

## Research Article

# Inhibitory Effect of Arctigenin from Fructus Arctii Extract on Melanin Synthesis via Repression of Tyrosinase Expression

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To identify the active compound arctigenin in Fructus Arctii (dried seed of medicinal plant *Arctium lappa*) and to elucidate the inhibitory mechanism in melanogenesis, we analyzed melanin content and tyrosinase activity on B16BL6 murine melanoma and melan-A cell cultures. Water extracts of Fructus Arctii were shown to inhibit tyrosinase activity in vitro and melanin content in  $\alpha$ -melanocyte stimulating hormone-stimulated cells to similar levels as the well-known kojic acid and arbutin, respectively. The active compound arctigenin of Fructus Arctii displayed little or no cytotoxicity at all concentrations examined and decreased the relative melanin content and tyrosinase activity in a dose-dependent manner. Melanogenic inhibitory activity was also identified in vivo with zebrafish embryo. To determine the mechanism of inhibition, the effects of arctigenin on tyrosinase gene expression and tyrosinase promoter activity were examined. Also in addition, in the signaling cascade, arctigenin dose dependently decreased the cAMP level and promoted the phosphorylation of extracellular signal-regulated kinase. This result suggests that arctigenin downregulates cAMP and the tyrosinase enzyme through its gene promoter and subsequently upregulates extracellular signal-regulated kinase activity by increasing phosphorylation in the melanogenesis signaling pathway, which leads to a lower melanin content.

## 1. Introduction

Researches on natural products, including traditional medicine and herbs, have been increasing recently due to an increase in demand for complementary and alternative medicines with less side effect and increased safety [1]. In dermatological research, many studies have been performed to determine the mechanisms behind the regulation of melanogenesis and to identify clinically useful hypopigmenting agents [2, 3]. Agents from medicinal herbs and natural resources have been reported in many previous studies [4, 5]. In addition to dermatological research, many other fields have shown increased interest in identifying agents from natural resources, including

cosmetics, functional foods, beverages, and feed additives in livestock [6–9]. In the cosmetic field, many studies have attempted to identify medicinal herbs and traditional medicines that can inhibit melanin and tyrosinase activity, which could be used as skin-whitening agent [10]. Many types of natural resources and products including various plants (whole plant or part) and extracts of various solvents have been screened with this goal in mind [11, 12]. In regard to skin, great efforts to develop materials that inhibit melanin biosynthesis and/or tyrosinase have been made for the development of skin-whitening agents [13].

Melanin exists elsewhere in nature and is found in almost all of the living organisms, including humans, where the high melanin content is found in skin, hair, and eyes [14].

It is now well known as a key determinant of color and tone of each tissue, which is dependent on the amount and distribution of melanin. Melanin is synthesized through a complex biochemical process, that is, melanogenesis, which typically occurs in melanosomes of epidermal melanocyte [15]. In this process, tyrosinase converts tyrosine to 3,4-dihydroxyphenylalanine (DOPA) through hydroxylation, which is then oxidized into dopaquinone (4-(2-carboxy-2-aminoethyl)-1,2-benzoquinone) and indole-5,6-quinone. This compound then leads to the synthesis of melanin. Tyrosinase functions as a hydroxylase and oxidase and can convert tyrosine into DOPA and can convert DOPA into dopaquinone [16, 17]. Melanogenesis is stimulated by UV irradiation and  $\alpha$ -melanocyte stimulating hormone ( $\alpha$ -MSH), which increase tyrosinase expression and melanin content by binding to melanocortin 1 receptor (MC1R). Activated MC1R and chemical compound forskolin and isobutylmethylxanthine (IBMX) also stimulate melanogenesis by activating cAMP in the melanocyte-specific pathway, and cAMP can in turn increase melanogenic signaling enzymes through protein kinase A (PKA), which induces microphthalmia-associated transcription factor (MITF) expression [18–20].

Among medicinal herbs, Fructus Arctii (FA) has been traditionally used for the treatment of inflammatory sore throats, urticaria, and furuncle. Arctiin and arctigenin (ATG) in *Arctium lappa* (AL) have been shown to inhibit leukemic B-cell hybridoma proliferation, human keratinocyte cell growth, and allergies, improve aging skin, and have ameliorative effects on experimental glomerulonephritis [21–25]. Also, these compounds have been reported to be novel cytotoxic and cancer chemopreventive agents in the medicinal herb *bardanae fructus* and *Saussurea medusa* [26, 27]. Arctiin and ATG were identified as the major compounds in *Centaurea sphaerocephala* ssp. *Polyacantha* and were also found in the AL used in this study [28, 29].

In this study, we investigated the inhibitory effect of FA extract (FAE) and ATG on tyrosinase and melanin synthesis and identified melanogenic inhibitory effect in both cell lines of B16BL6 and melan-A and also in zebrafish embryo. We found that these extracts downregulate cAMP, tyrosinase enzyme, and its gene promoter and promote extracellular signal-regulated kinase (ERK) phosphorylation in murine melanoma B16BL6 cell cultures to levels that were comparable with kojic acid (KA) and arbutin, which are widely used melanogenesis inhibitors.

