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Increased Dickkopf-1 expression in patients with unexplained recurrent spontaneous miscarriage

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Summary

Wnt pathways play an important role in pre-implantation embryo development, blastocyst implantation, and post-implantation uterine decidualisation. However, little is known about the potential role that Wnt signaling plays in patients with unexplained recurrent spontaneous miscarriage (URSM), and no single biomarker with a high predictive value of maternally caused URSM has been identified. We aim to study the molecular mechanisms by which the Wnt pathway controls the progression of early pregnancy by investigating the expression of Dickkopf-1 (DKK1), one of the Wnt agonists, in URSM patients. Plasma and fresh decidual tissues samples were collected from 59 subjects (29 patients with URSM and 30 patients with normal, early pregnancy). Time-resolved immunofluorometric assay system and quantitative real-time RT-PCR were used to determine the serum levels of DKK1 and DKK1 mRNA in the deciduas, respectively. Western blot and immunohistochemistry were used to measure DKK1 protein levels in the deciduas. Serum DKK1 levels were significantly higher in URSM patients compared to the control group (P < 0.001); the expression of DKK1 mRNA and protein in URSM patients were higher relative to healthy controls (P = 0.013). Glandular epithelium from decidual tissues demonstrated cytoplasmic signals for DKK1 in URSM patients, and DKK1 did not stain in healthy controls. Furthermore, serum DKK1 levels significantly correlated with those in the decidual tissues. Our study suggests that DKK1 may be a valuable biomarker of URSM; it can be reliably and conveniently detected in serum, thus obviating the need for decidual tissue analysis.

Keywords: Dickkopf-1, recurrent spontaneous miscarriage, wnt signaling pathway

Introduction

Recurrent spontaneous miscarriage (RSM), defined as two or more consecutive pregnancy losses before the 20th week of gestation, occurs in 1%-5% women of reproductive age, and has a negative effect on human reproductive health [1]. The aetiology of RSM remains partially unknown, although chromosomal, anatomic, endocrinological, infectious, and auto-immunologic abnormalities have been implicated [2]. Almost 50% of such cases, called unexplained recurrent spontaneous miscarriage (URSM), remain classified as 'of unknown aetiology', and usually occur during the first trimester [3]. Although numerous signalling factors and pathways have been found to be important for a successful early

pregnancy, the molecular basis of reciprocal interactions between the mother and foetus during pregnancy still remains largely unknown. Recent analyses regarding human endometrial gene expression within the window of implantation demonstrate the expression and regulation of select members of the Wnt signalling pathway, raising the question regarding the function of this pathway during human reproduction and suggest that it may be involved in this molecular network [4,5].

The Wnt signalling pathway is one of a handful of evolutionarily conserved signal transduction pathways used extensively during animal development, from Hydra to humans [6,7], and controls multiple aspects of development, including cell proliferation, fate specification,

polarity, and migration. Wnt signals are transduced in at least two distinct ways: a well-established canonical or Wnt/ β -catenin pathway, and a non-canonical β -catenin independent pathway or pathways [8]. B-Catenin is an intracellular mediator of canonical Wnt signalling, which is phosphorylated and stabilized because of activation by Wnt ligand, and subsequently transported to the nucleus, where, as part of a transcriptional complex, it regulates gene expression. The Wnt signal is extracellularly regulated by two classes of antagonists. The first group consists of Wnt ligand-binding proteins, including Wnt-inhibitory factor (WIF-1), the secreted frizzled-related protein (sFRP) family, and cerberus. Dickkopf (DKK) family of proteins belong to the second group [9], and DKK1, the most wellcharacterised member of the family, promotes the internalisation of LRP5/6, making it unavailable for Wnt reception by forming a triplet complex with LRP5/6 and Kremens [10,11].

Since the intricate interplay between the embryo and uterus during implantation shares similar features with the reciprocal cell-cell communication involving Wnt signaling occurring during organogenesis, the same pathway may drive embryogenesis. Chen *et al.* [12] have briefly summarised recent progress on the patho-physiological significance of differential Wnt pathways during pre-implantation embryonic development, blastocyst implantation, and postimplantation uterine decidualisation.

To date, however, little is known about the potential role played by Wnt signaling in URSM patients, and no single biomarker with a high predictive value of maternally caused URSM has been identified. Non-genetic biomarkers in URSM may reflect conditions in the gravid uterus, and we rarely know whether they are the causes or consequences of miscarriage. Studies of specific biomarkers are probably the best way to reveal the pathogenesis underlying URSM [13]. In this study, we examined DKK1 expression in both normal-pregnancy women and in women with URSM. Our results indicate abnormal expression of DKK1 in women with USRM, which suggests that interference with the Wnt signalling pathway may results in URSM.

