Published in final edited form as:
J Am Chem Soc. 2013 March 27; 135(12): 4584–4587. doi:10.1021/ja310929s.

Ligand conjugation of chemically exfoliated MoS_2

Stanley S. Chou[+], **Mrinmoy De**[+], **Jaemyung Kim**[+], **Segi Byun**[+,||], **Conner Dykstra**[+], **Jin Yu**[||], **Jiaxing Huang**[+,*], and **Vinayak P. Dravid**[+,*]

[+]Department of Material Science & Engineering, International Institute for Nanotechnology, Northwestern University, Evanston, Illinois 60208, United States

[||]Department of Material Science and Engineering, Korea Advanced Institute Science and Technology, Yuseonggu, Daejeon, Republic of Korea, 305-701

Abstract

MoS_2 is a two-dimensional material that is gaining prominence due to its unique electronic and chemical properties. Here, we demonstrate ligand conjugation of chemically exfoliated MoS_2 using thiol chemistry. Using this method, we modulate the zeta-potential and colloidal stability of MoS_2 sheets through ligand designs, thus enabling its usage as a selective artificial protein receptor for β-galactosidase. The facile thiol functionalization route opens the door for surface modifications of solution processable MoS_2 sheets.

The recent excitement over two dimensional atomically-layered materials has been fueled in large part by desires to exploit their low-dimensional properties, which can be distinct from their bulk counter-parts. One of these emerging materials, MoS_2, has received a fair share of attention due to its applicability in areas ranging from catalysis[1] and electronics[2–5] to biomedicine.[6] Nevertheless, to fully harness the capabilities of such new materials, ligand conjugations were often necessity. For example, it is typically a crucial step for solution processing, which enables the usage of nanoparticles as functional assembly blocks in bio-imaging,[7] electronics,[8] sensing,[9] and photovoltaics.[10] Previously, attempts to functionalize MoS_2 have been limited to bulk-like materials characterized by hydrophobic cleavage planes.[11,12] Specifically, Tremel and coworkers[13] have modified hydrophobic MoS_2 particles using nickel-nitrilotriacetic acid chelation.[13] However, colloidal surface modification of water dispersible, chemically exfoliated MoS_2 (ce-MoS_2) sheets has not been demonstrated.

To obtain large quantities of single-layer MoS_2 sheets, solution based exfoliation methods are often used. Most typically, this involves chemical exfoliation using lithium intercalation, which can produce single layer MoS_2 sheets in scalable quantities.[14–16] In this method, lithium is inserted between MoS_2 layers and then reacted with water to produce hydrogen gas at the interface. This, in combination with ultrasoncation, then enables the high yield production of water dispersible ce-MoS_2 sheets.[14] Due to the violent nature of this reaction however, MoS_2's crystal structure becomes deformed,[3,17] and internal edges (tears, pinholes and defects) can become visible.[18] Previous work has suggested these edge sites to possess higher molecular affinities,[19] with theoretical suggestions of thiol edge absorption.[20] It is thus possible that ce-MoS_2, which pos- sesses defects in both internal edges and perimeter

---

Corresponding Authors: v-dravid@northwestern.edu, jiaxing-huang@northwestern.edu.

Author Contributions

All authors have approved the final version of the manuscript.

Synthetic protocols, experimental procedures, and additional characterizations are included in the supporting information. This material is available free of charge at http://pubs.acs.org.

edges, may be amenable to thiol ligand modifications. As this has not been explicitly demonstrated, we show conjugation of ce-MoS_2 sheets by thiol-terminated ligands (Figure 1, Supplemental Information S1-5). This was used to conveniently tune ce-MoS_2 and its biomolecular interactions. This route opens pathways for making solution processable MoS_2 with tunable colloidal properties and can potentially facilitate improvements in its processing.

