

# Effect of Ketoconazole, a Cytochrome P450 Inhibitor, on the Efficacy of Quinine and Halofantrine against *Schistosoma mansoni* in Mice

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**Abstract:** The fear that schistosomes will become resistant to praziquantel (PZQ) motivates the search for alternatives to treat schistosomiasis. The antimalarials quinine (QN) and halofantrine (HF) possess moderate antischistosomal properties. The major metabolic pathway of QN and HF is through cytochrome P450 (CYP) 3A4. Accordingly, this study investigates the effects of CYP3A4 inhibitor, ketoconazole (KTZ), on the antischistosomal potential of these quinolines against *Schistosoma mansoni* infection by evaluating parasitological, histopathological, and biochemical parameters. Mice were classified into 7 groups: uninfected untreated (I), infected untreated (II), infected treated orally with PZQ (1,000 mg/kg) (III), QN (400 mg/kg) (IV), KTZ (10 mg/kg)+QN as group IV (V), HF (400 mg/kg) (VI), and KTZ (as group V)+HF (as group VI) (VII). KTZ plus QN or HF produced more inhibition ( $P < 0.05$ ) in hepatic CYP450 (85.7% and 83.8%) and CYT b5 (75.5% and 73.5%) activities, respectively, than in groups treated with QN or HF alone. This was accompanied with more reduction in female (89.0% and 79.3%), total worms (81.4% and 70.3%), and eggs burden (hepatic; 83.8%, 66.0% and intestinal; 68%, 64.5%), respectively, and encountering the granulomatous reaction to parasite eggs trapped in the liver. QN and HF significantly ( $P < 0.05$ ) elevated malondialdehyde levels when used alone or with KTZ. Meanwhile, KTZ plus QN or HF restored serum levels of ALT, albumin, and reduced hepatic glutathione (KTZ+HF) to their control values. KTZ enhanced the therapeutic antischistosomal potential of QN and HF over each drug alone. Moreover, the effect of KTZ+QN was more evident than KTZ+HF.

**Key words:** *Schistosoma mansoni*, quinine, halofantrine, ketoconazole, CYP3A4, glutathione, malondialdehyde

## INTRODUCTION

Schistosomiasis is a chronic and debilitating disease, caused by the blood-dwelling flukes of the genus *Schistosoma* that exacerbates poverty [1,2]. Although close to 800 million individuals are at risk of contracting the disease and over 200 million people are thought to be infected, schistosomiasis is often neglected [3]. The global burden of schistosomiasis has been estimated at 1.7 to 4.5 million disability adjusted life years (DALYs), but even the highest estimate might be an underestimation of the true burden [1,4].

Praziquantel (PZQ) is the only widely used drug for treat-

ment and control of this parasitic infection. Although no clinically relevant resistance to praziquantel has been described to date, development of drug-resistance remains a growing threat, particularly in view of mounting PZQ pressure [5]. Accordingly, in light of emerging resistance against PZQ, and in spite of the challenges associated with developing new antischistosomal drugs, alternatives to PZQ should be identified. There is no dedicated drug discovery and development program pursued for schistosomiasis, either by the pharmaceutical industry or through public-private partnerships. However, despite the paucity of concerted efforts to develop novel antischistosomal drugs, a number of compounds with promising antischistosomal properties have been identified, such as the synthetic trioxolanes [6], the cysteine protease inhibitor K11777 [7], alkylaminoalkanethiosulfuric acids [8], PZQ analogs [9], and the oxadiazoles [10].

Quinine (QN), a quinolone methanol, is the main alkaloid of *Cinchona* species [11]. Quinine has been used clinically in

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parenteral treatment of severe malaria and oral treatment of resistant *falciparum* malaria [12]. Quinine is a fast-acting drug with a short elimination half-life that has been recommended for the treatment of uncomplicated malaria in pregnant women and for drug-resistant malaria for almost 400 years [13]. Quinine is eliminated mainly by hepatic metabolism through cytochrome P450 (CYP) 3A4 with very little (< 15%) excreted unchanged in urine [14]. Seven metabolites have been identified with 3-hydroxyquinine (3-HQN) being the major metabolite [15]. Halofantrine (HF), a phenanthrenemethanol derivative of aminoalcohol, was first marketed in 1988. It was used for oral treatment of uncomplicated chloroquine- and multi-drug-resistant *Plasmodium falciparum* malaria in both adults and children [16,17]. Halofantrine is metabolized into *N*-debutylhalofantrine (HFM) essentially by CYP3A4. Maximal HF concentrations of plasma are reached at 2  $\mu$ M and its bioavailability is very weak and erratic [17], and hepatic clearance represents a large part of total clearance.

Interestingly, the artemisinins (artemether and artesunate), which are essential components of malaria treatment and control [18], also possess antischistosomal properties [19-21]. Recently, the antischistosomal properties in vitro and in the mouse model of several quinoline antimalarials, of which QN, HF, and mefloquine, with the highest worm reductions observed with mefloquine (> 80%) have been reported [22-24]. Meanwhile, Keiser et al. [22] supposed that the relatively lower worm reductions for QN (55%) and HF (51%) and other quinolines could be due to their reduced bioavailability.

Ketoconazole (KTZ), an imidazole derivative, is an orally active, broad-spectrum systemic antifungal agent extensively used in veterinary medicine. Ketoconazole is well known to potently inhibit CYP3A4 activity in human liver microsomes [25]. Since the major metabolic pathway of both QN and HF is through CYP3A4 subfamily [26,27], inhibition of the isoenzyme by KTZ could account for the increased plasma QN and HF levels. In the present study, and in the light of the above-mentioned data, we examined the inhibitory effects of KTZ on the antischistosomal potential of QN and HF against adult *S. mansoni* infection in mice, by evaluating parasitological, histopathological, and biochemical parameters.

## MATERIALS AND METHODS

### Chemicals and treatments

Bovine serum albumin (BSA), Tween-80, ethanol, 5,5'-di-

thiobis (2-nitrobenzoic acid) [DTNB, (Ellman's reagent)], ethylene diamine tetraacetic acid (EDTA), nicotinamide adenine dinucleotide phosphate reduced (NADPH), nitro blue tetrazolium (NBT), phenazine methosulphate (PMT), potassium dihydrogen phosphate ( $\text{KH}_2\text{PO}_4$ ), reduced glutathione (GSH), sodium dithionite, trichloroacetic acid (TCA), and thiobarbituric acid TBA were purchased from Sigma-Aldrich Chemical Co. (St. Louis, Missouri, USA). Ketoconazole tablets (Nizoral®; JASSEN-CILAG SpA., Italy, for Janssen Pharmaceutica N.V., Belgium) was given in a dose of 10 mg/kg/day prepared in ultrapure water containing 0.01 M hydrochloric acid for 3 consecutive days (on the 47<sup>th</sup>, 48<sup>th</sup> and 49<sup>th</sup> day post-infection) before QN or HF administration. Quinine tablets (Quinine Sulphate®; SmithKline Beecham Pharmaceuticals, Bentford, UK) and halofantrine tablets as halofantrine-HCl (HALFAN®; GlaxoSmithKline, Mayenne, France) were given in a single oral dose of 400 mg/kg as a fresh suspension in 7% (v/v) Tween-80 and 3% (v/v) ethanol. Treatment with either QN or HF started 2 hr after the last dose of KTZ. Praziquantel tablets (Distocide®; EIPICO, El-Asher Men Ramadan, Egypt) were given orally as a suspension in 2% cremophore-El (Sigma-Aldrich) in a dose of 1,000 mg/kg divided equally on 2 consecutive days at day 49 post-infection (PI).

