Effect of Lycopene Supplementation on Oxidative Stress: An Exploratory Systematic Review and Meta-Analysis of Randomized Controlled Trials

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ABSTRACT Lycopene is a potentially useful compound for preventing and treating cardiovascular diseases and cancers. Studies on the effects of lycopene on oxidative stress offer insights into its mechanism of action and provide evidence-based rationale for its supplementation. In this analysis, randomized controlled trials of the effects of oral lycopene supplementation on any valid outcomes of oxidative stress were identified and pooled through a search of international journal databases and reference lists of relevant publications. Two reviewers extracted data from each of the identified studies. Only studies of sufficient quality were included. Twelve parallel trials and one crossover trial were included in the systematic review, and six trials provided data for quantitative meta-analysis. Our results indicate that lycopene supplementation significantly decreases the DNA tail length, as determined using comet assays, with a mean difference (MD) of -6.27 [95% confidence interval (CI) -10.74, -1.90] (P=.006) between the lycopene intervention groups and the control groups. Lycopene supplementation does not significantly prolong the lag time of low-density lipoprotein (MD 3.76 [95% CI -2.48, 10.01]; P=.24). Lycopene possibly alleviates oxidative stress; however, biomarker research for oxidative stress needs be more consistent with the outcomes in lycopene intervention trials for disease prevention.

KEY WORDS: • lycopene • meta-analysis • oxidative stress • randomized controlled trial • systematic review

INTRODUCTION

CARDIOVASCULAR DISEASES AND CANCERS are main contributors to human morbidity and mortality, and the pivotal role of oxidative stress in the etiology of both diseases is well known. Oxidative stress is defined as disturbances in the prooxidant–antioxidant balance that favors oxidation. Thus, oxidative stress is essentially an imbalance between the production of various reactive species and the ability of the natural protective mechanisms of organisms to cope with these reactive compounds and to prevent adverse effects.¹ Oxidative status is assessed in terms of the overall oxidative/reductive potency of a given specimen (*e.g.*, blood or urine) or the susceptibility of various oxidizable components to *ex vivo* peroxidation.^{1–3}

Lycopene is a potentially powerful antioxidant because of its conjugated double bonds. *In vitro* evidence suggests that lycopene protects lipoproteins and vascular cells from oxidation, but *in vivo* evidence is limited.^{2,4} Lycopene reportedly has the strongest singlet oxygen-quenching capacity among carotenoids,⁵ and is the most effective quencher in plasma, low-density lipoprotein (LDL), and human lymphoid cells in vitro.^{4,6} Epidemiologic studies have reported that the consumption of lycopene-rich foods decreases the risk of cardiovascular diseases and cancers.7-9 Several clinical trials have provided evidence that lycopene lowers oxidative stress, particularly by preventing LDL oxidation.^{4,10,11} However, in 2000, the panel on Dietary Reference Intake evaluated the potential health effects of β -carotene and other carotenoids and concluded that no convincing evidence indicates that substantially increasing the carotenoid intake above current levels significantly affects the antioxidant status; however, lycopene was not specifically addressed.¹² Considering that lycopene metabolism has not been fully elucidated and antioxidative nutrients interact with each other during gastrointestinal absorption and metabolism, the function of lycopene in vivo possibly differs from that in vitro.13-16

Decades of clinical and preclinical studies have broadened our understanding of lycopene and its use in numerous chronic, oxidative stress-induced pathologies; however, the results of these studies have shown mixed results, although with promising implications on the efficacy of lycopene *in vivo*.^{13,17,18} To provide more convincing estimates, this systematic review and meta-analysis assesses the effect of lycopene supplementation on oxidative stress. A wide variety of functional assays for oxidative stress is covered,

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including various measures of lipid oxidation, DNA oxidation, and protein oxidation, for illustrative purposes. This article is the first systematic review on this topic.

METHODS AND MATERIALS

Search strategy

OVID MEDLINE, EMBASE, Web of Science, Springer, CNKI, and CENTRAL (The Cochrane Library) were systematically searched for relevant studies published until June 2012. Further searches were done by scanning the reference lists of original studies, meta-analyses, and review publications on lycopene and tomato. No language or other limitations were imposed. Figure 1 illustrates the selection process. Terms used for MEDLINE (and adopted for other databases) in both subject heading and body text were:

- 1. Lycopene, carotenoids, antioxidants, tomato, or dietary
- 2. LDL oxidation, LDL peroxidation, DNA damage, or oxidative stress
- 3. Both (1) and (2)
- 4. Limit (3) to humans

Studies were included if they met the following criteria: (1) randomized controlled trials (RCTs) on human subjects; (2) biomarkers of oxidative stress as outcomes were reported; (3) intervention included oral lycopene supplementation compared with a concurrent controlled group of placebo or no intervention treatments that were initiated at any time during the subject's lifetime; (4) the trials lasted at least 7 days. To reduce heterogeneity across interventions, trials of foods enriched with lycopene and/or unknown lycopene doses were excluded from this review. Supple-

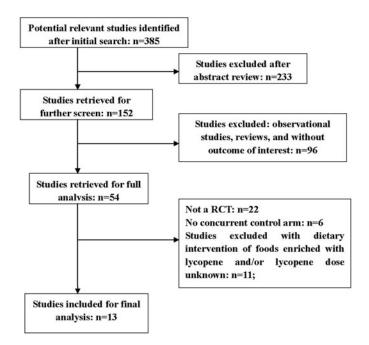


FIG. 1. Search strategy flow chart.

mentation with purified lycopene or formulation with small quantities of other carotenoids and/or flavonoids was included because of their usual inclusion in lycopene supplements. Further, each of the included trials was required to state explicitly the word "random" in the description of treatment assignment. Further details regarding randomization methods used (blocking, random-number generation, etc.) were not required. Missing data were dealt with by contacting the investigators of the original study to request the missing data, or to provide further clarification on data. Analysis was performed on the available data in cases where the missing data were irretrievable. The selection of these biomarkers was based on previous research and reviews, which specified that DNA damage, antioxidant capacity of plasma, lipid peroxidation, protein oxidation, and antioxidative enzymes could be used as biomarkers for evaluating oxidative stress in vivo.^{19,20} Various functional in vivo and ex vivo assays were covered for illustrative purposes. The assays were further categorized into:

- LDL oxidation: LDL lag-time, LDL oxidation rate
- Lipid and protein peroxidation products: plasma malondialdehyde (MDA), thiobarbituric acid–reactive substances (TBARS), urinary 8-iso-PGF_{2α}, plasma thiols.
- Plasma antioxidative capability: total antioxidant capability (TAC), ferric-reducing ability of plasma (FRAP), oxygen radical absorbance capacity (ORAC), P/S fatty acid ratio.
- Antioxidative enzymes: superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx).
- DNA damage, DNA products (comet assay results, 8-OHdG, *etc.*).