In this report, the as-made ce-$MoS_2$was first purified using exhaustive dialysis (7 days under continuous water flow). Survey of the material using X-Ray Diffraction (XRD) showed primary peak at $2\theta = 14$, thus indicating absence of mixed lithium phase (Figure 2a).[3,21] This was confirmed using Inductive Coupled Plasma-Mass Spectrometry (ICP-MS), which showed molecular ratios of samples obtained through dialysis to contain more than ten-fold less Li than the typical centrifugation purification process (vide infra). Atomic Force Microscopy (AFM) revealed sheets with average diameters of 0.7μm, and average thicknesses between ~0.8 and 1.5 nm, consistent with reported values of monolayers.[22] Additionally, the internal edges mentioned previously become are visible (Figure 2b, S6).[18] Transmission Electron Microscopy (TEM) Selective Area Diffraction (SAD) showed six-fold symmetrical diffraction spots (2H phase) with additional points in between those diffraction points (1T phase, Figure S7). This is consistent with previous reports by Dungey, et al,[17] for ce-MoS_2, thus indicating a mixed 1T and 2H material. Separately, Dynamic Light Scattering (DLS) show native ce-MoS_2 sheets to have a ζ-potential of −50 mV (Figure 3a). To this end, there are different models in literature regarding the origin of this negative charge. Principally, Divigalpitiya, et al,[19] proposed a model with charges at edges. Separately Heising, et al,[23] has a model based on partially oxidized Mo atoms on the basal plane due to Li intercalation. However, both agree that the sheets are defective, thus there is the possibility of chemical modification.

To illustrate this process, we first sought to understand the ligand affinity of chemically exfoliated MoS_2. For this, we synthesized a series of polyethylene glycol (PEG) ligands with head-groups of different charge (Figure 1, S1–S5). By including and excising the thiol moiety, we show that the thiol group is responsible for the ce-MoS_2 modification (**ligand 1a vs. 1b**).

We first examined the conjugation results using **ligand 1a** and **ligand 1b**. As can be seen in Figure 1, these ligands are nearly identical except for the thiol group on **ligand 1a**. Despite their similarities however, only the ce-MoS2 samples incubated with **ligand 1a** showed significant ζ-potential shifts after purification. The resultant ζ-potential of −7 mV represents a 43 mV shift from native ce-MoS_2, and is consistent with a neutral PEG functionalized colloid.[19] Comparatively, **ligand 1b** produced no ζ-potential change, indicating no modification (Figure 3a). Due to the attenuated ζ-potential, **ligand 1a** conjugate was found to be less stable in water.

To expand upon the conjugations, **ligand 2** and **ligand 3** were synthesized. Here, the conjugates also show appropriate changes in ζ-potential as well, with **ligand 2** producing a ζ-potential of −29 mV and **ligand 3** a ζ-potential of +36 mV. Overall, **ligand 3** conjugates exhibited the greatest colloidal stability. We believe this to be attributed to the pH independent nature of the charged NMe_3^+group on **ligand 3**.

Further characterizations were then performed. First, Fourier Transform Infrared spectroscopy (FT-IR, Figure 3b, 3c, S9) revealed the exposed S-H band from free ligands at 2563 cm^{-1}. This peak becomes absent after conjugation with ce-MoS_2, giving indication that the thiol moiety becomes buried on to the ce-MoS_2 surface (Figure 3b).[24,25] Presence of the ligands on the conjugates were also validated by monitoring the C–H aliphatic bands

2854 and 2930 cm^{-1}. Here, these aliphatic bands appear on the conjugates at 2864 and 2935 cm^{-1} (Figure 3c).[26] Unique pegylated ligand signature on conjugated ce-MoS$_2$ surfaces were also verified using X-Ray Photoelectron Spectroscopy (XPS). For example, ether (C-O-C) bonds characteristic of PEG ligands appeared in all conjugated samples (286.6 eV, Figure S8). These results further corroborate presence of PEG ligands on the ce-MoS$_2$, and are consistent with thiol-PEG functionalized materials.[27–29]