## 2. Materials and Methods

**2.1. Chemicals and Reagents.** Arbutin,  $\alpha$ -MSH, DOPA, dimethyl sulfoxide (DMSO), dithiothreitol (DTT), KA, mushroom tyrosinase, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), Triton X-100, TRI reagent, Tween 20, and phenylthiourea (PTU) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Lipofectamine 2000 reagent was obtained from Life Technologies (Carlsbad, CA, USA), and Dual Luciferase Assay System was from Promega (Madison, WI, USA).

Antibodies against ERK, phosphorylated-ERK (p-ERK), and tyrosinase were obtained from Cell Signaling Technology (Danver, MA, USA) and Abcam (Cambridge, UK), respectively.

**2.2. Preparation of Herb Extract.** Medicinal herb was obtained from local vendors in Daejeon, Korea. FA (50 g) was immersed in 1 L of water for 1 h at room temperature to enhance the extract yield and boiled for 3 h. The extract was then filtered through 106  $\mu$ m test sieve (Retsch, Germany), freeze dried, and stored at  $-20^{\circ}\text{C}$  until used. In the experiments, the extract was dissolved in Dulbecco's phosphate buffered saline (DPBS) at a concentration of 20 mg/mL, centrifuged (13,000 rpm, 10 min,  $4^{\circ}\text{C}$ ), and the acquired supernatant was filtered through 0.2  $\mu$ m syringe filter.

**2.3. Cell Culture and Viability Assay.** Murine melanoma cell line B16BL6 and non-neoplastic cell line melan-A were maintained in Dulbecco's Modified Eagle Medium (DMEM, Lonza, USA) and RPMI 1640, respectively, supplemented with 10% (v/v) fetal bovine serum and 1x penicillin-streptomycin mixture (Gibco, USA) in a humidified 5%  $\text{CO}_2$  incubator at  $37^{\circ}\text{C}$ . In the cell viability assay, cells were seeded on 96-well plate in 100  $\mu$ L of culture media ( $1.5 \times 10^4$  cells/well). The cells were further incubated for 24 h with or without ATG. The viability of cultured cells was determined by the reduction of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide to formazan, as described previously with slight modifications [30]. In brief, 10  $\mu$ L of the MTT solution (5 mg/mL) was added to each well and incubated for 1 h at  $37^{\circ}\text{C}$ . Following incubation, the medium was removed, cells were washed with DPBS, and 100  $\mu$ L of DMSO was added to dissolve formazan precipitated by reduced MTT. The absorbance at 570 nm of the negative control (without inhibitor) was measured and the percentage of inhibition was calculated as follows: inhibition (%) =  $[\text{OD } 570 \text{ nm (ATG)}/\text{OD } 570 \text{ nm (control)}] \times 100$ .

**2.4. In Vitro Tyrosinase Inhibition Assay.** Tyrosinase inhibition was determined spectrophotometrically as described previously, with minor modifications [31]. To examine the effect of FAE on tyrosinase activity in vitro, 50  $\mu$ L of DOPA (20 mM), KA (60  $\mu\text{g}/\text{mL}$ ), FAE, and 100  $\mu$ L of tyrosinase (250 U/mL) were mixed together in 96-well plate and incubated at  $37^{\circ}\text{C}$  for 10 min. The absorbance was then measured using a microplate reader at 475 nm. DOPA, KA, and tyrosinase were dissolved in 0.1 M potassium phosphate buffer (pH 6.6) at the concentrations indicated above. Concentrations and volumes of each reagent were determined based on preliminary optimization experiments performed in our laboratory (data not shown). For control, cells were treated with DOPA, potassium phosphate buffer and tyrosinase, with no tyrosinase inhibitor. All the experiments were carried in triplicate.

**2.5. Measurement of Melanin Contents.** The melanin content was measured as described previously [32]. B16BL6 murine

melanoma cells and melan-A cells (each  $5 \times 10^4$  cells/dish) were seeded on 6-well plate and treated with arbutin (positive control) or ATG. Melanogenesis stimulator  $\alpha$ -MSH (100 nM) was used to induce melanin synthesis in both cell lines. After 72 h, cells were harvested, washed with DPBS, and collected. Cell pellets were then completely lysed with 1N NaOH at 60°C for 1 h, and the absorbance at 405 nm of 200  $\mu$ L of the lysates was measured using microplate reader. The protein concentrations and melanin contents for each treatment were determined using the BCA protein assay kit (Thermo Scientific, USA) and were compared with the negative control (without arbutin and inhibitor).

**2.6. Cellular Tyrosinase Activity Assay.** Tyrosinase activity was measured as described previously, with minor modifications [33]. In brief, cells were seeded in 60 mm culture dish at a density of  $1 \times 10^5$  cells/dish and incubated for 24 h. After serum starvation for 24 h, the cells were further incubated for 72 h with or without ATG. Cells were harvested, washed in DPBS, and lysed with lysis buffer (1% Triton X-100 in 0.1M sodium phosphate buffer pH 6.8, containing protease inhibitor) and subjected to a freeze-thaw cycle (−80°C 30 min/25°C 25 min/37°C 5 min). After centrifugation (13,000 rpm, 10 min), protein concentrations were determined using the BCA protein assay kit and the concentrations were adjusted to 200  $\mu$ g/mL. Cell lysate supernatants (each 50  $\mu$ L) were transferred to 96-well plate and mixed with 100  $\mu$ L of DOPA (20 mM) and 50  $\mu$ L of DPBS. After incubation at 37°C overnight, the absorbance at 475 nm was measured using a microplate reader. Cellular tyrosinase activities were determined as the ratio of tyrosinase content over total protein content (ng/ $\mu$ g).