Data extraction

The following information was retrieved from each included study:

- Characteristics of study participants
- Alterations in plasma lycopene
- Outcome measurements
- Trial design and blinding methods
- Intervention measures, type of control, and trial duration
- Allocation of concealment, description of randomization, evaluation of losses to follow-up, and subject compliance
- Dietary advice before and during treatment

The inclusion and exclusion criteria for the study participants were also extracted (data not presented).

Quality assessment

All trials were objectively assessed for the risk of bias by two independent reviewers (J.C. and Y.S.) as recommended by the Cochrane guidelines.²¹ The Cochrane protocol explicitly discourages the use of scales to assess the quality of trials when empirical support was lacking; thus, the quality of each trial included in this review was assessed using seven criteria.²²

Statistical analysis

Data were extracted as baseline and endpoint means, standard deviations, and sample sizes for both intervention and placebo groups for each oxidative outcome. When testing was performed at multiple time points, only data from baseline and the final time point were extracted. For each trial following a parallel design, the effect was calculated as the difference in the baseline and end-trial levels between the intervention and the control groups; for crossover design, the effect was assessed using the data from the first period.^{23,24} A fixed-effect model was used to combine data where the studies are assumed to estimate the same underlying treatment effect. If clinical heterogeneity was sufficient to expect that the underlying treatment effects differed between trials, or if substantial statistical heterogeneity was detected, a random-effect model was adopted to produce an overall summary. The results of the metaanalysis are presented as forest plots.

Heterogeneity was assessed using both χ^2 and I^2 . The χ^2 test assesses whether the differences in the results are due to chance only. The data were considered heterogeneous when the *P*-value is low. I^2 assesses the percentage of variability in the estimated effect that is due to heterogeneity rather than chance. I^2 values higher than 50% indicate substantial heterogeneity.²¹ Sensitivity analysis was performed to verify the results of the meta-analysis. Publication bias was assessed by inspecting funnel plots. The analysis was performed using Review Manager 5.0 software.²⁵ Data reported in formats that did not allow entry into RevMan software were reported in tables or in text.

To investigate the differences in the results across studies, several prior hypotheses were developed as follows:

- Study population: normal or in a pathologic state, wherein larger treatment effects were found in the latter.
- Different methodological qualities: randomization concealed or unconcealed; blinded or unblinded; extent of losses to follow-up; dietary advice (whether taking lycopene-containing food or not; anticipating that effect size was overestimated by refraining from lycopene-containing food).
- Study duration: anticipating longer treatments induce stronger effects.
- Lycopene formulations and doses: hypothesizing that the magnitude of the effect varies with the formulation.
- Baseline plasma lycopene level: hypothesizing that the effect is more pronounced with lower baseline plasma lycopene levels.

RESULTS

Description of included studies

The characteristics of the 13 clinical trials²⁶⁻³⁸ meeting inclusion criteria are summarized in Table 1. We excluded 22 trials for not having an RCT design,³⁹⁻⁶⁰ 6 trials for the absence of concurrent controlled arms,⁶¹⁻⁶⁶ 11 trials for dietary intervention with food and/or no lycopene dose mentioned,⁶⁷⁻⁷⁷ and 2 trials for single-dose administration.^{78,79}

These 13 trials were conducted between 1998 and 2011, comprising 844 participants. The trials were conducted in the United States, Canada, Ireland, Spain, Germany, Italy, Iran, France, and Korea. The trial size ranged from 26 to 175 participants (mean age of 25.9-70 years; mean body-mass index [BMI] of 21.0-28.5 kg/m² across 10 trials). In terms of trial design, one of the 13 trials was designed as a crossover:³² placebo/washout/lycopene and lycopene/washout/ placebo for two groups, with an intermittent washout period of 26 days. The remaining 12 trials were designed as parallel trials, 26-31,33-38 particularly the trial by Talvas *et al.*, 36 which was designed as a two-phase RCT: red tomato extract versus vellow tomato extract as the first phase and lycopene capsule versus placebo as the second phase, with a washout period of 2 weeks. The second phase was included in this review as a parallel design. Among the included studies, eight trials were designed as double-blind,^{28,31-37} two trials were singleblind,^{29,36} and three trials reported no information regarding blinding,^{26,27,30} as described in Tables 1 and 2.

Six trials administered one lycopene dosage as the only treatment arm, ^{30–32,34,36,38} three trials designed different lycopene doses as several treatment arms, ^{26,35,37} whereas four trials designed treatment arms with other carotenoid supplements. ^{27–29,33} The lycopene formulations differed across the studies, as shown in Table 1, which could be categorized into the following four formulations: (1) purified lycopene, ^{33,34,36} (2) lycopene with small quantities of tocopherol;^{28,35} (3) lycopene capsules containing 10% β -carotene;^{27,29} and (4) lycopene with its metabolites (phytoene and phytofluene) and small quantities of tocopherol and β -carotene.^{26,30–32,37,38} Trial participants were heterogeneous, varying from healthy adults,^{26–29,31–33,36–38} the most common subjects, to other particular populations such as those with newly diagnosed prostate cancer,³⁰ with type 2 diabetes mellitus,³⁴ or with mildly elevated cholesterol levels.³⁵

For missing data, in the trial by Mackinnon *et al.*,²⁶ the three lycopene treatment arms were merged into one LYCOPENE group in the original article. We used the merged data in this systematic review, because the original data for each arm were unavailable. The formulation of lycopene supplementation in the trial of Neyestani *et al.*³⁴ was irretrievable, whereas the review by Erdman *et al.*¹³ identified the lycopene source in the same article as purified lycopene.

