Research Article

The Effect of a Silage Inoculant on Silage Quality, Aerobic Stability, and Meat Production on Farm Scale

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The effect of inoculation on nutrient content, fermentation, aerobic stability, and beef cattle performance for whole-plant corn silage treated with a commercial product (blend of homo- and heterofermentative lactic acid bacteria, BSM, blend of *Enterococcus faecium*, *Lactobacillus plantarum*, and *Lactobacillus brevis*, DSM numbers 3530, 19457, and 23231, resp.), was compared to a control treatment with no silage additives (CT). The material had a DM of 323 g/kg, crude protein, and water-soluble carbohydrate concentrations of 87.9 and 110.5 g/kg DM, respectively. BSM increased the fermentation rate with a significantly deeper pH (P < 0.01), a significant increase in the total organic acids concentration (P < 0.05), more lactic acid (P < 0.01), and numerically more acetic acid compared to CT. BSM significantly decreased the concentrations of butyric acid (P < 0.01), ethanol, and anmonia-N compared to the CT. BSM-treated silage decreased DM by 3.0 % (P < 0.01) and had a higher digestible energy and a higher metabolizable energy concentration by 2.3 (P < 0.01) and 1.00 % (P < 0.05), respectively, compared to untreated silage. Aerobic stability improved by more than 2 days in BSM silage. The DM intake of silage treated with BSM increased by 6.14 %, and improved weight gain and the feed conversion by 8.0 (P < 0.01) and 3.4%.

1. Introduction

Today, silage is the world's largest fermentation process, with an estimated 287 million tons produced in EU alone [1]. For dairy and beef cattle farmers, their purpose is to produce more high-quality silage, rich in energy and protein.

The key factors influencing the feeding value of silages include crop characteristics, stage of crop development at ensiling, and the extent and type of fermentation achieved within the silo. Silage additives have gained more and more interest over recent years. It is widely accepted that silage additives can increase animal intake and animal performance through their effect on silage quality [2]. However, the market became reluctant to use acid additives as they were considered corrosive to machinery and concrete, and dangerous to those farm operatives who had to use them.

Successful silage production depends on the promotion of fermentation by beneficial bacteria [3], and therefore

bacterial inoculants have been very popular, especially over the last 10 years.

Microbial inoculants have been added to silages to improve fermentation efficiency [4–6]. Microbial inoculants containing homofermentative lactic acid bacteria (LAB), in most of the cases Lactobacillus plantarum, are often added to silage because they very quickly produce large quantities of lactic acid, which lowers the pH of the silage [7, 8]. However, classic microbial inoculants often have no effect or can even make the aerobic stability of silages worse [7, 9, 10] because yeasts metabolize lactic acid to produce alcohol. Recently, the aerobic stability of a variety of silage crops has been markedly improved through inoculation with a heterolactic acid bacterium [11]. Improvements in aerobic stability brought about by this organism have been reported in corn silage [12]. Acetic acid produce from heterofermentative LAB inhibits the proliferation of yeasts in silage. Prevention of the growth of these organisms is crucial in restricting

Parameter	Method	Units	Average	Standard deviation
Dry matter (DM)	Oven drying at 67°C for 24 h, equilibrated to room humidity overnight, milled through a 1 mm sieve and further dried at 105°C to constant weight.	g/kg	322.8	8.954
Crude protein	Kjeldahl-AOAC 984.13. 10.5 g of catalyst is used. With Block Digestion and Tecator Kjeltec system 1002 Distilling Unit		87.9	2.124
Crude fat	Extraction by Soxtec System using petrol ether 40–600°C. Crude fat residue determined gravimetrically after drying		19.0	1.513
Crude fiber	With Fibercap (Foss Tecator) using sulphuric acid and Na hydroxide treatment	g/kg DM	213.2	10.725
Nitrogen-free extract	Calculated		633.7	14.467
Crude ash	AOAC Method 942.05. Ca—AOAC 968.08 dry ashing, atomic absorption Spectrophotometry		46.2	3.850
Water-soluble carbohydrates (WSC)	Using the anthrone reaction assay (MAFF, 1986), from the herbage or silage extracts obtaining from steeping fresh herbage or silage in water		110.5	4.799
Digestible energy	Calculated	MJ/kg	12.95	0.051
Metabolizable energy	Calculated	DM	10.69	0.102

TABLE 1: Chemical composition of whole plant corn at ensiling (average of 5 samples).

aerobic deterioration. A strain of heterofermentative LAB *Lactobacillus brevis* has been reported as a promising strain for improving the aerobic stability of silages [13].

