

Reciprocal Regulation of NF- κ B (Relish) and Subolesin in the Tick Vector, *Ixodes scapularis*

Victoria Naranjo^{1,2,3,4}, Nieves Ayllón^{2,5}, José M. Pérez de la Lastra^{2,5}, Ruth C. Galindo^{1,2}, Katherine M. Kocan¹, Edmour F. Blouin¹, Ruchira Mitra^{1,6}, Pilar Alberdi², Margarita Villar², José de la Fuente^{1,2,*}

1 Department of Veterinary Pathobiology, Center for Veterinary Health Sciences, Oklahoma State University, Stillwater, Oklahoma, United States of America, **2** SaBio, Instituto de Investigación en Recursos Cinegéticos IREC-CSIC-UCLM-JCCM, Ciudad Real, Spain

Abstract

Background: Tick Subolesin and its ortholog in insects and vertebrates, Akirin, have been suggested to play a role in the immune response through regulation of nuclear factor-kappa B (NF- κ B)-dependent and independent gene expression via interaction with intermediate proteins that interact with NF- κ B and other regulatory proteins, bind DNA or remodel chromatin to regulate gene expression. The objective of this study was to characterize the structure and regulation of *subolesin* in *Ixodes scapularis*. *I. scapularis* is a vector of emerging pathogens such as *Borrelia burgdorferi*, *Anaplasma phagocytophilum* and *Babesia microti* that cause in humans Lyme disease, anaplasmosis and babesiosis, respectively. The genome of *I. scapularis* was recently sequenced, and this tick serves as a model organism for the study of vector-host-pathogen interactions. However, basic biological questions such as gene organization and regulation are largely unknown in ticks and other arthropod vectors.

Principal Findings: The results presented here provide evidence that *subolesin/akirin* are evolutionarily conserved at several levels (primary sequence, gene organization and function), thus supporting their crucial biological function in metazoans. These results showed that NF- κ B (Relish) is involved in the regulation of *subolesin* expression in ticks, suggesting that as in other organisms, different NF- κ B integral subunits and/or unknown interacting proteins regulate the specificity of the NF- κ B-mediated gene expression. These results suggested a regulatory network involving cross-regulation between NF- κ B (Relish) and Subolesin and Subolesin auto-regulation with possible implications in tick immune response to bacterial infection.

Significance: These results advance our understanding of gene organization and regulation in *I. scapularis* and have important implications for arthropod vectors genetics and immunology highlighting the possible role of NF- κ B and Subolesin/Akirin in vector-pathogen interactions and for designing new strategies for the control of vector infestations and pathogen transmission.

Citation: Naranjo V, Ayllón N, Pérez de la Lastra JM, Galindo RC, Kocan KM, et al. (2013) Reciprocal Regulation of NF- κ B (Relish) and Subolesin in the Tick Vector, *Ixodes scapularis*. PLoS ONE 8(6): e65915. doi:10.1371/journal.pone.0065915

Editor: Olle Terenius, Swedish University of Agricultural Sciences, Sweden

Received: December 2, 2012; **Accepted:** April 30, 2013; **Published:** June 12, 2013

Copyright: © 2013 Naranjo et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This research was supported by grants BFU2008-01244/BMC and BFU2011-23896 to JF, the Oklahoma Agricultural Experimental Grant 1669 and the Walter R. Sitlington Endowed Chair for Food Animal Research to KMK, and the EU FP7 ANTIGONE project number 278976. V. Naranjo was funded by the European Social Fund and the Junta de Comunidades de Castilla-La Mancha (Program FSE 2007-2013), Spain. N. Ayllón was funded by Ministerio de Educación y Ciencia (MEC), Spain. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: jose_delafuente@yahoo.com

These authors contributed equally to this work.

