

REVIEW

Open Access

Hydrogen sulfide as a vasculoprotective factor

Eloise Streeter, Hooi H Ng and Joanne L Hart*

Abstract

Hydrogen sulfide is a novel mediator with the unique properties of a gasotransmitter and many and varied physiological effects. Included in these effects are a number of cardiovascular effects that are proving beneficial to vascular health. Specifically, H₂S can elicit vasorelaxation, prevention of inflammation and leukocyte adhesion, anti-proliferative effects and anti-thrombotic effects. Additionally, H₂S is a chemical reductant and nucleophile that is capable of inhibiting the production of reactive oxygen species, scavenging and neutralising reactive oxygen species and boosting the efficacy of endogenous anti-oxidant molecules. These result in resistance to oxidative stress, protection of vascular endothelial function and maintenance of blood flow and organ perfusion. H₂S has been shown to be protective in hypertension, atherosclerosis and under conditions of vascular oxidative stress, and deficiency of endogenous H₂S production is linked to cardiovascular disease states. Taken together, these effects suggest that H₂S has a physiological role as a vasculoprotective factor and that exogenous H₂S donors may be useful therapeutic agents. This review article will discuss the vascular effects and anti-oxidant properties of H₂S as well as examine the protective role of H₂S in some important vascular disease states.

Keywords: Hydrogen sulfide, Vasculoprotective, Atherosclerosis oxidative stress

Introduction

Hydrogen sulfide is now a recognised gaseous mediator and induces many and varied biological effects [1]. Several cardiovascular actions of H₂S have been described, including vasorelaxation, prevention of inflammation and leukocyte adhesion, anti-proliferative effects, anti-thrombotic effects, resistance to oxidative stress and protection against ischemia-reperfusion injury. These result in protection of endothelial function, resistance to vascular remodelling and maintenance of blood flow and organ perfusion. Taken together, these effects suggest that H₂S has a physiological role as a vasculoprotective factor. This review examines the evidence that H₂S is an important vascular regulator and protectant.

H₂S production, storage and metabolism

H₂S is produced endogenously via the metabolism of cysteine and/or homocysteine [2], by the enzymes cystathionine-β-synthase (CBS, EC 4.2.1.22) [3] and cystathionine-γ-lyase (CSE, EC 4.4.1.1) [4]. 3-mercaptopyruvate sulfurtransferase (3-MST, EC 2.8.1.2) can also generate H₂S acting in concert with cysteine aminotransferase (EC 2.6.1.75) to

metabolise cysteine, generating pyruvate and H₂S [5]. CBS is a major contributor to H₂S production in the brain, whilst CSE levels predominate in most peripheral tissues. 3-MST appears to contribute to H₂S production in both the periphery and central nervous system [5,6]. In the vascular system CSE is primarily expressed in vascular smooth muscle cells but there is also evidence that it is expressed in the endothelium [7,8].

H₂S is metabolized by mitochondrial oxidative modification that converts sulfide into thiosulfate, which is converted further into sulfite and finally sulfate, which is the major end product of H₂S metabolism [9]. H₂S consumption in the presence of O₂ is high [10], thus H₂S production is offset by rapid clearance, resulting in low basal levels of H₂S. In addition to high clearance H₂S may also be stored as acid-labile sulphur [11] or bound sulfane sulphur within cells [12]. The metabolic turnover of H₂S and concentrations of the gas generated *in vivo* during cell stimulation are yet to be fully elucidated and will be an area of importance in H₂S biology future research.

Gasotransmitter and chemical properties

Gaseous mediators or gasotransmitters are a relatively new class of signalling molecules, These gases share

* Correspondence: Joanne.hart@rmit.edu.au
School of Medical Sciences and Health Innovations Research Institute (HIRI),
RMIT University, PO Box 70, Bundoora, Vic 3083, Australia

many features in their production and action but differ from classical signalling molecules. Advantages of gases as signalling molecules include their small size which allows easy access to a variety of target sites that would not be accessible by larger molecules. They easily cross membranes, are labile with short half-lives and are made on demand. They are not stored in their native form as they can't be constrained by vesicles and need to be bound for storage or rely upon *de novo* synthesis. They can have endocrine, paracrine, autocrine or even intracrine effects. It is also interesting that all the molecules confirmed as gasotransmitters (nitric oxide (NO), carbon monoxide (CO), H₂S) were all considered only as toxic molecules until their endogenous production and effects were determined.

About 80% of H₂S molecules dissociate into hydrosulfide anion (HS⁻) at physiological pH 7.4 in plasma and extracellular fluids [13]. HS⁻ is a potent one-electron chemical reductant and nucleophile that is capable of scavenging free radicals by single electron or hydrogen atom transfer [14,15] Thus, H₂S should readily scavenge reactive nitrogen species (RNS) and reactive oxygen species (ROS) [16]. It is also now established that H₂S can signal via sulhydration of proteins [17], and much research is ongoing in this area.

H₂S effects on blood vessels

Endothelium derived substances that cause vasodilatation (eg NO, prostacyclin) are anti-proliferative and anti-thrombotic while constrictor factors (endothelin-1, thromboxane A₂) are proliferative and pro-coagulant. Thus the vasodilators can be considered vasculoprotective, as they protect and promote blood flow and a balance of endothelium-derived relaxing and contracting factors is required for a healthy vascular function [18]. H₂S is produced in blood vessels by both endothelial cells and vascular smooth muscle has these same vasculoprotective properties (Figure 1). These are further discussed below.

Vasorelaxation elicited by H₂S

H₂S induced vasorelaxation in peripheral vessels may be mediated by various mechanisms, including opening of potassium channels, blockade of voltage-gated Ca²⁺ channels, enhanced production or activity endothelial derived factors, such as NO, PGI₂ and EDHF and decreased pH_i. The vasorelaxant effect occurs in both large conduit [19-22] and small resistance-like blood vessels [7,23,24] and is physiologically relevant since an inhibition of CSE in isolated mouse aorta *in vitro* causes significant vascular contraction [19] and most importantly, mice deficient in CSE are hypertensive and have endothelial dysfunction [8].