**2.7. RNA Isolation and Quantitative RT-PCR.** B16BL6 cells were treated with ATG for 1 h and further incubated for 24 h with  $\alpha$ -MSH treatment. Total RNA was isolated using TRI reagent and 1  $\mu$ g of each samples was reverse-transcribed using AccuPower RT PreMix (Bioneer, Korea) according to the manufacturer's instructions, and aliquots of cDNA were subjected to real-time quantitative polymerase chain reaction (RT-qPCR) with AccuPower 2X Greenstar qPCR Master Mix (Bioneer, Korea). A forward primer sequence of 5'-GGCCAGCTTTCAGGCAGAGGT-3' and reverse primer sequence of 5'-TGGTGCTTCATGGGCAAATC-3' were used for tyrosinase exon 1. Thermal cycling of PCR amplification was carried out as follows: initial denaturation at 95°C for 10 min, followed by 40 cycles of denaturation at 95°C for 10 s, annealing at 60°C for 30 s, and elongation at 72°C for 30 s using Eco Real-Time PCR System (Illumina, USA). For  $\beta$ -actin, forward primer 5'-TGTCCACCTTCCAGCAGATGT-3' and reverse primer 5'-AGCTCAGTAACAGTCCGCCTAGA-3' were used in the same thermal cycling conditions as described for the tyrosinase PCR experiments.

**2.8. Construction of Mouse Tyrosinase Promoter Reporter.** Upstream region, including the tyrosinase promoter, was amplified and isolated from genomic DNA by PCR

using the following primers: forward 5'-GGTACCTTCAACCCTTTTCTATGTCC-3' (−2267 to −2242) and reverse 5'-CTCGAGCACAGACTTCTTTTCCAGCAAC-3' (−9 to +19). Genomic DNA information was obtained from <http://genome.ucsc.edu>. The underlined sequences of each primer were not present in the genomic DNA and were artificially attached to confer restriction enzyme recognition sites for DNA cloning. The PCR product (2,286 bp) was then introduced into the T-Blunt vector system. The inserted PCR product was digested with restriction enzyme *Xho* I and *Kpn* I and then subsequently cloned into the same enzyme site of the pGL3-basic vector, which contained a luciferase coding sequence in its upstream region. Cloned DNA was identified by sequencing and enzyme digestion followed by sizing on the agarose gel electrophoresis.

**2.9. Transfection and Luciferase Assay.** For the analysis of CRE (cAMP response element) promoter activity, murine melanoma B16BL6 cells in 24-well dishes were transfected with 0.2  $\mu$ g of CRE-Luc vector and 0.05  $\mu$ g of Rlu, using 2  $\mu$ g of lipofectamine 2000 reagent in a total reaction volume of 200  $\mu$ L. After 24 h, the transfection medium was changed and the cells were treated with ATG for 1 h and further incubated with  $\alpha$ -MSH for 24 h at 37°C. Cells were then washed with DPBS and lysed with Triton X-100 (1% in 25 mM Tris-phosphate buffer, pH 7.8) containing ethylenediaminetetraacetic acid (EDTA, 2 mM) and DTT (2 mM). Luciferase activity was determined using a Dual-Luciferase Assay system. All reagents were prepared as described in the manufacturer's instruction, and the data are represented as the ratio of firefly to Renilla luciferase activity (Fluc/Rluc).

**2.10. Western Blot Analysis.** Cells were treated with ATG or arbutin for 1 h, further incubated for 24 h at 37°C with  $\alpha$ -MSH, and lysed with Triton X-100 (1% in 50 mM sodium phosphate buffer pH 6.8, containing protease inhibitor). The proteins in the cell lysate were separated by 10% sodium dodecyl sulfate- (SDS-) polyacrylamide gel electrophoresis and blotted onto a nitrocellulose membrane. The blotted membrane was incubated for 1 h with 5% nonfat milk in Tris-buffered saline (pH 7.6) containing 0.05% Tween 20. Subsequently, the membrane was incubated overnight with the anti-goat tyrosinase antibody (1:1000), and further incubated for 1 h with the anti-goat antibody conjugated with horseradish peroxidase (1:10,000). For ERK, the membrane was incubated overnight with ERK and p-ERK antibody (1:1000), and then incubated for 1 h with the anti-rabbit antibody conjugated with horseradish peroxidase (1:10,000). The  $\beta$ -actin antibody (1:1,000) was used for normalization. The signals were visualized using the enhanced chemiluminescence reagent (ECL western blotting substrate, Thermo Scientific, USA).