Quality assessment

The quality of the trials was assessed, as shown in Table 2. The majority of trials did not report details regarding concealment allocation (12 trials) and randomization method (12 trials). Only one trial³⁸ reported an appropriate blinding method. Except for the trial by Carroll *et al.*,²⁹ the other 12 trials mentioned the methods for evaluating participant compliance with intervention protocol, and seven trials discussed losses to follow-up.^{26,29,31,35–38} With regard to dietary advice, 11 trials described a specific instruction;^{26,27,30–38} among them, three trials instructed subjects to

		IABLE I. CHA	TABLE I. CHARACTERISTICS OF INCLUDED STUDIES	
Study ID	Characteristics of population ^a	Study design ^b	Treatment/control ^c	Biomarkers of oxidative stress; other clinical outcomes
Collins et al. ²⁷	Spain; 40 healthy subjects (age 25–45, BMI n.a.)	Randomized placebo-controlled four-group parallel trial; 84 days	T1 ($n = 10$): 15 mg α - and β -carotene T2 ($n = 10$): 15 mg lutein T3 ($n = 10$): 15 mg lycopene (containing 10% β -carotene) C ($n = 10$): placebo (0 mg lycopene in corn oil)	DNA damage; n.a.
Carroll et al. ²⁸	Ireland; 58 elderly volunteers [age 70 ± 5 (63-83), BMI 26.5]	Randomized placebo-controlled double blind three-group parallel study; 84 days	T1 ($n = 16$): 13.3 mg lycopene (with 3.5 mg α -tocopherol and 2.5 mg γ -tocopherol) T2 ($n = 15$): 11.9 mg carotene C ($n = 16$): placebo 0 mg (0.5 g corn oil, 0.06 mg a-tocopherol, and 0.23 mg γ -tocopherol)	Lipid oxidation; n.a.
Hininger et al. ²⁹	Spain; 175 healthy nonsmoking males [age 32.8 (25–45), BMI 28.5]	Randomized single-blind placebo-controlled four-group parallel trial; 84 days	T1 ($n = 35$): 15 mg β -carotene T2 ($n = 45$): 15 mg lutein T3 ($n = 52$): 15 mg lycopene (containing 10% β -carotene) C ($n = 52$): placebo 0 mg	Lipid and protein oxidation, antioxidative enzymes; n.a.
Kucuk et al. ³⁰	United States; 26 men with newly diagnosed prostate cancer [age 61 (51–71), BMI n.a.]	Randomized two-arm parallel trial; 21 days	T ($n = 15$): 30 mg lycopene (Lyc-o-Mato® capsule) C ($n = 11$): no placebo capsule while increasing daily fruit and vegetable intake	DNA damage; prostatic pathologic evaluation, PSA
Briviba et al. ³¹	Germany; 30 nonsmoking and 25 smoking males (age 34.2, BMI n.a.)	Randomized double-blind placebo-controlled two-group parallel trial; 14 days	T ($n = 27$): 14.6 mg (each capsule contained 4.88 mg lycopene, 0.48 mg phytoene, 0.44 mg phytofluene, and 1.181 mg α -tocopherol.) C ($n = 28$): placebo 0 mg	DNA damage; immune response (lymphocyte proliferation, NK cells activity, IL-2, IL-4, and TNF-α production)
Riso et al. ³²	Italy; 26 healthy men and women (age 25.8, BMI 21.0)	Random placebo-controlled, double-blind two-group crossover study; 26 days for each period	T ($n = 13$): Lyc-o-Mato drink (5.7 mg of lycopene, 3.7 mg of phytoene, 2.7 mg of phytofluene, 1 mg of β -carotene, and 1.8 mg α -tocopherol) C ($n = 13$): placebo drink devoid of carotenoids	DNA damage; immune/ inflammatory factors (TNF- α and interferon- γ production)
Zhao et al. ³³	United States; 37 healthy, nonsmoking postmenopausal women [age 59.6 (50–70), BMI 25.5]	Randomized double-blind placebo-controlled five-group parallel trial; 56 days	T1 ($n=8$): 4 mg of lutein, β -carotene, lycopene T2 ($n=8$): 12 mg lycopene (pure lycopene beadlets) T3 ($n=8$): 12 mg lutein T4 ($n=7$): 12 mg β -carotene C ($n=6$): placebo 0 mg	DNA damage; n.a.
Neyestani et al. ³⁴	Iran; 35 patients with T2DM of both sexes (age 54±9, BMI 28.0)	Randomized placebo controlled double blind two-group parallel study, age- and sex-matched; 60 days	T ($n = 16$): 10 mg lycopene C ($n = 19$): placebo 0 mg	Plasma antioxidative capability; IgG

(continued)

