

Published in final edited form as:

Nat Rev Cancer. 2010 December ; 10(12): 819–823. doi:10.1038/nrc2962.

Sirt1: recent lessons from mouse models

Daniel Herranz¹ and Manuel Serrano^{1,*}

¹Tumor Suppression Group Spanish National Cancer Research Center (CNIO) Madrid, Spain

PREFACE

The family of protein deacetylases represented by yeast Sir2 has been the focus of intense investigation because of its longevity activity in yeast, worms and flies. Research in mammals has mainly focused on Sirt1, the closest homologue of Sir2. Emerging evidence from mouse models is yielding a sharper picture where Sirt1 is a potent protector from aging associated-pathologies, such as diabetes, liver steatosis, cardiovascular disease, neurodegeneration, osteoporosis and, importantly, various types of cancer.

INTRODUCTION

Over the past ten years, the aging research community has developed a growing interest in the mammalian sirtuin family (formed by paralogues Sirt1 to Sirt7), and more specifically in Sirt1, the closest homologue of yeast Sir2¹. This interest stems from pioneer reports linking Sir2 to lifespan regulation in lower organisms. In particular, genetic overexpression of Sir2 was shown to increase lifespan in yeast², worms³ and flies⁴. This remarkable effect was further corroborated in studies using purported Sir2-chemical activators, most notably resveratrol, that extended lifespan in yeast⁵, worms and flies⁶ in a Sir2-dependent manner. Moreover, calorie restriction (CR), which is the most effective dietary intervention to extend lifespan in different organisms¹, was shown to require Sir2, both in yeast⁷ and flies⁴, in order to produce lifespan extension. All these studies led to the appealing hypothesis that Sir2 is an evolutionary conserved longevity gene that mediates CR effects. If this was the case, then Sirt1 activation in mammals might improve aging without the necessity to reduce food intake. As discussed below, recent research in genetically-modified mice has demonstrated clear positive effects of Sirt1 on a variety of aspects related to mammalian health, including aging-associated diseases. Yet, current evidence does not support that Sirt1 can increase mammalian longevity.

Tidal changes

The story of Sirt1 has had its tidal changes, which should not be surprising when dealing with proteins that integrate and coordinate multiple stimuli and responses. Often, expectations run too high, and this is inevitably followed by counter expectations that go too low. In the case of Sirt1, the high expectations on its role in longevity were dimmed by studies that challenged the notion that *S. cerevisiae* Sir2 mediates CR-associated lifespan extension^{8,9} and, subsequently, by a report that failed to reproduce lifespan extension by resveratrol in *D. melanogaster* and *C. elegans*¹⁰. Regarding mammals, resveratrol was shown to have beneficial effects on health, including improved glucose tolerance and protection from fatty liver, but without extending lifespan¹¹⁻¹³. However, accumulating evidence indicates that resveratrol is not a direct activator of Sirt1¹⁴⁻¹⁶, but rather acts

*Correspondence: Manuel Serrano, Spanish National Cancer Research Center (CNIO), 3 Melchor Fernandez Almagro street, Madrid E-28029, Spain, Tel.: +34.91.732.8000, Fax: +34.91.732.8033, mserrano@cnio.es.

indirectly through AMPK^{17,18} (see Box1). Together, these conflictive results have casted a shadow on the relevance of Sirt1 in mammalian aging¹⁹.

Another aspect that has generated debate is the relationship between Sirt1 and cancer, which, in turn, has its roots at the interplay between Sirt1 and p53. Early studies with *in vitro* cultured cells showed that Sirt1 is able to interact with and deacetylate the tumour suppressor protein p53, therefore inhibiting its transactivation potential^{20,21}. In support of this, studies in Sirt1 knockout mice showed p53 hyperactivation, leading to increased thymocyte apoptosis²², although this could not be confirmed by other investigators²³. Moreover, Sirt1 is upregulated in several types of human tumours (reviewed in²⁴), which has further supported the idea that Sirt1 could be oncogenic. However, contrary to this, all the currently available data indicate that Sirt1 is a tumour suppressor *in vivo*²⁵⁻²⁸ and the possible consequences of Sirt1 on p53-mediated tumour suppression remain to be elucidated.

The above controversies highlight the importance of genetically-modified mouse models to address Sirt1 effects in the context of the organism. In this Progress, we discuss recent studies using genetic mouse models of Sirt1, which have shed new light into the field. For more comprehensive reviews on Sirt1, the reader is referred to recent reviews^{29,30}.

Sirt1 and metabolism

Sirt1 effects in metabolism are the best-characterized ones. A considerable amount of recent literature based on a variety of genetically-modified mouse models has solidly demonstrated a protective role for Sirt1 against pathologies associated to high dietary fat intake (HFD, high fat diet), such as glucose intolerance and liver steatosis (also known as fatty liver) (Table 1).