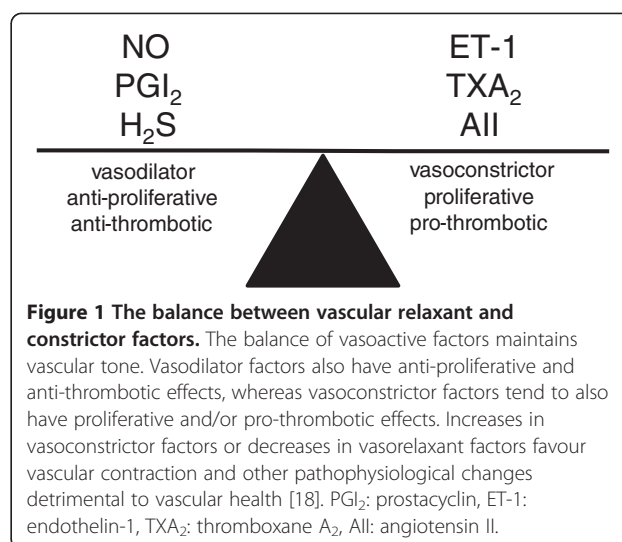


Figure 1 The balance between vascular relaxant and constrictor factors. The balance of vasoactive factors maintains vascular tone. Vasodilator factors also have anti-proliferative and anti-thrombotic effects, whereas vasoconstrictor factors tend to also have proliferative and/or pro-thrombotic effects. Increases in vasoconstrictor factors or decreases in vasorelaxant factors favour vascular contraction and other pathophysiological changes detrimental to vascular health [18]. PGI₂: prostacyclin, ET-1: endothelin-1, TXA₂: thromboxane A₂, All: angiotensin II.

Platelet inhibition

Limited data is available on the action of H₂S on platelets, although it has been reported that H₂S can decrease platelet aggregation [25]. A recent *in vitro* study showed that platelet adhesion to collagen and fibrinogen, the first step in platelet activation and aggregation, was significantly reduced by nanomolar concentrations of NaHS. Additionally, platelet superoxide production was also inhibited although the mechanism of this effect was not examined [26]. Whilst platelet adhesion and aggregation are important for vascular haemostasis in trauma, they are undesirable under conditions of vascular inflammation and atherosclerosis, so further investigation into the role of H₂S in platelet function is warranted.

H₂S as an anti-oxidant in the vasculature

Reactive oxygen species (ROS) can be divided into free radicals, such as superoxide (O₂^{•-}) and hydroxyl (OH[•]); non-radicals, such as hydrogen peroxide (H₂O₂); and reactive nitrogen species, such as NO (technically, NO[•], since it is a radical gas, with an unpaired electron) and peroxynitrite (ONOO⁻). In vascular cells, there are multiple sources for the generation of ROS, including mitochondria, cyclooxygenases and NADPH oxidases, xanthine oxidase, cyclo-oxygenase [27]. In mammalian tissues, reactive oxygen species (ROS) such as superoxide (O₂^{•-}) are produced under both pathological and physiological conditions. They are essential for the immunological defence mechanism of phagocytes, however, overproduction of ROS has detrimental effects on tissues including the vasculature. Excess ROS levels or oxidative stress are implicated in the pathology and progression of cardiovascular disease [28]. Excess levels of ROS can compromise the antioxidant defence mechanism of the cells and react with cellular macromolecules such as lipids, proteins, membrane bound polyunsaturated fatty acids and DNA leading

to irreversible cellular damage [29]. Furthermore, perhaps the best characterized mechanism by which oxidative stress can cause dysfunction and damage to vascular cells is via the scavenging of vasoprotective nitric oxide by O_2^- leading to a reduction its biological half-life [30].

Superoxide is the parent ROS molecule in all cells. It can be generated in vascular cells by NADPH oxidases (or "Nox oxidases"), uncoupled endothelial NO synthase (eNOS), the mitochondrial enzyme complexes, cytochrome P450 and xanthine oxidase [27]. The Nox oxidases are the only enzymes discovered to date that have the primary function of generating superoxide (Nox1-3) and hydrogen peroxide (Nox4). This family of enzymes comprises two membrane-bound subunits, the Nox catalytic subunit and p22phox as well as various combinations of cytoplasmic subunits [31]. In the aorta at least 3 isoforms of Nox oxidase are expressed, Nox1-, Nox2- and Nox4-containing Nox oxidases. Importantly, ROS are generated at low levels in cerebral vessels and act there as signalling molecules involved in vascular regulation [32]. Excessive production of ROS, in particular superoxide (O_2^-) from Nox oxidases is implicated as a key mediator of endothelial dysfunction (loss of NO bioavailability) associated with many cardiovascular diseases, including atherosclerosis, diabetic vascular disease and hypertension [33].

H₂S as a ROS scavenger

H₂S is a potent one-electron chemical reductant and nucleophile that is theoretically capable of scavenging free radicals by single electron or hydrogen atom transfer [14]. Thus, H₂S may participate in many reactions [34] and is reported to readily scavenge reactive oxygen and nitrogen species such as peroxyxynitrite [35], superoxide [36], hydrogen peroxide [37], hypochlorous acid [38] and lipid hydroperoxides [14]. However the kinetics, reactivity and mechanism of H₂S/HS⁻ interactions with ROS are poorly understood under physiological conditions [14]. H₂S has been reported to inhibit superoxide production in human endothelial cells [39] and vascular smooth muscle cells [40] by reducing Nox oxidase expression and activity. However it is not known if this activity is physiologically relevant, or whether H₂S can protect against oxidative-stress driven vascular dysfunction. In addition, H₂S is reported to increase glutathione levels and bolster endogenous anti-oxidant defences [41]. Collectively, these findings suggest that this molecule may be a useful vasoprotective agent.