TABLE 1. CHARACTERISTICS OF INCLUDED STUDIES

Study ID	Characteristics of population ^a	Study design ^b	Treatment/control ^c	Biomarkers of oxidative stress; other clinical outcomes
Devaraj et al. ³⁵	United States; 82 healthy subjects with mildly elevated cholesterol levels [age 50.6 (≥ 40) , BMI 25.78]	Randomized placebo-controlled four group parallel trial; 56 days	T1 ($n = 21$): 6.5 mg lycopene (purified lycopene with 5% ascorbyl palmitate and 1.5% dl- α -tocopherol) T2 ($n = 17$): 15 mg (as T1) T3 ($n = 21$): 30 mg (as T1) C ($n = 18$): placebo 0 mg (with 5% ascorbyl palmitate and 1.5% dl- α -tocopherol)	DNA damage, lipid oxidation; lipid profile
Talvas et al. ³⁶	France; 30 healthy, nonsmoking men [age 60.8 (50-70), BMI 25.8 (24-28)]	Randomized placebo-controlled two-group parallel trial; 7 days	T ($n = 15$): 16 mg (purified lycopene) C ($n = 15$): placebo 0 mg	Plasma antioxidative capability, DNA damage; PSA level
Mackinnon et al. ²⁶	Canada; 60 postmenopausal women [age 55.16 (50–60), BMI 25.85]	Randomized placebo-controlled four-group parallel trial; 120 days	T1 ($n = 15$): regular tomato juice T2 ($n = 15$): lycopene-rich tomato juice T3 ($n = 15$): 30 mg lycopene capsule (Lyc-OMato [®] capsules) C ($n = 15$): placebo 0 mg	DNA damage, lipid and protein oxidation, plasma antioxidative capability; bone resorption marker NTx
Kim et al. ³⁷	Korea: 126 healthy men [age 34.15 (22–57), BMI 24.76]	Randomized double-blind placebo-controlled three- group parallel trial; 56 days	T1 (n =41): 6 mg lycopene (Lyc-OMato® capsules, <0.2% β -carotene, >1.5% tocopherols, >1.0% phytoene and phytofluene) T2 (n =37): 15 mg lycopene (as T1) C (n =38): placebo 0 mg capsule of soybean oil	DNA damage, antioxidative enzymes; endothelial cell function (RH-PAT, hs-CRP, systolic blood pressure, sICAM-1, and sVCAM-1)
van Breemen et al. ³⁸	United States; 105 African American male veterans [age 66.9 ± 7.5 ($50-83$), BMI 28.5 ± 5.3]	Randomized, double-blind, placebo-controlled two-group parallel study; 21 days	T ($n = 54$): 30 mg lycopene (LycoRed capsule: 2% plant sterols, 1.5% tocopherols, 1.0% phytoene and phytofluene, and 0.2% β -carotene) C ($n = 51$): placebo 0 mg (capsule of soybean oil)	DNA damage, lipid oxidation; n.a.
^a Country whe	re study was conducted; number at	nd composition of subjects; age (years); F	^a Country where study was conducted; number and composition of subjects; age (years); BMI (kg/m^2). Values are reported as mean \pm SD and/or range.	

TABLE 1. (CONTINUED)

^bType of study; study duration (days). ^cSample sizes; intervention measures (daily dosages).

n.a., not available; T, treatment groups; C, control group; PSA, prostate specific antigen; NK, natural killer; IL, interleukin; TNF-a, tumor necrosis factor alpha; Ig, immunoglobulin; NTx, N-telopeptide; RH-PAT, reactive hyperemia peripheral arterial tonometry; hs-CRP, high sensitive C-reactive protein.

		TABLE 2. QUALITY	QUALITY ASSESSMENT OF INCLUDED STUDIES	
Study ID	$Description^{a}$	Loss to follow-up	Compliance	Dietary advice
Collins et al. ²⁷	Randomized (blinding not described)	n.a.	Compliance by measuring plasma lycopene	Four weeks of vitamin E supplement before initiation
Carroll <i>et al.</i> ²⁸	Randomized, double blinded	11/58 18.9%	Compliance by questioning volunteers, counting returned capsules monthly, and measuring plasma response	n.a.
Hininger et al. ²⁹	Randomized, single blinded	n.a.	n.a.	n.a.
Kucuk et al. ³⁰	Randomized (blinding not described)	n.a.	Compliance was assessed by performing remaining pill counts at the end of the intervention	Instructed to increase daily fruit and vegetable intake to five servings for control group
Briviba <i>et al.</i> ³¹	Randomized, double blinded	0	No participant excluded for health problems, compliance, or other reasons	Instructed to refrain from carrot and tomato product 4 weeks before and during treatment
Riso et al. ³²	Randomized, double blinded	n.a.	Compliance by weekly interviews, frequency of consumption of fruits and vegetables	Instructed to maintain habitual diet
Zhao <i>et al.</i> ³³	Randomized, double blinded	n.a.	Compliance by counting pills, checking food calendar, 3-day food records, and food- frequency questionnaires biweekly	Instructed to exclude foods rich in carotenoids and to limit fruit and vegetable intake to two servings/day of light-colored fruit and vegetables for 2 weeks before and during treatment
Neyestani et al. ³⁴	Randomized, double blinded	n.a.	Dietary assessment was carried out after washout and after intervention	Instructed to follow a lycopene-free diet for a 2-week washout period and instructed to keep their diets and physical activities constant during the experimental period
Devaraj <i>et al.</i> ³⁵	Randomized, double blinded	5/82 6.10%	Compliance (93.9%) by food diary, tablet counting, and plasma lycopene measurement	Instructed to refrain from lycopene-containing food for 2 weeks before and during treatment
Talvas <i>et al.</i> ³⁶	Randomized, single blinded	0	Compliance by an interview with dietitian	Instructed to avoid sources of tomato/lycopene 2 weeks before and during treatment
Mackinnon et al. ²⁶	Randomized (blinding not described)	5/65 7.69%	Compliance by using supplement counting	Instructed to refrain from lycopene-containing food 1 month before and during treatment and from vitamins containing antioxidants and nutritional supplements during the study
Kim et al. ³⁷	Randomized, double blinded	10/126 7.94%	Compliance (92.1%) at the end of the study using pill counting, food records, and measurement of plasma lycopene levels	Instructed to maintain usual lifestyle and dietary habits
van Breemen <i>et al.</i> ³⁸	Computer-generated pseudorandom number for randomization; the method of double blinding was described and it was appropriate. Allocation of concealment: subjects enrolled by the study manager	26/131 19.85%	Compliance (99%) by calendars to record capsules and capsule counting, reminded to take capsules during calls for 24-hour dietary recalls	Dietary intakes of Jycopene and other carotenoids and nutrients at baseline and during the study were determined on the basis of five 24-hour dietary recalls
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^aRandomization and blinding, as described by the literature. Allocation of concealment was not described for any of the studies except for van Breemen et al.³⁸

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avoid lycopene-containing food during the washout and treatment periods;^{26,35,36} one trial instructed subjects to refrain from lycopene-containing food during the washout period and to keep a constant diet during treatment period;³⁴ two trials asked subjects to refrain from food enriched with lycopene and carotenoids;^{31,33} two trials encouraged subjects to maintain their diets and lifestyle,^{32,37} which possibly inflated the effect size; one trial provided vitamin E supplement before initiation;²⁷ one trial instructed the controlled subjects to increase fruit and vegetable intake, possibly resulting in underestimation of the effect size;³⁰ and one trial instructed the subjects to keep records of lycopene and carotenoid intake.³⁸ Compliance and dietary monitoring were satisfactory in all evaluated trials.

