Ser Lys Asn Arg Met Asn Asn Tyr Arg \*\*\* 4487 AAGTTTGTACCGTGACGAACATAGCACGAACACAACATTTAATAGAAATAGCTTGGGAACCAAAATTTTAGAAATTTTTTCACTAATGAGAAGGAAAGCGTTGCAA

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FIG. 1. Nucleotide sequence of the SPT6 gene (GenBank accession number M34391) and predicted amino acid sequence of its gene product. Nucleotides are numbered on the left. Amino acids are numbered on the right. Asterisks indicate the termination codon. Acidic residues are underlined. The EcoRI restriction sites used to construct the trpE-SPT6 gene fusion used for antibody preparation are indicated. Additional EcoRI sites are located at positions 1389, 2596, 2973, 3059, 3163, and 3832. The HindIII site used to construct a null mutation (6) is at position 661. The complete sequence was determined on both strands with the M13mp18 and M13mp19 vectors (27). In four cases the restriction fragments were digested with Bal31 nuclease (IBI) prior to cloning, and in six cases 20-base oligomers (Biopolymers Laboratory, Department of Genetics, Harvard Medical School) complementary to the adjacent SPT6 sequence were used as primers.

dant in yeast cells carrying the SPT6 gene on a multicopy plasmid (Fig. 2, compare lanes A and B).

Nuclear localization of the SPT6 protein. The distribution of the SPT6 protein in yeast cells was determined by indirect immunofluorescence microscopy. Initially, we attempted to localize the SPT6 protein by using the affinity-purified SPT6-specific antibody. This antibody stained cells faintly throughout the cell (data not shown). However, because an *spt6* null mutation is lethal (6, 25), a yeast strain lacking SPT6 protein was not available as a control to determine whether the staining was due to nonspecific binding of the antibody.

To circumvent this problem, we used an epitope addition method (12, 23). A hybrid gene encoding an antigenic epitope from the hemagglutinin of influenza virus (HA1 [12, 27]) fused to the SPT6 protein was constructed by using the Muta-Gene M13 in vitro mutagenesis kit (Bio-Rad Laboratories). The template for the mutagenesis was pMS26, which contains the 1.8-kb *Hin*dIII fragment from pCC11 (6) cloned into the *Hin*dIII site of M13mp19 (28). The primer used for the mutagenesis was a 69-base oligonucleotide (Biopolymers Laboratory, Department of Genetics, Harvard Medical School) including the 17 nucleotides 5' to the SPT6 initiation

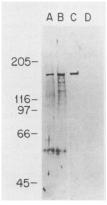


FIG. 2. Immunoblot analysis of the SPT6 protein. Total protein (40 μg in lanes A and B and 25 μg in lanes C and D) was prepared as described previously (4), separated by electrophoresis in 7.5% polyacrylamide, and electroblotted to nitrocellulose. Protein samples were prepared from (lane A) wild-type yeast strain MCRY1093, (lane B) an spt6 mutant strain carrying SPT6 on a multicopy plasmid (MCRY237), (lane C) wild-type yeast cells carrying the HA1-SPT6 hybrid gene on a centromere-containing plasmid (MS147), and (lane D) wild-type yeast cells carrying the SPT6 gene on a centromerecontaining plasmid (MS146). SPT6-specific antibody affinity-purified from rabbit and diluted 1:20 was used with the Promega Protoblot system at a 1:7,500 dilution of rabbit immunoglobulin G (IgG)specific antibody affinity-purified from goat serum and conjugated to alkaline phosphatase (lanes A and B). HA1-specific antibody was diluted 1:100 and used with a 1:7,500 dilution of mouse IgG-specific antibody affinity-purified from goat serum and conjugated to alkaline phosphatase (lanes C and D). The minor bands in lane B were not found consistently and are presumably degradation products. Numbers at right mark positions of protein standards (in kilodaltons).

codon, the initiation codon, the 27 nucleotides encoding the HA1 epitope, and then the 22 nucleotides 3' to the SPT6 initiation codon. The mutated 1.8-kb HindIII fragment was used to create pMS33, which encodes the HA1 epitope fused to the complete SPT6 protein on a centromere-containing plasmid. The hybrid protein was functional, as judged by its ability to complement the temperature-sensitive lethality and Spt<sup>-</sup> phenotype caused by an spt6-140 mutation (32) (data not shown).

To study the HA1-SPT6 hybrid protein, we used a specific monoclonal antibody against the HA1 epitope, called 12CA5, that was prepared and characterized previously (27). The specificity of the anti-HA1 antibody for the HA1-SPT6 hybrid protein in yeast cells was determined first by immunoblot analysis. The epitope-tagged SPT6 protein detected by the HA1-specific antibody (Fig. 2, lane C) comigrated with the wild-type 170-kDa SPT6 protein detected by SPT6-specific antibodies (Fig. 2, lanes A and B). No staining by anti-HA1 antibody was observed in a strain lacking the HA1 epitope (Fig. 2, lane D).