^{‡a} Current address: ProBioVet S.L., Instituto Regional de Investigación Científica Aplicada (IRICA), Laboratorio 3.06, Universidad de Castilla-La Mancha, Ciudad Real, Spain

^{‡b} Current address: Office of Research Compliance and Biosafety, Texas A&M University, College Station, Texas, United States of America

Introduction

Subolesin, initially called 4D8, was discovered as a tick protective antigen in a mouse model of *Ixodes scapularis* (Acari: Ixodidae) infestations [1]. Subolesin gene and protein sequences are highly conserved in ticks and are the ortholog of insect and vertebrate Akirins [2–6]. RNA interference (RNAi) studies demonstrated that *subolesin* knockdown profoundly affects tick biological processes such as feeding and fertility and resulted in degeneration of tick guts, salivary glands, reproductive tissues and

embryos [2–4,7–9]. In subsequent studies, Subolesin was found to control tick gene expression, impact the innate immune response, and affect tick infection by *Anaplasma phagocytophilum*, *A. marginale*, *Babesia bigemina* and *Borrelia burgdorferi* [2–4,7–9,10–18]. Additionally, vaccination with Subolesin/Akirin demonstrated promising results for the control of arthropod vector infestations and pathogen infection/transmission [16–18].

The evolution of *subolesin/akirin* has been characterized in metazoans but little is known about gene organization and function in ticks [2–6,13,17]. Furthermore, although functional

studies have been conducted in several species including ticks, little is known about *subolesin/akirin* gene regulation [2–5,13,17]. Subolesin/Akirin have been suggested to play a role in the immune response through regulation of nuclear factor-kappa B (NF- κ B)-dependent and independent gene expression through interaction with intermediate proteins that interact with NF- κ B and other regulatory proteins, bind DNA or remodel chromatin to regulate gene expression [3,5,13,17–21]. However, further experiments are required to characterize the role of Subolesin/Akirin in the immune response.

Recently, *subolesin* and *NF- κ B (Relish)* gene knockdown in ticks suggested that NF- κ B might participate in the regulation of *subolesin* expression while Subolesin may be involved in the regulation of *NF- κ B (Relish)* expression [3]. The NF- κ B transcription factor is a key molecule in various cellular processes such as proliferation, cell death, development and innate and adaptive immunity [19]. These results suggested the hypothesis that NF- κ B (Relish) may play a role in the regulation of *subolesin* expression, thus adding an additional complexity to the immune response in ticks.

The objective of this study was to characterize the structure and regulation of *subolesin* in the tick vector, *I. scapularis*. *I. scapularis* is a three-host tick with a telotropic host behaviour that transmits emerging pathogens such as *B. burgdorferi*, *A. phagocytophilum* and *Babesia microti* that cause human Lyme disease, anaplasmosis and

babesiosis, respectively [22–25]. *I. scapularis* is a model organism for the study of vector-host-pathogen interactions [26,27]. Recent developments in *I. scapularis* genomics, proteomics and functional genomics have advanced our understanding of tick biological processes [26,27–30]. However, basic biological aspects such as gene organization and regulation are largely unknown in ticks. The information obtained from these studies advances our understanding of gene organization and regulation in *I. scapularis* and other tick species.

Results and Discussion

The *subolesin* Gene Sequence and Structure are Highly Conserved

Five exons and four introns were identified in the *subolesin* gene. From 5' to 3', exons 1–5 had 120 bp, 249 bp, 291 bp, 72 bp and 2,041 bp, respectively, and introns 1–4 had 128 bp, 7,807 bp, 11,410 bp and 2,386 bp, respectively (Figs. 1A and 1B). The alignment of *subolesin* gene sequences identified a total of 186 candidate polymorphisms with 5 single-nucleotide polymorphisms (SNPs) and 3 insertions/deletions (INDELs) on exon sequences and 156 SNPs and 22 INDELs located on intron sequences (Table 1). These polymorphisms and particularly INDELs could be attributed to errors in the assembly of the *I. scapularis* genome and/or to inter-strain variations that are common in ticks [31–33].