### Animals and infection

A total of 56 male CD1 Swiss albino mice, average weight  $20 \pm 2$  g, were bred and maintained at the Schistosome Biology Supply Center (SBSC) of Theodor Bilharz Research Institute (TBRI), Giza, Egypt. Animals were housed in a controlled temperature and light environment, and were given water and commercial chow *ad libitum* (Tanta Company for Oils and Soap, Tanta, Egypt). After approval by the Institutional Review Board (IRB) of TBRI, the animal experiments were conducted according to the international ethical guidelines at the animal unit. Cercariae of *S. mansoni* (Egyptian strain) were obtained from infected intermediate host snails maintained at SBSC. Animals were infected with  $80 \pm 10$  cercariae/mouse using the body immersion technique [28].

### Animal grouping

Mice were grouped as follows: uninfected untreated control (I), infected untreated (only received drug vehicles) (II), infected and treated with PZQ (III), infected and treated with QN (IV), infected and treated with both KTZ and QN (V), infected and treated with HF (VI), and infected and treated with both

KTZ and HF (VII).

Mice were killed by rapid decapitation at day 63 PI (day 14 after treatment). After decapitation, blood was collected and sera were separated by centrifugation at 1,850 g for 10 min and stored frozen at -70°C pending assay.

#### Assessment of parasitological criteria

After killing, hepatic and portomesenteric vessels of mice were perfused according to Duvall and De Witt [29], with no resort to general anesthesia, for worm recovery and subsequent counting. Immediately after perfusion, the liver was isolated, chilled on ice and divided into 2 unequal pieces. The small piece of the liver with the middle part of the small intestine was used for determining the number of eggs per gram of liver or intestinal tissues [30], while the other piece of the liver was used for hepatic enzyme assessment. The oogram pattern of the different egg developmental stages in small intestine tissues was also examined [31], in which eggs at different stages of maturity were identified (in groups I-IV) according to the size of the embryo and were counted. In addition, mature eggs containing fully developed miracidia and dead eggs (granular, dark, and semitransparent) were also counted in 3 fragments of the small intestine and the mean number of each stage was calculated.

#### Histopathology and granuloma measurement

Pieces of the liver recovered from mice were fixed in 10% neutral buffered formalin and processed to paraffin blocks. Sections (4 µm thick) were cut into 250 µm in apart from the proceeding sections to avoid measuring the same granuloma. Five paraffin liver sections were prepared from each animal and stained with hematoxylin and eosin (H-E) and Masson trichrome stains to examine the histopathological changes and for measurement of hepatic granuloma diameters (30/mouse). Measurements of the granulomas were conducted on non-contiguous granulomas, each containing a single egg (with intact or degenerated miracidia), using an ocular micrometer. The mean diameter of each granuloma was calculated by measuring 2 diameters of the lesion at right angles to each other [32]. Egg viability was assessed microscopically in the same liver sections and cell composition of granulomas was also investigated.

#### Preparation of liver homogenates

Liver tissue was homogenized in 4 volumes (w/v) of ice-cold 0.1 M potassium phosphate buffer (pH 7.4) containing 1 mM

EDTA, followed by sequential centrifugation in a Christ cooling centrifuge at 4°C and 600 g for 10 min and then at 10,000 g for 20 min. The first half of the supernatant was collected and kept at -80°C for subsequent analysis for determination of liver content of GSH, superoxide dismutase (SOD), and lipid peroxidation as expressed by malondialdehyde formation. The second half of the supernatant was subjected to ultracentrifugation at 105,000 g for 60 min at 4°C in a Sorval ultracentrifuge to yield the microsomal pellet which was resuspended in 0.1 M potassium phosphate buffer (pH 7.4) and stored at -80°C [33] until assay of the cytochrome P-450 (CYP450) and cytochrome b5 (CYT b5) activities.

#### Glutathione (GSH) assay

The level of GSH was determined in the liver homogenate according to the method of Ellman [34]. A 0.5-ml aliquot of the homogenate was added to a tube with 0.5 ml of 10% TCA. The tubes were centrifuged at 3,000 g for 10 min. A 0.2-ml aliquot of the resulting supernatant was added to a tube containing 5 ml of 0.1 M phosphate buffer and 0.1 ml of DTNB (Ellman's reagent), and the absorbance was measured at 412 nm. With the help of the standard curve drawn using gradual concentrations of a standard GSH, the content of GSH in the liver homogenates of the experimental animals were calculated.

#### Superoxide dismutase (SOD) assay

SOD activity was assayed spectrophotometrically at 560 nm by the procedure of Winterbourn et al. [35]. The activity of SOD depends on its ability to inhibit phenazine methosulphate (PMT)-mediated reaction of nitroblue tetrazolium (NBT) dye. One unit of enzyme activity is defined as the amount of enzyme that causes half-maximal inhibition of NBT reduction. Activity was expressed in µmole/min/g liver.

#### Assessment of hepatic lipid peroxidation products

Degree of lipid peroxidation in the liver tissue homogenate of mice was determined in terms of thiobarbituric acid reactive substances (TBARS) formation [36]. One-ml of supernatant was mixed with 1-ml of TCA (10% w/v) in a centrifuge tube and centrifuged at 1,850 g for 15 min at room temperature. One-ml of TBA solution (0.67% w/v) was added to 1-ml of supernatant and kept in a boiling water bath for 45 min. Absorbance was read after cooling at 530 nm against a blank containing all the reagents except the tissue homogenate. As 99% of the TBARS is malondialdehyde, TBARS concentrations of the

samples were calculated using the extinction coefficient of malondialdehyde, which is  $1.56 \times 10^5 \text{ M}^{-1}\text{cm}^{-1}$ .

#### CYP450 and CYT b5 assay

CYP450 and CYT b5 activities were determined spectrophotometrically using a double-beam spectrophotometer (Lambda 3B, Perkin Elmer, Waltham, Massachusetts, USA) by the method of Omura and Sato [37]. In its reduced form, CYP450 binds with carbon monoxide (CO) gas to give a complex with a maximum absorbance in the solet region at 450 nm. Accordingly, it was determined by the CO-difference spectrum of sodium dithionite-reduced samples using an extinction coefficient of 91 mM per Cm for the difference between 450 and 490 nm. Cytochrome b5 was determined by measuring the difference in its spectrum between the sodium dithionite-reduced samples and oxidized form using an extinction coefficient of 185/mM per Cm for the difference between 409 and 424 nm. The specific activities of CYP450 and CYT b5 enzymes were expressed as units of the enzyme content/mg microsomal protein. Microsomal protein concentrations were assayed according to the method of Lowry [38].

#### Serum enzymes

Concentrations of alanine aminotransferase (ALT) and albumin in the collected sera were estimated using the available commercial kits.

#### Statistical analysis

Results were analyzed with the use of SPSS software (Version 9.0). Data are expressed as the mean  $\pm$  SD. Comparison between the mean values of different parameters in the studied groups was performed using one-way ANOVA test, with post-hoc using LSD test. The data were considered significant if  $P$ -

value was  $\leq 0.05$ .

## RESULTS

#### Parasitological studies

Data in Table 1 shows a significant ( $P < 0.05$ ) decrease in males, females, and total number of worms in all treated groups which was to a lower extent for total males in the groups treated with QN, KTZ+HF, and HF compared to infected group. In addition, all treatment regimens except HF-treated group decreased hepatic and intestinal ( $P < 0.05$ ) tissue egg loads significantly accompanied with an increase ( $P < 0.05$ ) in the percentage of dead eggs and a decrease ( $P < 0.05$ ) in the percentage of immature and mature eggs when compared to infected control (Table 2).

Treatment with KTZ+QN significantly enhanced the decrease ( $P < 0.05$ ) in female, total number of worms, hepatic and intestinal tissue egg loads, the percentage of immature and mature ( $P < 0.05$ ) eggs and the increase in the percentage of dead eggs ( $P < 0.05$ ) vs the group treated with QN alone.