With the conjugation assessed, we then evaluate the effect of residual lithium (Li) from the exfoliation process. Here, ce-MoS$_2$ purified using the 3X centrifugation process described by Heising[23] produced a Li:Mo ratio of 0.25, consistent with their report. ce-MoS$_2$ purified using the dialysis process described here reduced the Li:Mo ratio to 0.02, more than tenfold lower (Figure S10). However, the residual Li doesn't appear to effect conjugation results radically (Figure S11)

We also examined the effect of ligand conjugation on the hydrogen evolution reaction (HER) process. As previous studies suggest edges to dominate HER catalysis,[30] we expect that ligand conjugation to suppress HER activity. Indeed, decrease catalytic activity was clearly observed (Figure 3c, S12). It thus appears that the thiol ligands can at minimum functionalize the various edges in ce- MoS$_2$.

We then monitored the changes in colloidal stability with ligand conjugation. Most observably, native ce-MoS$_2$ irreversibly precipitated due to restacking within the first 21 days (Figure 5a).[14] Comparably, the conjugates maintained colloidal stability (**ligand 3**), or can be readily re-dispersed with shaking (**ligand 1a, 2**) (Figure 4a). This is likely due PEG ligand preventing complete restacking. Due to the pH independent nature of the NMe_3^+group on **ligand 3**, its conjugates are also capable of resisting flocculation in acidic media (Figure 4b). To compare, nativece-MoS$_2$ flocculated at low pH.[14] **Ligand 3** conjugates, on the other hand were able to maintain colloidal stability (Figure 4b).

The combination of ζ-potential tailoring and pegylation also enable the tuning of ce-MoS$_2$ for selective host-guest interactions with biomolecules. As we have recently shown ce-MoS$_2$ to be a promising photothermal agent,[31] ligand modifications shown here can have immediate applications. To demonstrate this, we show the ligands ability to change ce-MoS$_2$'s function as an artificial protein receptor. To this end, we use β-galactosidase (β-gal), a hydrolase enzyme. Here, β-gal, due to a ring of anionic residues (Asp and Glu) surrounding its active site does not interact with native ce-MoS$_2$ (Figure 5a).[32,33] By tuning ce-MoS$_2$ for the correct electrostatic host-guest interactions we thus enable selective complexation and inhibition of this enzyme.[34] For this, 62.5 pM of β-gal were incubated with various concentrations of ce-MoS$_2$-conjugates. After 30 minutes, *o*-nitrophenyl-β-D-galactopyranoside (ONPG), a chromogenic substrate was added to monitor β-gal activity. From this experiment, it can be seen that only the **ligand 3** conjugate, which is cationic, showed ability to modulate β-gal activity. Specifically, 50% of β-gal activity can be inhibited by 0.04 μg/ml of the conjugate (Figure 5b). These results demonstrate the possibility to tune ce-MoS$_2$ for selective host-guest interactions with enzymes. Further development of this demonstrated concept can thus be utilized for biosensor design. [9,35,36]

In summary, we have demonstrated ligand conjugation of chemically exfoliated MoS$_2$ sheets through thiol chemistry. In this manner, we were able to tune the ζ-potential and surface functionality of ce-MoS$_2$ sheets to enable its broad usage as artificial receptors for enzymes. Because these aforementioned assemblies have potential applications in sensing,[35,37] energy,[1,30] etc, and because devices using 2D materials can outperform their nanoparticles,[34,38] it is possible that the functionalized ce-MoS$_2$ assemblies may enjoy similar advantages. The ability to functionalize these sheets through facile thiol chemistry suggests that similar chemistry may be applied to other chemically exfoliated transition metal dichalcogenide sheets (Figure S11), and can therefore open the window for further applications and solution processing of these emerging two-dimensional materials.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This work was partly funded by the National Cancer Institute Center for Cancer Nanotechnology Excellence (CCNE) initiative at Northwestern University Award No. U54CA119341. JH acknowledges support from National Science Foundation (DMR CAREER 0955612) and the Alfred P. Sloan Research Foundation. S. B. was supported by Basic Science Research Program through the National Research Foundation of Korea funded by the MEST (Project no. 2012-000181). We thank NU-HTA and NU NU*ANCE and NU-QBIC* for characterization equipment availability and expertise. We also thank A. Yan and Y. Huang for assistance with EM microscopy and M. Rotz and Dr. K. MacRenaris assistance with ICP-MS. SC thanks US DHS for a graduate fellowship.