H₂S as an inhibitor of ROS formation

H₂S has also been shown to be important in regulating mitochondrial function [42] and can reduce mitochondrial ROS formation [43]. Hyperglycaemia induced overproduction of ROS was reversed with H₂S treatment

and furthermore, endogenously produced H₂S acts to protect endothelial function from hyperglycaemic oxidative stress [44]. NaHS protects rat aortic smooth muscle cells from homocysteine-induced cytotoxicity and reactive oxygen species generation, and furthermore NaHS-induced protective effects were synergistic with endogenous anti-oxidants [36]. This study suggests that H₂S is capable of reducing production of H₂O₂, ONOO⁻ and O_2^- in a time and concentration dependent manner. The mechanism of this effect was not established, however H₂S at nanomolar concentrations has been reported to inhibit superoxide formation in human endothelial cells [39] and vascular smooth muscle cells [40] by reducing Nox oxidase expression and activity.

H₂S effects on endogenous anti-oxidants

NaHS has been shown to protect neurons from oxidative stress by boosting glutathione levels [41] and others have also shown that NaHS increases the activity of endogenous anti-oxidants such as superoxide dismutase, glutathione peroxidase and glutathione reductase [36,43,45,46]. There is now increasing evidence that H₂S has a role in regulating the nuclear factor erythroid 2 (NF-E2)-related factor 2 (Nrf2) pathway. Nrf2 is a key transcription regulator of inducible cell defence. In the presence of electrophiles and/or reactive oxygen species, Nrf2 accumulates, translocates to the cell nucleus and binds with antioxidant response elements (AREs). These are located within the promoter regions of an array of cell defence genes, regulating both basal and inducible expression of anti-oxidant proteins, detoxification enzymes and other stress response proteins [47].

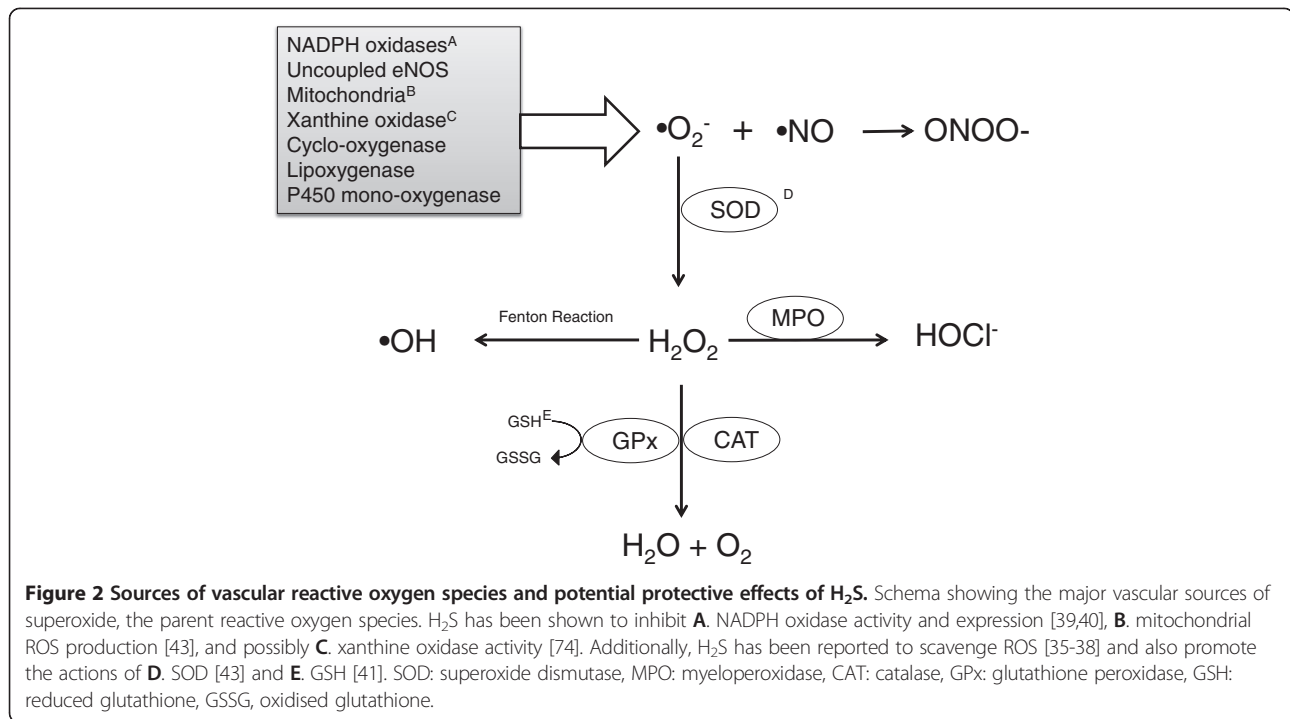
Recent studies have shown that H₂S donor treatment can induce Nrf2 expression [48,49] enhance Nrf2 translocation to the nucleus [50,51] and activate Nrf2 signalling [52], resulting in reduced oxidative stress and cardioprotection. The mechanism of the upregulation of Nrf2 by H₂S is under investigation with recent reports that H₂S inactivates the negative regulator of Nrf2, Keap1 [53,54] resulting in the Nrf2 mediated induction of cytoprotective genes.

Taken together, recent reports suggest that H₂S is capable of inhibiting the production of ROS, scavenging and neutralising ROS and boosting the efficacy of endogenous anti-oxidant molecules (Figure 2). The net effect is protection of vascular function and future work is needed to further examine the potential therapeutic benefits of the anti-oxidant effects of H₂S.

Studies in vascular disease states showing vasculoprotective effects of H₂S

Hypertension

Hypotensive effects of H₂S were first reported when administration of H₂S donors *in vivo* to anaesthetised rats was found to induce a transient hypotensive effect [55].