**Table 1.** Effects of QN or HF alone and in addition to KTZ on the number of male and female worms in *S. mansoni*-infected mice at week 9 post-infection (i.e., week 2 post-treatment)

Animal groups	Total no. males	Total no. females	Total no. worms
Infected	14.4 $\pm$ 6.1	10.1 $\pm$ 3.1	24.5 $\pm$ 4.8
PZQ	0.3 $\pm$ 0.3 <sup>a</sup> (97.8)	0.6 $\pm$ 0.7 <sup>a</sup> (94.0)	0.9 $\pm$ 0.5 <sup>a</sup> (96.2)
QN	5.8 $\pm$ 3.8 <sup>a</sup> (59.6)	2.7 $\pm$ 1.4 <sup>a</sup> (73.9)	8.5 $\pm$ 4.2 <sup>a</sup> (65.5)
KTZ + QN	3.5 $\pm$ 1.9 <sup>a</sup> (76.0)	1.1 $\pm$ 0.6 <sup>ab</sup> (89.0)	4.6 $\pm$ 1.6 <sup>ab</sup> (81.4)
HF	6.6 $\pm$ 6.0 <sup>a</sup> (54.4)	3.9 $\pm$ 2.8 <sup>a</sup> (61.8)	10.4 $\pm$ 3.8 <sup>a</sup> (57.5)
KTZ + HF	5.2 $\pm$ 4.7 <sup>a</sup> (64.0)	2.1 $\pm$ 2.2 <sup>a</sup> (79.3)	7.3 $\pm$ 3.3 <sup>a</sup> (70.3)

No. of animals in each group; 8 mice. Values are presented as the mean  $\pm$  SD. Numbers in the parentheses indicate the percentage of reduction from infected control.

<sup>a</sup>Significant difference from infected group at  $P < 0.05$ .

<sup>b</sup>Significant difference from QN group at  $P < 0.05$ .

**Table 2.** Effects of QN or HF alone and in addition to KTZ on the percentage of egg developmental stages and tissue egg load in *S. mansoni*-infected mice at week 9 post-infection (i.e., week 2 post-treatment)

Animal groups	Egg developmental stages (%)			Tissue egg load $\times 10^3$	
	Immature	Mature	Dead	Hepatic	Intestinal
Infected	56.14 $\pm$ 9.84	36.56 $\pm$ 9.28	7.30 $\pm$ 1.64	17.98 $\pm$ 5.40	26.02 $\pm$ 4.24
PZQ	0.00 $\pm$ 0.00 <sup>a</sup>	0.00 $\pm$ 0.00 <sup>a</sup>	100 $\pm$ 0.00 <sup>a</sup>	1.26 $\pm$ 1.16 <sup>a</sup> (93.0)	0.57 $\pm$ 0.31 <sup>a</sup> (97.8)
QN	12.26 $\pm$ 5.77 <sup>a</sup>	19.61 $\pm$ 6.11 <sup>a</sup>	68.13 $\pm$ 8.80 <sup>a</sup>	6.33 $\pm$ 3.22 <sup>a</sup> (64.8)	11.32 $\pm$ 3.17 <sup>a</sup> (56.5)
KTZ + QN	6.13 $\pm$ 3.25 <sup>ab</sup>	10.63 $\pm$ 6.02 <sup>ab</sup>	83.24 $\pm$ 12.05 <sup>ab</sup>	2.91 $\pm$ 0.65 <sup>ab</sup> (83.8)	8.32 $\pm$ 1.81 <sup>ab</sup> (68.0)
HF	22.14 $\pm$ 7.52 <sup>a</sup>	29.45 $\pm$ 8.63	48.41 $\pm$ 6.11 <sup>a</sup>	8.81 $\pm$ 2.23 <sup>a</sup> (51.0)	13.53 $\pm$ 3.65 <sup>a</sup> (48.0)
KTZ + HF	14.32 $\pm$ 3.56 <sup>ac</sup>	19.97 $\pm$ 7.44 <sup>ac</sup>	65.71 $\pm$ 10.38 <sup>ac</sup>	6.11 $\pm$ 3.31 <sup>a</sup> (66.0)	9.23 $\pm$ 3.08 <sup>ac</sup> (64.5)

No. of animals in each group; 8 mice. Values are presented as the mean  $\pm$  SD. Numbers in the parentheses indicate the percentage of reduction from infected control.

<sup>a</sup>Significant difference from infected group at  $P < 0.05$ ; <sup>b</sup>Significant difference from QN group at  $P < 0.05$ ; <sup>c</sup>Significant difference from HF group at  $P < 0.05$ .

Meanwhile, the treatment with KTZ+HF numerically decreased males, females, and total number of worms, but significantly ( $P < 0.05$ ) enhanced both the reduction in the percentage of

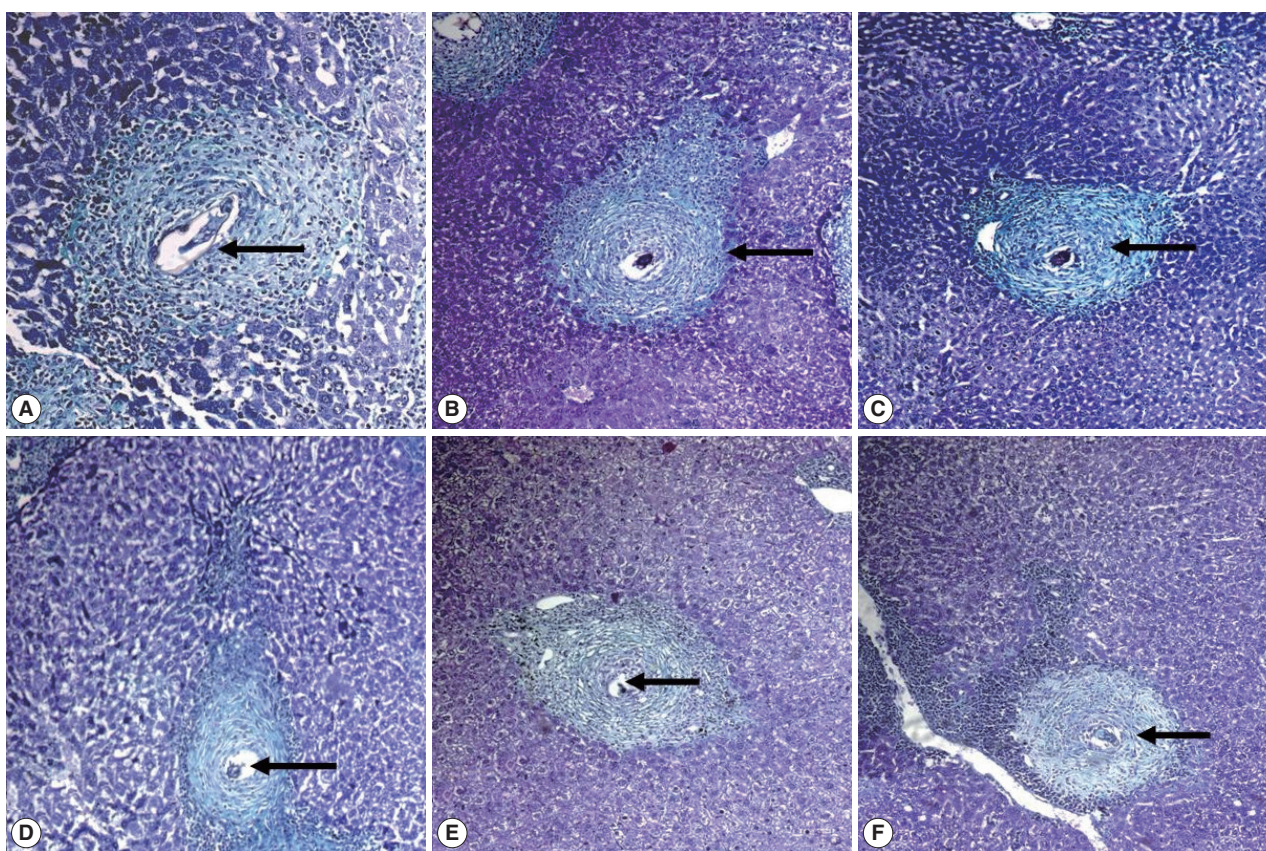
immature and mature eggs and intestinal tissue egg load and the increase in the percentage of dead eggs ( $P < 0.05$ ) vs the group treated with HF alone (Tables 1, 2).