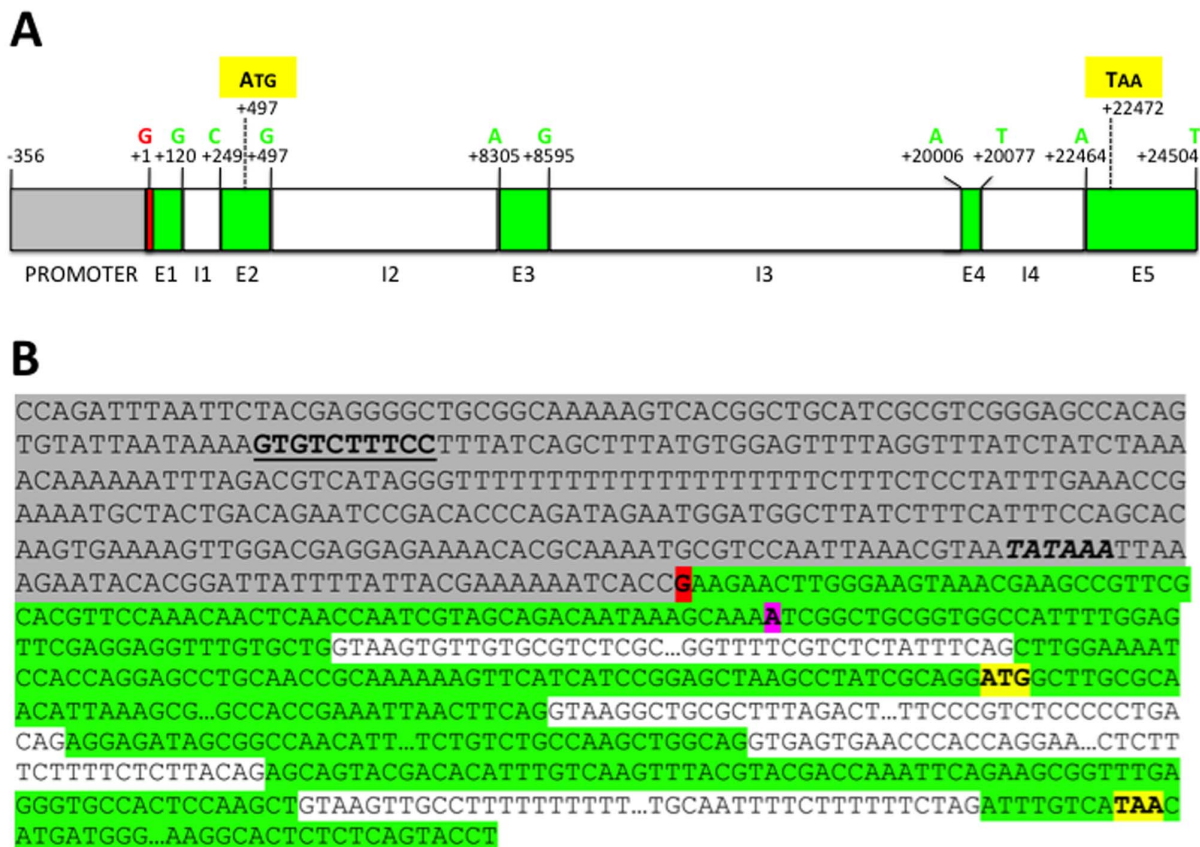


Figure 1. *subolesin* gene organization and sequence. (A) *subolesin* gene organization is shown containing promoter (grey), exon (E; green) and intron (I; white) regions. Major transcription start site at position +1 is shown in red. The first and last positions for exon sequences are shown in green. Translation start and stop codons are shown in yellow with positions marked for the first base of the codon. (B) *subolesin* gene sequence is shown using the same colors depicted in (A). The core promoter is shown with putative NF- κ B binding site (bold underlined), TATA box (bold italics), major (red) and minor (purple) transcription start sites indicated. Translation start and stop codons are shown in yellow. Only exon/intron junction sequences are shown. In the mRNA, polyA starts after the last nucleotide in exon 5. doi:10.1371/journal.pone.0065915.g001

Table 1. Characterization of polymorphisms in *subolesin* exon and intron sequences.

EXON/INTRON	SNPs	INDELS
Exon 1	0	0
Intron 1	1	0
Exon 2	2	0
Intron 2	60	7
Exon 3	4	0
Intron 3	59	7
Exon 4	0	0
Intron 4	36	8
Exon 5	0	1
Total	161	25

The number of SNPs and INDELS was recorded after *subolesin* sequence alignments performed between Wikel tick colony and ISE6 tick cells genomic sequences for introns and between Wikel tick colony genomic sequence and ISE6 tick cells and tick strains cDNAs for exons.
doi:10.1371/journal.pone.0065915.t001

Additionally, as in most *akirin* genes [6], the last tick *subolesin* exon was made mostly of untranslated sequences (Figs. 1A and 1B).

