

# Genome Sequence of *Xanthomonas arboricola* pv. *Corylina*, Isolated from Turkish Filbert in Colorado

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Previously, we reported the isolation of a bacterium producing leaf spots in Turkish filbert. Here, we present the draft genome assembly of the bacterium identified as *Xanthomonas arboricola* pv. *corylina*. To our knowledge, this is the first published genome of this pathovar of *X. arboricola*.

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*Xanthomonas arboricola* pv. *corylina* is a pathogen of *Corylus avellana* L. and it also infects other species (1). In a previous report (2), we identified *X. arboricola* pv. *corylina* from leaf spots of *Corylus colurna* L. Here, we sequenced the genome of *X. arboricola* pv. *corylina* isolate NCCB100457 with 100 cycles of paired-end reads using an Illumina HiSeq sequencer at the USC Epigenomic Center. More than 13.3 million 100-base-long reads were produced for each end. We performed two genome assemblies, one using the A5 pipeline (3) and another using SPAdes v.2.3 (4). The A5 assembly has a total length of 5,227,695 bp consisting of 48 scaffolds (281 contigs), 43 of which were >500 bp, with the longest being 840,880 bp; the N<sub>50</sub> is 263,170 bp and the G+C content is 65.46%. The SPAdes assembly has a total length of 5,398,516 bp placed in 810 nodes, 182 of which were >500 bp, with the longest being 263,848 bp; the N<sub>50</sub> is 77,809 bp and the G+C content is 65.35%. We annotated the assembled genomes using the RAST server (5) and detected 4,452 and 4,500 coding sequences representing 445 and 449 “subsystems” for the A5 and SPAdes assemblies, respectively. Those numbers are similar to those in *Xanthomonas axonopodis* pv. *citri* (5,274,174 bp and 4,489 coding sequences represented in 461 subsystems), which is the most closely related organism present in the RAST database, based on nucleotide similarity. To further check the robustness of the NCCB100457 genome assemblies, we performed a BLASTn (6) search against 12 genes of *X. arboricola* pv. *corylina* that are commonly used as markers and that are present in the GenBank database. The genes *acnB*, *dnaK*, *fstZ*, *fyuA*, *gapA*, *qumA*, *rpoB*, and *rpoD* were detected and have 100% identity with other isolates from the same pathovar, while *fstX*, *groEL*, and 16S rRNA genes have 99% identity and *gyrB* has 98% identity.

We confirmed the presence of all 11 *hrp2* type 3 secretion system (T3SS) genes that are ubiquitous to all pathovars of *X. arboricola*; we also detected 20 out of 21 corresponding effector protein (T3E) genes that are present in all other *X. arboricola* pv. *corylina* isolates, including ATCC 19313, collected from *Corylus maxima*

in the United States (7); the only exception is *avrBs3*, which is absent in NCCB100457. A PCR with *avrBs3*-specific primers (7) using the genomic DNA of NCCB100457 did not result in amplification, whereas amplicons were detected in the positive controls *Xanthomonas oryzae* pv. *oryzae* PXO99A and *X. oryzae* pv. *oryzicola* BLS256. The gene *xopH*, which is present in most *X. arboricola* pv. *corylina* isolates but not in ATCC 19313, is also absent in our isolate. We did not detect 31 T3E genes in the NCCB100457 genome that are also absent in other *X. arboricola* pv. *corylina* genomes (7). However, we detected a putative *avr* similar to *hpoG1* from *Xanthomonas campestris* pv. *vasculorum* (NCBI accession no. ZP\_06487712.1) that was not reported before in *X. arboricola* pv. *corylina*. Differences in the arsenal of secretion systems and effectors can account for the pathogenicity and host specificity in pathogenic bacteria (8), including in isolate NCCB100457.

**Nucleotide sequence accession numbers.** This Whole-Genome Shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. [APMC00000000](http://www.ncbi.nlm.nih.gov/nuccore/APMC00000000). The version described in this paper is the first version, accession no. [APMC01000000](http://www.ncbi.nlm.nih.gov/nuccore/APMC01000000).

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## REFERENCES

1. EPPO. 2004. Diagnostic protocols for regulated pests: PM 7/22, *Xanthomonas arboricola* pv. *corylina*. *Bull. OEPP/EPPO Bull.* 34:155–157.
2. Ibarra J, Snelling J, Alexander K, Tisserat N. May 2012. Leaf spotting of Turkish filbert in Colorado caused by *Xanthomonas arboricola* pv. *corylina* and *Pseudomonas syringae* pv. *syringae*. *Plant Health Prog.* <http://www.plantmanagementnetwork.org/php/elements/sum.aspx?id=10321&photo=5671>.
3. Tritt A, Eisen JA, Facciotti MT, Darling AE. 2012. An integrated pipeline for *de novo* assembly of microbial genomes. *PLoS One* 7:e42304.

4. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J. Comput. Biol.* 19:455–477.
5. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75.
6. Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res.* 25:3389–3402.
7. Hajri A, Pothier JF, Fischer-Le Saux M, Bonneau S, Poussier S, Boureau T, Duffy B, Manceau C. 2012. Type three effector gene distribution and sequence analysis provide new insights into the pathogenicity of plant-pathogenic *Xanthomonas arboricola*. *Appl. Environ. Microbiol.* 78: 371–384.
8. Dean P. 2011. Functional domains and motifs of bacterial type III effector proteins and their roles in infection. *FEMS Microbiol. Rev.* 35:1100–1125.