**2.11. Observation of Depigmenting Effect in Zebrafish.** Inhibitory effect of ATG on melanin pigmentation was observed in vivo using zebrafish embryo following the previous report with slight modification [34]. Zebrafish embryos were obtained from natural spawning of wild type

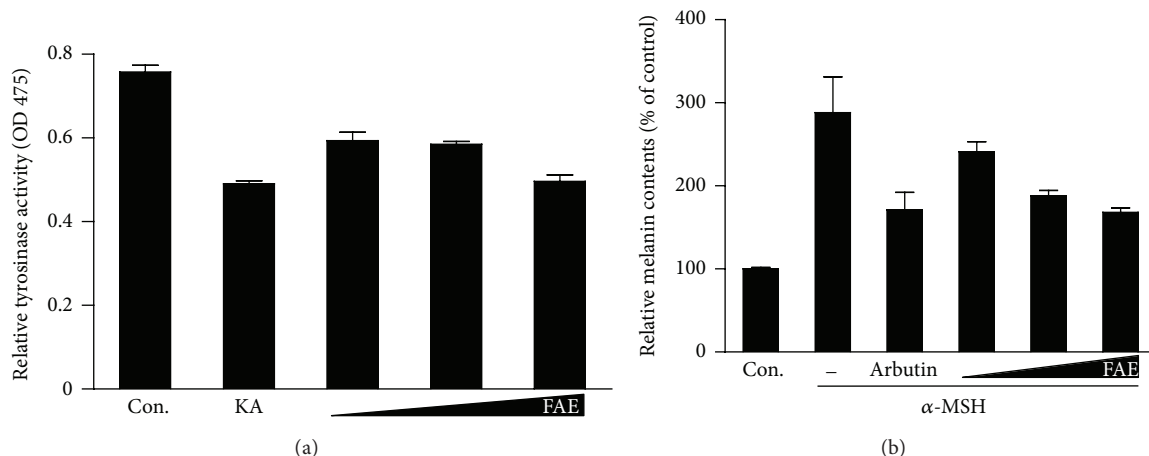


FIGURE 1: The inhibitory effect of FAE on tyrosinase activity in vitro (a) and melanin content in murine B16BL6 cells stimulated with 100 nM  $\alpha$ -MSH (b). Cells were treated with KA (60  $\mu$ g/mL) or arbutin (100  $\mu$ g/mL) and FAE at different doses (1, 100, 500  $\mu$ g/mL). In vitro tyrosinase activities were measured based on the absorbance at 475 nm, and melanin contents are depicted as a percentage (%) of control. Each value was expressed with the mean  $\pm$  SD of three independent experiments.

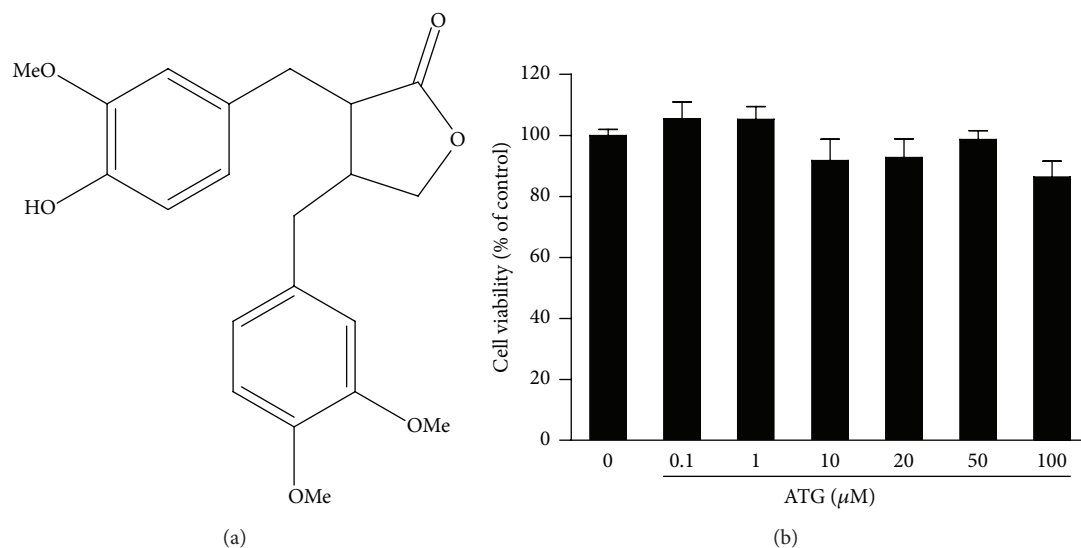


FIGURE 2: Chemical structure (a) and cytotoxicity of ATG at different treatment doses (b). Values of cytotoxicity were represented as a percentage (%) of control, with mean  $\pm$  SD of three independent experiments.

AB\*. The embryos were raised in standard fish water at 28.5°C. ATGs were dissolved in DMSO and added directly in fish water at the final concentration of 1, 10, and 100  $\mu$ M. As control, DMSO or PTU (well-known melanogenic inhibitor) treated embryos were analyzed together with the embryos of ATG treated. Live embryos at 40 hpf (hour post-fertilization) were classified and taken pictures.