The aim of this trial was to study the effect of a silage inoculant on the nutrient content, the silage quality, the aerobic stability, and the nutritive value of whole plant ensiled corn, as well as on the feed intake and growth performance of fattening young cattle.

2. Material and Methods

A trial was carried out with whole plant corn harvested at the milk/dough stage of maturity (32.3% DM, see Table 1) and used for ensiling, treated (BSM), or not (CT), with a silage inoculant.

The corn was chopped using a conventional forage harvester Massey Ferguson 5130 and ensiled directly after harvest in 2 horizontal silos with a capacity of 200 tones. The in oculant application rate of the commercial product was the rate recommended by the manufacturers (blend of Enterococcus faecium, Lactobacillus plantarum, and Lactobacillus brevis, DSM numbers 3530, 19457, and 23231 respectively; 4 g of product/ton of silage diluted in 41 of water) to guarantee a concentration of 1×10^5 cfu/g of material. The inoculant was applied uniformly using an applicator which was fixed on the harvester between the pick-up reel and cutting rollers. After weighing the untreated or inoculated chopped corn, it was transferred to one of two ferroconcrete trenches. Five control bags (made from four layers of cheesecloth) filled with 1 kg of ensiling mass were put in each silo to determine DM losses. The silos were filled within 48 hours and were covered with polythene sheet and weighted down with tires.

Five representative samples of harvested and chopped whole-plant corn were taken throughout the harvesting-ensilaging period. Silages were sampled every other week during the feeding experiment (14.12.2009 to 14.04.2010). At each sampling time two samples (approx. 500 g each) were taken 40–50 cm deep from the cut surface by coring vertically to the full depth of the silo using a 50 mm-silage corer.

Volatile fatty acid and lactic acid, as well as alcohol concentrations, were determined by gas-liquid chromatography on aqueous silage extracts obtained from steeping 30 g of fresh silage in 150 mL of deionized water for 16 hours at 4°C in a sealed container. This was followed by a preliminary filtering through 3 μ m filter paper. Deionized water (3 mL) from an internal standard solution (0.5 g 3-methyl*n*-valeric acid in 1000 mL 0.15 mol/l oxalic acid) was added to 1 mL of filtrate from the above, and the solution was filtered through a 0.45 μ m polyethersulfone membrane into a chromatographic sample vial for analysis. A gas-liquid chromatograph SHIMADZU 2010 was used with a wide-bore capillary column (Stabilwax-DA 30 m, 0.53 mm, ID, 0.5 μ m) according to the Gas Chromatography and Biochemistry Analyzer official methods.

Aerobic stability was measured using data loggers which recorded temperature readings once every 6 hours from thermocouple wires placed in three replicate 1.500–2.000 g silage representative samples, which were aerated in open plastic bags and placed into open-top polystyrene boxes (volume about 3 liters). Thermocouple wires were inserted into the silage. The boxes were constantly kept at room temperature (21°C). Aerobic deterioration was denoted by days (or hours) until the start of a sustained increase in temperature by more than 2°C above the ambient temperature.

TABLE 2: Effect of the treatment whole-plant corn.	nt with a commercial produc	t BSM on the chemical	composition and fe	ermentation characterist	ics of ensiled
Parameters	Unit	Treatm	nents	Standard error	Р

Darameters	Unit	IICati	lifeitts	Standard error	D
Taraniciers	Ollit	Control $X \pm SD$	BSM $X \pm$ SD	Standard error	1 1
Dry matter (DM)	g/kg	305.8 ± 4.30	312.2 ± 4.66	1.119	**
DM losses		70.2 ± 15.87	40.9 ± 2.60	5.946	**
Crude protein		80.2 ± 4.94	84.7 ± 3.24	0.954	*
Digestible protein		48.2 ± 2.96	52.5 ± 2.01	0.680	**
Crude fat		19.7 ± 1.71	19.8 ± 1.70	0.341	0.845
Crude fiber	g/kg DM	214.8 ± 4.59	210.2 ± 7.30	1.311	0.074
Nitrogen-free extract	0.0	640.1 ± 7.42	640.9 ± 11.38	1.919	0.833
Crude ash		45.2 ± 3.26	44.4 ± 4.10	0.744	0.622
Water-soluble carbohydrates (WSCs)		1.9 ± 1.36	2.1 ± 1.02	0.241	0.698
Neutral detergent fiber (NDF)		444.4 ± 11.73	439.1 ± 15.66	2.818	0.355
Acid detergent fiber (ADF)		238.3 ± 6.70	228.4 ± 12.24	2.221	*
Digestible energy (DE)	MI/kg DM	12.8 ± 0.06	13.1 ± 0.07	0.031	**
MJ/kg DM Metabolizable energy (ME)		10.8 ± 0.08	10.9 ± 0.13	0.024	*

* and ** denote significance at level 0.05 and 0.01, respectively.

