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Nontuberculous Mycobacterial Lung Disease Caused by *Mycobacterium lentiflavum* in a Patient with Bronchiectasis

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We report a rare case of lung disease caused by *Mycobacterium lentiflavum* in a previously healthy woman. A 54-year-old woman was referred to our hospital due to chronic cough and sputum, A computed tomography scan of the chest revealed bilateral bronchiectasis with bronchiolitis in the right middle lobe and the lingular division of the left upper lobe. Nontuberculous mycobacteria were isolated twice from three expectorated sputum specimens. All isolates were identified as *M. lentiflavum* by multilocus sequence analysis based on *rpoB*, *hsp65*, and 16S rRNA fragments. To the best of our knowledge, this is the first documented case of *M. lentiflavum* lung disease in an immunocompetent adult in Korea.

Key Words: Nontuberculous Mycobacteria; Bronchiectasis; Mycobacterium Infections, Nontuberculous

Introduction

Mycobacterium lentiflavum organisms are slow growing, tiny yellow nontuberculous mycobacteria (NTM) first identified in 1996¹. As with other NTM, M. lentiflavum has been isolated from soil and water samples around world². It is a rare cause of human disease and has been reported to have clinical importance primarily in young children with cervical lymphadenitis and in immunocompromised patients³⁻⁵. Recently, however, a few cases of chronic pulmonary disease caused by M.

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lentiflavum in immunocompetent patients have been reported^{2,6-8}.

Although the isolation of *M. lentiflavum* from clinical specimens has been reported in Korea, the infectious organisms seemed to be colonizers in the destructed lung due to previous tuberculosis⁹. Here, we report a very rare case of *M. lentiflavum* lung disease associated with bronchiectasis in an immunocompetent patient.

Case Report

A 54-year-old woman was referred to our hospital due to chronic cough and sputum, which developed three years earlier. She had been a healthy, non-smoker up to this point, with the exception of pulmonary tuberculosis 10 years prior. Laboratory results, including immunoglobulin levels, were normal with the exception of an elevated erythrocyte sedimentation rate to 82 mm/hr. She had no risk factors for human immunodeficiency virus infection.

A chest radiography revealed bilateral multifocal tram-track signs. A computed tomography scan of the

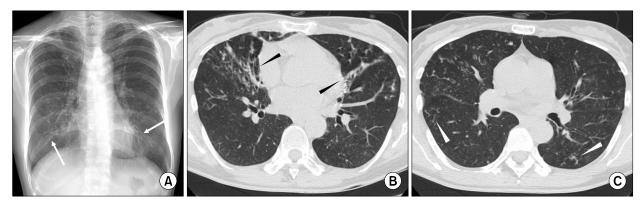


Figure 1. A 54-year-old woman with bronchiectasis and nontuberculous mycobacterial lung disease caused by *Mycobacterium lentiflavum*. (A) A chest radiography reveals bilateral multifocal tram-track signs (white arrows). (B) A transverse chest computed tomography (CT) scan (2,5-mm-section thickness) on the level with the right inferior pulmonary vein reveals bilateral bronchiectasis (black arrowheads) in the right middle lobe and the lingular segment of the left upper lobe. (C) A chest CT scan obtained on the level with the superior segmental bronchus of the left lower lobe reveals bilateral bronchiolitis in both lungs (white arrowheads).

chest revealed bilateral bronchiectasis and bronchiolitis in the right middle lobe and the lingular division of the left upper lobe (Figure 1). There was no evidence of cystic fibrosis or other common causes of bronchiectasis. NTM were isolated twice from three consecutively expectorated sputum specimens.

Initial identification of the isolated NTM was unsuccessful using the polymerase chain reaction (PCR)reverse blot hybridization assay (REBA) method based on the rpoB gene (REBA Myco-ID; M&D Inc., Wonju, Korea). No specific species could be assigned using this method due to a lack of reference sequences. Therefore, we used multilocus sequence analysis (MLSA) based on hsp65, rpoB, and 16S rRNA fragments to determine their taxonomic affiliations. Genomic DNA of the isolates was extracted using a commercially available kit (QIAamp DNA Mini kit; Qiagen, Hilden, Germany) and amplified using the appropriate pair of primers for each of the three genes. PCR conditions and primer sequences are described below. Partial amplification of the rpoB gene (723 bp) was performed using primers MycoF (5'-GGCAAGGTCACCCCGAAGGG-3') and MycoR (5'-AGCGGCTGCTGGGTGATCATC-3')¹⁰. Partial amplification of the hsp65 gene (439 bp) was performed using primers Tb11 (5'-ACCAACGATGGTG-TGTCCAT-3') and Tb12 (5'-CTTGTCGAACCGCATAC-

