www.nature.com/psp

ORIGINAL ARTICLE

A Nonlinear Mixed Effects Pharmacokinetic Model for Dapagliflozin and Dapagliflozin 3-*O*-glucuronide in Renal or Hepatic Impairment

J-S van der Walt¹, Y Hong², L Zhang², M Pfister², DW Boulton² and MO Karlsson¹

Dapagliflozin is a sodium–glucose co-transporter 2 inhibitor in development for the treatment of type 2 diabetes mellitus. A semi-mechanistic population pharmacokinetic (PK) model was developed for dapagliflozin and its inactive metabolite dapagliflozin 3-*O*-glucuronide (D3OG) with emphasis on renal and hepatic contribution to dapagliflozin metabolism. Renal and hepatic impairment decreased the clearance of dapagliflozin to D3OG and the clearance of D3OG. The fraction of D3OG formed via the renal route decreased from 40–55% in subjects with normal renal function (creatinine clearance (CLcr) > 80 ml/min) to 10% in subjects with severe renal insufficiency (CLcr = 13 ml/min). The model-based simulations suggested that the increase of systemic exposure (AUC_{ss}) of dapagliflozin and D3OG was less than twofold in subjects with mild or moderate renal impairment. This population modeling analysis presents a useful approach to evaluate the impact of renal and hepatic function on the PK of dapagliflozin.

CPT: Pharmacometrics & Systems Pharmacology (2013) 2, e42; doi:10.1038/psp.2013.20; advance online publication 8 May 2013

Dapagliflozin is a therapy in development for the treatment of type 2 diabetes mellitus (T2DM). Under normal conditions, ~180g of glucose is filtered through the renal glomerulus per day, and virtually all of this is reabsorbed into blood by the proximal tubules with the major facilitator of reabsorption being the sodium–glucose cotransporter.¹ Dapagliflozin acts reversibly and specifically on the sodium–glucose co-transporter 2 in the proximal tubules of the kidney to inhibit the reabsorption of glucose and to promote urinary glucose excretion.²

Dapagliflozin is rapidly absorbed following oral administration with an absolute oral bioavailability of 78%.3 The systemic exposures of dapagliflozin increase in a dose-dependent manner for doses ranging from 0.1 to 500 mg. The half-life of dapagliflozin (12.5h) and the corresponding sustained inhibition of urinary glucose reabsorption over 24-h after dose at the proposed clinical dose (10 mg) is amenable to oncedaily dosing.² Dapagliflozin is metabolized by uridine diphosphate glucuronosyltransferase (UGT) 1A9 to dapagliflozin 3-O-glucuronide (D3OG identified as M₁₅ in chromatography), which is 2600-fold less potent than the parent drug with regard to sodium-glucose co-transporter 2 inhibition. Following a single 50-mg dose of [14C]-dapagliflozin, D3OG and dapagliflozin represented >95% of the total radioactivity in all plasma samples. In healthy subjects, ~61% of the administered dose of dapagliflozin is recovered in urine as D3OG.4 Less than 2% of the administered dose of dapagliflozin is excreted in urine as unchanged dapagliflozin.2

Approximately 40% of adults with T2DM have comorbid chronic renal disease, and T2DM is widely regarded as the leading cause of end-stage renal disease.⁵ Individuals with T2DM have a higher incidence of liver function test abnormalities than those without T2DM, and therefore, hepatic impairment (HI) is also an important consideration in this population.⁶

A pharmacokinetic (PK)/pharmacodynamic (PD) study of dapagliflozin in renal impairment (NCT00554450) and a PK study in HI have been described.7-9 In the renal impairment study, systemic exposure of both dapagliflozin and D3OG increased incrementally with decreased renal function. The PK of dapagliflozin in patients with T2DM and normal renal function are comparable to that observed in healthy subjects.² Compared with healthy subjects and T2DM subjects with normal renal function (creatinine clearance $(CL_{rr}) > 80 \text{ ml/min}$), the dapagliflozin exposure (area under the concentration time curve from time zero to infinity (AUC₀₋₋₋)) increased by 28% in patients with T2DM and mild renal impairment (CL_{cr} > 50 and CL_x ≤ 80 ml/min), by 52% in patients with T2DM and moderate renal impairment ($CL_{cr} \ge 30$ and $CL_{cr} \le 50$ ml/min), and by 75% in patients with T2DM and severe renal impairment not on hemodialysis (CL_{cr} < 30 ml/min). The categorization of various stages of renal impairment was trial defined; however, the Kidney Disease Outcomes Quality Initiative clinical practice guidelines for chronic kidney disease define mild, moderate, and severe renal impairment as glomerular filtration rate of 60-89, 30-59, and 15-29 ml/min, respectively.10

Glucuronide conjugation is usually preserved in patients with hepatic insufficiency until the disease is severe (Child-Pugh Class C). In these cases, increased systemic exposure of dapagliflozin and decreased D3OG would be expected. In the HI study, patients with mild, moderate, and severe HI showed increases in the D3OG AUC_{0-∞} by 6, 100, and 29%, respectively, compared with age-, weight-, gender-, and smoking status-matched healthy subjects. These values were highly dependent on the calculated CL_{cr} of each group.

Furthermore, as UGT1A9 is expressed in both the kidney and the liver,¹¹ we postulated that the UGT1A9-mediated metabolic clearance of dapagliflozin occurs in both of these

¹Department of Pharmaceutical Biosciences, Uppsala University, Uppsala, Sweden; ²Discovery Medicine and Clinical Pharmacology, Bristol-Myers Squibb, Princeton, New Jersey, USA. Correspondence: MO Karlsson (mats.karlsson@farmbio.uu.se)

Received 19 December 2012; accepted 3 March 2013; advance online publication 8 May 2013. doi:10.1038/psp.2013.20

organs, and that both hepatic and renal impairment may impact the metabolic clearance of dapagliflozin. Therefore, we developed a semi-mechanistic nonlinear mixed effects model to gain further insights into the effects of renal and HI on the population PK of dapagliflozin and D3OG.