The CSE-L-cysteine pathway is downregulated in spontaneously hypertensive rats and treating them with a H₂S donor is protective, reducing blood pressure and vascular remodelling [56]. The most compelling evidence for the importance of H₂S in blood pressure regulation is that mice deficient in CSE develop endothelial dysfunction and hypertension within 8 weeks of birth and that H₂S replacement decreases systolic blood pressure in both CSE^{-/-} and CSE^{+/-} mice [8]. H₂S is also reported to regulate plasma renin levels [57] and inhibit angiotensin converting enzyme (ACE) activity in endothelial cells [58]. Inhibitory effects on ACE could also contribute to the anti-remodelling effects, which involve H₂S inhibition of collagen synthesis and smooth muscle proliferation in spontaneously hypertensive rats [59].

Angiogenesis

H₂S is implicated in the control of angiogenesis as NaHS treatment caused endothelial cell proliferation, adhesion, migration and tubule formation [60,61], with further work showing that vascular endothelial growth factor (VEGF) induced angiogenesis is mediated via H₂S [61] and that H₂S treatment *in vivo* increases collateral vessel growth, capillary density and blood flow in a hind-limb ischaemia model [62].

Atherosclerosis

Atherosclerosis is a chronic immune-inflammatory, fibro-proliferative disease caused by lipid accumulation, affecting large and medium-sized arteries [63]. Atherosclerosis is the

most common underlying cause in the development of coronary artery disease. It has a multifactorial pathogenesis, involving vascular inflammation, recruitment and infiltration of monocytes, differentiation of monocytes to foam cells. This leads to increased reactive oxygen species generation resulting in an impairment of vascular endothelial function, by reducing NO bioavailability [64]. Further accumulation of foam cells and vascular smooth muscle cell proliferation lead to the formation of vascular lesions or plaques, which disrupt blood flow and reduce vessel compliance. A number of studies have indicated that H₂S has many properties that may lead to the inhibition of atherogenesis (for review see [65]).

H₂S donors have been shown to reduce inflammatory mediators, an effect that is dose-dependent and also influenced by delivery of H₂S. Rapid delivery via NaHS is more likely to induce pro-inflammatory effects, whereas a more controlled delivery via the newer H₂S donor GYY4137 produces mostly anti-inflammatory effects [66]. H₂S treatment leads to decreased chemokine signalling [67] due to H₂S-donor dependent downregulation of macrophage CX3CR1 receptor expression, and CX3CR1-mediated chemotaxis [67]. NaHS inhibited leukocyte adhesion in mesenteric venules, and importantly, inhibiting CSE enhanced leukocyte adherence and infiltration [68]. NaHS treatment reduced ICAM-1 levels in ApoE^{-/-} mice [69]. This adhesion molecule participates in adhesion strengthening, monocyte spreading and transendothelial migration thus contributes to the infiltration of inflammatory cells into the vessel wall [70].

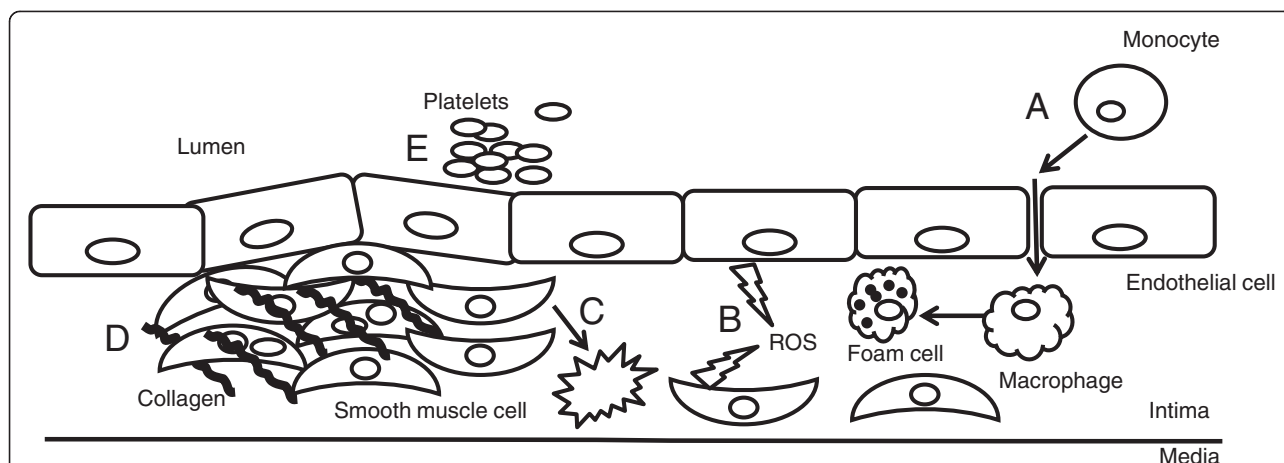


Figure 3 Potential sites of vasculoprotective effects of H₂S. Cartoon depicting a cross section of the vascular wall showing the endothelium, intima containing smooth muscle cells overlaying the vascular media. **A.** H₂S has been shown to decrease leukocyte adhesion and migration [60] and differentiation to foam cells [64]. **B.** H₂S can inhibit the production of ROS [39,40] as well as scavenge ROS [35-38], protecting endothelial function. **C.** H₂S prevents proliferation [66] and promotes apoptosis of vascular smooth muscle cells [67]. **D.** H₂S prevents collagen deposition [51] and neo-intima formation [65]. **E.** H₂S can inhibit platelet adhesion [26] and aggregation [25].

Once leukocytes have traversed the vessel wall the next stage in atherogenesis is foam cell formation. H₂S has been shown to inhibit hypochlorite induced atherogenic modification of purified LDL *in vitro* [71] and further studies have revealed that NaHS treatment inhibits macrophage expression of scavenger receptors (CD36 and scavenger receptor A) and acyl-coenzyme A:cholesterol acyltransferase-1, key proteins required for uptake of oxidized lipoproteins and subsequent cholesterol esterification required for foam cell production [72].