References

1. Li Y, Wang H, Xie L, Liang Y, Hong G, Dai HJ. Am Chem Soc. 2011; 133:7296.
2. Radisavljevic B, Radenovic A, Brivio J, Giacometti V, Kis A. Nat Nano. 2011; 6:147.
3. Eda G, Yamaguchi H, Voiry D, Fujita T, Chen M, Chhowalla M. Nano Lett. 2011; 11:5111. [PubMed: 22035145]
4. Zhang H, Loh KP, Sow CH, Gu H, Su X, Huang C, Chen ZK. Langmuir. 2004; 20:6914. [PubMed: 15274603]
5. Liu KK, Zhang W, Lee YH, Lin YC, Chang MT, Su CY, Chang CS, Li H, Shi Y, Zhang H, Lai CS, Li LJ. Nano Lett. 2012; 12:1538. [PubMed: 22369470]
6. Wu H, Yang R, Song B, Han Q, Li J, Zhang Y, Fang Y, Tenne R, Wang C. ACS Nano. 2011; 5:1276. [PubMed: 21230008]
7. Medintz IL, Uyeda HT, Goldman ER, Mattoussi H. Nat Mater. 2005; 4:435. [PubMed: 15928695]
8. Nakanishi H, Walker DA, Bishop KJM, Wesson PJ, Yan Y, Soh S, Swaminathan S, Grzybowski BA. Nat Nano. 2011; 6:740.
9. De M, Rana S, Akpinar H, Miranda OR, Arvizo RR, Bunz UHF, Rotello VM. Nat Chem. 2009; 1:461. [PubMed: 20161380]
10. Yaacobi-Gross N, Soreni-Harari M, Zimin M, Kababya S, Schmidt A, Tessler N. Nat Mater. 2011; 10:974. [PubMed: 21983889]
11. Rapoport L, Fleischer N, Tenne RJ. Mater Chem. 2005; 15:1782.
12. Yarar B, Kaoma J. Colloids Surf. 1984; 11:429.
13. Tahir MN, Zink N, Eberhardt M, Therese HA, Kolb U, Theato P, Tremel W. Angew Chem, Int Ed. 2006; 45:4809.
14. Joensen P, Frindt RF, Morrison SR. Mater Res Bull. 1986; 21:457.
15. Schumacher A, Scandella L, Kruse N, Prins R. Surf Sci. 1993; 289:L595.
16. Zeng Z, Yin Z, Huang X, Li H, He Q, Lu G, Boey F, Zhang H. Angew Chem, Int Ed. 2011; 50:11093.
17. Dungey KE, Curtis MD, Penner-Hahn JE. Chem Mater. 1998; 10:2152.
18. Eda G, Fujita T, Yamaguchi H, Voiry D, Chen M, Chhowalla M. ACS Nano. 2012; 6:7311. [PubMed: 22799455]
19. Divigalpitiya WMR, Frindt RF, Morrison SR. Science. 1989; 246:369. [PubMed: 17747918]
20. Raybaud P, Hafner J, Kresse G, Toulhoat H. Phys Rev Lett. 1998; 80:1481.
21. Joensen P, Crozier ED, Alberding N, Frindt RF. J Phys C. 1987; 20:4043.
22. Late DJ, Liu B, Matte HSSR, Rao CNR, Dravid VP. Adv Funct Mater. 2012; 22:1894.
23. Heising J, Kanatzidis MG. J Am Chem Soc. 1999; 121:11720.