RESULTS

Base model

The structural model that best described the plasma and urine data of dapagliflozin and D3OG simultaneously is shown in **Figure 1**. The parameter estimates of the base model are provided in **Table 1**.

The medians (interquartile range) of fraction of dapagliflozin excreted in urine (FE) and fraction of dapagliflozin metabolized to D3OG (FM_{M15}) were 1.6% (1.2–2.1%) and 60% (53–72%), respectively. In the base model, FE and FM_{M15} were reduced in subjects with severe renal impairment (**Supplementary Figure S1** online). The CLP_{M15} and the volume of distribution of D3OG (V2M) were found to be correlated (*r* = 0.6). Overall, the single-dose base model described the observed data well but underpredicted the population D3OG concentrations in plasma in subjects with CL_{cr} < 50 ml/ min (data not shown).

The estimates of the proportional components of the residual variability in plasma dapagliflozin and D3OG were 20.8 and 19.5%, respectively. Replicate residual variability in plasma accounted for 47.9% of the variability. As expected, residual variability in urine was larger: 54.2 and 51.9% for dapagliflozin and D3OG, respectively. Replicate residual variability in urine accounted for 80.4% of the variability.

Covariate model

Significant covariate relationships identified during the general additive modeling analysis and included in the stepwise covariate model (SCM) building are summarized in Table 2 and Figure 1. The parameter estimates of the single-dose covariate model are shown in Table 1. Renal function influenced the CLP_{M15} . The fraction of D3OG formed via the renal route decreased from 46% in a typical subject with normal renal function ($CL_{cr} \sim 100 \text{ ml/min}$) to 10% in a typical subject with $CL_{cr} = 13 \text{ ml/min}$, the lowest value in the study population. This relationship is illustrated in Figure 2.

The covariate model suggested that both formation (CLP_{M15}) and elimination (CLM) of D3OG were altered with HI. With severe HI (Child-Pugh Class C), CLP_{M15} decreased by 41% and V2M increased by 134%. Moderate or severe HI (Child-Pugh Class B or C) decreased CLM and the peripheral volume of distribution of dapagliflozin (V3P) by 29 and 60%, respectively.

During the backward elimination, the effects of age on the renal clearance of unchanged dapagliflozin to urine (CLP_{renal}), CLM, and V3P, and the effects of gender on CLM and the central volume of distribution of dapagliflozin (V2P) were removed. The effect of age on the metabolic clearance of dapagliflozin to unmeasured metabolites (CLP_{other}) remained significant, and the CLP_{other} decreased by 2% for every year of increasing age above 54 years.

The covariate model described the single- and multipledose data best and improved the population-predicted plasma D3OG concentrations compared with the base model (data not shown).

Final model

The final model incorporated gender effects on CLM and total dapagliflozin clearance (CLP) and resulted in a statistically significant improvements in model fit (Δ OFV = -15.828 points). In females, CLP and CLM were 16.7 and 19.6% lower, respectively. The parameter estimates of the final model are provided in Table 1.



Figure 1 The structural and covariate model to describe the renal and nonrenal elimination of dapagliflozin and D3OG in healthy subjects, T2DM subjects with normal or impaired renal function, and patients with hepatic impairment. BCL_c,IBW, baseline creatinine clearance calculated using ideal body weight; CLM, renal clearance of D3OG; BIO, bioavailability; CLP_{M15}, metabolic clearance of dapagliflozin to D3OG; CLP_{other} metabolic clearance of dapagliflozin to unmeasured metabolites; CLP_{renal}, renal clearance of dapagliflozin to unine; MTT, mean transit time; *N*, number of transit compartments; QP, intercompartmental clearance of dapagliflozin; T2DM, type 2 diabetes mellitus; V2P, central volume of distribution of dapagliflozin; V2M, central volume of distribution of D3OG; V3P, peripheral volume of distribution of adapagliflozin; shaded areas, added based on previous modeling experience. Covariates connected to compartments affect the relevant volume, those connected to pathways affect the relevant clearance.

3

Table 1 Parameter estimates

Parameter estimates	Single deep be		Single doop gove	riata madal	Final may		
Name (units)	Typical value ^a	Typical value ^a BSE% ^b					
Structural fixed effects		HSE /6		NGL /0	Typical value	NGL /0	
	6.67	9.7	6.88	12.5	7.54	12.0	
CLP (l/b)	3.83	12.0	4.85	12.5	5 35	12.0	
CLP_{other} (1/h)/(ml/minuto))	0.00288	10.0	4.00	9.0	0.00210	0.0	
	0.00200	10.9	0.00200	0.0	0.00310	9.9	
	57.7	4.0	50.2	4.3	39.0	4.4	
	6.09	8.0	6.91	7.3	7.07	7.1	
	01	9.7	09.0	7.1	/ 1.5	7.0	
	0.479	7.0	0.47	1.2	0.475	1.2	
	5.69	18.1	5.47	C.01	5.45	IO.D	
	0.823	NR ³	0.84	NR [®]	0.858	NR ³	
	0.0716	6.6	0.0723	6.1	0.0799	7.6	
V2M (I)	2.55	6.5	2.22	5.8	2.26	5.6	
	N14		0.00475	40.0	0.00500	40.0	
CLP _{M15} -Baseline CL _{cr} IBW	NA	NA	0.00475	49.3	0.00502	42.6	
CLP _{M15} -Child-Pugh Class C	NA	NA	-0.407	42.8	-0.422	38.6	
CLP _{other} – age	NA	NA	-0.0215	24.0	-0.0204	22.1	
V2M- Child-Pugh Class C	NA	NA	1.34	16.3	1.33	17.3	
V3P- Child-Pugh Class B,C	NA	NA	-0.596	12.4	-0.600	12.6	
CLM- Child-Pugh Class B,C	NA	NA	-0.292	25.4	-0.293	23.3	
CLP-gender	NA	NA	NA	NA	-0.167	64.1	
CLM-gender	NA	NA	NA	NA	-0.196	38.9	
Random effects ^a							
IIV. CLP _{M15}	51.6	15.0	38.0	17.4	36.8	16.5	
IIV.CLP _{other}	52.6	50.0	32.9	20.4	31.3	22.3	
IIV.CLP _{renal}	62.1	38.7	63.1	18.8	61.9	18.4	
IIV.V2P	21.9	32.3	22.6	20.0	23.6	20.5	
IIV.QP	42.5	39.5	38.8	41.1	39.1	35.9	
IIV.V3P	57.4	29.5	46.5	15.8	46.3	17.4	
IIV.MMT	59.1	28.0	60.1	20.5	60.3	21.0	
IIV.NN	195	13.2	189	10.9	189	10.6	
IIV. BIO ^g	13.3	NAd	12.7	NAd	11.1	15.2	
IIV.CLM	33.3	36.7	27.9	20.4	24.9	23.6	
IIV.V2M	66.2	12.8	46.5	13.3	44.7	15.5	
Correlation CLP _{M15} : V2M ^h	<i>r</i> = 0.60	35.7	<i>r</i> = 0.56	47.4	<i>r</i> = 0.562	49.9	
Residual error ⁱ							
Prop, dapa_plasma	0.208	7.8	0.207	6.4	0.207	6.4	
Add, dapa_plasma (ng/ml)	0.424	87.5	0.463	31.7	0.465	29.9	
Prop, M15_ plasma	0.195	74.4	0.195	6.5	0.195	6.4	
Add, M15_plasma (ng/ml)	0.633	98.1	0.579	53.2	0.585	51.3	
Prop, dapa_urine	0.542	8.4	0.565	10.1	0.564	9.9	
Add, dapa urine (ng/ml)	0.667	506.7	2.21	290.5	1.784	640.4	
Prop, M15_urine	0.519	8.1	0.564	8.7	0.567	8.9	
Add, M15_urine (ng/ml)°	0.02 fix	0 (0)	0.02 fix	0 (0)	0.02 fix	0 (0)	
Plasma, replicate	0.479	12.8	0.473	12.4	0.475	12.5	
Urine, replicate	0.804	6.1	0.820	6.0	0.821	6.2	