The *subolesin/akirin* sequences are highly conserved in ticks and other invertebrates and vertebrates [2–6]. Coding sequence identity at the nucleotide level was greater than 99% for all tick sequence pairwise comparisons (Fig. 2A). As reported by Macqueen and Johnston [6], our results support the conservation of gene organization (5 exons and 4 introns) in tick *subolesin* and insect and vertebrate *akirins* (Fig. 2B).

The exon/intron/exon junction sequences are highly conserved in all organisms with a consensus (<exon><GT–intron–AG><exon>) sequence [34] (Fig. 2B). In tick *subolesin*, all intron sequences at the <exon><intron><exon> junctions had the consensus sequence (Fig. 1B), which was conserved in human, tick and *Drosophila melanogaster* fly intron 2–4 sequences (Fig. 2B). Although tick <exon><intron> or <intron><exon> junction sequences diverged from consensus in exons 1, 2 and 4, these polymorphisms were common in other organisms (Fig. 2B). In general, *subolesin/akirin* <exon><intron><exon> junction sequences were conserved between different organisms and highly similar to the consensus (Fig. 2B). Remarkably, <exon 4><intron 4> junction sequence diverged from consensus but was conserved in human, tick and fly genes (Fig. 2B).

The *subolesin* Core Promoter Region is Located within 356 Nucleotides Upstream of the Transcription Start Site

Two putative transcription start sites were identified in tick *subolesin* (Fig. 1B). The strongest peak after mRNA start site analysis by automated capillary electrophoresis suggested that the major transcription start site was located at a guanine, 316 bases before the translation start site (Fig. 1B). The rapid amplification of cDNA ends (5′-RACE) analysis of *subolesin* cDNA corroborated both major and minor transcription start sites in 60% and 20% of sequenced clones, respectively, suggesting the possibility of more than one transcript for *subolesin* gene in tick cells.

Seven fragments (51–56 and SIN56) of the *subolesin* 5,503 pb promoter region were amplified by PCR, cloned and sequenced (Fig. 3A). The 51 to 56 fragments shared the same 3′ end with increasing deletions at the 5′ end, while the SIN56 fragment had the same 5′ end as the full-length 5,503 bp fragment 51, but with a

769 bp deletion at the 3′ end (Fig. 3A and Table 2). These promoter fragments were characterized for their ability to direct the expression of DsRed fluorescent marker in ISE6 tick cells. Although transfection efficiency of ISE6 tick cells was very low (approximately 0.1%; C+ in Figs. 3A and 3B), FACS analysis of transfected tick cells was consistent in two different experiments and showed that between $0.1\pm 0.0\%$ and $1.5\pm 0.6\%$ of the cells were positive for the DsRed marker in all constructs but SIN56 (Figs. 3A and 3B). These results suggested that the core promoter region [35] necessary and sufficient for *subolesin* expression in tick cells was located on the 56 fragment (Figs. 3A and 3B), containing 356 bp upstream of the major transcription start site (Fig. 1B). To further confirm these results, the reporter construct containing the 56 fragment with the *subolesin* core promoter region was transfected into mosquito Aag2 cells in which transfection efficiency was higher than in tick cells (35–40% transfected cells). The results confirmed the activity of the identified *subolesin* core promoter region in Aag2 cells (Figs. 4A–4C). In the *subolesin* core promoter region, a putative TATA box and NF- κ B-binding sites were predicted at positions –41 to –46 and –279 to –270 with respect to the major transcription start site, respectively (Fig. 1B).

NF- κ B Transcription Factors are Present in ISE6 Tick Cell Nuclear Extracts

NF- κ B (RelA (p65) and RelB)-binding activity was characterized in nuclear extracts of ISE6 tick cells under different treatments (Fig. 5A). The results demonstrated the presence of RelA and RelB-like factors in tick cell nuclear extracts (Fig. 5A). Competition with the consensus NF- κ B-binding site oligonucleotide suggested that binding activity in tick cells extracts was specific and similar to that obtained with Raji cell nuclear extracts (Fig. 5A). As in previous experiments with *Relish* [3], *subolesin* knockdown resulted in the inhibition of RelB but not RelA binding activity (Fig. 5A), probably reflecting a reduction in tick NF- κ B protein levels.