Plasma/tissue lycopene levels

Except for one trial,³² the other 12 studies reported alterations in plasma lycopene levels, among which seven trials provided explicit data on pretreatment and posttreatment.^{26,28–31,36,38} Plasma lycopene at baseline ranged from 0.053 to 0.69 μ M, as shown in Table 3. Among the 12 studies, two reported prostate lycopene levels^{30,38} and one reported the LDL and high-density lipoprotein concentrations of lycopene.²⁸

Plasma and tissue lycopene in the lycopene intervention groups of all trials was significantly elevated after supplementation, which further supported the validity of these clinical trials (Table 3).

Oxidative biomarker assessment

The oxidative parameters assessed in each study were categorized and are presented in Table 4. In total, at least 20 oxidative parameters were applied across these studies, as summarized below.

LDL oxidation. Three trials assessed the LDL lag time, 28,29,35 and one reported the LDL oxidation rate.³⁵ These data were considered appropriate for meta-analysis (Table 5). With a total pooled sample size of 167 subjects, the lycopene treatment did not significantly prolong the LDL lag time (MD 3.76 [95% confidence interval (CI) -2.48, 10.01]; P = .24). A considerable heterogeneity was observed ($I^2 = 100\%$; P < .00001).

Lipid and protein oxidation and peroxidation products. Lipid oxidation products were investigated in five trials^{26,32,35,36,38} with the same test principle for plasma MDA, hydroxyl nonenal (HNE), and TBARS, and two trials for urinary 8-iso-PGF_{2x}.^{32,36} Except for one trial,²⁶ which reported increased TBARS versus baseline in the LYCO-PENE group, no other significant effect of lycopene on lipid peroxidation products compared with the placebo or baseline level was reported. For protein oxidation products, two trials reported thiol groups in different measurement units, as μ mol/g proteins in one trial²⁹ and μ M in plasma in another.²⁶ A significantly positive effect was found between the post-treatment level with the baseline level of the LY-COPENE group in the latter trial. Antioxidative enzymes and plasma antioxidative capability. Antioxidative enzymes were investigated in three trials.^{26,29,37} No significant effect was reported in GPx, glutathione disulfide (GSSG), glutathione (GSH), and SOD, except that one trial³⁷ reported increased SOD versus baseline in an intervention arm of 15 mg/day lycopene (P<.05).

FRAP,³⁶ ORAC,³⁶ TAC,^{26,34} and P/S fatty acid ratio²⁹ were adopted to evaluate plasma antioxidative capability. One trial²⁶ reported increased TAC versus baseline in the LYCOPENE group. However, no significant effect was found in other parameters.

DNA damage (by Comet assay or products). Three trials^{32,33,35} assessed the lymphocyte DNA tail length, and the data were considered appropriate for meta-analysis (Table 5). With a pooled sample size of 115 subjects, lycopene was found to decrease the DNA tail length significantly, which was measured using a comet assay (MD -6.27 [95% CI -10.74, -1.90]; P = .006). Considerable heterogeneity ($I^2 = 100\%$; P < .00001) was observed across the three studies. The urinary 8-OHdG,³⁵ plasma 8-OHdG,²⁷ and 8-oxo-dG/ 10^6 dG levels in prostate tissue,³⁸ as well as peripheral blood lymphocyte level of 5-OHmdU³⁰ were determined in four trials. Except for Devaraj *et al.*,³⁵ who found decreased urinary 8-OHdG versus baseline at 30 mg/d lycopene, no other significant effect was reported in the other four trials.

Clinical outcomes

The clinical outcomes assessed in these trials were also analyzed, and they are listed in Table 1. Clinical outcomes were evaluated in eight trials:

- Lipid profile: No significant differences were found.³⁵
- Prostate cancer-related pathologic status: In the trial by Kucuk *et al.*,³⁰ the pathologic stage of prostate cancer in the lycopene intervention group was less advanced and had lower PSA. In the trial by Talvas *et al.*,³⁶ the PSA levels in the healthy men did not change after lycopene intervention.
- Inflammatory/immune factors: IL-4 production was reduced by lycopene intervention,³¹ TNF- α production was reduced by lycopene intervention,³² and IgG increased after lycopene intervention.³⁴
- Bone resorption marker (NTx): NTx was reduced after lycopene intervention.²⁶
- Endothelial cell function: An increase in reactive hyperemia peripheral arterial tonometry (RH-PAT) index from baseline, whereas high-sensitive C-reactive protein (hs-CRP), systolic blood pressure, soluble intercellular adhesion molecule-1 (sICAM-1), and soluble vascular cell adhesion molecule-1 (sVCAM-1) were significantly decreased in the 15 mg/d group.³⁷

Sensitivity analysis and subgroup analysis

Sensitivity analysis was performed by excluding certain studies to examine the influence of the omitted study on the pooled effect. Thus, the overall effects were 1.15 (-7.37 to)