Materials and methods

Patients

All participants enrolled were outpatients at the Department of Gynaecology, Shanghai First Maternity and Infant Hospital. Twenty-nine patients who had experienced at least two consecutive first-trimester losses (9.40 ± 2.25 weeks of gestation) of unexplained aetiology, with an age (mean \pm standard deviation) of 31.13 ± 1.07 years (age range, 26–38 years) were recruited as participants. 'Unexplained' RSM was diagnosed as per previously described guidelines, to exclude any verifiable causes [14]. For a relevant control group, we selected 30 healthy women who were undergoing elective termination of apparently normal pregnancies. The elective terminations were conducted during the first trimester $(9.23 \pm 1.89$ weeks of gestation). All these women had at least one living child, and no history of spontaneous miscarriage, ectopic pregnancy, preterm delivery, or stillbirth. In all 30 cases, foetal heart activity had been identified within two weeks of sample collection.

The fresh decidual tissues collected from all patients under aseptic conditions, after receiving their consent, was immediately delivered to the laboratory.

This study was approved by the ethics review board of Shanghai First Maternity and Infant Hospital, and informed consent was obtained after the study was explained in detail to all the participants.

Serum DKK1 and progesterone measurement

Serum levels of DKK1 were measured by time-resolved immunofluorometric assay system [15]. A monoclonal anti-DKK1 antibody and an anti-DKK1 antibody labelled with Eu³⁺ as a tracer were used to develop a non-competitive 'sandwich-type' assay. Fluorescence can be measured using a time-resolved fluorometer after the immunoreaction. The sensitivity of the assay was 0.7 μ g/l. Accuracy studies, specificity, parallelism, and precision data were found to be satisfactory. As progesterone plays a pivotal role in ovulation, implantation, and the establishment and maintenance of pregnancy, we measured progesterone levels in the same set of serum samples from all subjects. Serum progesterone was detected using electrochemiluminescent assays by using Beckman Coulter, Inc., reagent sets and UniCel DxI 800 Immunoassay System.

Quantitative real-time RT-PCR

Total RNA from patient decidual tissue samples was prepared using Rneasy Mini or Micro Kits (QIAGEN, Valencia, USA) according to the manufacturer's instructions. Total RNA was quantified with a BioPhotometer plus (Eppendorf AG, Hamburg, Germany), and $2 \mu g$ of each sample was reverse transcribed using Random Hexamer Primers (Promega Corporation, Madison, WI, USA) and M-MLV Reverse Transcriptase (Promega Corporation) in a final volume of 25 µl. Subsequently, cDNA samples were used as PCR templates, along with specific primers specifically designed for real-time PCR.

For quantitative real-time RT-PCR, amplification was performed in LightCyclert system (Roche, Mannheim, Germany), and visualized with SYBR Green Kit (Quantitectt SYBR Green PCR, QIAGEN). Standard curves for each gene were established to quantify the copy number for each sample, and expression was considered only when the sample Ct was within the limit of each standard curve. Fol-

Table 1. Characteristics of URSM and control subjects (mean + SD and range of values).

Group	n	Age (year)	No. of miscarriage	Gestational age(weeks)
URSM	29	31·13 ± 1·07 (26–38)	4·47 ± 0·21 (2-8)	9·40 ± 2·25 (7-12)
Control	30	30·0 ± 8·59 (25–36)	0	9·23 ± 1·89 (7–12)
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URSM *versus* Control: Age, P = 0.14; Gestational age, P = 0.62.

lowing primer sequences were used for real-time PCR: β -actin, 5': AAGGCCAACCGCGAG; 3': TAATGTCACGC ACGATTCCCG; DKK1 5': CTCGGTTCTCAATTCCAACG; 3': GCACTCCTCGTCCTCTG. The cycling conditions were 15 min at 95°C, 40 cycles of 15 s at 94°C, 30 s at 55°C, 30 s at 72°C and 5 s at 82°C (β -actin) or 84°C (DKK1). Gene expression was presented as the ratio between the target gene and β -actin. Each experiment was done in triplicate and the mean value was calculated.