The search for NF- κ B-like sequences in *I. scapularis* databases resulted in three sequences (XP_002399379, XP_002434504, EW791150; the last sequence is likely an EST derived from XP_002434504) with homology to NF- κ B transcription factors (Fig. 5B). All these sequences contained the IPT-NF- κ B domain (cd01177), highly conserved in all species NF- κ B transcription factors such as RelA, RelB, p50 and Relish (Fig. 5B). The XP_002399379 sequence encodes for a Dorsal-Dif (Dorsal-related immunity factor)-like factor probably involved in the tick Toll pathway while the XP_002434504 sequence likely encodes for Relish that may be required for the IMD pathway described in other organisms [21]. These results showed that NF- κ B transcription factors are present in *I. scapularis* tick cells with binding site sequences similar to those found in other organisms.

NF- κ B Transcription Factors Bind to *subolesin* Core Promoter

A putative NF- κ B-binding site was predicted at positions –279 to –270 on the *subolesin* core promoter with a sequence (GTGTCTTTCC), 80% identical to the consensus NF- κ B-binding site (GGGACTTTCC) (Fig. 1B). NF- κ B-binding sites (1,742 and 1,912 hits for sequences identical to GGGACTTTCC and GTGTCTTTCC, respectively) were also mapped to *D. melanogaster* chromosome 3L (NT_037436.3) where *akirin* gene is located (3L: 7362900,7366958). Some of these putative NF- κ B-binding sites were located at less than 25 kb of the *akirin* gene (Fig. S1).

Electrophoretic mobility shift assays demonstrated that NF- κ B transcription factors RelA (p65) and p50 bind to *subolesin*