BIO, oral bioavailability of dapagliflozin; CLM, renal clearance of D3OG; CLP, metabolic clearance of dapagliflozin to D3OG; CLP_{other}, metabolic clearance of dapagliflozin to unmeasured metabolites; CLP, enal, renal clearance of unchanged dapagliflozin to urine; FOCE, first-order condition method; IIV, interindividual variability; MTT, mean transit time; N, number of transit compartments; QP, inter-compartmental clearance of dapagliflozin; V2M, central volume of distribution of D3OG; V2P, central volume of distribution of dapagliflozin; V3P, peripheral volume of distribution of dapagliflozin. ^aRandom Effects parameter estimates are shown as %CV = $\sqrt{e^{\frac{\omega_c}{U}} - 1}$. ^bRSE% is the relative standard error (SE as a percentage of estimate) obtained from

Monte Carlo Importance Sampling EM Assisted by Mode a Posteriori estimation (IMPMAP) with Monte Carlo samples per individual = 10,000. °Renal clearances reported as I/h per mI/min creatinine clearance. ^dNot reported as this random effect was not MU_parameterized, a requirement for NONMEM7 IMPMAP method. "Fixed value. 'Covariates effects are identified by structural fixed effect then by covariate $\omega_{i,i}$ "Raw $\omega_{i,1}$ (logit scale) with coefficient of variance calculated using $SD = \theta_{TVBIO} \cdot (1 - \theta_{TVBIO}) \cdot \sqrt{\omega_{BIO}^2}$. ^hcorrelation $r = \omega_{i,i}^2 / \sqrt{\omega_{i,j}^2 \cdot \omega_{i,j}^2}$. ^hResidual error parameters are identified by type (proportional (prop) or additive (add)) then type of observation (dapa plasma, dapa urine, D3OG plasma, and D3OG urine).

Table 2 Summary of baseline covariates and PK–covariate relationships included *a priori*, retained (\checkmark) or removed (\times) during backwards deletion of the stepwise covariate model building (P = 0.01)

				Reference value of the							
Baseline covariates	Median (ra	ange) or cate	gory <i>N</i> /total	relationship ^{e,f}	CLP _{renal}	CLP M15	CLP	CLM	V2P	V3P	V2M
Study	MB102007	MB102027	MB102029								
Age (years)	63 (25–76)	43 (31–64)	67 (32–92)	53.5		×	\checkmark	×		×	
$CL_{_{cr,IBW}}$ (ml/min) ^b	60 (13–116)	103 (58–143)	40 (20–80)	80.14	a priori	✓°		a priori			
Body weight (kg)	81.2 (51.8–140.4)	86.3 (58.0– 126.0)	92.0 (55.4–148.3)	70 ^a	×	a priori	a priori		a priori	a priori	a priori
Body mass index (kg/m²) ≥30	19/40	11/24	108/163	N/A							
Male	24/40	16/24	107/163	Male				×	×		
Hepatic impairment ^c											
None/mild	40/40	12/24 ^d	N/A	None/mild			1		1		
Moderate/severe	0/40	12/24	N/A	None/mild			1		1		
Not severe	40/40	18/24	N/A	Not severe		1			×		1
Severe	0/40	6/24	N/A	Not severe		1			×		1

CL_{cr}IBW, creatinine clearance calculated using ideal body weight; CLM, renal clearance of D3OG; CLP_{M15}, metabolic clearance of dapagliflozin to D3OG; CLP_{other}, metabolic clearance of dapagliflozin to unmeasured metabolites; CLP_{renal}, renal clearance of unchanged dapagliflozin to urine; N/A, not applicable; QP, inter-compartmental clearance of dapagliflozin; V2M, central volume of distribution of D3OG; V2P, central volume of distribution of dapagliflozin; V3P, peripheral volume of distribution of dapagliflozin; NA, not used during the model development.