**Table 3.** Effect of QN or HF alone and in addition to KTZ on granuloma diameter and associated histopathological changes in *S. mansoni*-infected mice 9 weeks post-infection (i.e., 2 weeks post treatment)

Animal groups	Granuloma diameter	Types of granuloma (%)			State of <i>S. mansoni</i> eggs (%)	
		Cellular	Fibrocellular	Fibrous	Intact	Degenerated
Infected	297.1 ± 26.7	14.3	85.7	0	75.7 ± 10.4	24.3 ± 10.4
PZQ	201.3 ± 18.1 <sup>a</sup> (32.3)	0.0	100.0	0	27.4 ± 10.3 <sup>a</sup>	72.1 ± 10.3 <sup>a</sup>
QN	176.9 ± 19.8 <sup>a</sup> (40.5)	25.0	75.0	0	38.1 ± 8.4 <sup>a</sup>	61.9 ± 8.4 <sup>a</sup>
KTZ + QN	148.1 ± 10.7 <sup>a,b</sup> (50.2)	50.0	50.0	0	23.1 ± 6.4 <sup>a,b</sup>	76.9 ± 6.4 <sup>a,b</sup>
HF	188.8 ± 24.7 <sup>a</sup> (36.5)	25.0	75.0	0	46.4 ± 8.6 <sup>a</sup>	53.6 ± 8.6 <sup>a</sup>
KTZ + HF	177.5 ± 16.2 <sup>a</sup> (40.3)	37.5	62.5	0	42.5 ± 13.1 <sup>a</sup>	57.5 ± 13.1 <sup>a</sup>

No. of animals in each group; 8 mice. Values are presented as the mean ± SD. Numbers in the parentheses indicate the percentage of reduction from infected group.

<sup>a</sup>Significant difference from infected group at  $P < 0.05$ ; <sup>b</sup>Significant difference from QN group at  $P < 0.05$ .



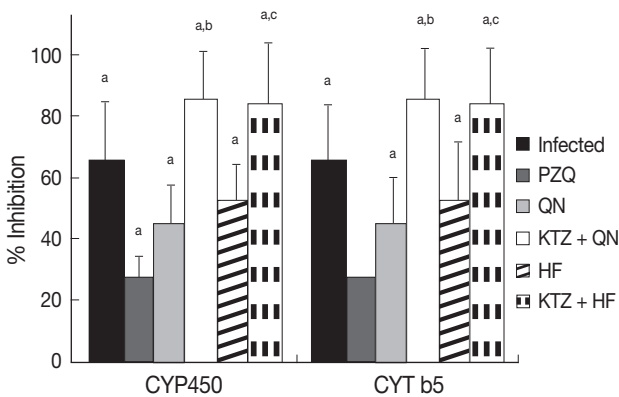
**Fig. 1.** Masson's trichrome stained liver sections of (A) *S. mansoni*-infected untreated mice showing irregularly outlined large fibrocellular granuloma formed of a central egg (with a living miracidium; arrow), surrounded mainly by lymphocytes, eosinophils, and extensive collagen depositions as concentric collagen fibers. (B) PZQ-treated group showing medium circumscribed fibrocellular granuloma (arrow) with eggs starting degeneration. (C, D) QN and KTZ+QN-treated groups, respectively, showing the smallest size and well-demarcated fibrocellular granuloma, formed of central degenerated egg (D; arrow), surrounded by lymphocytes, eosinophils, neutrophils, scattered macrophages, and fewer collagen depositions (C; arrow). (E, F) HF and KTZ+HF-treated groups, respectively, showing small fibrocellular granuloma formed of central degenerated egg (E; arrow), surrounded mainly by lymphocytes, eosinophils, neutrophils, and few concentric collagen fibers (F; arrow) ( $\times 200$ ).

**Histopathological studies**

Liver sections of infected control and treated groups were studied for granuloma size and cellular profiles. All treated groups showed significant reductions ( $P < 0.05$ ) in the mean granuloma diameters when compared to the infected group. KTZ+QN-treated group recorded the smallest granuloma diameter (50.2% reduction) followed by QN-treated group (40.5% reduction) and KTZ+HF-treated group (40.3% reduction) (Table 3; Fig. 1). Moreover, a significant reduction in granuloma diameter ( $P < 0.05$ ) was observed in KTZ+QN-treated group vs QN-treated group. The percentages of degenerated eggs were significantly ( $P < 0.05$ ) higher in all treated groups compared with the infected group. In addition, there was a significant increase ( $P < 0.05$ ) in the percentages of degenerated eggs in KTZ+QN-treated group vs QN-treated group

(Table 3).

At week 9 PI, the inflammatory granulomatous lesions were mainly seen in the hepatic parenchyma and to a lesser extent in the portal tracts. Granulomas were fibrocellular (85.7%) and cellular (14.3%) consisting of central eggs (most of the eggs inside the granulomas exhibited intact cellular miracidia; Fig. 1A) surrounded by numerous eosinophils, neutrophils, lymphocytes, few epitheloid cells, macrophages, and concentric or scattered collagen fibers. In contrast, granulomas of KTZ+QN, KTZ+HF, QN and HF-treated groups were smaller in size and fibrocellular (50.0%, 62.5%, 75.0%, and 75.0%, respectively) and cellular (50.0%, 37.5%, 25.0%, and 25.0%, respectively) formed of central eggs where the majority of miracidia were dead surrounded by lymphocytes and peripherally located eosinophils, epitheloid cells, and macrophages (Table 3; Fig. 1C-F).



**Fig. 2.** Effect of QN or HF alone and in addition to KTZ on percent inhibitions in hepatic CYP450 and CYT b5 relative to uninfected group in *S. mansoni*-infected mice at week 9 PI (i.e., week 2 post treatment). No. of animals in each group; 8 mice. Values are presented as the mean  $\pm$  SD. <sup>a</sup>Significant difference from uninfected group at  $P < 0.05$ . <sup>b</sup>Significant difference from QN group at  $P < 0.05$ . <sup>c</sup>Significant difference from HF group at  $P < 0.05$ .