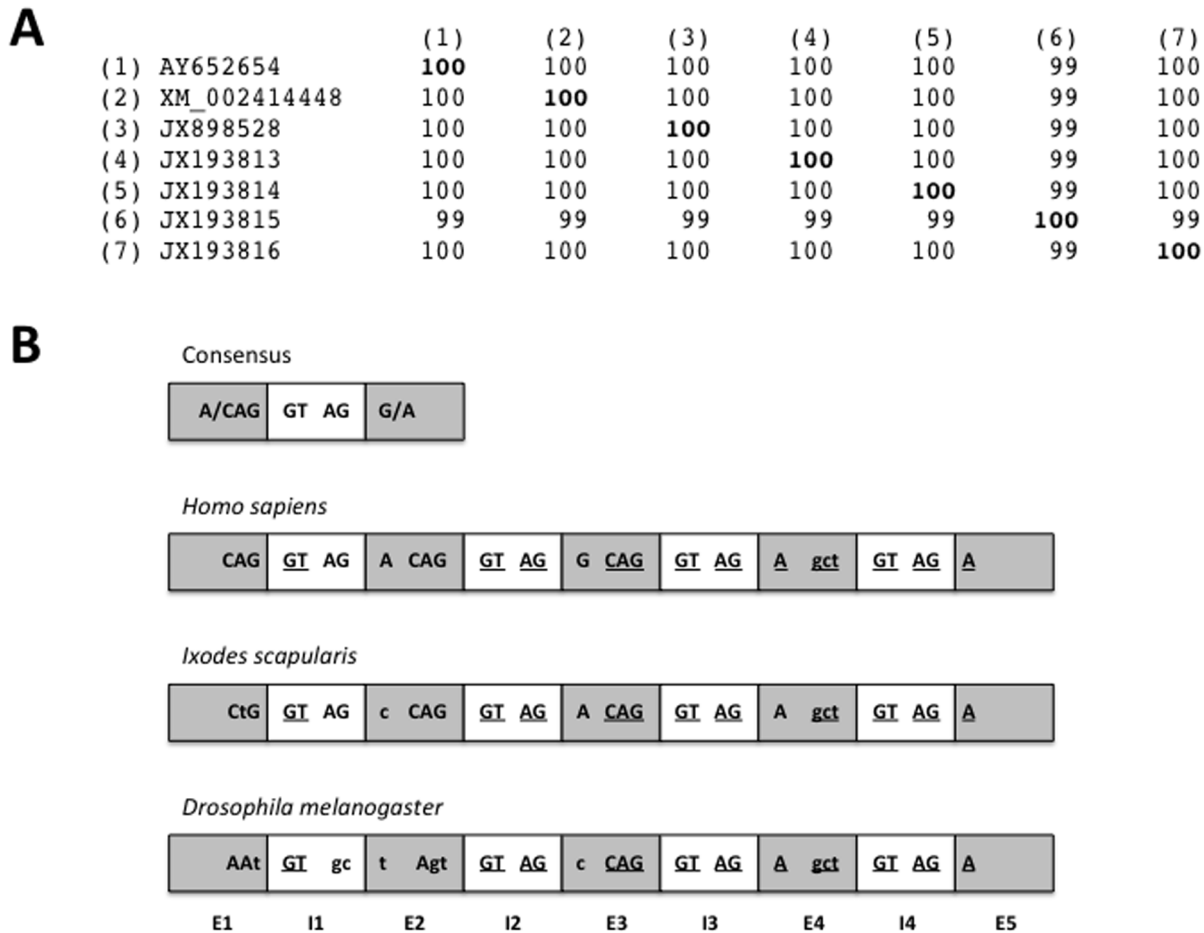


Figure 2. *subolesin* gene sequence and structure are highly conserved. (A) Percent identity matrix created by Clustal 2.1 for *subolesin* coding sequences in *I. scapularis* tick cell lines (1, IDE8; 3, ISE6) and tick strains (2, Wikel; 4, OSU; 5, NY; 6, RI1; 7, RI2). (B) Exon/intron/exon junction sequences in *subolesin* and *akirin* genes. The exon/intron/exon junction sequences are highly conserved in all organisms (consensus sequence). Human (*akirin2*; AC_000138), tick (JX898528) and fly (AE014296) *subolesin/akirin* gene exon/intron/exon junction sequences are shown. Nucleotides that differ from consensus sequence are shown in lower case letters. Nucleotides conserved among all three sequences are underlined. Abbreviations: E, exon; I, intron.

doi:10.1371/journal.pone.0065915.g002

promoter (Fig. 5C), recognizing the predicted GTGTCTTTCC binding site at positions -279 to -270 (Fig. 1B). Binding activity of p50 was higher than that of p65 (data not shown), reflecting the differences in DNA binding affinity of the NF- κ B subunits found in mammals [36].

NF- κ B Transcription Factors are Involved in the Regulation of *subolesin* Gene Expression in Ticks

RNAi experiments in *I. scapularis* female ticks suggested that NF- κ B (Relish) was involved in the regulation of *subolesin* gene expression [3]. In ISE6 tick cells, RNAi experiments did not affect cell viability and showed that NF- κ B (*Relish*) knockdown resulted in *subolesin* downregulation while *subolesin* knockdown also reduced *Relish* mRNA levels (Fig. 6A). Flow cytometry analysis showed that *subolesin* or *Relish* knockdown by RNAi resulted in lower Subolesin and NF- κ B protein levels (Fig. 6B).

The results from *subolesin* and *Relish* gene knockdown in ticks and ISE6 tick cells together with the p65 and p50 binding experiments shown herein in tick cells support the hypothesis that NF- κ B participates in the transcription of *subolesin* while Subolesin may be involved in the regulation of NF- κ B expression [3,17]. These results agreed with previous experiments in *D. melanogaster*

and mice suggesting that Akirins are involved in the regulation of NF- κ B and other transcription factors [3,37–39].

To further explore this hypothesis, chemical induction and repression of NF- κ B was conducted in ISE6 tick cells to characterize Subolesin mRNA and protein levels. The results showed that NF- κ B induction and repression did not affect cell viability and were accompanied with Subolesin induction and repression, respectively at the mRNA (Fig. 6C) and protein levels (Fig. 6D). These results strongly suggested that NF- κ B transcription factor is involved in the regulation of *subolesin* expression in ticks.

Subolesin and NF- κ B are Involved in Tick Cell Response to Pathogen Infection

The infection of ISE6 tick cells with *A. phagocytophilum* increased NF- κ B RelA (p65) and RelB binding activity (Figs. 5A and 5B) and NF- κ B (p50) protein levels (Figs. 7A and 7B) in infected ISE6 tick cells. Furthermore, although as in previous experiments *subolesin* mRNA levels did not change in response to *A. phagocytophilum* infection of ISE6 tick cells [40,41], protein levels were higher in infected than uninfected cells (Figs. 7A and 7B), probably reflecting regulation at the post-transcriptional level. Recently,