To determine the cellular location of the SPT6 protein, tetraploid cells (Table 1), whose large size facilitated microscopic analysis, were prepared for immunofluorescence microscopy. Experiments with the HA1-specific antibody and a strain containing the HA1-SPT6 fusion protein demonstrated that the epitope-tagged SPT6 protein was located in the nucleus of cells, as evident from the coincident fluorescein and DAPI (4',6'-diamidino-2-phenylindole dihydrochloride) staining (Fig. 3A to F). In the control experiment, no staining was detected in cells lacking the HA1 epitope (Fig. 3G to I). Since the HA1-SPT6 fusion protein provided SPT6 function, it is very likely that the wild-type SPT6 protein is also located in the nucleus.

To address the possibility that the HA1 epitope itself directs proteins to the nucleus, we determined the cellular location of an HA1- $\beta$ -galactosidase fusion protein. Previous

TABLE 1. Yeast strains

Strain	Genotype <sup>a</sup>
MCRY1093	MATa his4-539 lys2-801 ura3-52
MCRY237	MATa his4-539 ura3-52 ssn20-1 [pCE204]
MS146	MATa his4-912δ lys2-128δ ura3-52 trp1Δ63 [pCC11]
MS147	MATa his4-912δ lys2-128δ ura3-52 trp1Δ63 [pMS33]
MS149	MATa/MATa ura3-52/ura3-52 his4-9128/his4-9128
	$lys2-128\delta/lys2-128\delta leu2\Delta1/+ trp1\Delta63/+ [pMS33]$
MS150	MATα/MATα ura3-52/ura3-52 his4-9128/his4-9128
	$lys2-128\delta/lys2-128\delta leu2\Delta 1/+ trp1\Delta 63/+ spt6-140/+$
MS148	$MS149 \times MS150$
BM330	MATα/MATα ura3-52/ura3-52 his4-9128/his4-9128
	$lys2-128\delta/lys2-128\delta$ $trp1\Delta63/trp1\Delta63$
BM331	MATa/MATa ura3-52/ura3-52 his4-9128/his4-9128
	$lys2-128\delta/lys2-128\delta$ $trp1\Delta63/trp1\Delta63$
BM339	BM330 × BM331
MS157	BM339 [pMS35]

<sup>&</sup>lt;sup>a</sup> Plasmids are indicated in brackets. Diploid strains homozygous at the MAT locus were generated by UV-irradiating diploid cells (300 ergs/mm<sup>2</sup>) and then screening for diploids that could mate.

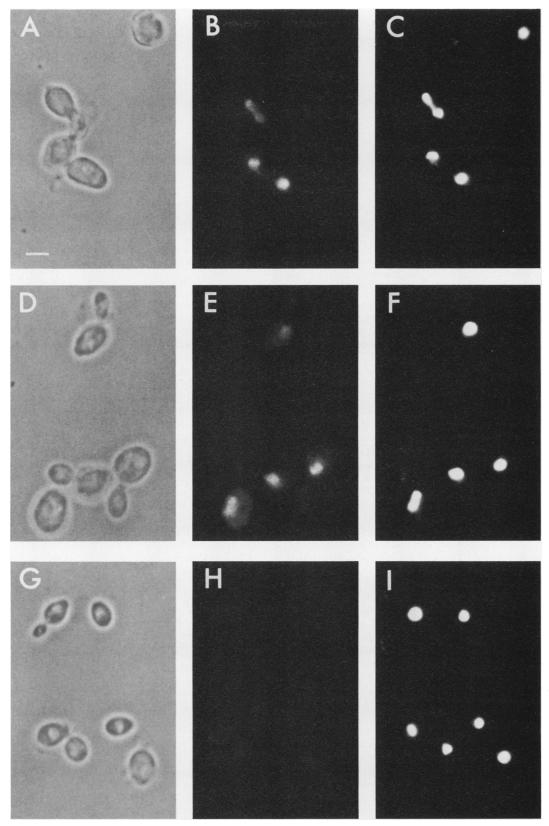
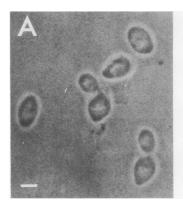
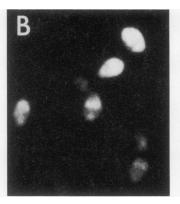


FIG. 3. Nuclear localization of SPT6 protein by indirect immunofluorescence. A tetraploid yeast strain containing the HA1-SPT6 fusion protein (MS148) (A to F) and a congenic control strain lacking the HA1 epitope (BM339) (G to I) were prepared essentially as described by Kilmartin and Adams (18), except that cell cultures were fixed in 4% paraformaldehyde (Fisher). The cells were stained with HA1-specific antibody diluted 1:400 and affinity-purified mouse IgG-specific F(ab')₂ fragment conjugated to fluorescein diluted 1:2,000. Micrographs shown are phase contrast (A, D, and G), fluorescein fluorescence (B, E, and H), and DAPI fluorescence, which indicates DNA (C, F, and I). Cells were examined with a Zeiss Photomicroscope III equipped for epi-illumination fluorescence with a Zeiss Neofluar 63× lens, N.A. 1.25. Cells were photographed with Kodak TMAX-400 film. Exposure times of 40 s were used for the immunofluorescence micrographs of the experimental and control samples. Bar, 5 μm.