The first mouse models that linked Sirt1 to improved glucose tolerance were studies in which Sirt1 overexpression was restricted, in one case, to pancreatic β -cells³¹ and, in the other one, to brain and adipose tissue³². The effect of Sirt1 on metabolism has been further explored in mice with whole-body transgenic expression of moderately increased levels of Sirt1 (2-3 x-fold) under its own endogenous transcriptional regulation^{33,34}. Importantly, these Sirt1 transgenic mice were protected from diabetes associated to diet-induced obesity^{33,34} or to genetically-induced obesity³⁴. The impact of moderate systemic Sirt1 overexpression on protection from high dietary fat goes beyond glucose tolerance. In fact, whole body Sirt1 transgenic mice are remarkably protected from developing liver steatosis³³. Consistent with this, mice deficient in Dbc1, a negative regulator of Sirt1, are also protected from fatty liver³⁵ and, conversely, Sirt1 heterozygous mice are prone to develop this disease³⁶. Mice with tissue-specific deletion of Sirt1 have further refined the picture and have indicated that Sirt1 exerts its protective metabolic effects through its actions on multiple tissues. Namely, myeloid lineage-specific deletion of Sirt1 resulted in insulin resistance³⁷; brain-specific Sirt1 deletion increased aging-associated glucose intolerance³⁸; Sirt1 deletion in the pro-opiomelanocortin (POMC) neurons, that control food intake and energy expenditure, resulted in leptin resistance in these neurons and increased susceptibility to diet-induced obesity³⁹; and, finally, liver-specific deletion of Sirt1 protected mice from steatosis⁴⁰. However, regarding the latter, it should be mentioned that other investigators have reported the opposite phenotype using the same genetic model of Sirt1 deficiency in the liver⁴¹; the basis for these conflictive results are unclear. A separate note must be made of the few metabolic studies using whole-body Sirt1-null mice because of their paradoxical results. In particular, Sirt1-null presented higher glucose tolerance⁴² and protection from fatty liver induced by LXR agonists⁴³, which is in sharp contrast with all above-described evidence indicating that Sirt1 protects from glucose intolerance and fatty liver. However, interpretation of the results obtained with whole-body Sirt1-null mice is

highly complex given that the majority of these mice die in the perinatal period and the survivors have developmental defects^{22,44} that may obscure the role of Sirt1 in a normal physiological context. In summary, as listed in Table 1, most studies using a variety of mouse models have concluded that Sirt1 is a robust metabolic protector acting at several systems, including not only the liver, but also the brain and the immune system.

A recurrent mechanistic finding in the above-described mouse models linking Sirt1 to protection from diabetes and fatty liver is a reduced inflammatory response associated to lower levels of NF κ B activity^{33,35-37,40} (Figure 1). This fully supports in mice the earlier finding in cultured cells that Sirt1 binds, deacetylates and inhibits NF κ B⁴⁵. Another recurrent mechanism is the deacetylation and activation of key transcription factors involved in lipid metabolism, such as Foxo1³⁴ and the nuclear receptors PPAR α ⁴⁰, LXR⁴³, and FXR⁴⁶, as well as, their general co-activator PGC1 α ^{33,40,47} (Figure 1). Numerous additional mechanisms implicating Sirt1 in metabolism have been also reported, but their detailed discussion goes beyond the scope of this Progress article. Briefly, these additional mechanisms include increased insulin production by repression of Ucp2^{31,42}; deacetylation and inhibition of SREBP^{33,40,48,49}; repression of the *Pparg* promoter⁵⁰; activation of the *Sirt6* promoter⁵¹ which, in turn, may contribute to protect from metabolic damage as demonstrated in Sirt6-overexpressing mice⁵²; and deacetylation and activation of Lkb1^{53,54}.

Diabetes and fatty liver are diseases that often occur concurrently, and in association with other pathologies, notably including cardiovascular disease. This group of associated pathologies is referred to as the metabolic syndrome⁵⁵. In this regard, it is worth mentioning that there is also evidence from Sirt1-mouse models indicating that Sirt1 can improve cardiovascular function⁵⁶⁻⁵⁹. The metabolic syndrome is characteristic of aged individuals chronically exposed to high caloric intake and low physical activity, a lifestyle of increasing prevalence that explains the high incidence of metabolic syndrome-associated diseases worldwide⁵⁵. Based on the above-mentioned data in genetically-modified mice, Sirt1 activation is an attractive target for pharmaceutical interventions aimed to delay or ameliorate the pathologies associated to the metabolic syndrome.