### 3. Results and Discussion

**3.1. Inhibitory Effect of FAE on In Vitro Tyrosinase Activity and  $\alpha$ -MSH-Mediated Melanogenesis in B16BL6 Cells.** *Arctium lappa* (common name Burdock) has been used as a therapeutic herb in Asia, Europe, and North and South America

for hundreds of years. It has also long been cultivated and used as a dietary and nutritional food, especially in Asia [35]. Here, we investigated the effect of FAE on tyrosinase activity and melanin contents using an in vitro screening system. These results were also compared with positive controls, KA, and arbutin. FAE was shown to inhibit tyrosinase activity in a dose-dependent manner (Figure 1(a)). We also examined the effect of FAE on cellular melanin synthesis (Figure 1(b)). Consistent with tyrosinase inhibition, melanin synthesis was also significantly inhibited by FAE in a dose-dependent manner. These results indicate that FAE has a significant inhibitory effect against not only tyrosinase activity but also cellular melanin synthesis, showing a similar activity as the positive control KA and arbutin. This suggests that FAE might

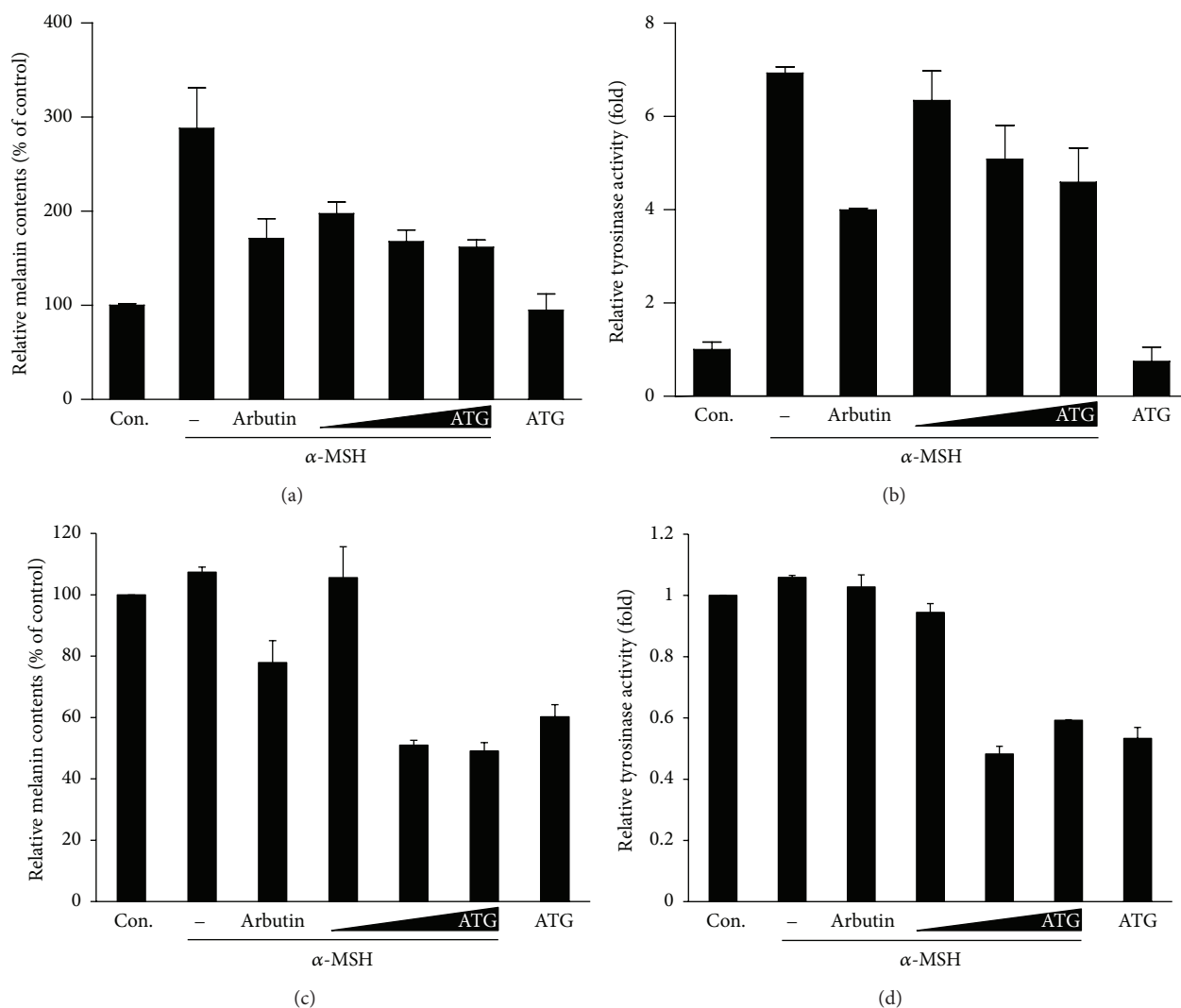


FIGURE 3: Effect of ATG (1, 10, and 50  $\mu$ M) on  $\alpha$ -MSH (100 nM) stimulated murine B16BL6 melanoma cells ((a) and (b)) and melan-A cells ((c) and (d)). Data are represented as relative melanin contents and tyrosinase activity compared with arbutin (100  $\mu$ g/mL). ATG concentration without  $\alpha$ -MSH was 10  $\mu$ M. All values were represented from three independent experiments with mean  $\pm$  SD values.

be a useful natural compound for inhibition of tyrosinase activity and melanin synthesis.