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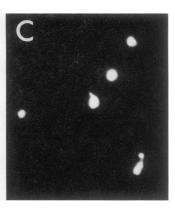


FIG. 4. Localization of HA1- $\beta$ -galactosidase by indirect immunofluorescence. Tetraploid yeast cells containing the HA1- $\beta$ -galactosidase fusion protein (MS157) were stained with HA1-specific antibody as described in the legend to Fig. 3. An exposure time of 25 s was used for the immunofluorescence micrograph. Bar, 5  $\mu$ m.

work has shown that β-galactosidase expressed in yeast cells is distributed throughout the cytoplasm and nucleus (13). We first constructed an HA1-lacZ hybrid gene under control of the ADH1 promoter. This construct was made by cloning the 1.5-kb BamHI-SalI fragment from pAD5, which contains the ADH1 promoter and the HA1 epitope-coding sequences (gift of J. Field, L. Rogers, and M. Wigler), into YEp353, which contains the coding region of E. coli lacZ (24), to create pMS35, which was then used to transform yeast cells. Immunofluorescence experiments demonstrated that the HA1-β-galactosidase fusion protein was located throughout yeast cells (Fig. 4). In the control experiment, no staining was detected in a strain that did not contain the epitopetagged β-galactosidase fusion protein (data not shown). Our observation that the HA1 epitope did not direct \(\beta\)-galactosidase to the nucleus is consistent with the fact that the HA1 amino acid sequence (YPYDVPDYA) does not resemble known nuclear signal sequences (9) and with the observation that addition of the epitope to adenylate cyclase, a plasma membrane-associated protein, did not alter the function of this enzyme in vivo (12).

The amino terminus of the predicted SPT6 protein included three regions that may serve as nuclear localization signals (see reference 9 for review): KLVPR (residues 8 to 12), KKRRKHKRR (residues 77 to 85), and KRLKRV (residues 120 to 125). The actual role of the these regions will have to be determined experimentally, since a variety of primary sequences have been shown to function as nuclear localization signals (9).

Nuclear localization of the SPT6 protein is consistent with the spt6 mutant phenotypes characterized previously. Genetic analysis has demonstrated that SPT6 affects transcription of a variety of genes and suggested that the SPT6 protein may act negatively at a downstream promoter element (8, 25). SPT6 protein may interact directly with DNA via a previously unidentified DNA-binding motif. Alternatively, SPT6 protein may play a role in chromatin assembly or modification. Many studies have correlated alterations in chromatin structure with transcriptional activation (for reviews, see references 22 and 29). Like SPT6, proteins believed to interact with chromatin have extremely acidic regions (10). Furthermore, the SPT6 gene dosage effects are similar to those caused by histone genes. Either increased or decreased dosage of some histone genes also suppresses δ insertion mutations (5) and deletions of the SUC2 upstream regulatory region (J. Hirschhorn and F. Winston, unpublished observations). Han and Grunstein (14) have shown that, like *spt6* mutations (25), nucleosome depletion restores mRNA expression from alleles with upstream activation sequence deletions.

One approach we are taking to determine the function of SPT6 is to identify genes encoding proteins that are functionally related to SPT6. Since SPT6 is an essential gene, it is conceivable that mutations in functionally related genes would cause lethality when combined with temperaturesensitive spt6 mutations. Two candidate genes are SPT4 and SPT5, which were also identified in genetic selections for extragenic suppressors of Ty insertion mutations (11, 32). The combination of any two of the missense mutations spt4-3, spt5-194, and spt6-140 causes lethality (32). Moreover, some combinations of spt4, spt5, and spt6 alleles exhibit unlinked noncomplementation (M. S. Swanson and F. Winston, unpublished observations), a phenotype observed for mutations in some proteins that are known to interact (15, 31). By genetic and molecular analysis of genes that appear to be functionally related to SPT6, such as SPT4, SPT5, and some histone genes, we hope to elucidate the role of SPT6 in gene expression in yeast.

We thank John Celenza for advice and assistance in sequencing the SPT6 gene and preparation of the TrpE-SPT6 antigen and antiserum and Laura Davis and Linda Marshall-Carlson for helpful suggestions regarding immunofluorescence methods.

This work was supported by Public Health Service grants GM34095 to M.C. and GM32967 to F.W. from the National Institutes of Health (NIH). At Columbia University, M.S. was supported by NIH training grant T32-GM07088 to the Department of Genetics and Development. At Harvard Medical School, M.S. was supported by NIH training grants to the Cellular and Developmental Biology Program (T32-GM07226) and the Genetics Program (T32-GM07196) and by the Markey Charitable Trust.

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