Sirt1 and cancer

The analysis of cancer in mice with genetically-modified Sirt1 levels has consistently supported the concept that Sirt1 possesses strong tumour suppressive activity (Table 1). In particular, increased or decreased Sirt1 expression results, respectively, in delayed or accelerated sarcoma and lymphoma development in a p53-heterozygous background^{27,28}. Interestingly, both studies obtained supporting *in vitro* evidence indicating that this tumour suppressive activity reflected the ability of Sirt1 to preserve genomic integrity in the face of p53 deficiency^{27,28}. Upon DNA damage, chromatin-bound Sirt1 relocates from repeated elements to the actual DNA damaged sites, presumably repressing transcription²⁷ (Figure 1).

Following on the reported functions of Sirt1 in protection from DNA damage and in protection from fatty liver, whole-body Sirt1 transgenic mice were subjected to a novel carcinogenic treatment that models metabolic syndrome-associated liver cancer. Feeding mice with a high fat diet (HFD) dramatically increases the incidence of liver carcinomas in mice previously treated with a hepatic carcinogen (diethylnitrosamine, DEN) which on its own has poor carcinogenic activity^{26,60}. Importantly, Sirt1 transgenic mice were remarkably protected from metabolic syndrome-driven liver carcinogenesis²⁶. This protection was due to the combined effects of Sirt1, first, in reducing DNA damage by the hepatic mutagen and, then, in preventing inflammation and fatty liver by HFD²⁶ (Figure 1). Dietary obesity leads to liver inflammation by enhancing the expression of IL-6 and TNF α , well known targets of NF κ B, and this inflammation promotes tumorigenesis⁶⁰. Sirt1 transgenic mice show decreased NF κ B activation and, concomitantly, decreased inflammation in the liver

resulting in the observed protection from liver tumorigenesis^{26,33}. Of note, human liver cancers present decreased levels of Sirt1 compared to normal liver²⁸. It remains to be elucidated whether Sirt1 would protect from inflammation-associated tumorigenesis in general or just from carcinogenesis associated to dietary-induced inflammation.

On another line of research, Sirt1 overexpression in enterocytes protected from intestinal tumours in the *Apc*^{+/*min*} model²⁵. This observation led to discovery that Sirt1 deacetylates and inhibits β -catenin in the intestine of these mice²⁵ (Figure 1). In further support of this, subsequent studies showed that Sirt1 suppresses the growth of colon tumour xenografts assays using human colon cancer lines⁶¹. Of note, recent studies using Sirt1-null mice revealed no difference in tumour development when combined with the *Apc*^{+/*min*} mutation⁶². It is formally possible that Sirt1 overexpression could protect from intestinal tumorigenesis, while in the absence of Sirt1, compensatory mechanisms could supply the missing activity of Sirt1. As briefly alluded above, caution must be taken when interpreting the phenotypes observed in whole-body Sirt1-null strains due to their developmental defects^{22,44}.

Finally, regarding aging-associated spontaneous cancer, whole-body Sirt1 transgenic mice displayed a global decrease in cancer incidence²⁶. This protection, however, was restricted to carcinomas and sarcomas, while lymphoma development was not affected by Sirt1 dosage. This differential effect is currently not well understood and it contrasts with the previously mentioned Sirt1-dependent protection from p53-deficient lymphomas. The aging-associated lymphomas developed in a p53-functional context are conceivably less aggressive and less genetically unstable than the lymphomas developed in the absence of p53. Hence, the ability of Sirt1 to preserve genomic stability could be less relevant in the context of aging-associated lymphomas. In addition to aging-associated lymphomas, Sirt1 was also found to be neutral regarding chemically-induced fibrosarcomas²⁶. Apart from these two cancer models in which Sirt1 levels do not seem to affect tumorigenesis, the general finding in the above-summarized studies is that Sirt1 protects from cancer through a number of mechanisms, including protection from DNA damage, protection from diet-induced inflammation, and inhibition of the oncogenic activity of β -catenin (Figure 1). At present, there are no reports of mouse models of cancer in which Sirt1 plays an oncogenic role (Table 1).

Sirt1 and aging

The effect of Sirt1 on mammalian aging is a long-sought result in the field. Importantly, mice moderately overexpressing Sirt1 (3x-fold) under its own transcriptional regulatory elements do not live longer under standard diet conditions, but they show a healthier aging²⁶. In particular, aged Sirt1 transgenic mice showed improved glucose homeostasis, better preservation of bone mineralization, reduced incidence of sarcomas and carcinomas, and reduced levels in molecular markers of aging, such as p16^{Ink4a} or DNA damage²⁶. Moreover, recent studies in mice have also demonstrated a protective role for Sirt1 against Alzheimer's disease and amyotrophic lateral sclerosis^{63,64}. The emerging picture is one in which Sirt1, through its well-established protective activities against diet-induced metabolic damage, genomic instability, and cancer, improves some aspects of aging and delays or ameliorates a number of aging-associated diseases such as metabolic syndrome, Alzheimer or some types of cancer. In summary, although current evidence does not allow to categorize Sirt1 as a "longevity" gene, it clearly is a beneficial gene for aging and aging-associated diseases.