Western blotting

Western blotting using total tissue lysates was performed as previously described [16]. Briefly, we subjected 30 μ g of crude decidual tissue protein extract to 12% sodium dodecyl sulphate-polyacrylamide gel electrophoresis, and then transferred the proteins to a polyvinylidenedifluoride (PVDF) membrane (Bio-Rad, USA). Membranes were incubated overnight at 4°C in 1:1000 dilution of antihuman DKK1 antibody (Perkin Elmer, USA) or 1:1000 dilution of anti- β -actin antibody (Perkin Elmer, USA) then for 3 h with the corresponding secondary antibody. We visually estimated the band intensities for both proteins.

Immunohistochemistry

To detect DKK1 protein in decidual tissue samples embedded in paraffin blocks, we stained the sections as previously described [16]. Briefly, 3.3 µg/ml of rabbit polyclonal antihDKK1 antibody (Perkin Elmer, USA) was added to each slide after blocking endogenous peroxidase and proteins, and the sections were incubated with HRP-labelled Anti-Rabbit IgG (Perkin Elmer, USA) as the secondary antibody. Substrate chromogen was added, and the specimens were counterstained with haematoxylin. Normal rabbit IgG was used as a substitute for the primary antibody as negative controls. All sections were observed and photographed with an Axioskop 2 microscope (Carl Zeiss, Oberkochen, Germany). Quantitative analysis of immunohistochemistry was done as follows: 1, weak; 2, moderate; and 3, strong. Positive cells were quantified as a percentage of the total number of decidual cells, and were assigned to one of the five categories: 0, <5%; 1, 5%-25%; 2, 26%-50%; 3, 51%-75%; and 4, >75%. The percentage of positive decidual cells and the staining intensity were then multiplied to generate the immunoreactive score (IS) for each decidual specimen. A sample was defined as DKK1+ if it had an IS of \geq 1. An IS value of ≥ 2 was regarded as high level of expression. Two independent pathologists (Liu J and Huang WM) were involved in assessing protein expression.

Statistical analysis

Statistical analyses were performed using GraphPad Prism (GraphPad Software, Inc., La Jolla, CA, USA). The data regarding serum DKK1 levels are presented as the mean \pm standard deviation values unpaired student *t*-tests and the rank-sum test (Wilcoxon) were used to evaluate differences among unpaired observations within the two groups. A *P*-value of <0.05 was considered significant.

Results

Patient characteristics

Twenty-nine patients with URSM were recruited in the study and divided into two groups: 17 patients with two occurrences of miscarriages, and 12 patients with more than two occurrences of miscarriages. Thirty healthy women undergoing elective termination of apparently normal pregnancies were selected to form the control group (age, 30.0 ± 8.59 ; range, 25-36 years). No significant differences were observed in the age and gestational period between the two groups (Table 1).

Comparison of serum DKK1 and progesterone levels in URSM patients and control group

Serum DKK1 levels had a normal distribution in control subjects with a mean value of 11.98 µg/l, and they ranged from 5.43 to 18.5 µg/l. The 95th percentile of DKK1 levels for the control group, i.e. 14.43 µg/l, was used as the cut-off value for differentiating between patients and controls. For USRM patients, the mean value was $24.99 \,\mu g/l$, and the range was 15.39-34.59 µg/l. Serum DKK1 levels were greater (P < 0.005; Fig. 1a) for USRM patients. Moreover, no significant difference were observed in the serum DKK1 levels between women with two or more than two incidences of USRM (P = 0.45; Fig. 1b). We found no significant differences between the serum progesterone levels in the USRM and the control subjects (P = 0.36, Fig. 1c). Moreover, no correlations were observed between the levels of DKK1 and progesterone, with a Pearson's coefficient of 0.05 (P = 0.699, Fig. 1d).

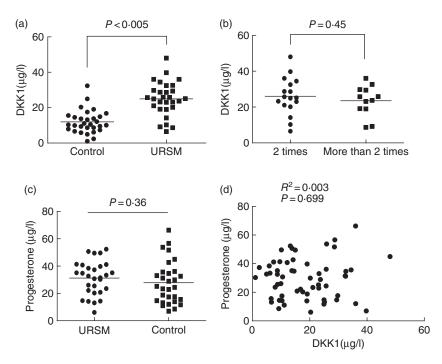


Fig. 1. A, DKK1 levels in serum samples from patients with URSM and healthy pregnancies determined by using time-resolved immunofluorometric assay system. B, Point plot of serum DKK1 levels in URSM patients with two or more occurrences of miscarriages. C, Serum progesterone levels in URSM patients and control subjects. No difference was noted in the serum progesterone levels between the two groups. D, Correlation between serum DKK1 and progesterone levels in patients with URSM and control subjects.