		Plasma lycopene (µM)	ene (µM)		
	Control groups	roups	Lycopene-treated groups	ted groups	
Study ID	Pretreatment	Post-treatment	Pretreatment	Post-treatment	Tissue lycopene
Mackinnon et al. ²⁶	0.264 ± 0.372^{a}	0.281 ± 0.771^{a}	0.288 ± 0.289^{a}	0.979 ± 0.485^{a}	
Carroll <i>et al.</i> ²⁸	0.10 ± 0.07	0.08 ± 0.07	0.10 ± 0.05	0.27 ± 0.10	LDL and HDL concentrations of lycopene were significantly higher ($P < .05$) than in the other groups and than the baseline level
Hininger et al. ²⁹	0.69 ± 0.13	0.63 ± 0.21	0.69 ± 0.13	1.17 ± 0.15	
Kucuk et al. ³⁰	0.53 ± 0.13^{b}	0.33 ± 0.006	0.53 ± 0.13^{b}	0.44 ± 0.04	0.36 ± 0.06 ng/g for placebo group vs. 0.53 ± 0.03 ng/g for lycopene-treated group in prostate tissue after intervention ($P < .05$)
Briviba et al. ³¹	0.223 ± 0.139	0.244 ± 0.44	0.275 ± 0.194	0.785 ± 0.290	
	for nonsmokers; 0.145±0.87 for smokers	for nonsmokers; 0.181 ± 0.162 for smokers	for nonsmokers; 0.228±0.101 for smokers	for nonsmokers; 0.828±0.204 for smokers	
Talvas et al. ³⁶	$0.43 \pm 0.07^{\rm b}$	0.30 ± 0.03	0.43 ± 0.07	0.87 ± 0.10	
van Breemen et al. ³⁸	0.599 ± 0.373	0.588 ± 0.392	0.741 ± 0.388	1.428 ± 0.613	0.45 ± 0.53 pmol/mg for placebo group vs. 0.59 ± 0.47 pmol/mg for lycopene-treated group in prostate tissue after intervention ($P < .05$)
Collins et al. ²⁷	Increase in plasma lycopene vs. baseline ($P < .05$) in lycopene-treated group	e vs. baseline ($P < .05$) in l	ycopene-treated group		
Zhao et al. ³³	Increase in plasma lycopene vs. baseline $(P < .05)$ in lycopene-treated group	e vs. baseline $(P < .05)$ in l	ycopene-treated group		
Neyestani et al.34	Increase in serum lycopene vs. baseline ($P < .001$) in lycopene-treated group	vs. baseline ($P < .001$) in 1	ycopene-treated group		
Devaraj <i>et al.</i> ³⁵	Increased plasma and LDL standardized all group compared with baseline, but not pl	creased plasma and LDL standardized all <i>trans</i> -lycopen group compared with baseline, but not placebo $(P < .05)$	<i>trans</i> - lycopene levels in each supplemented acebo ($P < .05$)	mented	
Talvas <i>et al.</i> ³⁶	Increase in plasma lycopene vs. baseline ($P < .05$) in lycopene-treated group (0.45 ± 0.03 for all subjects)	e vs. baseline $(P < .05)$ in I; subjects)	ycopene-treated		
Kim <i>et al.</i> ³⁷	Increase in serum and lipid standardized lycopene levels in each supplemented group compared with baseline ($P < .05$), but not placebo	standardized lycopene leve $(P < .05)$, but not placebo	els in each supplemented g	roup	
^a Represents all- <i>trans</i> -lycopene.	vcopene.				

TABLE 3. PLASMA/TISSUE LYCOPENE OF INCLUDED STUDIES

^aRepresents all-*trans*-lycopene. ^bRepresents plasma lycopene of subjects of all control and lycopene-treated groups. HDL, high-density lipoprotein; LDL, low-density lipoprotein.

	TABLE 4.	CATEGORIZED	OXIDATIVE	PARAMETERS	OF EACH STUDY
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Parameters of oxidative stress	Study ID
LDL oxidation LDL oxidation lag time	Carroll <i>et al.</i> ²⁸ ($P > .05$); Hininger <i>et al.</i> ²⁹ ($P > .05$); Devaraj <i>et al.</i> ³⁵ ($P > .05$)
LDL oxidation rate	Devaraj et al. ³⁵ (P > .05)
Lipid oxidation/peroxidation products	Riso <i>et al.</i> ³² ($P > .05$); Talvas <i>et al.</i> ³⁶ ($P > .05$)
Urinary 8-iso-PGF _{2α} Plasma malondialdehyde (MDA) and hydroxyl nonenal (HNE)	Devaraj <i>et al.</i> ³⁵ ($P > .05$)
Plasma MDA	van Breemen <i>et al.</i> ³⁸ (P >.05)
Plasma thiobarbituric acid reactive substances (TBARS)	Mackinnon <i>et al.</i> ²⁶ (increased vs. baseline in lycopene-treated groups $P < .05$)
Protein oxidation SH groups (µmol/g proteins)	Hininger <i>et al.</i> ²⁹ ($P > .05$)
Plasma thiols	Mackinnon <i>et al.</i> ²⁶ (decreased vs. baseline in lycopene-treated groups $P < .05$)
Antioxidative enzyme	······································
Glutathione peroxidase (GPx), glutathione disulfide (GSSG), and glutathione (GSH)	Hininger <i>et al.</i> ²⁹ ($P > .05$) in GSH-Px, GSSG, and GSH; Mackinnon <i>et al.</i> ²⁶ ($P > .05$) in GPx
Superoxide dismutase (SOD)	Kim <i>et al.</i> ³⁷ (increased vs. baseline in lycopene-treated group $P < .05$); Mackinnon <i>et al.</i> ²⁶ ($P > .05$); Hininger <i>et al.</i> ²⁹ ($P > .05$)
Catalase (CAT)	Mackinnon <i>et al.</i> ²⁶ ($P > .05$)
Plasma antioxidative capability	
Ferric reducing ability of plasma (FRAP) and oxygen radical absorbance capacity (ORAC)	Talvas <i>et al.</i> ³⁶ ($P > .05$)
Total antioxidant capacity (TAC) to MDA	Neyestani et al. ³⁴ (P >.05)
Total antioxidant capability (TAC)	Mackinnon <i>et al.</i> ²⁶ (increased vs. baseline in lycopene treated groups $P < .05$)
P/S fatty acid ratio	Hininger et al. ²⁹ ($P > .05$)
DNA damage (by comet assay)	Collins <i>et al.</i> ²⁷ ($P > .05$)
Lymphocyte DNA comet assay visual score DNA tail intensity	Collins <i>et al.</i> ²¹ ($P > .05$) Briviba <i>et al.</i> ³¹ ($P > .05$)
Lymphocyte DNA tail length	Riso et al. ³² ($P > .05$); Zhao et al. ³³ (decreased lymphocyte DNA tail
	length vs. baseline: $P < .01$); Devaraj <i>et al.</i> ³⁵ (decreased lymphocyte DNA tail length vs. baseline: $P < .05$)
Tail DNA(%) and tail moment	Kim <i>et al.</i> ³⁷ (decreased lymphocyte DNA tail length, tail DNA (%), and tail moment vs. baseline $P < .05$)
DNA damage (by products)	
Urinary 8-OHdG/plasma 8-OHdG	Collins <i>et al.</i> ²⁷ (P > .05 in plasma 8-OHdG); Devaraj <i>et al.</i> ³⁵ (decreased urinary 8-OHdG vs. baseline P < .05)
Prostate tissue level of 8-oxo-dG/10 ⁶ dG Peripheral blood lymphocyte level of 5-OHmdU	van Breemen <i>et al.</i> ³⁸ ($P > .05$) Kucuk <i>et al.</i> ³⁰ ($P > .05$)