^aAllometric scaling (BBWT_i/70 kg)^{3/4}. ^bCreatinine clearance calculated using ideal body weight (IBW) was selected based on goodness-of-fit during development of the base model. ^cNormal, mild (Child-Pugh A), moderate (Child-Pugh B), severe (Child-Pugh C). ^dIncluding age, gender, smoking-status matched healthy controls. ^cContinuous covariates (i.e., age and CL_{cr,IBW}) were modeled as a proportional change model centered on the mean covariate (see supplemental NONMEM code). ^fFor categorical covariates (i.e., sex and hepatic impairment) the indicator variable for the most common category was set to 1 and the proportional change for the less common category was estimated (see supplemental NONMEM code).



Figure 2 Renal contribution to the metabolism of dapagliflozin to D3OG in healthy subjects, and T2DM subjects with normal or impaired renal function. Solid squares, MB102007; open squares, MB102027; Line, population predictions.

Model evaluation

In the final model, all the structural parameters were estimated with good precision (% relative standard error < 20%). The addition of gender effect on CLP and CLM improved the precision in the estimate of the effects of baseline CL_{cr} and severe HI (Child-Pugh Class C) on CLP_{M15} .

The visual predictive checks confirmed that the model had good simulation properties (Supplementary Figure S3

online). As seen in **Figures 3** and **4**, there was good agreement in model predicted and observed plasma and urine dapagliflozin and D3OG concentrations for different categories of renal function: $CL_{cr} < 30, 30-45, 45-60, 60-90$, and >90 ml/min. However, given the high variability in urine concentrations and the paucity of urine data in subjects with $CL_{cr} > 90$ ml/min, the model predictions in this stratum should be interpreted with caution. In an external evaluation, the final model accurately predicted the plasma dapagliflozin and D3OG concentrations in T2DM subjects with moderate renal impairment (**Supplementary Figure S2** online).

Model application

Simulations with the final model were performed to gauge the effect of decreasing renal function on the exposure of dapagliflozin and D3OG (**Table 3**). Compared with T2DM subjects with normal renal function (CL_{cr} of 80–100 ml/min), the exposure of dapagliflozin (realized as AUC at steady state (AUC_{ss})) was 1.26 and 1.35 times higher than subjects with CL_{cr} of 50–79 and 30–49 ml/min, respectively. The increase in D3OG exposure exceeds the increase in dapagliflozin exposure (1.32 times at CL_{cr} of 50–79 ml/min and 1.57 times at CL_{cr} of 30–49 ml/min). Similar changes were seen when patient characteristics of the two phase I studies were used for simulations.

DISCUSSION

Main findings

The semi-mechanistic model indicated that the influence of both renal and hepatic function on the ${\rm CLP}_{_{\rm M15}}$ was the most



Figure 3 Visual predictive check: plasma dapagliflozin and D3OG by renal function. CL_{cr}, creatinine clearance. Points, observed data; solid line, median of the observations; dotted lines, 5th and 95th percentiles of the observations; shaded areas, 95% confidence interval for simulated data (1,000 simulated data sets) for the corresponding percentiles.



Figure 4 Visual predictive check: urine dapagliflozin and D3OG by renal functions. Points, observed data; solid line, median of the observations; dotted lines, 5th and 95th percentiles of the observations; shaded areas, 95% confidence interval for simulated data (1,000 simulated data sets).

likely explanation of the altered dapagliflozin and D3OG PK observed in the subjects with renal or HI. The fraction of D3OG formed via the renal route decreased from 40–55% in subjects with normal renal function (CL_{cr} >80 ml/min) to ~10% in subjects with severe renal insufficiency (CL_{cr} = 13 ml/min) (**Figure 2**).With severe HI, reduction of 42% in the CLP_{M15} and 29% in the CLM resulted in D3OG exposure comparable to that in healthy subjects. The subjects with moderate HI had nearly double the exposure to D3OG. Therefore, differences in systemic exposure to D3OG between the HI and healthy subject groups did not correlate directly with the severity of HI. The possible explanation for these findings is that D3OG is mainly cleared via renal excretion, and subjects with severe HI may concomitantly present renal dysfunction.¹²

Effect of renal impairment

Metabolism (primarily glucuronidation via UGT1A9) is the major elimination pathway of dapagliflozin: ~60%, and <2% of an administered dapagliflozin dose is recovered from urine as D3OG and dapagliflozin, respectively.⁴ The tissue mRNA expression of UGT1A9 in humans has been reported in the literature. On the basis of the reverse transcriptase polymerase chain reaction data, the expression of UGT1A9 was found to be about eightfold higher in the human kidney as compared with that of the liver.¹³ These data suggest the involvement of kidney in the metabolism of dapagliflozin. Our analysis suggested that the impact of renal impairment on the metabolism of dapagliflozin was manifested by the reduced metabolic clearance of dapagliflozin. Although there is evidence in the

Renal	Dapagliflozin AUC _{ss} (ng*h/ml)⁰				D3OG AUC _{ss} (ng*h/ml)°				
function	nction Phase I ^e		Phase III ^f		Phase I ^e		Phase III ^r		
CL_a ^a	Median (95%CI) ^d	Ratio⁵	Median (95%CI) ^d	Ratio ^b	Median (95%CI) ^d	Ratio ^₅	Median (95%CI) ^d	Ratio⁵	
30–49	711 (660, 777)	1.26	793 (644, 987)	1.35	1696 (1600, 1864)	1.57	2053 (1573, 2645)	1.57	
50–79	567 (485,647)	1.07	697 (563, 839)	1.26	1133 (1012, 1317)	1.36	1283 (1010,1535)	1.32	
80–100	526 (260,822)	-	515 (396,614)	-	727 (319,1292)	-	873 (664, 1118)	-	

 Table 3 Effects of renal function on steady-state dapagliflozin and D3OG exposure

AUC, area under the plasma concentration-time curve; CI, confidence interval.