Clinical significance of serum DKK1 as a serologic biomarker for URSM

The utility of serum DKK1 levels in the detection of URSM was evaluated using receiver-operator characteristic curve (ROC) analysis. At a cut-off level of 14·43 μ g/l, the positive rate in URSM was 82·76% (24/29) and 30% (9/30) in the control. The sensitivity and specificity of serum DKK1 for URSM were 82·76% and 73·33%, respectively. The area under the ROC curves was 0·8609.

DKK1 expression in decidual tissues

To investigate whether high serum DKK1 levels coincided with DKK1 expression in the decidual tissues, 32 frozen tissues (18 from URSM group and 14 from control group) were randomly chosen from 59 cases for analysis by quantitative real-time RT-PCR. As shown in Fig. 2a, DKK1 mRNA expression in URSM patients was higher than that in the control group (P = 0.013). Significant correlations were noted between DKK1 mRNA expression in individual decidual tissue samples and DKK1 concentrations in serum samples (P < 0.001, Fig. 2b). Using Western bloting analysis, we confirmed that DKK1 protein expression in URSM patients was higher than that in the control group (P < 0.001, Fig. 2c and d). Among 59 study subjects, high expression of DKK1 protein was significantly correlated with the serum DKK1 levels (P < 0.001, Fig. 2e).

Local expression of DKK1

To further characterise the identified markers and localise DKK1 in the decidual tissue sections, we examined its

expression in several decidual tissue samples by immunohistochemistry using specific antibodies against DKK1. Negative controls without the primary antibody or with non-specific immunoglobulins had no signal. Decidual glandular epithelial tissues demonstrated cytoplasmic signals for DKK1 in URSM patients. However, DKK1 expression was lower, or even absent in the control deciduas. Two or more than two occurrences of miscarriages did not produce a significant difference (Fig. 3a). Overall, cytoplasmic DKK1 was present in 27 of 29 URSM cases (93.1%), and DKK1 cytoplasmic staining was present in 10 of 30 healthy pregenacies (33.3%). The intensity of staining and the representative tissues from all 59 cases are presented in Fig. 3b. The lack of DKK1 expression was more prevalent in the control group compared with the URSM group (Fig. 3c). A significant correlation was noted between elevated cytoplasmic expression of DKK1 in the decidual tissues and serum DKK1 levels (P < 0.001). Data were analysed using Spearman's correlation test (2-tailed).

Discussion

It has recently been clarified that secreted Wnt antagonists play important roles in regulating Wnt signalling [9]. DKK proteins are secreted antagonists of the Wnt cell- signalling molecules that possess a novel mode of action. The DKK family consists of four main members in vertebrates (DKK1, 2, 3, 4), and DKK1, the founding member of the family, was originally identified as the embryonic head inducer and Wnt antagonist in Xenopus species [17]. DKK1 binds to the LRP5/6 Wnt co-receptor and prevents the formation of active Wnt-Frizzled-LRP5/6 receptor

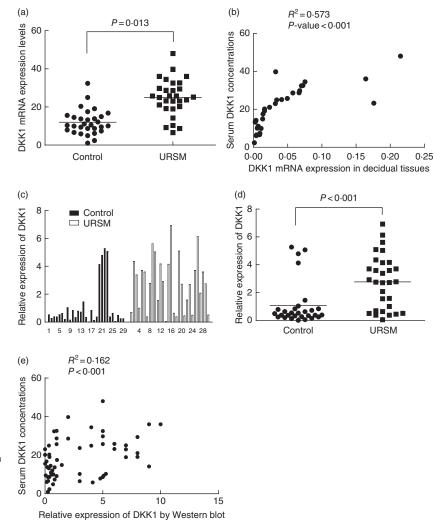


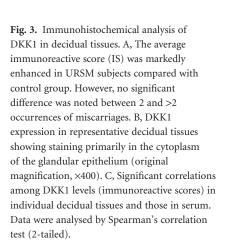
Fig. 2. Expression of DKK1 in individual tissues from patients with URSM and healthy pregnancy. A, DKK1 mRNA expression as measured by quantitative real-time RT-PCR in patients with URSM and control group. B, Correlative analysis of DKK1 mRNA levels in individual decidual tissue samples and serum DKK1 levels by Spearman's correlation test (2-tailed). C, Each DKK1 intensity was divided by the corresponding intensity of β -actin from the same tissue sample to adjust for the sample variation. D, Increased DKK1 protein expression was present in URSM decidual tissues than the control. E, Among 59 study subjects, high expression of DKK1 protein was significantly correlated with the serum DKK1 levels.