CCT-31)11. PCR conditions for these genes were as follows: 2 minutes at 95°C followed by 35 cycles of 94°C for 30 seconds, 60°C for 30 seconds, and 72°C for 2 minutes, with a final extension step at 72°C for 5 minutes. A 564 bp 5'-region of the 16S rRNA gene was amplified using primers 16MycF (5'-CGTGCTTAACACATGCAAG-TCG-3') and 16MycR (5'-GTGAGATTTCACGAACAA-CGC-3')¹². Conditions for 16S rRNA gene amplification were as follows: 2 minutes at 95°C followed by 35 cycles of 94°C for 30 seconds, 52°C for 30 seconds, and 72°C for 1 minute. The amplicons were cloned into the pGEM-TEasy vector (Promega, Madison, WI, USA), transformed into DH5 \alpha Escherichia coli and amplified (Invitrogen, Karlsruhe, Germany). Clones were sequenced using the ABI Prism d-Rhodamine dye terminator cycle sequencing ready reaction kit (Applied Biosystems, Foster City, CA, USA). Sequencing reactions were recorded with an ABI Prism 3100 DNA sequencer following the standard protocol of the supplier (Applied Biosystems). The sequences were compared by referring to publically available sequence in the National Center for Biotechnology Information GenBank database¹³ and the Ribosomal Differentiation of Medical Micro-organisms (RIDOM) database¹⁴. Homogeneity values above 99% were considered significant. M. lentiflavum was subsequently identified as the most closely

related species for which the sequences of the three genes were available in GenBank (accession nos. EU109300 for *rpoB*, AF547851 for *hsp65*, and GU142924 for 16S rRNA).

Five months after initial sputum cultures, the patient continued to have a mild cough and sputum. Additional sputum culture 4 months after the initial sputum examinations was negative. She was reluctant to receive long-term antibiotic therapy and refused further follow-up. Therefore, the long-term outcome of this patient is not available.

Discussion

NTM are a well-known cause of chronic pulmonary disease. Among the most commonly isolated species are the *M. avium* complex, *M. abscessus*, and *M. kansasii*, but other NTM species have also been described as the cause of lung disease. To the best of our knowledge, this is the first documented case of *M. lentiflavum* lung disease in an immunocompetent adult in Korea, in which the etiologic organism was confirmed using MLSA based on *rpoB*, *hsp65*, and 16S rRNA fragments.

M. lentiflavum is a recently described slowly growing NTM¹. It has been isolated primarily from lymph nodes of young children³, while isolations from other sites, including respiratory specimens, have been frequently described in immunocompromised patients⁴. *M. lenti-flavum* lung disease in immunocompetent patients has rarely been described in the literature^{2,6-8}, and the isolation of NTM always raises doubts about clinical significance. Our case confirms the potential pathogenicity of *M. lentiflavum* in chronic pulmonary disease.

However, the clinical significance of the isolation of *M. lentiflavum* in our case may be controversial, because of relatively short follow-up periods and small number of sputum acid-fast bacilli (AFB) analysis. As the diagnostic criteria for NTM lung disease are not proven pertinence to less common and low-virulence NTM, the patients with repeated isolations of low-virulence NTM require careful clinical evaluation and collection of multiple specimens for AFB analysis over time ¹⁵.

Considering that *M. lentiflavum* is known to have low-virulence, the patients may need to be followed with additional sputum examinations for longer time.

M. lentiflavum is phylogenetically related to other slow-growing NTM species such as M. avium, M. genavense, and M. simiae¹. Traditional biochemical and/or phenotypic features of M. lentiflavum are not distinguishable from those of these closely related NTM species. Therefore, M. lentiflavum may have been previously misidentified as M. avium complex or M. simiae^{1,4}. Accurate identification of the isolated NTM species is very important when initiating a treatment and selecting an antimicrobial regimen. However, M. lentiflavum cannot be identifiable by commercial kit for the identification of NTM species, such as PCR-restriction fragment length of polymorphism analysis or REBA Myco-ID based on the rpoB gene, because there were not included newly defined or infrequently species, especially for M. lentiflavum. Therefore, the prevalence of M. lentiflavum lung disease in Korea may be underestimated.

In conclusion, *M. lentiflavum* should be considered a possible etiologic pathogen of the nodular bronchiectatic form of NTM lung disease. We demonstrate here that diagnosis of *M. lentiflavum* can be verified by multilocus sequence analysis of *rpoB*, *hsp65*, and 168 rRNA fragments in clinical isolates.

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