**3.2. Effect of ATG on the Cytotoxicity of B16BL6 Cells.** Arctiin and ATG (Figure 2(a)), the main compounds of AL, have been reported to contain a variety of biological activities [36]. Among those compounds, we selected ATG because it has been shown to have beneficial biopharmaceutical properties and activities in the treatment of many diseases [37, 38]. However, the potential of using ATG for dermatological applications has not been examined until now, except one study that examined the photoprotective effect of fermented FA in terms of its effects on decreasing the expression of matrix metalloproteinase 1 (MMP-1) mRNA [39]. To investigate whether the ATG activity is related to the inhibitory effect on tyrosinase and melanin synthesis, we analyzed cellular tyrosinase activity and melanin contents in vitro. The cytotoxicity of ATG was also examined and we found that

there was no significant cytotoxicity at concentrations up to 100  $\mu$ M (Figure 2(b)). This is consistent with other studies, which reported that ATG did not show any cytotoxic effect against B16BL6 cells at concentrations up to 100  $\mu$ M [40].

**3.3. Inhibitory Effect of ATG on Melanin Content and Tyrosinase Activity.** To further confirm that ATG is an inhibitory active compound, the effect of ATG on melanin content and tyrosinase activity was analyzed in B16BL6 melanoma cells and nonneoplastic melan-A cells. Cellular melanin contents and tyrosinase activities were significantly reduced by ATG treatment in a dose-dependent manner in both B16BL6 and melan-A cells, with similar inhibition pattern between two cell lines (Figures 3(a), 3(b), 3(c), and 3(d)). As shown in the result of melanin contents, ATG exhibited inhibitory activity on melanin contents similar to the positive control arbutin at the concentrations of 10 and 50  $\mu$ M in both B16BL6 and melan-A cells. The inhibitory effect of ATG on tyrosinase

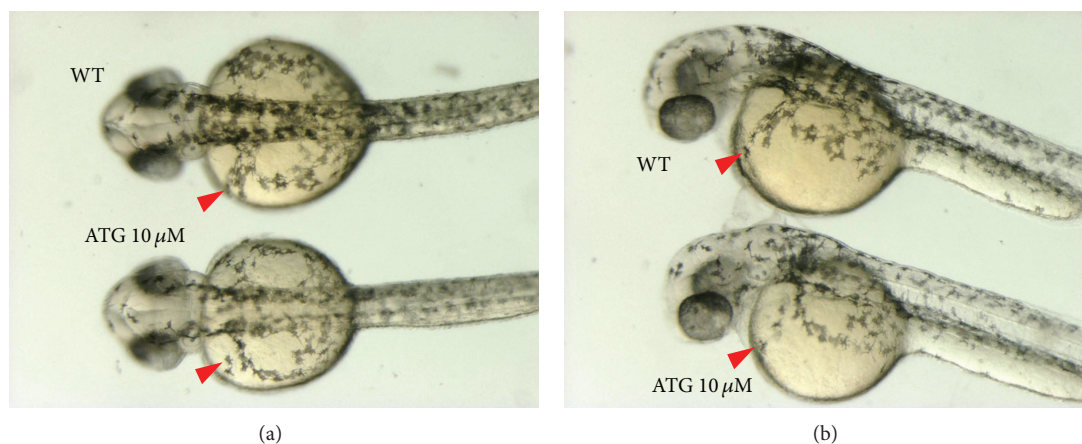


FIGURE 4: ATG treatment suppressed pigment deposition without affecting cell migration. Moderate concentration of ATG ( $10 \mu\text{M}$ ) treated embryos showed reduced embryonic pigmentation without causing any discernible pigment cells migration defect (red arrowhead). Treatment of high dose ATG ( $100 \mu\text{M}$ ) caused severe embryonic developmental retardation and growth was stalled at 18–20 somite stage (data not shown). All live embryos were treated with ATG for different dosages at 15 hpf (12–14 somite stage) followed by taken picture at 40 hpf. Pictures were shown by overview (a) and lateral view (b).

activity was also significant in both two cell lines at concentrations of 10 and  $50 \mu\text{M}$  similar with arbutin. Particularly in melan-A cells, inhibitory effect on melanin contents and tyrosinase activity was significantly stronger than that of arbutin control, at the concentrations of 10 and  $50 \mu\text{M}$ . For in vivo animal model, ATG treatment ( $10 \mu\text{M}$ ) prior to the emergence of pigmentation in 15 hpf developing zebrafish embryos showed the moderate reduction of pigment deposition (40%,  $n = 52$ , Figure 4). The embryos were normally developed without any discernible developmental retardation as well as morphological defects. However, 10-fold higher dose treatment of ATG became harmful to the embryos since the ATG treated ( $100 \mu\text{M}$ ) embryos did not undergo normal development but were instead stalled at between 18 and 20 somite stage (100%,  $n = 47$ , data not shown). Lower dose of ATG ( $1 \mu\text{M}$ ) failed to suppress the body pigmentation (100%,  $n = 54$ ). Analysis of tyrosinase activity has been widely used as a measure of melanin synthesis in cell culture systems [41]. For this reason, measuring both tyrosinase activity and melanin contents is usually conducted when evaluating the melanogenesis inhibitory effect of natural products or certain compounds. Therefore, we measured the effect of ATG on  $\alpha$ -MSH-mediated tyrosinase activity. Consistent with the results observed in the melanin content assay, ATG treatment significantly suppressed tyrosinase activity. Other natural herbs and medicines have been shown to inhibit tyrosinase activity and melanin contents. For example, Zhong et al. screened 90 traditional Chinese herb extracts to identify extracts displaying depigmentation activity [42]. Among these, the water extract of *Galla Chinensis* and ethanol extract of *Radix Clematidis* showed the highest activity in the cell culture assay. Although they did not identify the active compound in each extracts further investigations are needed for the development of useful dermatological agents. Lin et al. isolated biochanin A from the methanol extract of *Trifolium pretense* and found that it inhibited melanogenesis and tyrosinase activity [43]. They also confirmed its activity

after dermatological application to mouse skin. In addition, 7-methoxycoumarin was isolated as an active compound for antimelanogenesis from the methanol extract of the leaves of *Eupatorium triplinerve* Vahl [44].