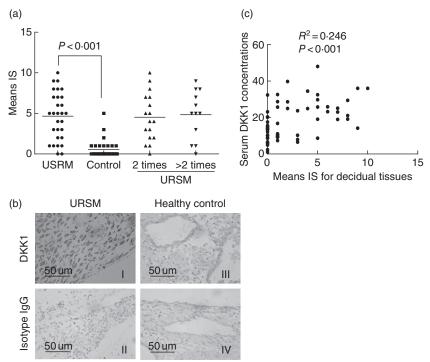
complexes, thus blocking the canonical Wnt/ β -catenin pathway [18,19].

In this study, we found that serum DKK1- levels increase in URSM, and this increase appears to be independent of instances of miscarriage, which suggests that the Wnt/ β catenin pathway may be down-regulated by the induction of DKK1 expression. Concurrently, the sensitivity and specificity of serum DKK1 for URSM were found to be 82.76% and 73.33%, repectively, by using ROC analysis; thus, DKK1 may be a useful indicator for URSM. However, future studies are necessary to identify the molecular mechanisms underlying DKK1 actions in URSM.

In mice, the expression of several Wnt genes fluctuates in adult mouse uteri during the oestrous cycle [20,21]. In addition, some antagonists of the Wnt signalling pathway, such as sFRP family members, showed dynamic changes during this process [22,23]. During the human endometrial menstrual cycle, although the expression of Wnt ligands, receptors, or downstream effectors does not vary significantly, dramatic changes in the expression of DKK1 and FrpHE(frizzled-related protein, an inhibitor of Wnt signalling) were evident [5,24]. Some researchers suggested that DKK1 mRNA regulation is dependent on progesterone, and its expression increases during the early-secretory phase, peaks during the mid-secretory phrase, and decreases during the late-secretory phase [25]. However, there is insufficient evidence of DKK1 staining during pregnancy. Our data showed that the expression of DKK1 mRNA and protein in the deciduas in URSM was higher relative to the control group. To investigate whether high serum DKK1 levels, coincides with DKK1 expression in the decidual tissues, we used Spearman correlation analysis. Significant correlations were noted between decidual DKK1 mRNA expression and serum concentrations.

Consistent with these findings, DKK1 showed positive immunostaining in the maternal decidual compartment in 'unexplained' URSM. A significant correlation was noted between elevated cytoplasmic expression decidual DKK1 and the serum DKK1 levels. However, evidence for DKK1 staining in the decidual tissue during pregnancy is inadequate. Our results indicate higher expression of DKK1 mRNA and protein in the decidua from the USRM patients





compared to the control patients. These results are not in agreement with other findings [26] regarding progesteroneregulated DKK1 expression, which could be regulated by other factors, since there were no differences in the concentrations of progesterone in the serum samples of USRM and control subjects in the present study. Macdonald *et al.* [27] reported on DKK1 expression in normal early pregnancy which appears to be in contrast to our findings, maybe all participants in our study were 'Unexplained' RSM, and were different from patients having luteal phase defection (LPD), one of the major causes of RSM. Different molecular mechanism exists in RSM due to endocrinological disorder and 'Unexplained' RSM.

Moreover, some investigators have found that the activation of the canonical Wnt pathway was essential in regulatory T (Treg) homeostasis, and may play a key role in anergy induction in mature T cells [28]. The vital role of Treg cell in maintaining self-tolerance is quite clear [29]. Defects in Treg cell development lead to systemic autoimmunity and early death in both mice and humans. Many researchers suggest that CD4+CD25^{bright} Treg cells might play a role in the maintaining pregnancy, and a decreased number of CD4+CD25^{bright} Treg cells in URSM women may induce maternal lymphocyte activation in the foetal allograft [30]. Thus, we speculate that increased DKK1 levels in the deciduas may affect paracrine as well as autocrine Wnt signalling by inhibiting the Wnt pathway, and thereby regulating signalling at a specific time in URSM patients by reducing the number of CD4+CD25^{bright} Treg cells. However, further work is needed to explore this possibility.

Our study suggests that DKK1 may be a valuable biomarker of URSM; it can be reliably and conveniently detected in the serum, thus obviating the need for decidual tissue analysis. Our findings may shed light on a new mechanism that exists in URSM. However, the precise mechanisms of how DKK1 regulates decidual and trophoblast function, as well as the mechanisms underlying Wnt regulation and action during early pregnancy in humans, are not well understood. Our future studies will address in detail the molecular mechanisms by which the Wnt pathway controls the progression of early pregnancy.

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Disclosure

The authors declare that they have no conflicts of interest.

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