**Enzyme assessments**

Infection of mice with *S. mansoni* significantly decreased ( $P < 0.05$ ) tissue GSH content, CYP450, CYT b5, SOD activities, and serum albumin, while increased ( $P < 0.05$ ) tissue malondialdehyde and serum ALT levels compared to uninfected group (Table 4). Treatment with PZQ alone approximately recovered the contents of GSH, the activities of SOD, CYT b5, and the levels of serum ALT and albumin relative to uninfected group. Treatment of *S. mansoni*-infected mice with QN or HF alone did not produce any significant change in hepatic antioxidant enzymes, CYP450, and CYT b5 activities, or in serum ALT and albumin levels vs those of uninfected group. Meanwhile, treatment with either QN or HF alone and in addition to KTZ elevated malondialdehyde levels. Moreover, the combined treatment regimens with either KTZ+QN or KTZ+HF produced further inhibition in the activities of CYP450 ( $P < 0.05$ ; 85.7% and 83.8%) and of CYT b5 ( $P < 0.05$ ; 75.5% and

**Table 4.** Effect of QN or HF alone and in addition to KTZ on liver antioxidant capacity, CYP450 and CYT b5 activities and hepatic function in *S. mansoni*-infected mice 9 weeks post-infection (i.e., 2 weeks post treatment)

Animal groups	GSH	SOD	MDA	CYP450	CYT b5	ALT	Albumin
Uninfected	4.3 $\pm$ 0.6	386.5 $\pm$ 17.6	29.8 $\pm$ 5.2	2.2 $\pm$ 0.5	3.0 $\pm$ 0.8	11.7 $\pm$ 2.8	3.4 $\pm$ 0.6
Infected	2.9 $\pm$ 0.5 <sup>a</sup>	245.6 $\pm$ 20.1 <sup>a</sup>	61.9 $\pm$ 6.1 <sup>a</sup>	0.8 $\pm$ 0.2 <sup>a</sup>	1.1 $\pm$ 0.4 <sup>a</sup>	26.7 $\pm$ 11.6 <sup>a</sup>	2.4 $\pm$ 0.4 <sup>a</sup>
PZQ	3.8 $\pm$ 0.5	372.2 $\pm$ 23.4	38.3 $\pm$ 6.8 <sup>a</sup>	1.6 $\pm$ 0.4 <sup>a</sup>	2.4 $\pm$ 0.6	14.0 $\pm$ 2.8	3.3 $\pm$ 0.5
QN	3.1 $\pm$ 0.5 <sup>a</sup>	312.8 $\pm$ 19.5 <sup>a</sup>	64.3 $\pm$ 7.4 <sup>a</sup>	1.2 $\pm$ 0.3 <sup>a</sup>	1.6 $\pm$ 0.5 <sup>a</sup>	17.5 $\pm$ 2.3 <sup>a</sup>	2.7 $\pm$ 0.5 <sup>a</sup>
KTZ+QN	3.7 $\pm$ 0.4 <sup>a,b</sup>	341.7 $\pm$ 24.2 <sup>a,b</sup>	59.4 $\pm$ 10.4 <sup>a</sup>	0.3 $\pm$ 0.1 <sup>a,b</sup>	0.7 $\pm$ 0.3 <sup>a,b</sup>	14.8 $\pm$ 3.1	3.2 $\pm$ 0.6
HF	3.4 $\pm$ 0.6 <sup>a</sup>	293.7 $\pm$ 16.2 <sup>a</sup>	69.4 $\pm$ 8.2 <sup>a</sup>	1.0 $\pm$ 0.2 <sup>a</sup>	1.4 $\pm$ 0.5 <sup>a</sup>	14.9 $\pm$ 3.1 <sup>a</sup>	2.8 $\pm$ 0.5 <sup>a</sup>
KTZ+HF	3.8 $\pm$ 0.4	314.4 $\pm$ 17.9 <sup>a,c</sup>	61.5 $\pm$ 10.7 <sup>a</sup>	0.4 $\pm$ 0.1 <sup>a,c</sup>	0.8 $\pm$ 0.3 <sup>a,c</sup>	14.4 $\pm$ 3.4	3.3 $\pm$ 0.6

Units: GSH,  $\mu$ mole/g liver; SOD,  $\mu$ mole/min/g liver; MDA, nmole/g liver; CYP450 and CYT b5, nmole/mg protein; ALT, U/L; Albumin, g/100 ml. No. of animals in each group; 8 mice. Values are presented as the mean  $\pm$  SD.

<sup>a</sup>Significant difference from uninfected group at  $P < 0.05$ ; <sup>b</sup>Significant difference from QN group at  $P < 0.05$ ; <sup>c</sup>Significant difference from HF group at  $P < 0.05$ .

73.5%), respectively, relative to groups treated with QN or HF alone (Table 4; Fig. 2). Furthermore, they restored tissue GSH (KTZ+HF) and the serum levels of ALT and albumin (KTZ+QN or KTZ+HF) to their control values.

## DISCUSSION

To our knowledge, this is the first experimental investigation documenting the inhibitory effects of the CYP3A4 inhibitor KTZ on the antischistosomal potentials of QN and HF against adult Egyptian *S. mansoni* infection. In the current work, infection of mice with *S. mansoni* produced a significant decrease in the hepatic activities of CYP450 and CYT b5 (65.3% and 62.1%, respectively). These findings are in agreement with previous investigations [39-41] and are attributed to denaturation of the CYP450 to its inactive form CYP422 as a result of different pathological reactions that follow egg deposition in the liver, including granulomatous, immunologic, and fibrogenic reactions [42]. Moreover, administration of KTZ in the present investigation before QN or HF enhanced the inhibition in the activities of CYP450 (85.7% and 83.8% vs 45.4% and 52.8%) and CYT b5 (75.5% and 73.5% vs 46.0% and 53.7%, respectively) than observed in mice treated with QN or HF alone. It is likely that this pronounced reduction is the augmenting effect of both KTZ and *S. mansoni* infection. Previous studies reported that KTZ potently inhibit CYP3A4 activity in human liver microsomes and modifies considerably the pharmacokinetics of CYP3A4 substrates such as tacrolimus [25]. Furthermore, KTZ competitively inhibits CYP3A12, a major isoenzyme of the CYP3A subfamily, and at a therapeutic dose, decreases the total body clearance of the CYP3A substrates midazolam [43] in beagle dogs. In addition, KTZ has inhibitory potency for not only CYP3A but also P-glycoprotein (P-gp) in the rat liver [44]. Likely, our work suggests that oral KTZ affected the hepatic CYP3A4 in mice.

In the present study, the efficacy of QN or HF against adult *S. mansoni* infection following their administration alone or in addition to KTZ was assessed. Administration of QN or HF alone produced significant reductions in males (59.6% and 54.4%, respectively), females (73.9% and 61.8%, respectively) and total number of worms (65.5% and 57.5%, respectively) compared to infected group. In addition, they decreased hepatic, intestinal tissue egg loads, the percentage of immature and mature eggs and increased the percentage of dead eggs. In this context, Xiao et al. [45] found 80.9-90.3% reduction in

total worm burden of adult *S. japonicum* in mice after a single oral dose of 400 mg/kg of mefloquine, quinine, and quinidine, while halofantrine and lumefantrine showed moderate and poor effects with total worm reductions of 67.5% and 38.4%, respectively. On the other hand, Corrêa Soares et al. [46] recorded that treatment of *S. mansoni*-infected female Swiss mice with daily intraperitoneal injections of QN (75 mg/kg/day) from the 11<sup>th</sup> to 17<sup>th</sup> day after infection caused significant decreases not only in worm burden (39%) but also in female worm burden (40%) and egg production (42%). Moreover, Keiser et al. [22] reported that QN and HF administered orally at a single dose of 400 mg/kg to mice harboring adult *S. mansoni* (Liberian strain) resulted in total and female worm reductions of 54.7% and 51.7%, and 62.0% and 57.4%, respectively. In the present investigation, it seems that worms of the Egyptian strain of *S. mansoni* were more susceptible to QN and HF than the Liberian strain.