TABLE 3: Effect of the treatment with a commercial product BSM on the fermentation characteristics of ensiled corn.

Parameters	Unit	Treatr	nents	Standard error	or P	
	Ollit	Control $X \pm SD$	BSM $X \pm$ SD	Standard error	1	
pН	_	3.89 ± 0.09	3.71 ± 0.03	0.024	**	
Total organic acids		80.0 ± 4.33	93.3 ± 10.52	2.126	**	
Lactic acid		50.3 ± 2.60	61.4 ± 5.88	1.472	**	
Acetic acid	g/kg DM	29.0 ± 2.16	31.5 ± 4.87	0.797	0.116	
Butyric acid		0.4 ± 0.30	0.1 ± 0.11	0.055	**	
Ethanol		13.2 ± 2.10	9.3 ± 2.41	0.606	**	
Ammonia N	g/kg total N	51.0 ± 10.29	38.0 ± 7.77	2.271	**	

* and ** denote significance at level 0.05 and 0.01, respectively.

For the animal feeding trial, 40 young beef cattle (8-9 months old) with similar mean live weight were used and divided into two analogous groups (20 animals each). The preexperimental adaptation for these animals lasted 21 days. The experimental period lasted 100 days.

During the preexperimental period, all animals were fed with untreated silage (CT) and a diet (CT + 4 kg of a commercial compound feed + 1 kg of barley straw) as during the experiment. During the experiment, each group was divided again into four subgroups, of five bulls each, and placed in four separate pens.

The animals were bedded on straw and had free access to water. Fresh silages were offered ad libitum twice daily, allowing for at least 10% orts (as-fed basis). Every 10 days the amount of silage fed and the refused silage were weighed over 2 consecutive days in order to calculate the daily silage intake. Silage DM intake was calculated per group as the difference between the amount of silage supplied and the amount of silage remaining. The compound feed was individually offered to all animals twice per day in a fixed amount. Barley straw was included in the diet (1 kg/animal/day; 88% of DM, energy value of 3.9 MJ ME/kg DM).

The animals were individually weighed on the first day of the experimental period and then once per month, and on the final day of the experiment. The average weight gain and growth rates were calculated for each animal and for each group. Feed conversion ratio was calculated as the ratio between feed intake and body weight gain.

Data was analyzed using variance analysis to test for the effect of silage treatments (Genstat, 1987). For the feed intake and feed conversion rates, a subgroup, of 5 beef cattle, was considered as the experimental unit. For body weight and daily weight gain, respectively, each animal within a group was considered an experimental unit. The Fisher's least significant difference (LSD) procedure at the 5% significance level was used to determine statistical differences between treatments. A probability of 0.05 < P < 0.10 was considered a near-significant trend.

3. Results and Discussion

The use of BSM significantly improved the silage quality compared to the CT (Table 2). The silage treated with BSM showed statistically significant higher DM recovery and

Treatment/statistical parameter	14	Trial day (kg, $X \pm SD$)				
	n	0	31	63	100 (trial end)	
Control	20	220.2 ± 11.83	249.1 ± 10.71	280.5 ± 9.80	320.0 ± 11.58	
Commercial product BSM	20	220.3 ± 12.07	249.5 ± 12.81	283.5 ± 14.14	328.1 ± 14.55	
Standard error	_	1.865	1.843	1.913	2.153	
P level	—	0.973	0.912	0.440	0.058	

TABLE 4: Average live weights of the beef cattle during the trial.