24. Shi W, Sahoo Y, Swihart MT. Colloid Surfaces A. 2004; 246:109.
25. Thangadurai P, Balaji S, Manoharan PT. Nanotechnology. 2008; 19:435708. [PubMed: 21832712]
26. Porter MD, Bright TB, Allara DL, Chidsey CED. J Am Chem Soc. 1987; 109:3559.
27. Harder P, Grunze M, Dahint R, Whitesides GM, Laibinis PE. J Phys Chem B. 1998; 102:426.
28. Wagner, CD.; Muilenberg, GE. Handbook of x-ray photoelectron spectroscopy: a reference book of standard data for use in x-ray photoelectron spectroscopy. Perkin-Elmer Corp., Physical Electronics Division; 1979.
29. Weisbecker CS, Merritt MV, Whitesides GM. Langmuir. 1996; 12:3763.
30. Jaramillo TF, Jørgensen KP, Bonde J, Nielsen JH, Horch S, Chorkendorff I. Science. 2007; 317:100. [PubMed: 17615351]
31. Chou SS, Kaehr B, Kim J, Foley BM, De M, Hopkins PE, Huang J, Brinker CJ, Dravid VP. Angew Chem, Int Ed. 2013 n/a. 10.1002/anie.201209229
32. Verma A, Simard JM, Worrall JWE, Rotello VM. J Am Chem Soc. 2004; 126:13987. [PubMed: 15506760]
33. Jacobson RH, Zhang XJ, DuBose RF, Matthews BW. Nature. 1994; 369:761. [PubMed: 8008071]
34. De M, Chou SS, Dravid VP. J Am Chem Soc. 2011; 133:17524. [PubMed: 21954932]
35. Miranda OR, Chen HT, You CC, Mortenson DE, Yang XC, Bunz UHF, Rotello VM. J Am Chem Soc. 2010; 132:5285. [PubMed: 20329726]
36. Miranda OR, You CC, Phillips R, Kim IB, Ghosh PS, Bunz UHF, Rotello VM. J Am Chem Soc. 2007; 129:9856. [PubMed: 17658813]
37. Lee JS, Ulmann PA, Han MS, Mirkin CA. Nano Lett. 2008; 8:529. [PubMed: 18205426]
38. Chou SS, De M, Luo J, Rotello VM, Huang J, Dravid VP. J Am Chem Soc. 2012; 134:16725. [PubMed: 22962967]

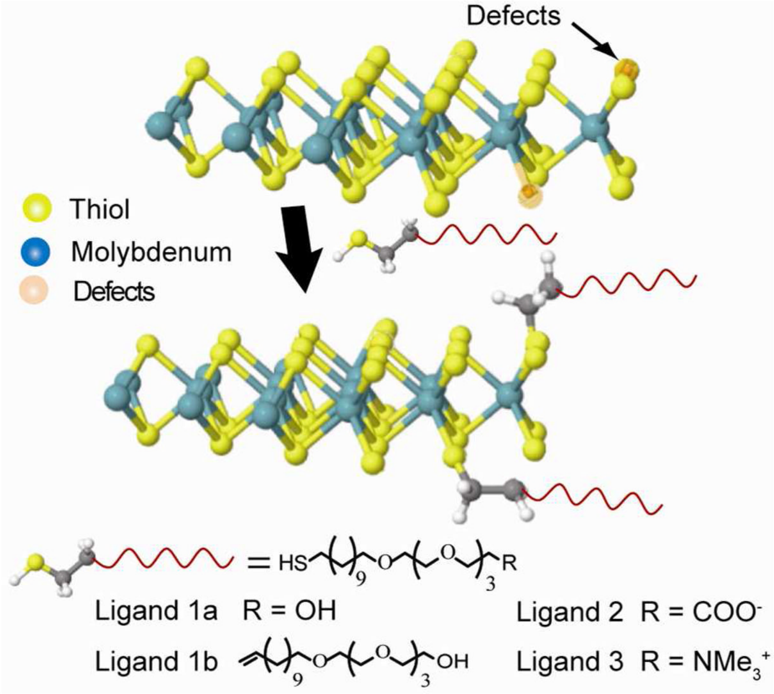

Figure 1.