A related issue is whether Sirt1 mediates the effects of calorie restriction (CR) on aging. Only one report has addressed this directly using Sirt1-null mice under CR⁶⁵. As mentioned above, most Sirt1-null mice die in the perinatal period and those that survive have

developmental defects^{22,44}, including a shortened lifespan (median lifespan of approximately 1 year)⁶⁵. Interestingly, CR started between 5-7 months of age did not have an impact on the longevity of Sirt1-null mice⁶⁵. Although suggestive, the abnormally short lifespan of Sirt1-null mice makes it difficult to interpret their lack of response to CR. Additionally, some of the beneficial effects of Sirt1 on metabolism are also characteristically produced by CR, such as improved insulin sensitivity. Finally, a number of studies have reported a variety of molecular and phenotypic effects of CR that are mediated by Sirt1 in mice^{38,65-69}. Overall, there is growing suggestive evidence indicating that Sirt1 participates in CR.

Conclusions and future perspectives

Recent work with genetically-modified mice has firmly established Sirt1 as a protector of metabolic syndrome and as a tumour suppressor in a wide range of cancers. Moreover, emerging evidence implicate Sirt1 in protection from cardiovascular disease⁵⁶⁻⁵⁹ and neurodegeneration^{63,64}. The fact that Sirt1 impinges on such a variety of aging-associated diseases suggests that it could be a longevity gene, although direct evidence for this is still lacking in mammals. The only available study of longevity in a mouse model with systemic Sirt1 overexpression (3x-fold) reported improved health during aging, but normal longevity²⁶. In this regard, it would be interesting to test whether higher levels of Sirt1 overexpression or contemporaneous overexpression of several sirtuins could extend longevity.

Regarding metabolism, it would be of critical importance to know if, once diabetes has developed, Sirt1 activation could be used as a treatment. To this end, and until potent Sirt1 activators are developed, Sirt1 inducible knock-in mice could be used to explore this possibility. If these activators showed a benefit in diabetic patients, and given the cancer protection activity of Sirt1, it is tempting to speculate that they could diminish cancer incidence concomitantly. Precedence for this can be found in the anti-diabetic drug metformin that decreases cancer incidence both in mice^{63,64} and humans^{63,64}.

With respect to cancer, genetic studies published so far support a protective role for Sirt1 in many types of cancer. However, this is not general and there are examples of mouse cancer types where Sirt1 does not play a role (Table 1). Besides, numerous studies based on *in vitro* cultured cancer cell lines have suggested that Sirt1 has oncogenic activities²⁴. Although this has not been confirmed yet in a more physiological setting, it remains possible that future studies could reveal oncogenic activities of Sirt1 *in vivo*. Finally, a key pending question is whether Sirt1 activators have therapeutic activity on cancer.

REFERENCES

1. Michan S, Sinclair D. Sirtuins in mammals: insights into their biological function. *Biochem J.* 2007; 404:1–13. [PubMed: 17447894]
2. Kaerberlein M, McVey M, Guarente L. The SIR2/3/4 complex and SIR2 alone promote longevity in *Saccharomyces cerevisiae* by two different mechanisms. *Genes Dev.* 1999; 13:2570–2580. [PubMed: 10521401]
3. Tissenbaum HA, Guarente L. Increased dosage of a sir-2 gene extends lifespan in *Caenorhabditis elegans*. *Nature.* 2001; 410:227–230. [PubMed: 11242085]
4. Rogina B, Helfand SL. Sir2 mediates longevity in the fly through a pathway related to calorie restriction. *Proc Natl Acad Sci U S A.* 2004; 101:15998–16003. [PubMed: 15520384]
5. Howitz KT, et al. Small molecule activators of sirtuins extend *Saccharomyces cerevisiae* lifespan. *Nature.* 2003; 425:191–196. [PubMed: 12939617]
6. Wood JG, et al. Sirtuin activators mimic caloric restriction and delay ageing in metazoans. *Nature.* 2004; 430:686–689. [PubMed: 15254550]