P > .05 means no significant difference with placebo groups or baseline.

9.68), 2.90 (-9.06 to 14.85), and 7.24 (3.81 to 10.27), respectively, with the exclusion of trials 28, 29, and 35 on the extension of the LDL lag time. With exclusion of trials 37, 32, and 33, the effects on the DNA tail length were -3.55 (-8.94 to 1.84), -9.05 (-14.53 to -3.57), and -6.32 (-17.19 to 4.55), respectively. No quantitative subgroup analysis was conducted because of the small number of included studies on the same parameter. Funnel plots were not constructed because of the small number of included studies.

DISCUSSION

Summary of main results

The present study is the first systematic review of the antioxidative effects of lycopene, in which all available RCTs on the subject were pooled. This review included 13 RCTs, which vary in participant characteristics, formula-

tions, and intervention measures of lycopene, parameters of oxidative stress, and study duration. Pooling the results of the three trials revealed no statistically significant difference in the LDL lag time between the lycopene treatment group and the controlled group. However, a meta-analysis of another three trials suggested that lycopene significantly decreases the DNA tail length as measured by comet assays. Other meta-analyses were impossible to carry out, because the contributing data for other outcomes came from only one trial. Comparisons of the effect of purified lycopene with other formulations and dosage/duration of lycopene is statistically impossible. However, the results showed no obvious discrepancy based on this systematic review.

Oxidative DNA damage is a potential marker for monitoring oxidative stress, as well a marker of oxidative disease risk,^{4,19,20} one of the major and well-characterized functions of lycopene antioxidative activity. Despite the uncertainty posed by the limited population size and the heterogeneity in

						ľ	100					100	
IA TAIL LENGTH	Moan difforence	IV, random [95% CI]	-		1	•	0 plement placebo control	-			•	0 50	pplement placebo control
D LYMPHOCYTE DN							-100 -50 lycopene supplement					-100 -50	lycopene supplement
lementation on Low-Density Lipoprotein Lag Time and Lymphocyte DNA Tail Length	Mean difference	[95% CI]	[8.49, 9.51]	[-3.48, -2.92]	[5.46, 5.54]	[-2.48, 10.01]		[- 13 28 - 10 50]	[-0.85, -0.75]	[-6.41, -6.19]	[-10.74, -1.80]		
)ensity Lipopr	Mean	IV, random	0.00	-3.20	5.50	3.76		_ 11 80	-0.80	-6.30	-6.27		
ON LOW-L		Weight	33.3%	33.3%	33.4%	100.0%		37 606	33.7%	33.7%	100.0%		
INTATION		Total	16	18	45	6 2	$^{2} = 100\%$	38	13	9	57	$^{2} = 100\%$	
COPENE SUPPLEME	Control	$Mean \pm SD$	-1 ± 0.67	1.2 ± 0.462	-15.6 ± 0.093		$P < .00001$; $P^2 = 1$	- 161+765	0.3 ± 0.056	1.2 ± 0.128		P < .00001; $P = 1$	
т оғ Lyg	lement	Total	16	21	51	88	, df=2 (. 06)	37	13	8	58	, $df = 2$ (,	4)
TABLE 5. EFFECT OF LYCOPENE SUPPL	Lycopene supplement	$Mean\pm SD$	8±0.79	-2 ± 0.437	-10.1 ± 0.088		5.41; $\chi^2 = 8057.54$ x: $Z = 2.75$ ($P = .00$	uil length 16 5 + 3 43	-0.5 ± 0.076	-5.1 ± 0.059		0.45; $\chi^2 = 3750.86$	ct: $Z = 1.18$ ($P = .2^{\circ}$
		Study ID	LDL lag time Carroll <i>et al.</i> ²⁸	Devaraj et al. ³⁵	Hininger et al. ²⁹	Total	Heterogeneity: $\tau^2 = 15.41$; $\chi^2 = 8057.54$, df = 2 (<i>P</i> < .00001); I^2 Test for overall effect: $Z = 2.75$ (<i>P</i> = .006)	Lymphocyte DNA tail length <i>k</i> im <i>at al</i> 37 – 16 5-	Riso et al. ³²	Zhao et al. ³³	Total	Heterogeneity: $\tau^2 = 30.45$; $\chi^2 = 3750.86$, df = 2 (P < .00001); I^2	Test for overall effect: $Z=1.18$ ($P=.24$)

df, degree of freedom.

the quantitative meta-analysis for DNA tail length, the positive effect observed still offers an insight into the *in vivo* mechanism of action of lycopene and provides perspectives for consumers and researchers.