TABLE 5: Average dail	y body weight	gain of the beef	cattle in differe	nt trial periods.
	1 1			

Treatment/statistical	ท		Trial period in d	ays (kg, $X \pm SD$)	
parameter	11	0-31	32-63	64–100	0-100
Control	20	0.931 ± 0.124	0.981 ± 0.129	1.068 ± 0.074	0.998 ± 0.087
Commercial product BSM	20	0.940 ± 0.081	1.062 ± 0.129	1.206 ± 0.089	1.078 ± 0.078
Standard error		0.016	0.021	0.017	0.014
P level		0.778	0.055	**	**

** denotes significance at level 0.01.

digestible protein, coinciding with Merry et al. [14], lower DM losses (P < 0.01 for all) and higher crude protein content (P < 0.05). Its ADF content was significantly lower, and the metabolizable energy was higher (P < 0.05), whereas the digestible energy content was highly significant in the treated silage compared to the untreated silage. There were no significant differences between untreated and treated silages in crude fiber, NFE, WSC, and NDF content.

BSM treatment increased fermentation rates in wholecrop corn silages, resulting in a significant pH decrease (P < 0.01) and a significant increase in total organic acids concentration (P < 0.05) compared to the CT (Table 3). The lactic acid content in the BSM treatment was also significantly higher (P < 0.01) since homofermentative LAB were used [15]. The acetic acid content of the BSM treatment was numerically higher than that of the CT. Silage inoculation with BSM significantly decreased concentrations of butyric acid, ethanol, and ammonia-N (P < 0.01) of corn silage compared to the CT. Homofermentative silage inoculants by improving silage fermentation can reduce wasteful end products such as ammonia-N and volatile fatty acids, which result in poorer feed conversion efficiency and higher in-silo dry matter losses [16–18].

The use of silage inoculants containing homofermentative lactic acid bacteria, to increase lactic acid production and enhance the rate and extent of pH decline [19–21], can also lead to a reduction in protein breakdown [14]. As shown in Table 2, the BSM silage treatment decreased DM losses by 3.0% (P < 0.01) and had higher digestible energy (DE) and metabolic energy (ME) concentrations by 2.3 and 1.00% (P < 0.01 and P < 0.05), respectively, compared to the untreated CT silage.

During aerobic exposure after opening the silos, the CT (Figure 1)

(a) started to heat after 66 hours,

- (b) had a temperature increase of more than 2°C above the ambient temperature after 84 hours, and
- (c) reached a temperature of more than 14°C above the ambient temperature after 186 hours.

The temperature rise in the BSM treatment of more than 2° C above the ambient temperature occurred after 156 hours and reached a maximum temperature (+6°C above the ambient temperature) after 234 hours.

Therefore, BSM silage was more stable by 72 hours (3 days) compared to the CT. Recently, silage studies with whole-crop corn silages using obligatory heterofermentative LAB *L. buchneri* as an inoculant showed a 20-fold increase in the aerobic stability of the silage, which increased from approximately 40 hours for untreated silages to more than 790 hours for the inoculated silages [22]. Other studies [12, 23] provide more definitive evidence for the existence of certain LAB strains with the power to inhibit yeast and molds growth, and to improve aerobic stability. Some authors have described the positive aspect of the formation of acetic acid by heterofermentative lactic acid bacteria, which inhibits spoilage organisms [24, 25].

No statistical differences were found between animals fed with BSM or CT silages at 0, 31, and 63 trial days in the live weight (Table 4). From the data presented it is obvious that, throughout the whole trial, animals fed with BSM silage achieved higher average weights compared to those from the CT. At the end of the experiment the difference in body weight reached 8.1 kg/animal, and this was considered a near-significant trend (0.05 < P < 0.10).

Average daily weight gains (ADWGs) for BSM and CT are shown in Table 5. Between 0 and 31 trial days neither statistically, nor numerically marked, differences in ADWG were found between the treatments; however, in the trial period between 32 and 63 days the differences in ADWG



FIGURE 1: Temperature rise in a silage treated with a commercial product BSM (BSM) or not (CT). (Superscripts * and ** denote statistical differences of means at 0.05 and 0.01 levels, resp.).

TABLE 6: The effect of the treatment with the commercial product BSM on silage DM, energy intake, and feed conversion rate.

Daramatar	Unit	Treatment		CE	D
rarameter	Olin	Control $X \pm SD$	BSM $X \pm$ SD	5E	Γ
Silage DM intake		3.74 ± 0.12	3.97 ± 0.17	0.065	0.065
Compound feed DM intake	kg DM/animal/day	1.74 ± 0.0	1.74 ± 0.0	0.000	0.000
Total DM intake ¹		6.36 ± 0.12	6.59 ± 0.17	0.066	0.065
Total metabolizable energy (ME) intake	MJ/animal/day	69.27 ± 1.33	72.34 ± 1.97	0.799	*
Feed conversion rate	MJ of ME/kg gain	69.52 ± 3.49	67.15 ± 2.26	1.062	0.298

^{*} denotes statistical significance at level 0.05.