Administration of H₂S donors lead to a number of effects on vessel remodelling. In one study, CSE expression was reduced, and endogenous H₂S production decreased in blood vessels with balloon-injury induced neointima. The neointima formation was attenuated in animals treated with NaHS [73]. H₂S is known to cause inhibition of proliferation [74], and induction of apoptosis [75] in human aortic vascular smooth muscle cells, and reduce collagen deposition [59]. CSE over-expression in human embryonic kidney cells inhibits proliferation [76] and importantly, a recent study showed that CSE-deficient mice have increased neointima formation, that was reversed with NaHS treatment [77].

NaHS treatment of ApoE^{-/-} mice on a high fat diet reduced atherosclerotic lesion area [69]. NaHS treatment has been shown to inhibit vascular smooth muscle cell calcification in both cell culture [78] and in a rat model of vascular calcification [79]. Additionally, NaHS treatment in fat fed ApoE^{-/-} mice improved endothelial function and reduced vascular oxidative stress. Plasma H₂S levels are correlated with higher HDL and adiponectin levels and lower triglycerides and LDL/HDL ratio [80] in healthy human subjects, suggesting that increasing

sulfide consumption may have cardiovascular benefits. Overall H₂S has been shown to impede atherogenesis at all stages of the disease process (Figure 3). Taken together these effects all point towards an atheroprotective effect of endogenous H₂S, that is elicited by endogenous H₂S and that exogenous H₂S application may be a useful therapeutic strategy to prevent vascular remodelling.

Changes in expression of CSE in disease states

Altered expression of CSE and reduced endogenous H₂S are observed in inflammation [68], atherosclerosis [69], diabetes [81], hypertension [56] and treatment with H₂S donors has been repeatedly shown to be beneficial. The inverse relationship between plasma H₂S levels and vascular disease strongly suggests a role for endogenous H₂S in maintaining normal vascular functions.

Conclusions

The field of H₂S biology is new and exciting with regular reports of new developments in the literature. It is clearly an important mediator in the vascular system, contributing to vascular regulation and protection of cells from oxidative stress and the vascular injury that result from this and leads to vascular dysfunction. There is good evidence that H₂S donor treatment has potential as a vasculoprotective agent for the prevention and reversal of cell damage that is implicit in many vascular disease states.

Abbreviations

CBS: Cystathionine-β-synthase; CSE: Cystathionine-γ-lyase; MST: 3-mercaptopyruvate sulfurtransferase; PGI₂: Prostacyclin; ET-1: Endothelin-1; Ang: Angiotensin II; EDHF: Endothelium-derived hyperpolarising factor; NADPH: Nicotinamide adenine dinucleotide phosphate; Nox: NADPH

oxidase; ROS: Reactive oxygen species; SOD: Superoxide dismutase; CAT: Catalase; MPO: Myeloperoxidase; GPx: Glutathione peroxidase; GSH: Reduced glutathione; GSSG: Oxidized glutathione; ACE: Angiotensin converting enzyme; VEGF: Vascular endothelial growth factor; LDL: Low density lipoprotein; HDL: High density lipoprotein.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

JH, HN and ES wrote the manuscript. All authors have read and approved the final manuscript.