7. Lin SJ, Defossez PA, Guarente L. Requirement of NAD and SIR2 for life-span extension by calorie restriction in *Saccharomyces cerevisiae*. *Science*. 2000; 289:2126–2128. [PubMed: 11000115]
8. Kaeberlein M, Kirkland KT, Fields S, Kennedy BK. Sir2-independent life span extension by calorie restriction in yeast. *PLoS Biol*. 2004; 2:E296. [PubMed: 15328540]
9. Fabrizio P, et al. Sir2 blocks extreme life-span extension. *Cell*. 2005; 123:655–667. [PubMed: 16286010]
10. Bass TM, Weinkove D, Houthoofd K, Gems D, Partridge L. Effects of resveratrol on lifespan in *Drosophila melanogaster* and *Caenorhabditis elegans*. *Mech Ageing Dev*. 2007; 128:546–552. [PubMed: 17875315]
11. Baur JA, et al. Resveratrol improves health and survival of mice on a high-calorie diet. *Nature*. 2006; 444:337–342. [PubMed: 17086191]
12. Lagouge M, et al. Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1alpha. *Cell*. 2006; 127:1109–1122. [PubMed: 17112576]
13. Pearson KJ, et al. Resveratrol delays age-related deterioration and mimics transcriptional aspects of dietary restriction without extending life span. *Cell Metab*. 2008; 8:157–168. [PubMed: 18599363]
14. Kaeberlein M, et al. Substrate-specific activation of sirtuins by resveratrol. *J Biol Chem*. 2005; 280:17038–17045. [PubMed: 15684413]
15. Behr D, et al. Resveratrol is not a direct activator of SIRT1 enzyme activity. *Chem Biol Drug Des*. 2009; 74:619–624. [PubMed: 19843076]
16. Pacholec M, et al. SRT1720, SRT2183, SRT1460, and resveratrol are not direct activators of SIRT1. *J Biol Chem*. 2010; 285:8340–8351. [PubMed: 20061378]
17. Canto C, et al. AMPK regulates energy expenditure by modulating NAD⁺ metabolism and SIRT1 activity. *Nature*. 2009; 458:1056–1060. [PubMed: 19262508]
18. Hawley SA, et al. Use of cells expressing gamma subunit variants to identify diverse mechanisms of AMPK activation. *Cell Metab*. 2010; 11:554–565. [PubMed: 20519126]
19. Garber K. A mid-life crisis for aging theory. *Nat Biotechnol*. 2008; 26:371–374. [PubMed: 18392009]
20. Luo J, et al. Negative control of p53 by Sir2alpha promotes cell survival under stress. *Cell*. 2001; 107:137–148. [PubMed: 11672522]
21. Vaziri H, et al. hSIR2(SIRT1) functions as an NAD-dependent p53 deacetylase. *Cell*. 2001; 107:149–159. [PubMed: 11672523]
22. Cheng HL, et al. Developmental defects and p53 hyperacetylation in Sir2 homolog (SIRT1)-deficient mice. *Proc Natl Acad Sci U S A*. 2003; 100:10794–10799. [PubMed: 12960381]
23. Kamel C, Abrol M, Jardine K, He X, McBurney MW. SirT1 fails to affect p53-mediated biological functions. *Aging Cell*. 2006; 5:81–88. [PubMed: 16441846]
24. Deng CX. SIRT1, is it a tumor promoter or tumor suppressor? *Int J Biol Sci*. 2009; 5:147–152. [PubMed: 19173036]
25. Firestein R, et al. The SIRT1 deacetylase suppresses intestinal tumorigenesis and colon cancer growth. *PLoS One*. 2008; 3:e2020. [PubMed: 18414679]
26. Herranz. Sirt1 improves healthy ageing and protects from metabolic syndrome - associated cancer. *Nature Communications*. 2010; 1:3
27. Oberdoerffer P, et al. SIRT1 redistribution on chromatin promotes genomic stability but alters gene expression during aging. *Cell*. 2008; 135:907–918. [PubMed: 19041753]
28. Wang RH, et al. Impaired DNA damage response, genome instability, and tumorigenesis in SIRT1 mutant mice. *Cancer Cell*. 2008; 14:312–323. [PubMed: 18835033]
29. Brooks CL, Gu W. How does SIRT1 affect metabolism, senescence and cancer? *Nat Rev Cancer*. 2009; 9:123–128. [PubMed: 19132007]
30. Haigis MC, Sinclair DA. Mammalian sirtuins: biological insights and disease relevance. *Annu Rev Pathol*. 2010; 5:253–295. [PubMed: 20078221]
31. Moynihan KA, et al. Increased dosage of mammalian Sir2 in pancreatic beta cells enhances glucose-stimulated insulin secretion in mice. *Cell Metab*. 2005; 2:105–117. [PubMed: 16098828]
32. Bordone L, et al. SIRT1 transgenic mice show phenotypes resembling calorie restriction. *Aging Cell*. 2007; 6:759–767. [PubMed: 17877786]

