

# Complete Genome Sequence of a European Genotype Porcine Reproductive and Respiratory Syndrome Virus in China

Zhi Zhou,<sup>a</sup> Qi Liu,<sup>a</sup> Dongmei Hu,<sup>a</sup> Xiaojuan Yue,<sup>a</sup> Xiuling Yu,<sup>a</sup> Qian Zhang,<sup>a</sup> Xiaoxue Gu,<sup>a</sup> Jianqiang Ni,<sup>a</sup> Xiangdong Li,<sup>b</sup> Xinyan Zhai,<sup>a</sup> Kegong Tian<sup>a</sup>

Veterinary Diagnosis Center and OIE Porcine Reproductive and Respiratory Syndrome Laboratory, China Animal Disease Control Center, Chaoyang District, Beijing, People's Republic of China<sup>a</sup>; Department of Anatomy and Physiology, College of Veterinary Medicine, Kansas State University, Manhattan, Kansas, USA<sup>b</sup>

**Here, we report a novel European genotype porcine reproductive and respiratory syndrome virus (PRRSV) strain with 15 nucleotide deletions in the nonstructural protein 2 region and 3 nucleotide deletions in the overlapping regions of the open reading frame 3 (ORF3) and ORF4 regions. This study will aid in further exploration of the genetic and antigenic diversity of the European genotype of the PRRSV in China.**

Received 10 March 2013 Accepted 22 March 2013 Published 9 May 2013

**Citation** Zhou Z, Liu Q, Hu D, Yue X, Yu X, Zhang Q, Gu X, Ni J, Li X, Zhai X, Tian K. 2013. Complete genome sequence of a European genotype porcine reproductive and respiratory syndrome virus in China. *Genome Announc.* 1(3):e00175-13. doi:10.1128/genomeA.00175-13.

**Copyright** © 2013 Zhou et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Kegong Tian, tiankg@263.net, or Xinyan Zhai, zhaixy2010@sina.cn.

Porcine reproductive and respiratory syndrome virus (PRRSV), a member of the *Arteriviridae* family in the order *Nidovirales*, is an enveloped, positive-sense, single-stranded RNA virus that causes reproductive failure in pregnant sows and respiratory distress in young pigs (1, 2). PRRS occurs in most pig-producing countries worldwide and causes enormous economic losses in the swine industry (3). A significant characteristic of PRRSV is the extensive genetic variation within the different isolates in the field (4). Based on genetic and antigenic differences, the PRRSVs have been divided into two major genotypes, the European genotype (EU) (type 1, prototype Lelystad virus) and the North American genotype (NA) (type 2, prototype VR-2332). The two genotypes share only about 60% identity on the nucleotide level (2, 5). In China, the North American genotype PRRSV and the European genotype PRRSV were first reported in 1996 and 2006 (6, 7), respectively. To further explore the genetic diversity of the European genotype PRRSV in China, here we report the complete genome sequence of the European genotype PRRSV NVDC-NM1-2011.

The NVDC-NM1-2011 PRRSV was isolated from naturally infected swine herds in 2011. The virus total RNA was extracted from sera of infected swine, and the complete genome of NVDC-NM1-2011 was generated with reverse transcriptase PCR (RT-PCR) using 16 pairs of primers amplifying 16 overlapped fragments of PRRSV (6). The 16 amplified RT-PCR products were purified and cloned into a pGEM-T Easy vector (Promega) and sequenced with an automated sequencer (Genetic analyzer 3730XL; Applied Biosystems). The complete genomic sequence of NVDC-NM1-2011 is 15,081 nucleotides (nt), excluding the 3' poly(A) tail. Comparison of NVDC-NM1-2011 with the EU PRRSV prototypic strain Lelystad virus by whole-genome BLAST techniques showed that there are 18-nt discontinuous deletions, which are 15-nt deletions in the Nsp2 region (12-nt deletions at positions 2224 to 2235 and 3-nt deletions at positions 2374 to 2376) and 3-nt deletions within the 252-nt overlapping regions of

the open reading frame 3 (ORF3) and ORF4 regions (at positions 202 to 204). Previous studies have indicated the nonstructural Nsp2, ORF3, and ORF5 sequences are most variable among the different strains within the genome of PRRSV, which are likely to be associated with the virulence of PRRSV (8–10). In the future, investigations should be undertaken to determine whether these mutations of the NVDC-NM1-2011 isolate will lead to pathogenic changes in swine. In addition, PRRSV shares 90.77% nucleotide identity with the genome of the prototype Lelystad virus.