^aBaseline creatinine clearance corrected for ideal body weight (ml/min). ^bRatio of the median AUC compared with the median AUC for baseline creatinine clearance of 80–100 ml/min. ^cSteady-state AUC. ^d95% CI for 100 simulated trials. ^eCovariates (baseline body weight, age, creatinine clearance corrected for ideal body weight and sex) from studies MB102007 and MB102027 (were used for simulations with a dose of dapagliflozin 10 mg daily. [']Covariates (baseline body weight, age, creatinine clearance corrected for ideal body weight and sex) from study MB102029 were used for simulations with a dose of dapagliflozin 10 mg daily. [']Covariates (baseline body weight, age, creatinine clearance corrected for ideal body weight and sex) from study MB102029 were used for simulations with a dose of dapagliflozin 10 mg daily.

literature to suggest that chronic kidney disease contributes to decreased nonrenal metabolism,¹⁴ the impact of chronic kidney disease on the liver metabolism of dapagliflozin is not known. Nevertheless, our simulation results along with the literature findings suggest that the kidney plays an important role in the metabolism of dapagliflozin via the UGT1A9 pathway, and this may also explain the greater systemic exposure of dapagliflozin in subjects with moderate and severe renal impairment at steady state as compared with subjects with normal renal function.

Effect of HI

The model assigned the effects of HI on the formation (CLP_{M15}) and clearance of D3OG (CLM) during the stepwise covariate modeling. For technical reasons (linearized SCM), hepatic dysfunction was tested as two separate binary covariates (i.e., mild vs. moderate/severe, and mild/moderate vs. severe). The selection of mild/moderate vs. severe on CLP_{M15} and mild vs. moderate/severe on CLM should be seen in context of the flexibility of the model, limited data, and the loss of information when categorizing HI.¹⁵ Despite that all subjects contributed data on renal function, only six (9%) informed the model regarding moderate or severe HI. The results suggest that metabolic clearance of dapagliflozin to D3OG (CLP_{M15}) is preserved in patients with moderate HI.

Absorption and oral bioavailability

Misspecification of dapagliflozin absorption also resulted in misfit of D3OG data during early sampling times. Therefore, considerable effort was made to characterize the absorption of dapagliflozin. A transit compartment model estimating both the mean transit time and number of transit compartments resulted in a flexible absorption model that well described the rich data during the absorption phase. The drawback was considerably longer computation time, especially when incorporating the multiple-dose data.

Absolute oral bioavailability and nonrenal clearance can be determined from modeling oral data when plasma and urine data are available in subjects with varying renal function.¹⁶⁻¹⁸ Estimating the absolute oral bioavailability, and thereby the absolute value of clearance, reduced the risk of biased estimates of apparent nonrenal clearance when only oral data were available.¹⁹ The model-estimated absolute bioavailability of dapagliflozin was 82% (base model), 84% (covariate model), and 86% (final model), which are in good agreement with the absolute oral bioavailability (geometric mean (90% Cl): 78% (73–83%)) determined in healthy subjects following the concomitant administration of single intravenous ¹⁴C-labeled microdose at time of maximum concentration (T_{max}) of the extravascular (unlabeled) therapeutic dose.³

Model application

Mild renal impairment is common in T2DM and presents a significant proportion of the population treated with dapagliflozin. Given the interplay in body weight, sex, and renal function, simulations were performed (using the covariates reported for each subject) to assess the effects of mild and moderate renal impairment on dapagliflozin and D3OG exposure. As seen in Table 3, the simulations based on the subjects who participated in the phase I PK studies do not differ substantially from those for the subjects in the phase III study. The increases in D3OG AUC see of 1.36 and 1.32 for mild (50 \leq Cl_{cr} < 80 ml/min), and 1.57 and 1.57 for moderate $(30 \le Cl_{r} < 50 \text{ ml/min})$ renal impairment, are comparable. However, the simulations using the characteristics of phase I subjects could have underpredicted the increase in dapagliflozin exposure compared with using the characteristics from the phase III study. Differences in body weights of subjects in the phase I and III studies should be considered when a model developed with mostly normal weight subjects is used to simulate into a new and obese population.

In conclusion, the semi-mechanistic model developed to estimate the renal and nonrenal clearance of dapagliflozin presented a simplification of the interplay among biological systems; however, it provided a useful approach to evaluate the impact of renal and HI on the population PK of dapagliflozin. The results emphasized the role of renal metabolism of dapagliflozin to D3OG, which is the major elimination pathway of dapagliflozin. The model estimated that the kidneys contributed 40–55% of the metabolism of dapagliflozin with normal renal function. In renal impairment and severe HI, both formation and elimination of D3OG were decreased. In T2DM subjects, D3OG systemic exposure (AUC_{ss}) increased more than dapagliflozin exposure.

METHODS

Study design

Dapagliflozin and D3OG plasma and urine concentration-time data from a single- and multiple-dose (7 days of q.d. dosing) renal impairment study (NCT00554450),^{7,8} a 5-day single-dose HI study,⁹ and a 52-week, phase III (NCT00663260) clinical trial²⁰ in patients with T2DM and moderate renal impairment were available for analysis.

The renal impairment PK/PD study evaluated the PK, PD, and safety and tolerability of oral dapagliflozin in eight healthy adult subjects (50 mg single dose) and in 32 T2DM subjects with normal, mild, moderate, and severe renal impairment (50 mg single dose followed by 20 mg daily dose for 7 days).^{7,8} The HI study compared the safety and serial PK of a single dose of dapagliflozin (10 mg) in 18 subjects with HI and 6 healthy subjects.⁹ The phase 3 clinical trial investigated the glycemic efficacy, renal safety, and PK/PD of dapagliflozin in 252 subjects with T2DM and moderate renal impairment.²⁰

Each clinical trial was conducted in accordance to local and international ethics standards and was monitored for compliance with current good clinical practice. The appropriate ethics and regulatory approvals were obtained before study initiation, and written informed consent was obtained from all participants.

Analysis of dapagliflozin and D3OG concentrations

Analysis of dapagliflozin and D3OG plasma and urine concentrations was performed using high-pressure liquid chromatography with tandem mass spectrometry detection within the known period of stability. All determinations of dapagliflozin and D3OG concentrations in human plasma and urine were generated in analytical runs using appropriate calibration curves and quality control samples that met preestablished acceptance criteria. The standard curve ranges for dapagliflozin in plasma and urine were 1–500 and 1–1000 ng/ml, respectively. The standard curve ranges for D3OG in plasma and urine were 1–500 and 10–5000 ng/ml, respectively. The between-run and within-run coefficients of variation for the analytical quality controls of both analytes in either matrix were \leq 10%.