¹1 kg/animal/day of barley straw (88% of DM, 3.9 MJ ME/kg DM) was included in the diet for both treatments.

show a near-significant trend (0.05 < P < 0.10) with a *P* value of 0.055. The ADWG in the last third of the feeding trial period (from 64 to 100 days), and throughout the whole trial period (0 to 100 days), showed a statistically significant difference (*P* < 0.01) of 138 and 80 g, respectively.

In order to avoid differences due to different moisture contents, the intake is shown in Table 6 on the DM basis. Each mean is based on 4 observations. Randomized complete block where one pen is treated as a replication.

The silage DM intake for BSM was higher by 6.14% compared to the CT (3.97 versus 3.74 kg DM/animal/day) and had a near-significant trend (P = 0.065). As expected, because of the restricted feeding, no differences were found in compound feed DM intake. These results were similar to those reported by Muck and Kung Jr. [7]; however, some researchers found that feeding microbial inoculated silage to cattle does not affect dry matter intake compared to noninoculated silage [26, 26]. A combination of increased DM intake and higher energy, in the silage treated with BSM, led to a significant increase (P < 0.05) in metabolizable

energy intake compared to those animals fed with the CT. The animals receiving BSM had a better conversion of energy into body weight compared to that of the CT because they needed 2.37 MJ of ME (3.4%) less to increase the body weight by 1 kg. However, this difference was not statistically proven.

4. Conclusions

The microbial silage inoculant which was tested, based on homo- and heterofermentative lactic bacteria, had a significant positive effect on whole-crop corn silage quality in terms of

- (i) lowering pH and shifting fermentation towards lactic acid,
- (ii) suppressing butyric acid, ethanol, and ammonia-N formation,
- (iii) significantly reducing DM losses,
- (iv) statistically increasing digestible and metabolizable energy,

- (v) statistical significant improvement of aerobic stability, and
- (vi) improvement of the silage intake and the performance of beef cattle, and a positive effect on the utilization of feed energy.

Therefore, it is concluded that using such a silage inoculant improves the whole-plant corn silage characteristics and the nutritive value for beef cattle.

References

- R. J. Wilkins, L. Syrjala-Qvist, and K. K. Bolsen, "The future role of silage in sustainable animal production," in *Proceedings* of the 12th International Silage Conference, pp. 23–40, Uppsala, Sweden, 1999.
- [2] R. J. Merry, R. Jones, and K. M. Theodorou, "The conservation of grass," in *Grass. Its Production and Utilisation*, A. Hopkins, Ed., Blackwell Science, Oxford: UK, 3rd edition, 2000.
- [3] D. Ziggers, "Good or bad guys determine silage quality," *Dairy and beef*, vol. 2, pp. 27–29, 2003.
- [4] J. A. Rooke, F. M. Maya, J. A. Arnold, and D. G. Armstrong, "The chemical composition and nutritive value of grass silages prepared with no additive or with the application of additives either Lactobacillus plantarum or formic acid," *Grass and Forage Science*, vol. 43, pp. 87–95, 1988.
- [5] E. Saarisalo, E. Skyttä, A. Haikara, T. Jalava, and S. Jaakkola, "Screening and selection of lactic acid bacteria strains suitable for ensiling grass," *Journal of Applied Microbiology*, vol. 102, no. 2, pp. 327–336, 2007.
- [6] J. R. Weddell, R. Agnew, and B. Cottrill, "The UK forage additives approval scheme—developments and products approvals," in *Proceedings of the 13th International Silage Conference*, pp. 230–231, Auchincruive, UK, 2002.
- [7] R. E. Muck and L. Kung Jr., "Effects of silage additives on ensiling," in *Proceedings of the Silage: Field to Feed Bunk North American Conference*, no. NRAES -99, pp. 187–199, 1997.
- [8] E. Muck, F. E. Contreras, and D. R. Mertens, "Silage inoculant effects on in vitro rumen fermentation," *Journal of Animal Science*, vol. 85, p. 284, 2007.
- [9] I. Filya, A. Karabulut, and E. Sucu, "The effect of Lactobacillus plantarum and Lactobacillus buchneri on the fermentation, aerobic stability and ruminal degradability of corn silage in warm climate," in *Proceedings of the 13th International Silage Conference*, 2002.
- [10] N. J. Moon, "Inhibition of the growth of acid tolerant yeasts by acetate, lactate and propionate and their synergistic mixtures," *Journal of Applied Bacteriology*, vol. 55, no. 3, pp. 453–460, 1983.
- [11] Y. Tyrolová and A. Výborná, "Effect of the stage of maturity on the leaf percentage of lucerne and the effect of additives on silage characteristics," *Czech Journal of Animal Science*, vol. 53, no. 8, pp. 330–335, 2008.
- [12] N. K. Ranjit and L. Kung, "The effect of Lactobacillus buchneri, Lactobacillus plantarum, or a chemical preservative on the fermentation and aerobic stability of corn silage," *Journal* of Dairy Science, vol. 83, no. 3, pp. 526–535, 2000.
- [13] H. Danner, M. Holzer, E. Mayrhuber, and R. Braun, "Acetic acid increases stability of silage under aerobic conditions," *Applied and Environmental Microbiology*, vol. 69, no. 1, pp. 562– 567, 2003.
- [14] R. J. Merry, R. Jones, and M. K. Theodorou, "The conservation of grass," in *Grass. Its Production and Utilisation*, A. Hopkins, Ed., Blackwell Science, Oxford, UK, 3rd edition, 2000.