Mackinnon et al.26 reported increased antioxidative capacity with lycopene supplementation and decreased lipid and protein oxidation products. Devaraj et al.³⁵ reported decreased urinary DNA damage products, whereas Kim et al.³⁷ reported increased plasma SOD activity. No other significant effect was reported in these studies. Several factors contributed to the conflicting results of antioxidative activity studies. The null results might have resulted from the very low basal oxidative damage among the young healthy subjects who participated in several studies.^{27,29,31,32,37} Thus, basal variations are likely negligible and have little biological significance. Subjects with low baseline lycopene levels, such as those in the trial of Mackinnon *et al.*,²⁶ where the baseline lycopene level was 0.264 μ M, may respond better than the subjects with elevated baseline lycopene levels. The dietary advice of the trials by Mackinnon et al.²⁶ and Devaraj et al.³⁵ both included refraining from lycopene-containing food for both the washout and the treatment period. The former instructed all participants to refrain from any vitamins and antioxidant supplementation, which could overestimate the magnitude of the effect.

Epidemiologic evidence and dietary intervention studies have associated tomato and lycopene with lower incidence rates of cardiovascular diseases and cancers.^{3,4,8} However, the effect of lycopene supplementation remains unconfirmed. Tomatoes contain significant amounts of β -carotene, vitamin C, folate, and potassium.⁸⁰ Thus, studies on the health benefits of tomatoes should consider other bioactive compounds that could contribute to the beneficial effects investigated.

Oxidative stress and clinical outcomes

Oxidative stress is often associated with high levels of reactive oxygen and oxidation products, particularly hydroperoxides and DNA fragments.¹ Pinchuk et al.⁸¹ proposed several problems in the commonly used assays for ranking antioxidants, including that (1) the assays usually evaluate the effects of antioxidants that quench free radicals, which constitute only a part of the antioxidative network, and (2) antioxidative capacity and the potency of antioxidants, as determined by various methods, do not necessarily correlate with each other. Pinchuk et al.⁸¹ and Dotan et al.⁸² further suggested that the term oxidative stress of any given type is context dependent, and that the evaluation should be based on the use of the most sensitive probe or combination for oxidative damage. As shown in Table 1, we included five trials that measured DNA damage as the only biomarker,^{27,30–33} whereas one trial measured DNA damage, lipid and protein oxidation, and TAC.²⁶ The contrasting findings on the particular oxidative stress parameters in chronic diseases highlight the importance of validating and standardizing biomarkers for oxidative stress.

Among our included studies, Kucuk et al.³⁰ recruited men with newly diagnosed prostate cancer. After intervention, the subjects in the lycopene intervention group had smaller tumors, less involvement of surgical margins and/or extraprostatic tissues with cancer, less diffuse involvement of the prostate by high-grade prostatic intraepithelial neoplasia, and lower PSA levels compared with subjects in the control group. However, no significant difference was found in DNA oxidation products, which implied that DNA oxidation might not be as critical and indicative in the progression of prostate cancer as it is in the occurrence of prostate cancer. Lycopene may also exert its anticarcinogenic effect via another pathway, such as by alteration of cell gap communication, as the authors suggested. Talvas et al.³⁶ found that the PSA levels in healthy men were unchanged after lycopene intervention. This result also suggests the fact that the efficacy of lycopene might manifest more in the pathologic state. Kim et al.³⁷ found that the beneficial effects of lycopene supplementation on endothelial function (i.e., RH-PAT, hs-CRP, and sVCAM-1) were remarkable among subjects with relatively impaired endothelial cell function. To some extent, this finding confirms the assumption that lycopene is more effective in the oxidative impaired population. However, the small number of qualified samples limits the correlation of oxidative stress and outcomes of cancer and cardiovascular diseases in this meta-analysis.

It is well documented that subjects with risk factors such as hypertension, diabetes, and smoking have increased levels of oxidative stress based on the rate of oxidant production and by elevated plasma markers for molecular oxidation such as lipid hydroperoxides.^{19,20} However, these oxidation products or biomarkers have not yet been adequately validated as markers for the onset, progression, and regression of any chronic diseases, which presents a challenge to the interpretation and integration of current evidence and justifies the ongoing research on oxidative biomarkers and antioxidant efficacies.^{12,83}

Strengths and limitations of this review

The present systematic review has several advantages. This is the first systematic review that examines the effects of lycopene supplementation on oxidative stress *in vivo*. Also, a detailed research protocol was included, with prior specification and evaluation of potential study design considerations without language exclusion. The strengths of the study also include the analysis of only randomized, controlled clinical trials and the objective assessment of trial quality. Furthermore, variable biomarkers for oxidative stress were categorized for analysis.

Except for the uncertainty in evaluating oxidative stress biomarkers as discussed above, the following limitations warrant further discussion. First, the RCTs of lycopene supplementation with small quantities of other micronutrients were included, and not all lycopene supplements are equivalent. However, the same is true for all the manufactured lycopene supplements, whether purified or formulations. Thus, this review would still offer insights into the consumption of lycopene supplements. Second, this review includes one crossover trial; while in the presence of carryover, a common strategy is to base the analysis on the first period. The corresponding data were not a source of bias, but it usually decreases the weight of trials in metaanalyses.²⁵ We also did not perform subgroup analysis because of the limited number of studies. Therefore, we were unable to confirm whether lycopene has different antioxidative effects among subjects with variable physiologic states, or whether lycopene may act differently in short-term versus long-term application, which remains an important area for future research.

Conclusion and implication for practice and research

The included trials used variable methods to measure oxidative stress, which caused incomparability across studies. Understanding the effectiveness of antioxidants in vivo requires identifying the markers or combinations of markers that are predictive of human diseases. Thus, biomarker research needs to move in parallel with lycopene intervention trials for disease prevention. In retrospect, dietary interventions using antioxidant supplements, lycopene in this case, may have been premature because of the need for more fundamental information on the mechanisms of action of these compounds. Given the current evidence, the supposition that lycopene is efficient as an antioxidant in vivo needs further examination. With the elucidation of the oxidative stress pathways, understanding the efficacy of lycopene could be clarified. Therefore, until further research establishes significant health benefits for lycopene supplementation in humans, it should be concluded that the consumption of natural carotenoid-rich fruits and vegetables is preferential to purified lycopene supplementation.

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AUTHOR DISCLOSURE STATEMENT

The authors declare that there is no conflict of interest.

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