Pharmacometric analysis

Modeling approach The single/first-dose data were used to develop a semi-mechanistic (base) model to estimate the renal and nonrenal clearances of dapagliflozin. SCM building was used to evaluate the effects of renal function markers, HI, and other baseline covariates on the structural parameters (covariate model). After confirming that the single-dose models adequately described the multiple-dose data in subjects with T2DM and moderate renal impairment, the final model incorporated important covariates identified in the previous analyses.²¹ The NONMEM code (ICON Development Solutions, Ellicott city, MD), example data set, and summary of data used in the model development and evaluation are provided in the **Supplementary Material**.

The compartmental models were parameterized as clearance and volume terms using a parameterization that is stable for different types of estimation methods. The structural model parameters were described as follows (Eqs. 1 and 2):

$$\mathsf{MU}_i = \mathsf{log} P_{\mathsf{TV}} \tag{1}$$

$$\boldsymbol{P}_{i} = \boldsymbol{e}^{(\mathsf{MU}_{i} + \eta_{\boldsymbol{P}_{i}})} \tag{2}$$

 P_{TV} is the typical value of parameter *P*, MU_i is a function of P_{TV} *P_i* is the value of parameter *P* for the *i*th individual, and

 $\eta_{P_i} \sim N(0, \omega_P^2)$ is a realization of a normally distributed random variable with zero mean and variance ω_P^2 . Interindividual variability was reported as approximate percent coefficient of variation, and significant correlations between random effects (identified in the base model using nonparametric estimation) were retained in the subsequent models.

Plasma and urine concentrations of dapagliflozin and D3OG were analyzed simultaneously, but four separate residual error models were developed to describe the differences between the individual predictions and the observed dapagliflozin and D3OG concentrations in plasma or urine. The correlation in residual errors for dapagliflozin and D3OG, when measured in either plasma or urine at the same time, was estimated (i.e., replicate residual error).²² As an example, the residual error model for urine dapagliflozin is shown (Eqs. 3 and 4).

$$W_{dapa_{U}} = \sqrt{\sqrt{\hat{Y}}_{ij,dapa_{U}} \cdot \theta_{prop,dapa_{U}} + \left(\theta_{add,dapa_{U}}\right)^{2}}$$
(3)

$$Y_{ij,dapa_{ij}} = \widehat{Y}_{i,j,dapa_{ij}} + W_{dapa_{ij}} \cdot \varepsilon_{dapa_{ij}} \cdot \sqrt{1 - \theta_{replicate_{ij}}} + \sqrt{\theta_{replicate_{ij}}} \cdot \varepsilon_{replicate_{ij}} (4)$$

 $Y_{ij,dapa_{ij}}$, the *j*th dapagliflozin observation in urine in the *i*th individual, was described by the corresponding model prediction ($\hat{Y}_{i,j,dapa_{ij}}$), the standard deviation of dapagliflozin in urine ($W_{dapa_{ij}}$), $\theta_{prop,dapa_{ij}}$, $\theta_{add,dapa_{ij}}$, and the proportion of residual error common to both dapagliflozin and D3OG in urine ($\theta_{replicate_{ij}}$) $\cdot \varepsilon_{dapa_{ij}} \sim N(0,1)$ and $\varepsilon_{replicate_{ij}} \sim N(0,1)$ are residual random variables that are normally distributed with a mean of 0 and variance of 1 (fixed). $W_{dapa_{ij}}$ is parameterized in terms of proportional ($\theta_{prop,dapa_{ij}}$) and additive ($\theta_{add,dapa_{ij}}$) components.

The first-order condition estimation method with η - ϵ interaction in NONMEM (version 7.1.2) was used to obtain the parameter estimates. Standard errors were generated using Monte Carlo importance sampling EM assisted by mode a posteriori estimation. Xpose (version 4.3.2) (GNU Lesser Public License) was used to process the NONMEM output.

Base model

The absorption of dapagliflozin was described by a transit compartment model.²³ k_{TRANSIT} was calculated from the estimate of the mean transit time (i.e., the time for a drug molecule to transit from the first to the last absorption site) and number of transit compartments (N + 1) (Eq. 5). The bioavailability was described using a logit model to ensure that the posterior individual estimates of bioavailability did not exceed 100% (Eq. 6).

$$k_{\text{TRANSIT}_i} = \frac{N_i + 1}{\text{MTT}_i}$$
(5)

$$\mathsf{BIO}_{i} = \frac{e^{\log\left(\frac{\mathsf{TVBIO}}{1-\mathsf{TVBIO}}\right)+\eta_{\mathsf{BIO},i}}}{1+e^{\log\left(\frac{\mathsf{TVBIO}}{1-\mathsf{TVBIO}}\right)+\eta_{\mathsf{BIO},i}}} \tag{6}$$

The availability of plasma and urine dapagliflozin and D3OG supported three identifiable elimination pathways of dapagliflozin: (1) CLP_{renal} , (2) CLP_{M15} , and (3) CLP_{other} (Eq. 7). CLP_{renal} and CLM were proportional to the CL_{cr} and reported

as $[L/h]/[mL/min[CL_{cr}]]$. Metabolic clearances (i.e., CLP_{M15} and CLP_{other}) were scaled by the individual baseline body weight (WT_j) using an allometric model, (wT_i/70 kg), to account for the size of metabolizing organ(s). The FE and FM_{M15} were obtained from Eqs. 8 and 9.

$$CLP_{i} = CLP_{renal,i} + CLP_{M15,i} + CLP_{other,i}$$
(7)

$$\mathsf{FE} = \frac{\mathsf{CLP}_{\mathsf{renal},i}}{\mathsf{CLP}_i} \tag{8}$$

$$\mathsf{FE}_{\mathsf{M15},i} = \frac{\mathsf{CLP}_{\mathsf{M15},i}}{\mathsf{CLP}_i} \tag{9}$$

Covariate model building

Covariate relationships with structural parameters were evaluated visually and statistically using empirical Bayes estimates vs. covariates and stepwise general additive modeling. Only the covariate relationships significant in the stepwise general additive modeling were included in the SCM. The covariate–PK relationships that were evaluated are listed in **Table 1**. The linearization SCM tool was used for forward and backward selection.²⁴ In this method, the individual predictions were linearized with respect to the ε values (random effects). To implement the linearized SCM method, categorical covariates (e.g., Child-Pugh classification) were dichotomized (**Table 2**).