**3.4. Regulatory Effect of ATG on Promoter Activity and cAMP Signaling.** Since ATG inhibited cellular melanin synthesis and tyrosinase activity, we examined whether ATG could affect the expression of tyrosinase and intracellular signaling pathway. ATG significantly inhibited  $\alpha$ -MSH-induced tyrosinase mRNA expression and tyrosinase protein expression was also greatly repressed to levels greater than the arbutin control (Figures 5(a) and 5(b)). These results imply that ATG downregulates tyrosinase gene expression, which would decrease tyrosinase enzyme activity and ultimately inhibit melanogenesis. ATG may lower the tyrosinase activity by regulating posttranslational modification and degradation of tyrosinase protein in the cells. To confirm the effect of ATG on tyrosinase gene expression, we analyzed tyrosinase promoter activity using luciferase reporter assay, which encompasses the promoter region (–2267 to +19). As shown in Figure 5(c), we found that ATG reduced the promoter activity in a dose-dependent manner. This result demonstrated that this promoter region contains an important regulatory element that is involved in cAMP. In addition, it was reported that the tyrosinase mRNA level can be regulated by  $\alpha$ -MSH and cAMP in mouse melanoma cells [45, 46]. They observed increased tyrosinase activity and mRNA expression and suggested that  $\alpha$ -MSH promotes tyrosinase expression and activity by acting through cAMP.

Recently, Bertolotto et al. demonstrated that cAMP level was closely related with the transcriptional activity of tyrosinase gene [47]. They showed that an elevated cAMP level can be a strong stimulator of the tyrosinase promoter. They also reported that the M-box and E-box, which is located up- and downstream of the TATA-box ahead of the initiation site, are involved in the regulation of tyrosinase promoter