Structural models illustrating ligand conjugation of ce-MoS_2 sheets.

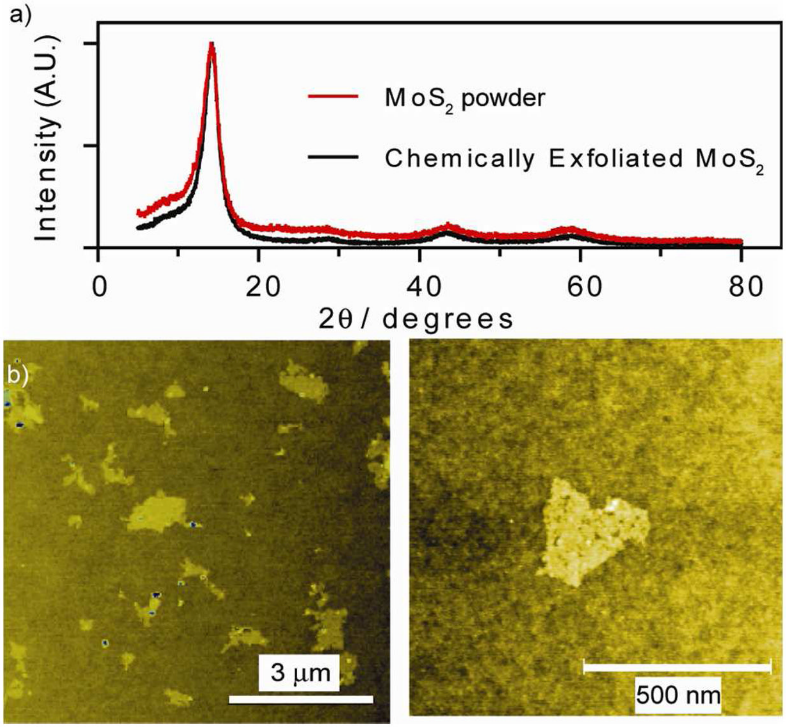

Figure 2.

(a) XRD spectra of ce-MoS_2. (b) Low and high magnification AFM micrograph of ce-MoS_2.