33. Pfluger PT, Herranz D, Velasco-Miguel S, Serrano M, Tschop MH. Sirt1 protects against high-fat diet-induced metabolic damage. *Proc Natl Acad Sci U S A*. 2008; 105:9793–9798. [PubMed: 18599449]
34. Banks AS, et al. SirT1 gain of function increases energy efficiency and prevents diabetes in mice. *Cell Metab*. 2008; 8:333–341. [PubMed: 18840364]
35. Escande C, et al. Deleted in breast cancer-1 regulates SIRT1 activity and contributes to high-fat diet-induced liver steatosis in mice. *J Clin Invest*. 2010; 120:545–558. [PubMed: 20071779]
36. Xu F, et al. Lack of SIRT1 (Mammalian Sirtuin 1) activity leads to liver steatosis in the SIRT1+/- mice: a role of lipid mobilization and inflammation. *Endocrinology*. 2010; 151:2504–2514. [PubMed: 20339025]
37. Schug TT, et al. Myeloid deletion of SIRT1 induces inflammatory signaling in response to environmental stress. *Mol Cell Biol*. 2010
38. Cohen DE, Supinski AM, Bonkowski MS, Donmez G, Guarente LP. Neuronal SIRT1 regulates endocrine and behavioral responses to calorie restriction. *Genes Dev*. 2009; 23:2812–2817. [PubMed: 20008932]
39. Ramadori G, et al. SIRT1 deacetylase in POMC neurons is required for homeostatic defenses against diet-induced obesity. *Cell Metab*. 2010; 12:78–87. [PubMed: 20620997]
40. Purushotham A, et al. Hepatocyte-specific deletion of SIRT1 alters fatty acid metabolism and results in hepatic steatosis and inflammation. *Cell Metab*. 2009; 9:327–338. [PubMed: 19356714]
41. Chen D, et al. Tissue-specific regulation of SIRT1 by calorie restriction. *Genes Dev*. 2008; 22:1753–1757. [PubMed: 18550784]
42. Bordone L, et al. Sirt1 regulates insulin secretion by repressing UCP2 in pancreatic beta cells. *PLoS Biol*. 2006; 4:e31. [PubMed: 16366736]
43. Li X, et al. SIRT1 deacetylates and positively regulates the nuclear receptor LXR. *Mol Cell*. 2007; 28:91–106. [PubMed: 17936707]
44. McBurney MW, et al. The mammalian SIR2alpha protein has a role in embryogenesis and gametogenesis. *Mol Cell Biol*. 2003; 23:38–54. [PubMed: 12482959]
45. Yeung F, et al. Modulation of NF-kappaB-dependent transcription and cell survival by the SIRT1 deacetylase. *EMBO J*. 2004; 23:2369–2380. [PubMed: 15152190]
46. Kemper JK, et al. FXR acetylation is normally dynamically regulated by p300 and SIRT1 but constitutively elevated in metabolic disease states. *Cell Metab*. 2009; 10:392–404. [PubMed: 19883617]
47. Rodgers JT, et al. Nutrient control of glucose homeostasis through a complex of PGC-1alpha and SIRT1. *Nature*. 2005; 434:113–118. [PubMed: 15744310]
48. Walker AK, et al. Conserved role of SIRT1 orthologs in fasting-dependent inhibition of the lipid/cholesterol regulator SREBP. *Genes Dev*. 2010; 24:1403–1417. [PubMed: 20595232]
49. Ponugoti B, et al. SIRT1 deacetylates and inhibits SREBP-1C activity in regulation of hepatic lipid metabolism. *J Biol Chem*. 2010
50. Picard F, et al. Sirt1 promotes fat mobilization in white adipocytes by repressing PPAR-gamma. *Nature*. 2004; 429:771–776. [PubMed: 15175761]
51. Kim HS, et al. Hepatic-specific disruption of SIRT6 in mice results in fatty liver formation due to enhanced glycolysis and triglyceride synthesis. *Cell Metab*. 2010; 12:224–236. [PubMed: 20816089]
52. Kanfi Y, et al. SIRT6 protects against pathological damage caused by diet-induced obesity. *Aging Cell*. 2010; 9:162–173. [PubMed: 20047575]
53. Hou X, et al. SIRT1 regulates hepatocyte lipid metabolism through activating AMP-activated protein kinase. *J Biol Chem*. 2008; 283:20015–20026. [PubMed: 18482975]
54. Lan F, Cacicedo JM, Ruderman N, Ido Y. SIRT1 modulation of the acetylation status, cytosolic localization, and activity of LKB1. Possible role in AMP-activated protein kinase activation. *J Biol Chem*. 2008; 283:27628–27635. [PubMed: 18687677]
55. Guarente L. Sirtuins as potential targets for metabolic syndrome. *Nature*. 2006; 444:868–874. [PubMed: 17167475]