Received: 11 February 2013 Accepted: 24 April 2013

Published: 29 April 2013

References

1. Wang R: Physiological implications of hydrogen sulfide: a whiff exploration that blossomed. *Physiol Rev* 2012, **92**:791–896.
2. Moody BF, Calvert JW: Emergent role of gasotransmitters in ischemia-reperfusion injury. *Med Gas Res* 2011, **1**:3.
3. Kimura H: Hydrogen sulfide: its production and functions. *Exp Physiol* 2011, **96**:833–835.
4. Renga B: Hydrogen sulfide generation in mammals: the molecular biology of cystathionine-beta- synthase (CBS) and cystathionine-gamma-lyase (CSE). *Inflamm Allergy Drug Targets* 2011, **10**:85–91.
5. Shibuya N, Mikami Y, Kimura Y, Nagahara N, Kimura H: Vascular endothelium expresses 3-mercaptopyruvate sulfurtransferase and produces hydrogen sulfide. *J Biochem* 2009, **146**:623–626.
6. Shibuya N, Tanaka M, Yoshida M, Ogasawara Y, Togawa T, Ishii K, Kimura H: 3-Mercaptopyruvate sulfurtransferase produces hydrogen sulfide and bound sulfane sulfur in the brain. *Antioxid Redox Signal* 2009, **11**:703–714.
7. Streeter E, Hart J, Badoer E: An investigation of the mechanisms of hydrogen sulfide-induced vasorelaxation in rat middle cerebral arteries. *Naunyn Schmiedebergs Arch Pharmacol* 2012, **385**:991–1002.
8. Yang G, Wu L, Jiang B, Yang W, Qi J, Cao K, Meng Q, Mustafa AK, Mu W, Zhang S, et al: H₂S as a physiologic vasorelaxant: hypertension in mice with deletion of cystathionine gamma-lyase. *Science* 2008, **322**:587–590.
9. Kimura H: Metabolic turnover of hydrogen sulfide. *Front Physiol* 2012, **3**:101.
10. Furne J, Saeed A, Levitt MD: Whole tissue hydrogen sulfide concentrations are orders of magnitude lower than presently accepted values. *Am J Physiol Regul Integr Comp Physiol* 2008, **295**:R1479–R1485.
11. Ogasawara Y, Isoda S, Tanabe S: Tissue and subcellular distribution of bound and acid-labile sulfur, and the enzymic capacity for sulfide production in the rat. *Biol Pharm Bull* 1994, **17**:1535–1542.
12. Ishigami M, Hiraki K, Umemura K, Ogasawara Y, Ishii K, Kimura H: A source of hydrogen sulfide and a mechanism of its release in the brain. *Antioxid Redox Signal* 2009, **11**:205–214.
13. Olson KR: Is hydrogen sulfide a circulating "gasotransmitter" in vertebrate blood? *Biochim Biophys Acta* 2009, **1787**:856–863.
14. Carballal S, Trujillo M, Cuevasanta E, Bartsaghi S, Moller MN, Folkes LK, Garcia-Bereguain MA, Gutierrez-Merino C, Wardman P, Denicola A, et al: Reactivity of hydrogen sulfide with peroxynitrite and other oxidants of biological interest. *Free Radic Biol Med* 2010, **50**:196–205.
15. King SB: Potential biological chemistry of hydroge sulfide (H₂S) with the nitrogen oxides. *Free Radic Biol Med* 2013, **55**:1–7.
16. Nagy P, Winterbourn CC: Rapid reaction of hydrogen sulfide with the neutrophil oxidant hypochlorous acid to generate polysulfides. *Chem Res Toxicol* 2010, **23**:1541–1543.
17. Paul BD, Snyder SH: H₂(S) signalling through protein sulfhydration and beyond. *Nat Rev Mol Cell Biol* 2012, **13**:499–507.
18. Triggler CR, Samuel SM, Ravishanker S, Marei I, Arunachalam G, Ding H: The endothelium: influencing vascular smooth muscle in many ways. *Can J Physiol Pharmacol* 2012, **90**:713–738.
19. Al-Magableh MR, Hart JL: Mechanism of vasorelaxation and role of endogenous hydrogen sulfide production in mouse aorta. *Naunyn Schmiedebergs Arch Pharmacol* 2011, **383**:403–413.
20. Cheang WS, Wong WT, Shen B, Lau CW, Tian XY, Tsang SY, Yao X, Chen ZY, Huang Y: 4-aminopyridine-sensitive K⁺ channels contributes to NaHS-induced membrane hyperpolarization and relaxation in the rat coronary artery. *Vascular Pharmacol* 2010, **53**:94–98.
21. Lee SW, Cheng Y, Moore PK, Bian JS: Hydrogen sulphide regulates intracellular pH in vascular smooth muscle cells. *Biochem Biophys Res Commun* 2007, **358**:1142–1147.
22. Kiss L, Deitch EA, Szabo C: Hydrogen sulfide decreases adenosine triphosphate levels in aortic rings and leads to vasorelaxation via metabolic inhibition. *Life Sci* 2008, **83**:589–594.
23. Zhao W, Wang R: H₂(S)-induced vasorelaxation and underlying cellular and molecular mechanisms. *Am J Physiol Heart Circ Physiol* 2002, **283**:H474–H480.
24. Cheng Y, Ndisang J, Tang G, Cao K, Wang R: Hydrogen sulfide-induced relaxation of resistance mesenteric artery beds of rats. *Am J Physiol Heart Circ Physiol* 2004, **287**:2316–2323.
25. Zagli G, Patacchini R, Trevisani M, Abbate R, Cinotti S, Gensini GF, Masotti G, Geppetti P: Hydrogen sulfide inhibits human platelet aggregation. *Eur J Pharmacol* 2007, **559**:65–68.
26. Morel A, Malinowska J, Olas B: Antioxidative properties of hydrogen sulfide may involve in its antiadhesive action on blood platelets. *Clin Biochem* 2012, **45**:1678–1682.
27. Land WG: Emerging role of innate immunity in organ transplantation: part I: evolution of innate immunity and oxidative allograft injury. *Transplant Rev (Orlando)* 2012, **26**:60–72.
28. Forstermann U: Oxidative stress in vascular disease: causes, defense mechanisms and potential therapies. *Nat Clin Pract Cardiovasc Med* 2008, **5**:338–349.
29. Chen AF, Chen DD, Daiber A, Faraci FM, Li H, Rembold CM, Laher I: Free radical biology of the cardiovascular system. *Clin Sci (Lond)* 2012, **123**:73–91.
30. MacKenzie A, Martin W: Loss of endothelium-derived nitric oxide in rabbit aorta by oxidant stress: restoration by superoxide dismutase mimetics. *Br J Pharmacol* 1998, **124**:719–728.
31. Drummond GR, Selemidis S, Griendling KK, Sobey CG: Combating oxidative stress in vascular disease: NADPH oxidases as therapeutic targets. *Nat Rev Drug Discov* 2011, **10**:453–471.
32. Miller AA, Drummond GR, Sobey CG: Reactive oxygen species in the cerebral circulation: are they all bad? *Antioxid Redox Signal* 2006, **8**:1113–1120.
33. Brandes RP, Weissmann N, Schroder K: NADPH oxidases in cardiovascular disease. *Free Radic Biol Med* 2010, **49**:687–706.
34. Stasko A, Brezova V, Zalibera M, Biskupic S, Ondrias K: Electron transfer: a primary step in the reactions of sodium hydrosulphide, an H₂S/H₂(–) donor. *Free Radic Res* 2009, **43**:581–593.
35. Whiteman M, Armstrong J, Chu S, Jia-Ling S, Wong B, Cheung N, Halliwell B, Moore P: The novel neuromodulator hydrogen sulfide: an endogenous peroxynitrite 'scavenger'? *J Neurochem* 2004, **90**:765–768.
36. Yan SK, Chang T, Wang H, Wu L, Wang R, Meng QH: Effects of hydrogen sulfide on homocysteine-induced oxidative stress in vascular smooth muscle cells. *Biochem Biophys Res Commun* 2006, **351**:485–491.
37. Lu M, Hu LF, Hu G, Bian JS: Hydrogen sulfide protects astrocytes against H₂O₂-induced neural injury via enhancing glutamate uptake. *Free Radic Biol Med* 2008, **45**:1705–1713.
38. Whiteman M, Cheung N, Zhu Y, Chu S, Siau J, Wong B, Armstrong J, Moore P: Hydrogen sulphide: a novel inhibitor of hypochlorous acid-mediated oxidative damage in the brain? *Biochem Biophys Res Commun* 2005, **343**:303–310.
39. Muzaffar S, Jeremy JY, Sparatore A, Del Soldato P, Angelini GD, Shukla N: H₂S-donating sildenafil (ACS6) inhibits superoxide formation and gp91phox expression in arterial endothelial cells: role of protein kinases A and G. *Br J Pharmacol* 2008, **155**:984–994.
40. Muzaffar S, Shukla N, Bond M, Newby AC, Angelini GD, Sparatore A, Del Soldato P, Jeremy JY: Exogenous hydrogen sulfide inhibits superoxide formation, NOX-1 expression and Rac1 activity in human vascular smooth muscle cells. *J Vasc Res* 2008, **45**:521–528.
41. Kimura Y, Goto Y, Kimura H: Hydrogen sulfide increases glutathione production and suppresses oxidative stress in mitochondria. *Antioxid Redox Signal* 2010, **12**:1–13.
42. Modis K, Coletta C, Erdelyi K, Papapetropoulos A, Szabo C: Intramitochondrial hydrogen sulfide production by 3-mercaptopyruvate sulfurtransferase maintains mitochondrial electron flow and supports cellular bioenergetics. *FASEB J* 2012, **27**:601–611.
43. Sun WH, Liu F, Chen Y, Zhu YC: Hydrogen sulfide decreases the levels of ROS by inhibiting mitochondrial complex IV and increasing SOD activities in cardiomyocytes under ischemia/reperfusion. *Biochem Biophys Res Commun* 2012, **421**:164–169.

