

Complete Genome Sequence of the Probiotic *Bifidobacterium thermophilum* Strain RBL67

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***Bifidobacterium thermophilum* RBL67, an isolate from infant feces, exhibits bacteriocin-like antimicrobial activity against *Listeria* spp. and *Salmonella* spp. and protects HT29-MTX cells against *Salmonella* infection. Here, the complete genome sequence of the probiotic *B. thermophilum* strain RBL67 is presented.**

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Bifidobacterium thermophilum belongs to the *Bifidobacterium boum* group of bifidobacteria (1), which has not been studied extensively. Similar to the *Bifidobacterium longum* and *Bifidobacterium adolescentis* species, *B. thermophilum* strains have been isolated from bovine rumen, calf feces, sewage, and piglet feces (2). In contrast, *B. thermophilum* strain RBL67 was isolated from baby feces in a consortium with *Pediococcus acidilactici* UVA1 (3, 4). RBL67 is a moderately oxygen-tolerant strain that reaches high cell numbers in fermentation (2). Furthermore, it produces a bacteriocin-like inhibitory substance (BLIS) that is active against *Listeria* spp. (4, 5). Clear activity against *Salmonella enterica* subsp. *enterica* serovar Typhimurium establishment and infection was observed in a combined colonic fermentation using immobilized child fecal microbiota and an epithelial HT29-MTX cell model (6, 7). RBL67 increased the life span of the small soil nematode *Caenorhabditis elegans* during *S. enterica* subsp. *enterica* serovar Virchow N90 exposition (6) and reduced the severity of rotavirus-associated diarrhea in suckling mice (8). Its protective and antimicrobial effects, growth characteristics, and technological ability make RBL67 a promising microbe for enhancing gastrointestinal health.

The genome of RBL67 was sequenced using a combined Roche GS-FLX Titanium and Illumina HiSeq 2000 approach (GATC-Biotech, Konstanz, Germany). DNA was prepared with a lysozyme/mutanolysine-based cell lysis and subsequent purification using the Wizard genomic DNA purification kit (Promega, Madison, WI) (9).

RBL67 is the first strain of the species *B. thermophilum* to be completely sequenced, assembled, and publically available. The genomes of closely related *Bifidobacterium* spp., including *B. adolescentis* ATCC 15703 (accession no. AP009256), *B. animalis* ATCC 25527 (accession no. CP002567), and *B. longum* NCC2705 (accession no. AE014295), were used as references for alignment in Projector 2 (10). Sanger sequencing for subsequent gap closing was performed at GATC-Biotech (Germany). Highly repetitive regions, such as clustered regularly interspaced short palindromic repeat regions (CRISPRs), were validated by primer walking and Sanger sequencing. Assembly was performed with Lasergene Seq-

Man Pro 8.0.2 (DNASTAR, Madison, WI) and CLC Genomics workbench 6.0.1 (CLC bio, Aarhus, Denmark). The genome was annotated using the NCBI Prokaryotic Genomes Automatic Annotation Pipeline (PGAAP, <http://www.ncbi.nlm.nih.gov/genomes/static/Pipeline.html>) and the Rapid Annotation using Subsystems Technology (RAST) platform (11) with RAST-based and Glimmer-3-based gene calling. Annotations obtained through these pipelines were compared and subsequently curated manually.

The genome of *B. thermophilum* RBL67 consists of a 2,291,643-bp circular molecule. The G+C content of the genome is 60.1%, which is within the 59.2 to 60.5% range of its relatives *B. adolescentis*, *B. animalis*, and *B. longum*. RBL67 harbors 47 tRNA genes and 12 rRNA genes, including 4 copies of the 16S rRNA gene. A total of 1,845 coding sequences (CDS) were predicted in the genome, of which 50 CDS have not been found so far in other *Bifidobacterium* species. However, 25 out of these 50 CDS were <100 amino acids. The complete genome of RBL67 will provide insight in the evolution of bifidobacteria and contributes to the further understanding of the biology of the genus and properties of this particular species.

Nucleotide sequence accession number. The genome sequence of *B. thermophilum* RBL67 was deposited at GenBank under the accession no. [CP004346](https://ncbi.nlm.nih.gov/nucl/CP004346).

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REFERENCES

1. Turroni F, van Sinderen D, Ventura M. 2011. Genomics and ecological overview of the genus *Bifidobacterium*. *Int. J. Food Microbiol.* 149:37–44.
2. Biavati B, Mattarelli P. 2009. Genus I. *Bifidobacterium*, p 171–206. In Whitman WB, Kämpfer P, Goodfellow M, Garrity GM, Ludwig W (ed), *Bergey's manual of systematic bacteriology*, 2nd ed: vol 5, the *Actinobacteria*, part 1. Springer Verlag, New York, NY.

3. von Ah U, Mozzetti V, Lacroix C, Kheadr EE, Fliss I, Meile L. 2007. Classification of a moderately oxygen-tolerant isolate from baby faeces as *Bifidobacterium thermophilum*. *BMC Microbiol.* 7:79.
4. Touré R, Kheadr E, Lacroix C, Moroni O, Fliss I. 2003. Production of antibacterial substances by bifidobacterial isolates from infant stool active against *Listeria monocytogenes*. *J. Appl. Microbiol.* 95:1058–1069.
5. Von Ah U. 2006. Ph.D. thesis no. 16927. Identification of *Bifidobacterium thermophilum* RBL67 isolated from baby faeces and partial purification of its bacteriocin. ETH Zurich, Zurich, Switzerland.
6. Zihler A. 2010. Ph.D. thesis no. 19059. *In vitro* assessment of bacteriocinogenic probiotics for prevention and treatment of *Salmonella* in children using novel *in vitro* continuous colonic fermentation and cellular models. ETH Zurich, Zurich, Switzerland.
7. Zihler A, Gagnon M, Chassard C, Lacroix C. 2011. Protective effect of probiotics on *Salmonella* infectivity assessed with combined *in vitro* gut fermentation-cellular models. *BMC Microbiol.* 11:264.
8. Gagnon M. 2007. Ph.D. thesis. Rôle des probiotiques lors d' infections entériques d' origine bactérienne et virale: analyses *in vitro* et études *in vivo* chez des modèles murins. Université Laval, Québec, Canada.
9. Jans C, Gerber A, Bugnard J, Njage PM, Lacroix C, Meile L. 2012. Novel *Streptococcus infantarius* subsp. *infantarius* variants harboring lactose metabolism genes homologous to *Streptococcus thermophilus*. *Food Microbiol.* 31:33–42.
10. van Hijum SA, Zomer AL, Kuipers OP, Kok J. 2005. Projector 2: contig mapping for efficient gap-closure of prokaryotic genome sequence assemblies. *Nucleic Acids Res.* 33:W560–W566.
11. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75.