56. Alcendor RR, et al. Sirt1 regulates aging and resistance to oxidative stress in the heart. *Circ Res.* 2007; 100:1512–1521. [PubMed: 17446436]
57. Zhang QJ, et al. Endothelium-specific overexpression of class III deacetylase SIRT1 decreases atherosclerosis in apolipoprotein E-deficient mice. *Cardiovasc Res.* 2008; 80:191–199. [PubMed: 18689793]
58. Stein S, et al. SIRT1 reduces endothelial activation without affecting vascular function in ApoE^{-/-} mice. *Aging (Albany NY).* 2010; 2:353–360. [PubMed: 20606253]
59. Stein S, et al. SIRT1 decreases Lox-1-mediated foam cell formation in atherogenesis. *Eur Heart J.* 2010
60. Park EJ, et al. Dietary and genetic obesity promote liver inflammation and tumorigenesis by enhancing IL-6 and TNF expression. *Cell.* 2010; 140:197–208. [PubMed: 20141834]
61. Kabra N, et al. SirT1 is an inhibitor of proliferation and tumor formation in colon cancer. *J Biol Chem.* 2009; 284:18210–18217. [PubMed: 19433578]
62. Boily G, He XH, Pearce B, Jardine K, McBurney MW. SirT1-null mice develop tumors at normal rates but are poorly protected by resveratrol. *Oncogene.* 2009; 28:2882–2893. [PubMed: 19503100]
63. Kim D, et al. SIRT1 deacetylase protects against neurodegeneration in models for Alzheimer's disease and amyotrophic lateral sclerosis. *EMBO J.* 2007; 26:3169–3179. [PubMed: 17581637]
64. Donmez G, Wang D, Cohen DE, Guarente L. Sirt1 suppresses β -amyloid production by activating the α -secretase gene ADAM10. *Cell.* 2010; 142:320–332. [PubMed: 20655472]
65. Boily G, et al. SirT1 regulates energy metabolism and response to caloric restriction in mice. *PLoS One.* 2008; 3:e1759. [PubMed: 18335035]
66. Chen D, Steele AD, Lindquist S, Guarente L. Increase in activity during calorie restriction requires Sirt1. *Science.* 2005; 310:1641. [PubMed: 16339438]
67. Qin W, et al. Neuronal SIRT1 activation as a novel mechanism underlying the prevention of Alzheimer disease amyloid neuropathology by calorie restriction. *J Biol Chem.* 2006; 281:21745–21754. [PubMed: 16751189]
68. Satoh A, et al. SIRT1 promotes the central adaptive response to diet restriction through activation of the dorsomedial and lateral nuclei of the hypothalamus. *J Neurosci.* 2010; 30:10220–10232. [PubMed: 20668205]
69. Kume S, et al. Calorie restriction enhances cell adaptation to hypoxia through Sirt1-dependent mitochondrial autophagy in mouse aged kidney. *J Clin Invest.* 2010; 120:1043–1055. [PubMed: 20335657]
70. Zheng J, Ramirez VD. Inhibition of mitochondrial proton F₀F₁-ATPase/ATP synthase by polyphenolic phytochemicals. *Br J Pharmacol.* 2000; 130:1115–1123. [PubMed: 10882397]
71. Gledhill JR, Montgomery MG, Leslie AG, Walker JE. Mechanism of inhibition of bovine F₁-ATPase by resveratrol and related polyphenols. *Proc Natl Acad Sci U S A.* 2007; 104:13632–13637. [PubMed: 17698806]

Box1: Resveratrol and Sirt1

Resveratrol has been widely used as a Sirt1 activator, yet a number of studies have concluded that resveratrol is not a direct activator of Sirt1¹⁴⁻¹⁶. This does not invalidate the concept that resveratrol, through indirect mechanisms, can activate Sirt1 *in vivo*. In fact, earlier reports more than 10 years ago already indicated that resveratrol was an inhibitor of mitochondrial ATP synthase⁷⁰ and this was corroborated by the crystallographic structure of ATP synthase complexed with resveratrol⁷¹. Inhibition of ATP synthase is predicted to increase AMP levels and therefore activate AMPK. In fact, resveratrol has been recently demonstrated to activate AMPK¹⁸, and, finally, it is now known that AMPK increases NAD⁺ levels and this, in turn, activates Sirt1¹⁷. This chain of events now provides a rationale for how resveratrol, albeit indirectly, can activate Sirt1.