44. Suzuki K, Olah G, Modis K, Coletta C, Kulp G, Gero D, Szoleczky P, Chang T, Zhou Z, Wu L, et al: **Hydrogen sulfide replacement therapy protects the vascular endothelium in hyperglycemia by preserving mitochondrial function.** *Proc Natl Acad Sci USA* 2011, **108**:13829–13834.
45. Rossoni G, Sparatore A, Tazzari V, Manfredi B, Del Soldato P, Berti F: **The hydrogen sulphide-releasing derivative of diclofenac protects against ischaemia-reperfusion injury in the isolated rabbit heart.** *Br J Pharmacol* 2008, **153**:100–109.
46. Benetti LR, Campos D, Gurgueira SA, Vercesi AE, Guedes CE, Santos KL, Wallace JL, Teixeira SA, Florenzano J, Costa SK, et al: **Hydrogen sulfide inhibits oxidative stress in lungs from allergic mice in vivo.** *Eur J Pharmacol* 2013, **698**:463–469.
47. Copley IM: **The Keap1–Nrf2 cell defense pathway—a promising therapeutic target?** *Adv Pharmacol* 2012, **63**:43–79.
48. Han W, Dong Z, Dimitropoulou C, Su Y: **Hydrogen sulfide ameliorates tobacco smoke-induced oxidative stress and emphysema in mice.** *Antioxid Redox Signal* 2011, **15**:2121–2134.
49. Ganster F, Burban M, de la Bourdonnaye M, Fizanet L, Douay O, Loufrani L, Mercat A, Cales P, Radermacher P, Henrion D, et al: **Effects of hydrogen sulfide on hemodynamics, inflammatory response and oxidative stress during resuscitated hemorrhagic shock in rats.** *Crit Care* 2010, **14**:R165.
50. Calvert JW, Elston M, Nicholson CK, Gundewar S, Jha S, Elrod JW, Ramachandran A, Lefer DJ: **Genetic and pharmacologic hydrogen sulfide therapy attenuates ischemia-induced heart failure in mice.** *Circulation* 2010, **122**:11–19.
51. Li HD, Zhang ZR, Zhang QX, Qin ZC, He DM, Chen JS: **Treatment with exogenous hydrogen sulfide attenuates hyperoxia-induced acute lung injury in mice.** *Eur J Appl Physiol* 2013. in press.
52. Peake BF, Nicholson CK, Lambert JP, Hood RL, Amin H, Amin S, Calvert JW: **Hydrogen sulfide preconditions the db/db diabetic mouse heart against ischemia-reperfusion injury by activating Nrf2 signaling in an Erk-dependent manner.** *Am J Physiol Heart Circ Physiol* 2013. in press.
53. Yang G, Zhao K, Ju Y, Mani S, Cao Q, Puukila S, Khaper N, Wu L, Wang R: **Hydrogen sulfide protects against cellular senescence via S-sulfhydration of Keap1 and activation of Nrf2.** *Antioxid Redox Signal* 2013, **18**:1906–1919.
54. Hourihan JM, Kenna JG, Hayes JD: **The gasotransmitter hydrogen sulfide induces Nrf2-target genes by inactivating the Keap1 ubiquitin ligase substrate adaptor through formation of a disulfide bond between Cys-226 and Cys-613.** *Antioxid Redox Signal* 2013. in press.
55. Zhao W, Zhang J, Lu Y, Wang R: **The vasorelaxant effect of H(2)S as a novel endogenous gaseous K(ATP) channel opener.** *EMBO J* 2001, **20**:6008–6016.
56. Yan H, Du J, Tang C: **The possible role of hydrogen sulfide on the pathogenesis of spontaneous hypertension in rats.** *Biochem Biophys Res Commun* 2004, **313**:22–27.
57. Lu M, Liu YH, Goh HS, Wang JJ, Yong QC, Wang R, Bian JS: **Hydrogen sulfide inhibits plasma Renin activity.** *J Am Soc Nephrol* 2010, **21**:993–1002.
58. Laggner H, Hermann M, Esterbauer H, Muellner MK, Exner M, Gmeiner BM, Kapiotis S: **The novel gaseous vasorelaxant hydrogen sulfide inhibits angiotensin-converting enzyme activity of endothelial cells.** *J Hypertens* 2007, **25**:2100–2104.
59. Zhao X, Zhang LK, Zhang CY, Zeng XJ, Yan H, Jin HF, Tang CS, Du JB: **Regulatory effect of hydrogen sulfide on vascular collagen content in spontaneously hypertensive rats.** *Hypertens Res* 2008, **31**:1619–1630.
60. Cai WJ, Wang MJ, Moore PK, Jin HM, Yao T, Zhu YC: **The novel proangiogenic effect of hydrogen sulfide is dependent on Akt phosphorylation.** *Cardiovasc Res* 2007, **76**:29–40.
61. Papapetropoulos A, Pyriochou A, Altaany Z, Yang G, Marazioti A, Zhou Z, Jeschke MG, Branski LK, Herndon DN, Wang R, Szabo C: **Hydrogen sulfide is an endogenous stimulator of angiogenesis.** *Proc Natl Acad Sci USA* 2009, **106**:21972–21977.
62. Wang MJ, Cai WJ, Li N, Ding YJ, Chen Y, Zhu YC: **The hydrogen sulfide donor NaHS promotes angiogenesis in a rat model of hind limb ischemia.** *Antioxid Redox Signal* 2010, **12**:1065–1077.