In all, the sequence data of NVDC-NM1-2011 indicate that the EU genotype PRRSV variant prevails in China. This information will aid in further exploration of the genetic and antigenic diversity of the European genotype of PRRSV in China.

**Nucleotide sequence accession number.** The complete genome sequence of NVDC-NM1-2011 is available in GenBank under the accession number [JX187609](https://www.ncbi.nlm.nih.gov/nuccore/JX187609).

## ACKNOWLEDGMENTS

This work was supported by grants from the National Basic Research Program of China (grant 2008FY130100-2) and the modern agricultural technology and innovation program of Beijing (grant GWZJ-2009-05).

## REFERENCES

1. Benfield DA, Nelson E, Collins JE, Harris L, Goyal SM, Robison D, Christianson WT, Morrison RB, Gorcyca D, Chladek D. 1992. Characterization of swine infertility and respiratory syndrome (SIRS) virus (isolate ATCC VR-2332). *J. Vet. Diagn. Invest.* 4:127–133.
2. Meulenbergh JJ, Hulst MM, de Meijer EJ, Moonen PL, den Besten A, de Kluyver EP, Wensvoort G, Moormann RJ. 1993. Lelystad virus, the causative agent of porcine epidemic abortion and respiratory syndrome (PEARS), is related to LDV and EAV. *Virology* 192:62–72.
3. Neumann EJ, Kliebenstein JB, Johnson CD, Mabry JW, Bush EJ, Seitzinger AH, Green AL, Zimmerman JJ. 2005. Assessment of the economic impact of porcine reproductive and respiratory syndrome on swine production in the United States. *J. Am. Vet. Med. Assoc.* 227:385–392.
4. Meng XJ. 2000. Heterogeneity of porcine reproductive and respiratory

- syndrome virus: implications for current vaccine efficacy and future vaccine development. *Vet. Microbiol.* 74:309–329.
5. Nelsen CJ, Murtaugh MP, Faaberg KS. 1999. Porcine reproductive and respiratory syndrome virus comparison: divergent evolution on two continents. *J. Virol.* 73:270–280.
  6. Chen N, Cao Z, Yu X, Deng X, Zhao T, Wang L, Liu Q, Li X, Tian K. 2011. Emergence of novel European genotype porcine reproductive and respiratory syndrome virus in mainland China. *J. Gen. Virol.* 92:880–892.
  7. Guo BQ, Chen ZS, Liu WX, Cui YZ. 1996. Isolation and identification of porcine reproductive and respiratory syndrome (PRRS) virus from aborted fetuses suspected of PRRS. *Chin. J. Anim. Poult. Infect. Dis.* 26:1–5.
  8. Fang Y, Kim DY, Ropp S, Steen P, Christopher-Hennings J, Nelson EA, Rowland RR. 2004. Heterogeneity in Nsp2 of European-like porcine reproductive and respiratory syndrome viruses isolated in the United States. *Virus Res.* 100:229–235.
  9. Music N, Gagnon CA. 2010. The role of porcine reproductive and respiratory syndrome (PRRS) virus structural and non-structural proteins in virus pathogenesis. *Anim. Health Res. Rev.* 14:1–29.
  10. Ostrowski M, Galeota JA, Jar AM, Platt KB, Osorio FA, Lopez OJ. 2002. Identification of neutralizing and nonneutralizing epitopes in the porcine reproductive and respiratory syndrome virus GP5 ectodomain. *J. Virol.* 76:4241–4250.