The detailed mechanism of action of quinoline methanols on schistosomes remains to be investigated. It was demonstrated that the antimalarial chloroquine inhibits the formation of hemozoin (Hz), a heme detoxification aggregate in *S. mansoni* female homogenates [47]. It is interesting to note that adult female *S. mansoni* was more affected by QN or HF than male adult *S. mansoni*. Differences in drug susceptibility between male and female *S. mansoni* have been reported previously, e.g., following hycanthone treatment [48] and point to a sex-specific interference of the drug with the target, or different drug targets. In addition, it was shown that 4-aminoquinolines interact with "free" heme, hindering its crystallization into Hz. The "free" heme interacts with membranes and exerts severe toxic effects, ultimately killing the parasite through oxidative stress [49]. In this regard, Corrêa Soares et al. [46] reported that interference with heme crystallization in *S. mansoni* represents an important mechanism by which antimalarial quinoline methanols exerts their antischistosome action. From all of the 3 compounds the authors tested, QN and quinidine (QND) caused the most striking reductions in the Hz content (40-65%) on female worms, which are more susceptible to interference in this process [47]. Hz formation represents the main heme detoxification mechanism in *S. mansoni*, accounting for more than 50% of total heme content in female worms [50]. Moreover, it was concluded that reduction in total and female worm load, obtained after treatment with QN, QND, and quinacrine (QCR), was positively correlated with the percentage of Hz content reduction in female worms [46]. The

present work, together with previous evidence, describes schistosomicidal activity of QN and HF [22,46].

Herein, results revealed that administration of KTZ prior QN or HF significantly enhanced the therapeutic efficacy against *S. mansoni* infection vs those treated with QN or HF alone. These effects were more pronounced in KTZ+QN-treated group than in KTZ+HF-treated group. In the present study, the enhanced reduction in female and total number of worms in *S. mansoni*-infected mice treated with KTZ+QN or KTZ+HF was in accordance with the results on the activities of drug-metabolizing enzymes, because mice treated with QN or HF in addition to KTZ showed more inhibition in the activities of CYP450 and CYT b5. Lower metabolism as a result of lower levels of drug-metabolizing enzymes because KTZ possibly leads to increased QN or HF plasma concentrations and to a greater elimination half-life, which in turn increased the length of parasite exposure to the drug.

Schistosome eggs induce liver fibrosis, which is a common pathological process leading to the development of irreversible cirrhosis [51] and inability of the liver to perform its biochemical functions [52]. In the present investigation, the smallest granuloma diameter (50.2% reduction) was recorded in KTZ+QN-treated group followed by QN-treated group (40.5% reduction), KTZ+HF-treated group (40.3% reduction) and PZQ-treated group (32.3%) when compared to the infected group. Moreover, a significant reduction in granuloma diameter was observed in KTZ+QN-treated group vs QN-treated group. Such reductions in granuloma diameter may be attributed to elimination of adult worms by the treatment regimens leading to sustained diminution of egg-induced immunopathology [53] and consequent stopping of further egg deposition [24,54]. The combined treatment regimen of KTZ+QN after *S. mansoni* infection had an added merit of maximal amelioration of histopathological changes where the liver sections of this group showed remarkably better cellular architecture upon comparing with the *S. mansoni*-infected untreated and QN-treated groups. Moreover, granuloma in this group showed either partially or almost completely degenerated miracidia beside the minimal surrounding inflammatory reactions. This may be related to either the enhanced killing of the parasite, with marked reduction in egg deposition as evidenced by the parasitological results or because of immunological and anti-inflammatory effects of QN. This is in accordance with Corrêa Soares et al. [46] who observed that QN treatment promoted remarkable ultrastructural changes in male and female worms, particularly in

the gut epithelium and reduced the granulomatous reactions to parasite eggs trapped in the liver. Although nothing is known about the presence or expression of P-glycoprotein (P-gp) in *S. mansoni* eggs, P-gp has been observed in external layers of the egg shells of the red stomach worm, *Haemonchus contortus*, a parasitic nematode of ruminants, suggested that P-gp inhibition might contribute to elimination of parasite eggs [55].

Previous studies have reported that schistosomiasis is associated with free radical liberation and disturbance in the cellular antioxidant system [56,57]. The intracellular concentration of reactive oxygen species (ROS) is tightly regulated by multiple defense mechanisms involving ROS scavenging antioxidant enzymes and small antioxidant molecules. In the present study, *S. mansoni* infection induced a significant decrease in SOD activity, GSH content, serum albumin level, and enhanced lipid peroxidation (malondialdehyde) as well as the serum level of ALT, and these results are in agreement with those previously obtained [20,57-60]. Fernandez-Checa and Kaplowitz [61] reported that this alteration might influence the capability of the liver to provide protection against oxidative damage. The balance between biosynthesis, uptake, oxidation, and export maintains GSH homeostasis at the cellular level. Its reduction in *S. mansoni*-infected liver tissues is probably related to a reduced synthesis of the tripeptide by the infected liver, which was evidenced by the reduction in serum albumin in this study. It was manifested by increase in malondialdehyde level that lipid peroxidation was enhanced during infection. The decreased activity of SOD in *S. mansoni*-infected mice may be due to the enhanced lipid peroxidation or accumulation of superoxide radicals and hydrogen peroxide [62].

In the present work, the administration of PZQ alone approximately recovered the contents of GSH, the activities of SOD, and the levels of serum ALT and albumin relative to uninfected group. This could be due to the antischistosomal properties of PZQ as it eradicates parasites, prevents the possible formation of toxic metabolites of *S. mansoni* worms, abolishes further insults to the liver through interruption of egg deposition, and to its direct effect on mature miracidia within egg granulomas [63] and stopping emission of soluble egg antigens.

Herein, treatment of infected mice with QN or HF alone did not produce any significant change in hepatic antioxidant enzymes or in serum ALT and albumin levels vs those of uninfected group. Similarly, Corrêa Soares et al. [46] reported no changes in malondialdehyde levels in *S. mansoni*-infected mice



treated with QN, which may be attributed to the increased expression of aldehyde dehydrogenase (ADH) gene, which encodes an enzyme involved in detoxification of lipid peroxidation products such as reactive malondialdehyde and hydroxynonenal [64]. On the other hand, Farombi et al. [65,66] reported a decrease in GSH contents and an elevation of malondialdehyde levels upon HF treatment in rat liver microsomes. Moreover, Adaramoye et al. [67] documented that HF treatment significantly reduced GSH and SOD, while increased malondialdehyde in rat liver as well as their serum ALT. Meanwhile, the treatment with either QN or HF alone and in addition to KTZ elevated malondialdehyde levels. Nevertheless, the combined treatment regimens restored serum levels of ALT, albumin, and hepatic GSH (KTZ+HF) to their control values. These results indicate that the liver microsomal lipid peroxidation could be enhanced by quinoline methanols, QN, and HF, and they could have adverse effects on the antioxidant status of the animals. Under such a condition, the need for antioxidants as adjuvant to antimalarials is recommended, and their presence may be crucial to eliminate the products of oxidative reactions.

In conclusion, administration of KTZ, a CYP3A4 inhibitor, with QN or HF produced more inhibition in the activities of CYP450 and CYT b5 that leads to lower metabolism and clearance, which in turn increased the length of parasite exposure to the drug. Accordingly, KTZ enhanced the therapeutic antischistosomal potential of QN and HF over each drug alone. This was evident by increased female, total worm, and egg reductions and healing of hepatic granulomatous lesions, nevertheless, these effects were more obvious in KTZ+QN than in KTZ+HF. QN and HF have adverse effects on the antioxidant status of the animals; therefore, the need for antioxidants as adjuvant to antimalarials is recommended.