63. Weber C, Noels H: **Atherosclerosis: current pathogenesis and therapeutic options.** *Nat Med* 2011, **17**:1410–1422.
64. Judkins CP, Diep H, Broughton BR, Mast AE, Hooker EU, Miller AA, Selemidis S, Dusting GJ, Sobey CG, Drummond GR: **Direct evidence of a role for Nox2 in superoxide production, reduced nitric oxide bioavailability, and early atherosclerotic plaque formation in ApoE–/– mice.** *Am J Physiol Heart Circ Physiol* 2010, **298**:H24–H32.
65. Lynn EG, Austin RC: **Hydrogen sulfide in the pathogenesis of atherosclerosis and its therapeutic potential.** *Expert Rev Clin Pharmacol* 2011, **4**:97–108.
66. Whiteman M, Li L, Rose P, Tan CH, Parkinson D, Moore P: **The effect of hydrogen sulfide donors on lipopolysaccharide-induced formation of inflammatory mediators in macrophages.** *Antioxid Redox Signal* 2010, **12**(10):1147–54.
67. Zhang H, Guo C, Wu D, Zhang A, Gu T, Wang L, Wang C: **Hydrogen sulfide inhibits the development of atherosclerosis with suppressing CX3CR1 and CX3CL1 expression.** *PLoS One* 2012, **7**:e41147.
68. Zanardo RC, Brancaleone V, Distrutti E, Fiorucci S, Cirino G, Wallace JL: **Hydrogen sulfide is an endogenous modulator of leukocyte-mediated inflammation.** *FASEB J* 2006, **20**:2118–2120.
69. Wang Y, Zhao X, Jin H, Wei H, Li W, Bu D, Tang X, Ren Y, Tang C, Du J: **Role of hydrogen sulfide in the development of atherosclerotic lesions in apolipoprotein E knockout mice.** *Arterioscler Thromb Vasc Biol* 2009, **29**:173–179.
70. Mestas J, Ley K: **Monocyte-endothelial cell interactions in the development of atherosclerosis.** *Trends Cardiovasc Med* 2008, **18**:228–232.
71. Laggner H, Muellner MK, Schreier S, Sturm B, Hermann M, Exner M, Gmeiner BM, Kapiotis S: **Hydrogen sulphide: a novel physiological inhibitor of LDL atherogenic modification by HOCl.** *Free Radic Res* 2007, **41**:741–747.
72. Zhao ZZ, Wang Z, Li GH, Wang R, Tan JM, Cao X, Suo R, Jiang ZS: **Hydrogen sulfide inhibits macrophage-derived foam cell formation.** *Exp Biol Med (Maywood)* 2011, **236**:169–176.
73. Meng Q, Yang G, Yang W, Jiang B, Wu L, Wang R: **Protective effect of hydrogen sulfide on balloon injury-induced neointima hyperplasia in rat carotid arteries.** *Am J Pathol* 2007, **170**:1406–1414.
74. Du J, Hui Y, Cheung Y, Bin G, Jiang H, Chen X, Tang C: **The possible role of hydrogen sulfide as a smooth muscle cell proliferation inhibitor in rat cultured cells.** *Hear Vessel* 2004, **19**:75–80.
75. Yang G, Sun X, Wang R: **Hydrogen sulfide-induced apoptosis of human aorta smooth muscle cells via the activation of mitogen-activated protein kinases and caspase-3.** *FASEB J* 2004, **18**:1782–1784.
76. Yang G, Cao K, Wu L, Wang R: **Cystathionine gamma-lyase overexpression inhibits cell proliferation via a H2S-dependent modulation of ERK1/2 phosphorylation and p21Cip/WAK-1.** *J Biol Chem* 2004, **279**:49199–49205.
77. Yang G, Li H, Tang G, Wu L, Zhao K, Cao Q, Xu C, Wang R: **Increased neointimal formation in cystathionine gamma-lyase deficient mice: role of hydrogen sulfide in alpha5beta1-integrin and matrix metalloproteinase-2 expression in smooth muscle cells.** *J Mol Cell Cardiol* 2012, **52**:677–688.
78. Zavadzki E, Jeney V, Agarwal A, Zarjou A, Oros M, Katko M, Varga Z, Balla G, Balla J: **Hydrogen sulfide inhibits the calcification and osteoblastic differentiation of vascular smooth muscle cells.** *Kidney Int* 2011, **80**:731–739.
79. Wu SY, Pan CS, Geng B, Zhao J, Yu F, Pang YZ, Tang CS, Qi YF: **Hydrogen sulfide ameliorates vascular calcification induced by vitamin D3 plus nicotinic in rats.** *Acta Pharmacol Sin* 2006, **27**:299–306.
80. Jain SK, Micinski D, Lieblong BJ, Stapleton T: **Relationship between hydrogen sulfide levels and HDL-cholesterol, adiponectin, and potassium levels in the blood of healthy subjects.** *Atherosclerosis* 2012, **225**:242–245.
81. Jain SK, Bull R, Rains JL, Bass PF, Levine SN, Reddy S, McVie R, Bocchini JA: **Low levels of hydrogen sulfide in the blood of diabetes patients and streptozotocin-treated rats causes vascular inflammation?** *Antioxid Redox Signal* 2010, **12**:1333–1337.

doi:10.1186/2045-9912-3-9

Cite this article as: Streeter et al: Hydrogen sulfide as a vasculoprotective factor. *Medical Gas Research* 2013 3:9.