The renal contribution to the metabolism of dapagliflozin to D3OG ($FM_{M15 renal}$) is calculated using Eqs. 10–13.

The relative clearance for dapagliflozin to D3OG (Δ CLP_{M15}) for changes in baseline CL_{cr} was calculated using the relationship between CL_{cr} and CLP_{M15} as estimated in the covariate model:

$$\Delta CLP_{M15} = 1 + \theta_{CL_{cr}} CLP_{M15} \cdot (CL_{cr,i} - CL_{cr,mean})$$
(10)

.....

where $\theta_{\text{CL}_{cr}}$ CLP_{M15} is the proportional change in CLP_{M15}, CL_{cr,i} is the individual baseline CL_{cr}, and CL_{cr,mean} is the mean baseline CL_{cr}. The nonrenal contribution to Δ CLP_{M15} (Δ CLP_{M15,nonrenal}) was calculated using the estimate of $\theta_{\text{CL}_{cr}}$ CLP_{M15} from the final model (see Table 1) and the mean baseline CL_{cr} of 80.14 ml/min for the two clinical pharmacology studies based on the assumption that the kidneys do not contribute to the metabolism at zero level of CL_{cr}:

$$\Delta CLP_{\text{M15,nonrenal}} = 1 + \theta_{\text{CL}_{\text{cr}}} CLP_{\text{M15}} \cdot \left(0 - CL_{\text{cr,mean}}\right) = 0.598 \text{ (11)}$$

The renal component of ΔCLP_{M15} ($\Delta CLP_{M15,renal}$) was the difference between the ΔCLP_{M15} for the range of baseline $CL_{cr,i}$ values (Eq. 10) and the nonrenal effect (Eq. 11):

$$\Delta CLP_{M15,renal} = \Delta CLP_{M15} - \Delta CLP_{M15,nonrenal} = \Delta CLP_{M15} - 0.598$$
(12)

The contribution of the kidney to dapagliflozin metabolism to D3OG (FM_{M15,renal}) was the ratio of the renal component of Δ CLP_{M15} to the relative clearance Δ CLP_{M15}:

$$FM_{M15,renal} = \frac{\Delta CLP_{M15,renal}}{\Delta CLP_{M15}}$$
(13)

Model selection and evaluation

Improvements in goodness-of-fit were assessed by comparing the changes in the NONMEM objective function values (OFVs), improvements in diagnostic plots, reductions in interindividual variability of structural model parameters and residual error, and acceptable predictive performance.

The simulation properties of the models were evaluated by visual predictive checks, a graphical assessment of the agreement between simulations from the model and the observations. To confirm that the model could predict plasma and urine dapagliflozin and D3OG concentrations over the range of renal functions in the data, CL_{cr} was used as the independent variable for visual predictive checks. To account for the different dose and covariate effects of dapagliflozin, the prediction and variance corrected visual predictive checks were used.²⁵

The effects of impaired renal function on the steady-state exposure of dapagliflozin and D3OG were realized by calculating the AUC_{ss} using Eqs. 14 and 15. The actual baseline body weight, baseline CL_{cr} , and sex of the subjects in the phase I PK and III PK/PD studies were used in the simulations. In the simulations, a daily dose of dapagliflozin 10 mg was administered to all subjects.

$$Dapagliflozin AUC = \frac{Dose \cdot BIO_i}{CLP_i}$$
(14)

Dapagliflozin 3 – O – glucuronide AUC =
$$\frac{FM_{M15,i} \cdot Dose \cdot BIO_i}{CLM_i}$$
 (15)

Acknowledgments. This analysis was sponsored by Bristol-Myers Squibb Company. We thank Prema Gopalakrishnan for preparing the analysis data sets. Editorial support was provided by Shelley Narula of inScience Communications, Springer Healthcare and was funded by Bristol-Myers Squibb Company (BMS).

Author contributions. J.-S.V. and Y.H. wrote the manuscript; M.O.K., J.-S.V., L.Z., M.P., and D.W.B. designed the research; M.O.K., J.-S.V., and Y.H. analyzed data; and L.Z. contributed New Reagents/Analytical Tools.

Conflict of Interest. Y.H., L.Z., M.P., and D.W.B. were stockholders and/or employees of BMS when these analyses were performed. BMS commissioned the analysis from Uppsala University. J.-S.V. and M.O.K. performed this work as employees of Uppsala University.

WHAT IS THE CURRENT KNOWLEDGE ON THE **TOPIC?**

Dapagliflozin is a first-in-class, orally active inhibitor of the human renal sodium-glucose co-transporter 2 and is in development for the treatment of T2DM. Until now, no article has been published reporting the population PK of dapagliflozin and its major metabolite D3OG.

WHAT QUESTION THIS STUDY ADDRESSED?

This study evaluated the effects of renal or HI on the PK of dapagliflozin and D3OG and also identified that both renal and HI decreased CLP_{M15} and CLM.

WHAT THIS STUDY ADDS TO OUR KNOWLEDGE

The kidneys contributed 40–55% of dapagliflozin metabolism in subjects with normal renal function. This article is the first analysis using a semi-mechanistic model to determine the renal and hepatic contributions on dapagliflozin metabolism and presents a useful approach to evaluate the impact of renal and hepatic function on drug PK.