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## REFERENCES

- King CH, Dangerfield-Cha M. The unacknowledged impact of chronic schistosomiasis. *Chronic Illn* 2008; 4: 65-79.
- Davis A. Schistosomiasis. In Cook GC, Zumla AI eds. *Manson's Tropical Diseases*. 22nd edition. London, UK. Saunders. 2009, p 1413-1456.
- Utzinger J, Raso G, Brooker S, De Savigny D, Tanner M, Ombjerg N, Singer BH, N'goran EK. Schistosomiasis and neglected tropical diseases: towards integrated and sustainable control and a word of caution. *Parasitology* 2009; 136: 1859-1874.
- King CH, Dickman K, Tisch DJ. Reassessment of the cost of chronic helminth infection: a meta-analysis of disability-related outcomes in endemic schistosomiasis. *Lancet* 2005; 365: 1561-1569.
- Melman SD, Steinauer ML, Cunningham C, Kubatko LS, Mwangi IN, Wynn NB, Mutuku MW, Karanja DM, Colley DG, Black CL, Secor WE, Mkoji GM, Loker ES. Reduced susceptibility to praziquantel among naturally occurring Kenyan isolates of *Schistosoma mansoni*. *PLoS Negl Trop Dis* 2009; 3: e504.
- Xiao SH, Keiser J, Chollet J, Utzinger J, Dong Y, Endriss Y, Vennerstrom JL, Tanner M. In vitro and in vivo activities of synthetic trioxolanes on major human schistosome species. *Antimicrob Agents Chemother* 2007; 51: 1440-1445.
- Abdulla MH, Lim KC, Sajid M, McKerrow JH, Caffrey CR. Schistosomiasis *mansoni*: novel chemotherapy using a cysteine protease inhibitor. *PLoS Med* 2007; 4: e14.
- Luiz Oliveira Penido M, Resende DM, Vianello MA, Humberto da Silveira Bordin F, Jacinto AA, Dias WD, Montesano MA, Nelson DL, Marcos Zech Coelho P, Vasconcelos EG. A new series of schistosomicide drugs, the alkylaminoalkaneethiosulfuric acids, partially inhibit the activity of *Schistosoma mansoni* ATP diphosphohydrolase. *Eur J Pharmacol* 2007; 570: 10-17.
- Ronchetti F, Ramana AV, Chao-Ming X, Pica-Mattocchia L, Cioli D, Todd MH. Praziquantel derivatives I: Modification of the aromatic ring. *Bioorg Med Chem Lett* 2007; 17: 4154-4157.
- Sayed AA, Simeonov A, Thomas CJ, Inglese J, Austin CP, Williams DL. Identification of oxadiazoles as new drug leads for the control of schistosomiasis. *Nat Med* 2008; 4: 407-412.
- Gupta S, Thapar MM, Wernsdorfer WH, Björkman A. In vitro interactions of artemisinin with atovaquone, quinine and mefloquine against *Plasmodium falciparum*. *Antimicrob Agents Chemother* 2002; 46: 1510-1515.
- Merkus FW, Smit EH. The use of quinine in the treatment of malaria from the past to future. *Int J Pharmacy* 1988; 2: 47-50.
- Meshnick SR. Why does quinine still work after 350 years of use? *Parasitol Today* 1997; 13: 89-90.
- Mirghani RA, Hellgren U, Westerberg PA, Ericsson O, Bertilsson L, Gustafsson LL. The roles of cytochrome P450 3A4 and 1A2 in the 3-hydroxylation of quinine in vivo. *Clin Pharmacol Ther* 1999; 66: 454-460.
- Diczfalusy U, Miura J, Roh HK, Mirghani RA, Sayi J, Larsson H, Bodin KG, Allqvist A, Jande M, Kim JW, Akillu E, Gustafsson LL, Bertilsson L. 4Beta-hydroxycholesterol is a new endogenous CYP3A marker: relationship to CYP3A5 genotype, quinine 3-hydroxylation and sex in Koreans, Swedes and Tanzanians. *Pharmacogenet Genomics* 2008; 18: 201-208.
- Boudreau EF, Pang LW, Dixon KE, Webster HK, Pavanand K, Tosingha L, Somutsakorn P, Canfield CJ. Malaria: treatment effi-

- cacy of halofantrine (WR 171,669) in initial field trials in Thailand. *Bull World Health Organ* 1988; 66: 227-235.
17. Karbwang J, Na Bangchang K. Clinical pharmacokinetics of halofantrine. *Clin Pharmacokinet* 1994; 27: 104-119.
  18. White NJ. Qinghaosu (artemisinin): the price of success. *Science* 2008; 320: 330-334.
  19. Xiao SH, Booth M, Tanner M. The prophylactic effects of artemether against *Schistosoma japonicum* infections. *Parasitol Today* 2000; 16: 122-126.
  20. Seif el-Din SH. Prophylactic effect of artesunate with/without antioxidants on juvenile and adult Egyptian strain of *Schistosoma mansoni* in mice. *Az J Pharm Sci* 2007; 35: 80-93.
  21. Utzinger J, Xiao SH, Tanner M, Keiser J. Artemisinins for schistosomiasis and beyond. *Curr Opin Investig Drugs* 2007; 8: 105-116.
  22. Keiser J, Chollet J, Xiao S, Mei JY, Jiao PY, Utzinger J, Tanner M. Mefloquine—an aminoalcohol with promising Antischistosomal Properties in Mice. *PLoS Negl Trop Dis* 2009; 3: e350.
  23. Manneck T, Haggmüller Y, Keiser J. Morphological effects and tegumental alterations induced by mefloquine on schistosome and adult flukes of *Schistosoma mansoni*. *Parasitology* 2010; 137: 85-98.
  24. El-Lakkany NM, Seif el-Din SH, Sabra AN, Hammam OA. Pharmacodynamics of mefloquine and praziquantel combination therapy in mice harbouring juvenile and adult *Schistosoma mansoni*. *Mem Inst Oswaldo Cruz* 2011; 106: 814-822.
  25. Floren LC, Bekersky I, Benet LZ, Mekki Q, Dressler D, Lee JW, Roberts JP, Hebert ME. Tacrolimus oral bioavailability doubles with coadministration of ketoconazole. *Clin Pharmacol Ther* 1997; 62: 41-49.
  26. Zhao XJ, Ishizaki T. Metabolic interactions of selected antimalarial and non-antimalarial drugs with the major pathway (3-hydroxylation) of quinine in human liver microsomes. *Br J Clin Pharmacol* 1997; 44: 505-511.
  27. Baune B, Furlan V, Taburet AM, Farinotti R. Effect of selected antimalarial drugs and inhibitors of cytochrome P-450 3A4 on halofantrine metabolism by human liver microsomes. *Drug Metab Dispos* 1999; 27: 565-568.
  28. Liang YS, Bruce JI, Boyd DA. Laboratory cultivation of schistosome vector snails and maintenance of schistosome life cycles. *Proc First Sino-Am Symp* 1987; 1: 34-48.
  29. Duvall RH, DeWitt WB. An improved perfusion technique for recovering adult schistosomes from laboratory animals. *Am J Trop Med Hyg* 1967; 16: 483-486.
  30. Kamel IA, Cheever AW, Elwi AM, Mosimann JE, Danner R. *Schistosoma mansoni* and *S. haematobium* infections in Egypt. I. Evaluation of technique for recovery of worms and eggs at necropsy. *Am J Trop Med Hyg* 1977; 26: 696-701.
  31. Pellegrino J, Oliveira CA, Faria J, Cunha AS. New approach to the screening of drugs in experimental schistosomiasis mansoni in mice. *Am J Trop Med Hyg* 1962; 11: 201-215.
  32. von Lichtenberg. Host response to eggs of *Schistosoma mansoni*. I. Granuloma formation in the sensitized laboratory mouse. *Am J Pathol* 1962; 41: 711-731.
  33. Netter KJ. A method for the direct measurement of O-demethylation in liver microsomes and its use in studying microsome inhibition by SKF-525-A. *Naunyn Schmiedebergs Arch Exp Pathol Pharmacol* 1960; 238: 292-300.
  34. Ellman GL. Tissue sulfhydryl groups. *Arch Biochem Biophys* 1959; 82: 70-77.
  35. Winterbourn CC, Hawkins RE, Brian M, Carrell RW. The estimation of red cell superoxide dismutase activity. *J Lab Clin Med* 1975; 85: 337-341.
  36. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem* 1979; 95: 351-358.
  37. Omura T, Sato R. The carbon monoxide-binding pigment of liver microsomes. I. Evidence for its haemoprotein nature. *J Biol Chem* 1964; 239: 2370-2378.
  38. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem* 1951; 193: 265-275.
  39. El-Lakkany NM, Seif el-Din SH, Badawy AA, Ebeid FA. Effect of artemether alone and in combination with grapefruit juice on hepatic drug-metabolizing enzymes and biochemical aspects in experimental *Schistosoma mansoni*. *Int J Parasitol* 2004; 34: 1405-1412.
  40. Botros SS, Seif el-Din SH, El-Lakkany NM, Sabra AN, Ebeid FA. Drug-metabolizing enzymes and praziquantel bioavailability in mice harboring *Schistosoma mansoni* isolates of different drug susceptibilities. *J Parasitol* 2006; 92: 1344-1349.
  41. Botros S, El-Lakkany N, Seif el-Din SH, Sabra AN, Ibrahim M. Comparative efficacy and bioavailability of different praziquantel brands. *Exp Parasitol* 2011; 127: 515-521.
  42. Badawi AF, Mostafa MH. Possible mechanisms of alteration in the capacities of carcinogen metabolizing enzymes during schistosomiasis and their role in bladder cancer induction. *J Int Med Res* 1993; 21: 281-305.
  43. Kuroha M, Azumano A, Kuze Y, Shimoda M, Kokue E. Effect of multiple dosing of ketoconazole on pharmacokinetics of midazolam, a cytochrome P-450 3A substrate in beagle dogs. *Drug Metab Dispos* 2002; 30: 63-68.
  44. Kageyama M, Namiki H, Fukushima H, Ito Y, Shibata N, Takada K. In vivo effects of cyclosporin A and ketoconazole on the pharmacokinetics of representative substrates for P-glycoprotein and cytochrome P450 (CYP) 3A in rats. *Biol Pharm Bull* 2005; 28: 316-322.
  45. Xiao SH, Mei JY, Jiao PY. Further study on mefloquine concerning several aspects in experimental treatment of mice and hamsters infected with *Schistosoma japonicum*. *Parasitol Res* 2009; 106: 131-138.
  46. Corrêa Soares JB, Menezes D, Vannier-Santos MA, Ferreira-Pereira A, Almeida GT, Venancio TM, Verjovski-Almeida S, Zishiri VK, Kuter D, Hunter R, Egan TJ, Oliveira MF. Interference with hemozoin formation represents an important mechanism of schistosomicidal action of antimalarial quinoline methanols.