- [15] M. Marciňáková, A. Laukova, M. Simonova, V. Strompfova, B. Korenekova, and P. Naď, "Probiotic properties of Enterococcus faecium EF9296 strain isolated from silage," *Czech Journal of Animal Science*, vol. 53, pp. 336–345, 2008.
- [16] Davies D. R., "Silage inoculants—where next?" in *Proceedings* of the 14th International Symposium Forage Conservation, V. Jambor, S. Jamborova, B. Vosynkova, P. Prochacka, D. Vosynkova, and D. Kumprechtova, Eds., pp. 32–39, Mendel University, Brno, Czech Republic, 2010.
- [17] J. Jatkauskas and V. Vrotniakiene, "Fermentation characteristics and nutritive value of inoculated corn silage," in *Proceedings of the 20th general meeting of EGF*, pp. 1077–1079, Luzern, Switzerland, 2004.
- [18] G. Pahlow and H. Honig, "The role of microbial additives in the aerobic stability of silage," in *Proceedings of the 15th General Meeting of EGF*, pp. 149–152, The Netherlands, 1994.
- [19] L. Kung, M. R. Stokes, and C. J. Lin, "Silage additives," in *Silage Science and Technology*, D. R. Buxton, R. E. Muck, and J. H. Harison, Eds., Agronomy Series, no. 42, p. 305, Madison, Wis, USA, 360.
- [20] P. McDonald, A. R. Henderson, and S. J. E. Heron, Eds., *The Biochemistry of Silage*, Chalcombe, Bucks, UK, 2nd edition, 1991.
- [21] Z. G. Weinberg and R. E. Muck, "New trends and opportunities in the development and use of inoculants for silage," *FEMS Microbiology Reviews*, vol. 19, no. 1, pp. 53–68, 1996.
- [22] F. Driehuis, S. J. W. H. Oude Elferink, and S. F. Spoelstra, "Anaerobic lactic acid degradation during ensilage of whole crop maize inoculated with Lactobacillus buchneri inhibits yeast growth and improves aerobic stability," *Journal of Applied Microbiology*, vol. 87, no. 4, pp. 583–594, 1999.
- [23] R.A. Reis, G. R. Almeida, G. R. Siqueira, E. R. Bernardes, and E. Janusckiewicz, "Microbial changes and aerobic stability in high moisture corn silages inoculated with Lactobacillus buchneri," in *Proceedings of the 14th International Silage Conference*, R. S Park and M. D. Stronge, Eds., p. 223, Belfast, UK, 2005.
- [24] L. Cooke, "New strains slow silage spoilage," Agricultural Research, vol. 40, article 17, 1995.
- [25] J. A. Rooke, "Acetate silages: microbiology and chemistry. Landbauforsch," *Voelkenrode Sonderheft*, vol. 123, pp. 309– 312, 1991.
- [26] R. M. Luther, "Effect of microbial inoculation of whole-plant corn silage on chemical characteristics, preservation and utilization by steers," *Journal of Animal Science*, vol. 63, pp. 13–29, 1986.