HOW THIS MIGHT CHANGE CLINICAL PHARMACOLOGY AND THERAPEUTICS

- The study results may promote model-based analyses and the consideration of both the excretory and metabolic capacity of the kidney when investigating the effects of renal impairment on pharmacokinetics.
- Wright, E.M. Renal Na(+)-glucose cotransporters. Am. J. Physiol. Renal Physiol. 280, 1. F10-F18 (2001).
- Komoroski, B., Vachharajani, N., Feng, Y., Li, L., Kornhauser, D. & Pfister, M. Dapagliflozin, a novel, selective SGLT2 inhibitor, improved glycemic control over 2 weeks in patients with type 2 diabetes mellitus. Clin. Pharmacol. Ther. 85, 513-519 (2009).
- 3. Boulton, D.W. et al. Simultaneous oral therapeutic and intravenous 14C-microdoses to determine the absolute oral bioavailability of saxagliptin and dapagliflozin. Br. J. Clin. Pharmacol. 75, 763-768 (2013).
- Kasichayanula, S. et al. Disposition and Mass Balance of [14C]-Dapagliflozin After 4. Single Oral Doses in Healthy Male Volunteers. (American Association of Pharmaceutical Scientists, Atlanta, GA, 2008).

- Molitch, M.E., DeFronzo, R.A., Franz, M.J., Keane, W.F., Mogensen, C.E. & Parving, 5 H.H.; American Diabetes Association. Diabetic nephropathy. Diabetes Care 26 (suppl. 1), \$94-\$98 (2003)
- Fraser, A., Harris, R., Sattar, N., Ebrahim, S., Davey Smith, G. & Lawlor, D.A. Alanine 6. aminotransferase, gamma-glutamyltransferase, and incident diabetes: the British Women's Heart and Health Study and meta-analysis. Diabetes Care 32, 741-750 (2009).
- Kasichayanula, S. et al. Influence of renal function on dapagliflozin pharmacodynamics 7. in patients with type 2 diabetes mellitus [abstr. TH-PO525]. In American Society of Nephrology (Philadelphia, PA, 2011).
- Kasichayanula, S. et al. Influence of renal function on dapagliflozin pharmacokinetics 8. in healthy subjects and in patients with type 2 diabetes mellitus [abstr. TH-PO526]. In American Society of Nephrology. (Philadelphia, PA, 2011).
- Kasichayanula, S., Liu, X., Zhang, W., Pfister, M., LaCreta, F.P. & Boulton, D.W. Influence of hepatic impairment on the pharmacokinetics and safety profile of dapagliflozin: an openlabel, parallel-group, single-dose study, Clin. Ther. 33, 1798-1808 (2011).
- National Kidney Foundation. K/DOQI clinical practice guidelines for bone metabolism and 10 disease in chronic kidney disease. Am. J. Kidney Dis. 42, S1-201 (2003).
- 11. Kasichayanula, S. et al. The influence of kidney function on dapagliflozin exposure, metabolism, and efficacy in healthy subjects and in patients with type 2 diabetes mellitus. Br. J. Clin. Pharmacol. (2012); e-pub ahead of print (doi:10.1111/bcp.12056).
- 12. Betrosian, A.P., Agarwal, B. & Douzinas, E.E. Acute renal dysfunction in liver diseases. World J. Gastroenterol. 13, 5552-5559 (2007).
- Nishimura, M. & Naito, S. Tissue-specific mRNA expression profiles of human phase I 13 metabolizing enzymes except for cytochrome P450 and phase II metabolizing enzymes. Drug Metab. Pharmacokinet. 21, 357-374 (2006).
- 14. Nolin, T.D. Altered nonrenal drug clearance in ESRD. Curr. Opin. Nephrol. Hypertens. 17, 555-559 (2008).
- Durkalski, V. & Berger, V.W. Categorizing data. In Encyclopedia of Statistics in 15. Behavioral Science (John Wiley & Sons, Ltd., 2005); published online 15 October 2005 (doi:10.1002/0470013192.bsa077).
- 16. Hinderling, P.H. & Shi, J. Absolute bioavailability estimated from oral data. J. Pharm. Sci. 84, 385-386 (1995).
- 17 Hinderling, P.H. Evaluation of a novel method to estimate absolute bioavailability of drugs from oral data. Biopharm. Drug Dispos. 24, 1-16 (2003).
- Hamrén, B., Ericsson, H., Samuelsson, O. & Karlsson, M.O. Mechanistic modelling of 18 tesaglitazar pharmacokinetic data in subjects with various degrees of renal function evidence of interconversion. Br. J. Clin. Pharmacol. 65, 855-863 (2008).
- 19. Hinderling, P.H. Biased estimates of nonrenal clearance. J. Pharm. Sci. 90, 960-966 (2001).
- 20. Kohan, D.E. et al. Efficacy and safety of dapagliflozin in patients with type 2 diabetes and moderate renal impairment [abstr. TH-PO524]. In American Society of Nephrology (Philadelphia, PA, 2011).
- 21. Zhang, L., Feng, Y., List, J., Kasichayanula, S. & Pfister, M. Dapagliflozin treatment in patients with different stages of type 2 diabetes mellitus: effects on glycaemic control and body weight. Diabetes. Obes. Metab. 12, 510-516 (2010).
- 22. Karlsson, M.O., Beal, S.L. & Sheiner, L.B. Three new residual error models for population PK/PD analyses. J. Pharmacokinet. Biopharm. 23, 651-672 (1995).
- Savic, R.M., Jonker, D.M., Kerbusch, T. & Karlsson, M.O. Implementation of a transit 23. compartment model for describing drug absorption in pharmacokinetic studies. J. Pharmacokinet. Pharmacodyn. 34, 711-726 (2007).
- Khandelwal, A., Harling, K., Jonsson, E.N., Hooker, A.C. & Karlsson, M.O. A fast method 24. for testing covariates in population PK/PD Models. AAPS J. 13, 464-472 (2011).
- 25. Bergstrand, M., Hooker, A.C., Wallin, J.E. & Karlsson, M.O. Prediction-corrected visual predictive checks for diagnosing nonlinear mixed-effects models. AAPS J. 13, 143-151 (2011).



CPT: Pharmacometrics & Systems Pharmacology is an open-access journal published by Nature Publishing Group. This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivative Works 3.0 License. To view a copy of this license, visit http://creativecommons.org/licenses/by-nc-nd/3.0/

Supplementary information accompanies this paper on the Pharmacometrics & Systems Pharmacology website (http://www.nature.com/psp)