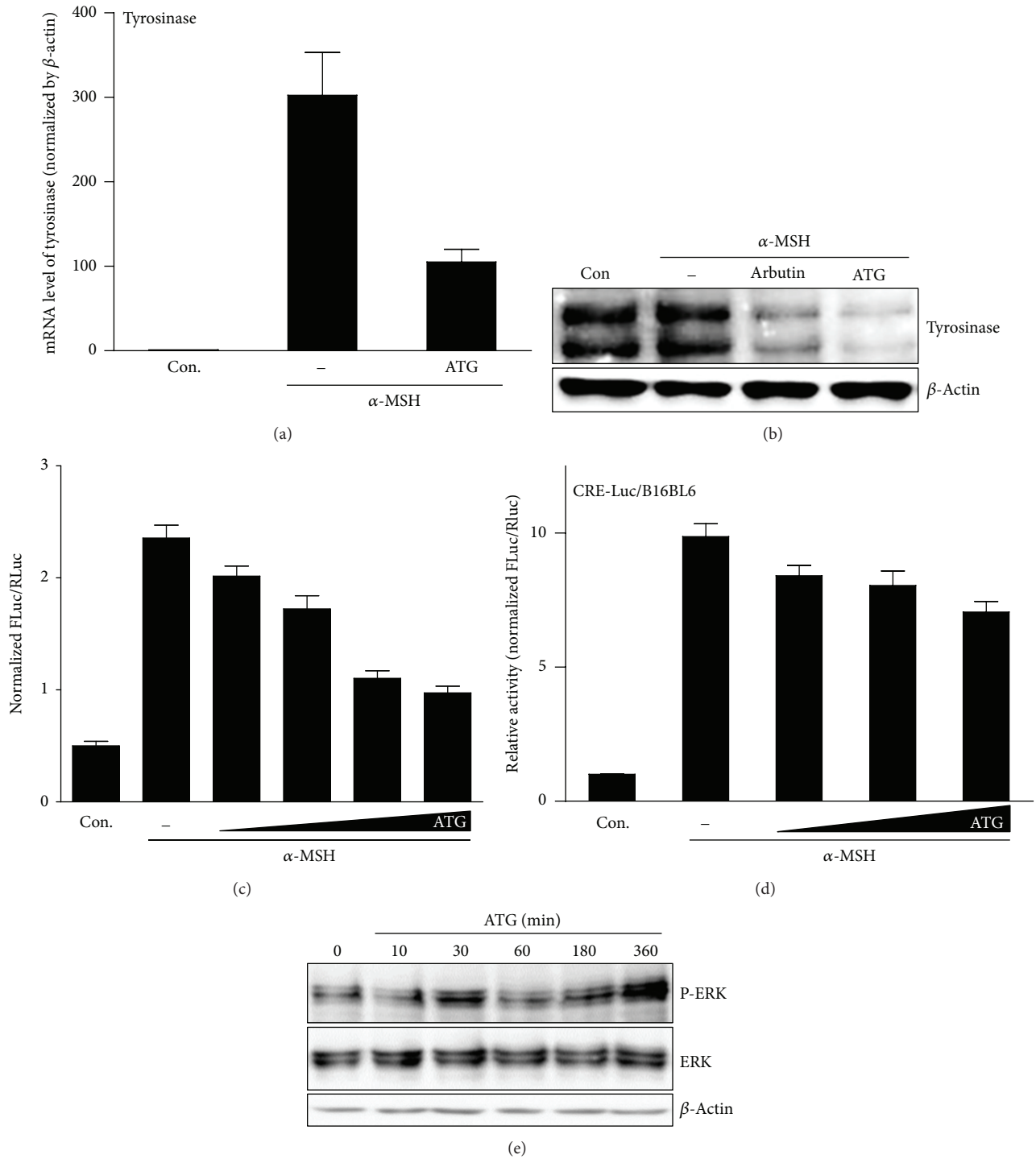


FIGURE 5: Inhibitory effect of ATG (10  $\mu$ M) on tyrosinase gene expression was analyzed by real-time qPCR (a) and western blot (b) in B16BL6 cells stimulated with  $\alpha$ -MSH. The effect of ATG (1, 10, 20, and 50  $\mu$ M) on tyrosinase promoter activity was determined using the luciferase reporter system (c). Dose-dependent inhibitory effect of ATG (1, 10, and 50  $\mu$ M) was observed in the CRE-luciferase assay (d), and the level of P-ERK increased in a time-dependent manner (e).

activity by cAMP. The tyrosinase promoter region of DNA they analyzed was very similar to the region analyzed in this study. These results imply that ATG may inhibit tyrosinase activity and melanogenesis by altering gene expression

through the formation of DNA-protein complexes and signaling pathways (especially with regard to cAMP).

Moreover, it is well known that  $\alpha$ -MSH stimulates melanin synthesis and also acts as a cAMP-elevating agent,

which indicates that cAMP plays an important role in the regulation of melanogenesis [48]. It was also shown that ERK activation (phosphorylation of ERK) can be induced by physical factors and natural compounds [49–51]. cAMP and ERK signaling are key regulatory elements in the pathways of cell proliferation and differentiation and also play an important role in melanogenesis [52]. In mammals, cAMP has an important function in pigmentation, where it was shown to increase the expression of MITF through the activation of protein kinase A, which in turn stimulates tyrosinase gene expression for melanin synthesis [53]. cAMP-elevating agents, such as  $\alpha$ -MSH, IBMX, and forskolin, also stimulate melanin synthesis. Moreover, it is known that  $\alpha$ -MSH, which is a signal transducer, potently induces MITF expression and increases melanin synthesis. cAMP-related cellular signaling and the molecular mechanisms involved in different biochemical pathways have been widely studied. cAMP increases the expression of MITF by activating PKA which phosphorylate its substrates, enzymes, and regulatory proteins, and then PKA phosphorylates cAMP response element binding protein (CREB). Tyrosinase gene expression and melanin synthesis are then stimulated by this signaling pathway, and  $\alpha$ -MSH stimulates cAMP upregulation. This then induces melanogenesis, which is closely linked to the level of tyrosinase [54, 55]. To check whether ATG has an effect on this signaling pathway mainly involving cAMP, the CRE-Luc reporter assay was performed. In this analysis, ATG was shown to reduce the cAMP level in B16BL6 cells of reporter transfected (Figure 5(d)). Consequently, we found that ATG regulates tyrosine activity and inhibits melanogenesis by inhibiting cAMP levels and even further contributes in suppressing melanin synthesis.

In a previous study, acteoside from the leaves of *Rehmannia glutinosa* was shown to inhibit melanogenesis by ERK activation and tyrosinase downregulation [56]. According to this report, we determined if ATG can act as ERK activating agent in repressing melanin and tyrosinase synthesis. As shown in the Figure 5(e), the level of p-ERK increased with ATG (10  $\mu$ m) treatment time and the effect was remarkably after 30 min. This result is in agreement with previous reports on the ERK pathway in melanogenesis [57]. ATG exerts significant inhibitory activity on tyrosinase and melanin synthesis by blocking the cAMP pathway and activating the ERK pathway. Consequently, ATG significantly inhibit tyrosinase activity, expression, and melanogenesis.

In conclusion, we first investigated the inhibitory effect of ATG on tyrosinase activity and melanin biosynthesis and elucidated the signaling mechanism. We found ATG has a significant inhibitory effect on them without producing any significant cytotoxicity. Those results indicate that ATG inhibits melanogenesis in murine melanoma B16BL6 cells by repressing cAMP and tyrosinase gene/protein expression and increasing ERK phosphorylation in the signaling pathway of melanogenesis. Likewise, inhibition of melanogenesis was also confirmed in nonneoplastic cells melan-A and in vivo zebrafish embryos. However, ATG of high concentration 100  $\mu$ M was shown to have toxicity in zebrafish embryos with developmental retardation. The results of this study imply that ATG may be used as a natural resource for the

treatment of melanogenesis, which is highly important given the current increase in demand for complementary and alternative medicines.

## Conflict of Interests

The authors declare that there is no conflict of interests.

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