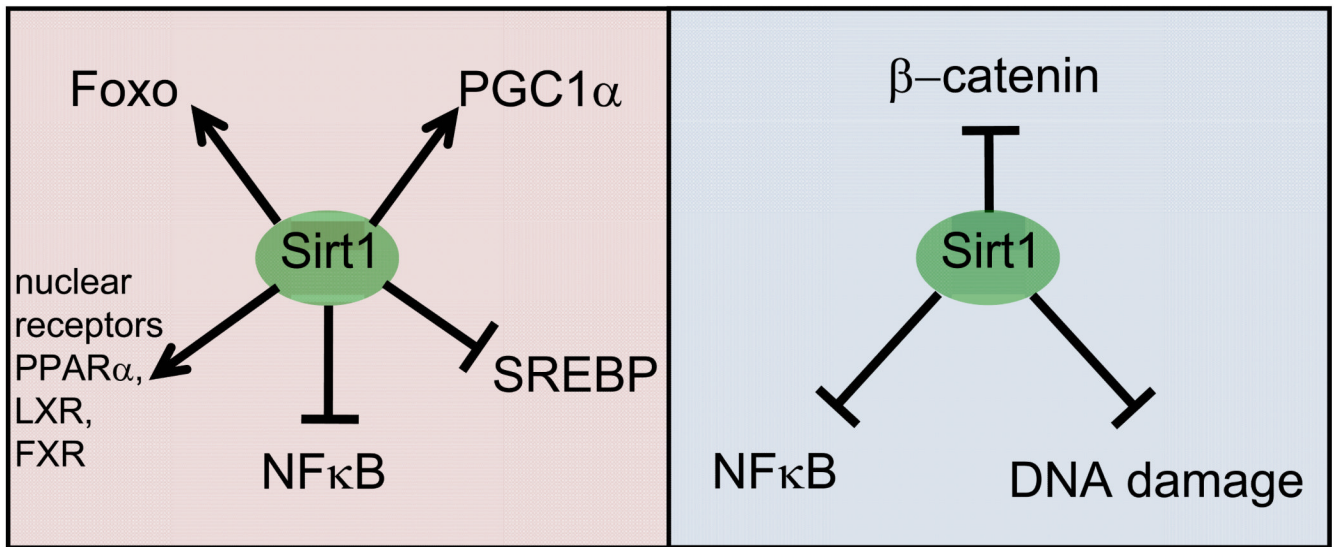


Figure 1. Summary of the main mechanisms through which Sirt1 protects from metabolic damage and cancer

The left panel shows the main mechanisms by which Sirt1, through direct deacetylation, protects from metabolic damage. Other mechanisms are mentioned in the text. The right panel shows the three mechanisms reported so far supporting cancer protection by Sirt1 in mice.

Table 1

Sirt1 mouse models and their effects on cancer and metabolism

Modification of Sirt1 levels	Mouse model	Cancer phenotypes ^(*)	Metabolic phenotypes ^(*)	Ref.
Whole-body overexpression	Whole-body Sirt1-tg	*Decreased aging associated carcinomas and sarcomas *Decreased metabolic-syndrome associated cancer *No effect in chemically-induced fibrosarcomas *No effect in aging associated lymphomas	*Protection from glucose intolerance (associated to aging, diet-induced obesity, and genetically-induced obesity) *Protection from fatty liver disease (induced by HFD) *Decreased inflammation in WAT and liver (induced by HFD)	26,33,34
Tissue-specific overexpression	β-cells Sirt1-tg	Not explored	*Increased insulin secretion	31
	Brain and adipose tissue Sirt1-tg	Not explored	*Increased glucose tolerance *Decreased body weight	32
	Intestinal Sirt1-tg combined with Apc ^{+/min}	*Decreased intestinal polyp formation	Not explored	25
Whole-body deficiency	Whole-body Sirt1-null	*No effect in Apc ^{+/min} -induced intestinal polyps *No effect in chemically-induced papillomas	*Decreased insulin production *Improved glucose tolerance *Protection from fatty liver (induced by LXR agonists)	42,43,62
	Whole-body Sirt1-het	Not explored	*Increased fatty liver disease (induced by HFD) *Increased body weight *Increased liver inflammation	36
	Whole-body Sirt1-het combined with p53-deficiency	*Increased incidence of sarcomas and lymphomas associated to p53 deficiency	Not explored	28
Tissue-specific deficiency	Liver Sirt1-null	Not explored	*Protection from glucose intolerance (induced by HFD) *Decreased liver weight with HFD *Decreased body weight with HFD	41
	Liver Sirt1-null	Not explored	*Increased fatty liver disease (induced by HFD) *Increased inflammation and ER-stress in liver (induced by HFD) *Increased body weight with HFD	40
	Myeloid lineage Sirt1-null	Not explored	*Increased glucose intolerance (induced by HFD) *Increased inflammation in WAT and liver (induced by HFD)	37
	Brain Sirt1-null	Not explored	*Increased glucose intolerance (associated to aging)	38
	POMC neurons Sirt1-null	Not explored	*Protection from diet-induced obesity	39

(*): Phenotypes in green colour are those that imply a beneficial effect of Sirt1, on the contrary, those in red imply a detrimental effect of Sirt1.