- PLoS Negl Trop Dis 2009; 3: e477.
47. Oliveira MF, d'Avila JC, Tempone AJ, Soares JB, Rumjanek FD, Ferreira-Pereira A, Ferreira ST, Oliveira PL. Inhibition of heme aggregation by chloroquine reduces *Schistosoma mansoni* infection. J Infect Dis 2004; 190: 843-852.
  48. Mattoccia LP, Lelli A, Cioli D. Sex and drugs in *Schistosoma mansoni*. J Parasitol 1982; 68: 347-349.
  49. Ginsburg H, Demel RA. The effect of ferriprotoporphyrin IX and chloroquine on phospholipid monolayers and the possible implications to antimalarial activity. Biochim Biophys Acta 1983; 732: 316-319.
  50. Oliveira MF, d'Avila JC, Torres CR, Oliveira PL, Tempone AJ, Rumjanek FD, Braga CM, Silva JR, Dansa-Petretski M, Oliveira MA, de Souza W, Ferreira ST. Haemozoin in *Schistosoma mansoni*. Mol Biochem Parasitol 2000; 111: 217-221.
  51. Xiong LJ, Zhu JF, Luo DD, Zen LL, Cai SQ. Effects of pentoxifylline on the hepatic content of TGF-beta1 and collagen in schistosomiasis *japonica* mice with liver fibrosis. World J Gastroenterol 2003; 9: 152-154.
  52. Rehermann B, Nascimbeni M. Immunology of hepatitis B virus and hepatitis C virus infection. Nat Rev Immunol 2005; 5: 215-229.
  53. Zwingenberger K, Richter J, Vergetti JG, Feldmeier H. Praziquantel in the treatment of hepatosplenic schistosomiasis: biochemical disease markers indicate deceleration of fibrogenesis and diminution of portal flow obstruction. Trans R Soc Trop Med Hyg 1990; 84: 252-256.
  54. Mahmoud MR, Zoheiry MM, Nosseir MM. Effect of combined chemotherapy and anti-inflammatory drugs on murine schistosomiasis. Arzneimittelforschung 2002; 52: 294-301.
  55. Riou M, Koch C, Delaleu B, Berthon P, Kerboeuf D. Immunolocalisation of an ABC transporter, P-glycoprotein, in the eggshells and cuticles of free-living and parasitic stages of *Haemonchus contortus*. Parasitol Res 2005; 96: 142-148.
  56. Abdallahi OM, Hanna S, De Reggi M, Gharib B. Visualization of oxygen radical production in mouse liver in response to infection with *Schistosoma mansoni*. Liver 1999; 19: 495-500.
  57. Gharib B, Abdallahi OM, Desein H, De Reggi M. Development of eosinophil peroxidase activity and concomitant alteration of the antioxidant defenses in the liver of mice infected with *Schistosoma mansoni*. J Hepatol 1999; 30: 594-602.
  58. Seif el-Din SH, Ebeid FA, Badawy AA, Ezzat AR. Protective effects of  $\beta$ -carotene, N-acetyl-L-cysteine with and without praziquantel treatment in *Schistosoma mansoni*-infected mice. Egypt J Schistosomiasis Infect Endem Dis 2006; 28: 67-90.
  59. Czeczot H, Scibior D, Skrzycki M, Podsiad M. Glutathione and GSH-dependent enzymes in patients with liver cirrhosis and hepatocellular carcinoma. Acta Biochimica Pol 2006; 53: 237-242.
  60. El-Lakkany N, Seif el-Din S, Ebeid F. The use of pentoxifylline as adjuvant therapy with praziquantel downregulates profibrogenic cytokines, collagen deposition and oxidative stress in experimental schistosomiasis *mansoni*. Exp Parasitol 2011; 129: 152-157.
  61. Fernandez-Checa JC, Kaplowitz N. Hepatic mitochondrial glutathione: transport and role in disease and toxicity. Toxicol Appl Pharmacol 2005; 204: 263-273.
  62. Mantawy MM, Ali HF, Rizk MZ. Therapeutic effects of *Allium sativum* and *Allium cepa* in *Schistosoma mansoni* experimental infection. Rev Inst Med Trop Sao Paulo 2011; 53: 155-163.
  63. Hirose Y, Kirinoki M, Matsuda H. Efficacy of administration of praziquantel on 2 days 2 weeks apart against *Schistosoma japonicum* eggs in mice. Parasitol Int 2003; 52: 141-146.
  64. Halliwell B, Gutteridge J. Free Radicals in Biology and Medicine. Fourth edition. Oxford. University Press. 2007.
  65. Farombi EO, Olowu BI, Emerole GO. Effect of three structurally related antimalarial drugs on liver microsomal components and lipid peroxidation in rats. Comp Biochem Physiol C Toxicol Pharmacol 2000; 126: 217-224.
  66. Farombi EO, Adoro S, Uahunmwangho S. Antimalarial drugs exacerbate rat liver microsomal lipid peroxidation in the presence of oxidants. Biosci Rep 2001; 21: 353-359.
  67. Adaramoye OA, Osaimoje DO, Akinsanya AM, Nneji CM, Fafunso MA, Ademowo OG. Changes in antioxidant status and biochemical indices after acute administration of artemether, artemether-lumefantrine and halofantrine in rats. Basic Clin Pharmacol Toxicol 2008; 102: 412-418.