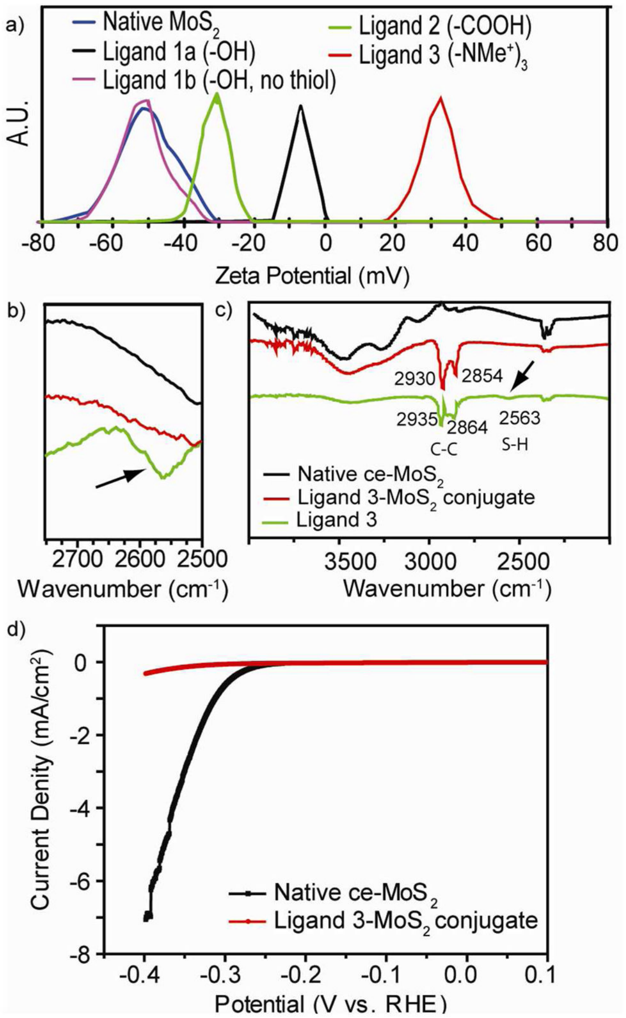

Figure 3.

(a) ζ-potential, (b) FT-IR spectra focused upon the thiol peak, (c) FT-IR spectra showing all peaks and (d) HER polarization curve before and after ligand conjugation

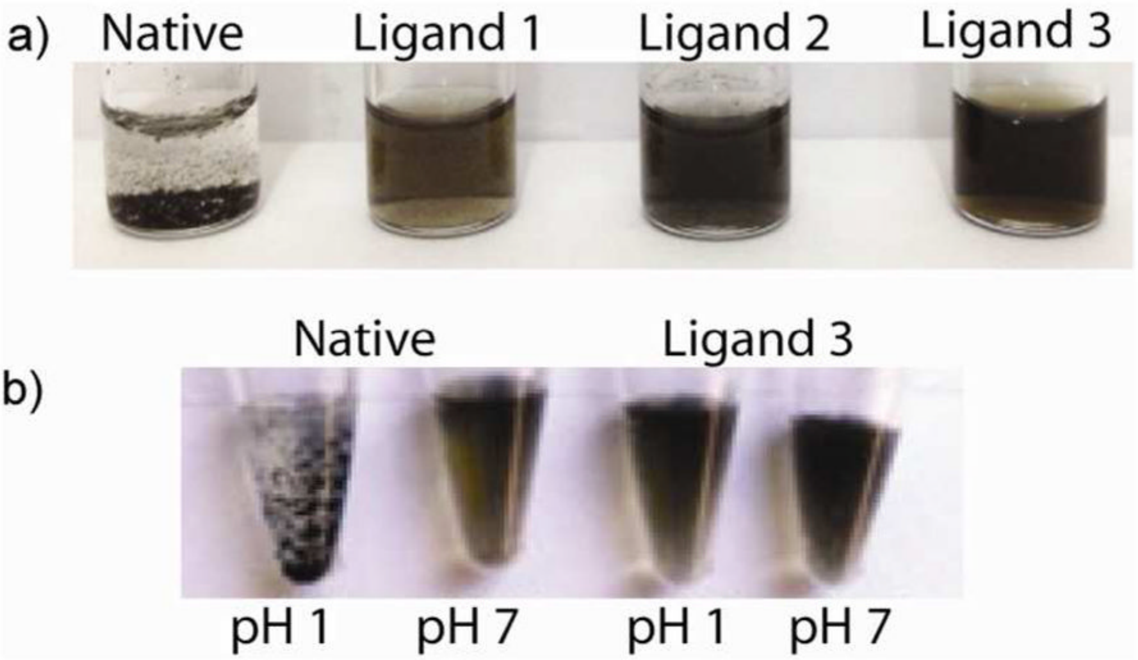

Figure 4.

(a) Colloidal stability of ce-MoS$_2$ samples after 21 days in water. (b) Colloidal stability of ce-MoS$_2$ functionalized with NMe$_3$ terminated thiol at various pH.

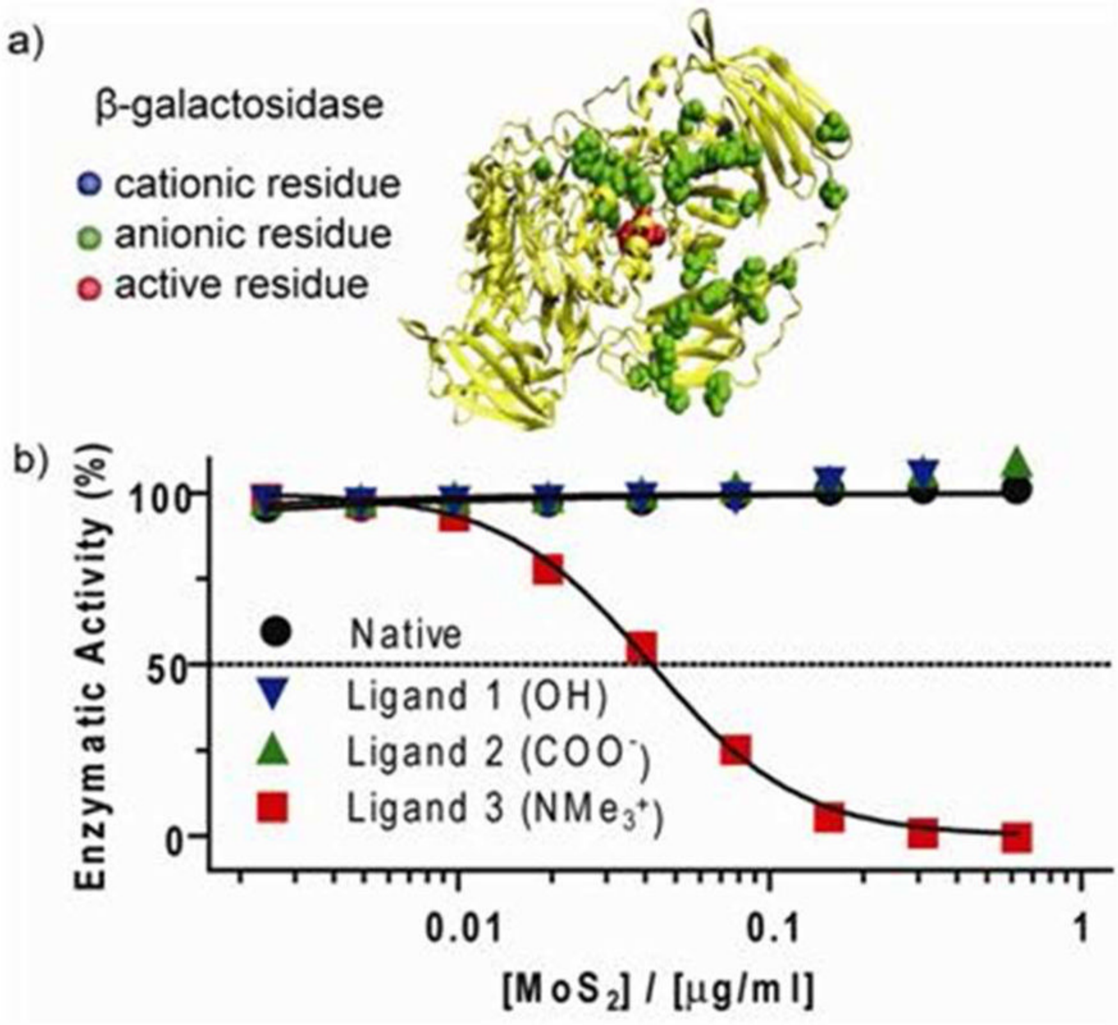

Figure 5.

(a) Structure of a single unit of a β-Gal tetramer. Anionic residues (Asp and Glu) surrounding β-gal's active site are highlighted (bottom) (b) Concentration dependence of β-gal inhibition with various ce-MoS_2 conjugate.