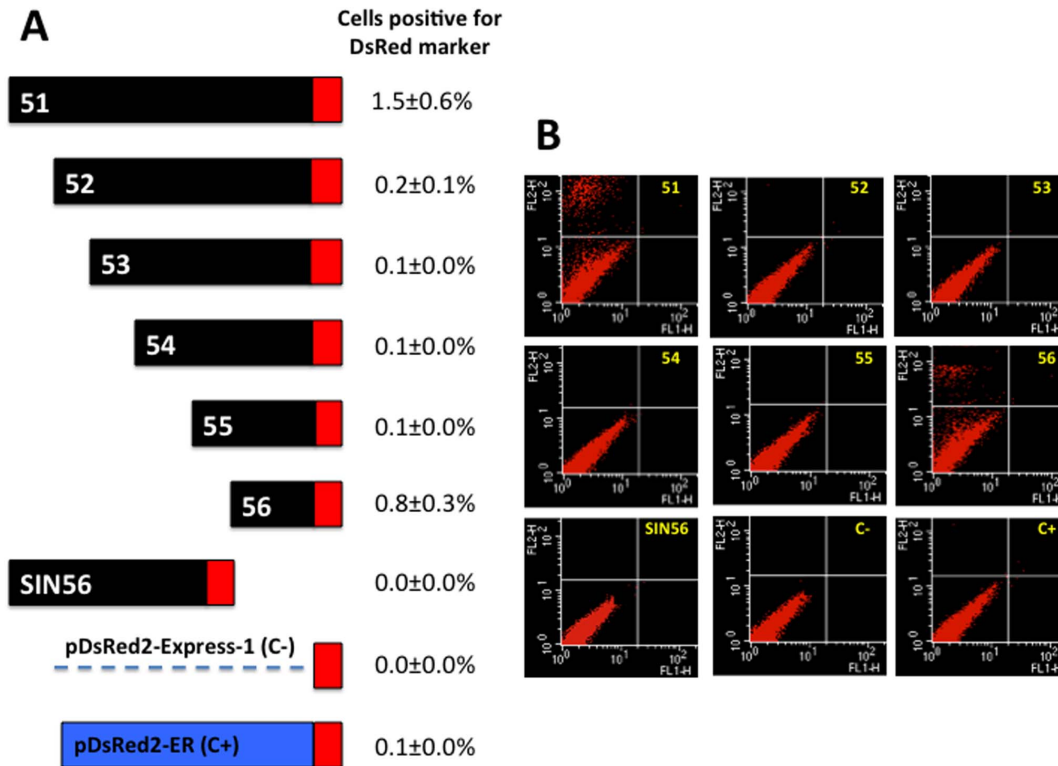


Figure 3. The *subolesin* core promoter is located in a fragment 356 bp upstream of the major transcription start site. (A) *subolesin* promoter fragments tested in ISE6 tick cells and percent positive cells for DsRed fluorescent marker (Ave±S.D.; N=2). (B) Flow cytometry profile of transiently transfected and control cells 24 h after transfection. doi:10.1371/journal.pone.0065915.g003

the proteome of ISE6 tick cells was characterized in response to *A. phagocytophilum* infection (unpublished results) and showed that one of the significantly increased proteins in infected cells was the ribosomal protein 3A (RP3A; XP_002402816; 3.1-fold increase, false discovery rate (FDR) = 0.004) which has been shown to be an integral subunit of the NF-κB transcription factor [19]. Together, these results showed that NF-κB and Subolesin protein levels increase in ISE6 tick cells in response to *A. phagocytophilum* infection.

Subolesin/Akirin are functionally conserved and have been proposed to be involved in host innate immune response to pathogen infection that affects the regulation of NF-κB-dependent gene expression [3,5,13–15,17,18,20,41–43] (Fig. 8A). In addition, Subolesin has a role in other molecular pathways including those required for tissue development and function that are important for pathogen infection and multiplication in ticks [2,14]. Consequently, a direct effect of *subolesin* knockdown in ticks is reduced

innate immunity, thereby increasing pathogen infection levels [44]. Lower pathogen infection levels may result from the effect of *subolesin* knockdown on tissue structure and function and the expression of genes that are important for pathogen infection and multiplication [11,15,16]. Collectively, these results support a role for Subolesin and NF-κB transcription factors in tick immune response, but Subolesin is also involved in the regulation of other molecular pathways in ticks.

Conclusions

Evolutionary and functional studies have shown that *subolesin/akirin* genes are conserved and essential to many physiological processes in metazoans [2–6,13,17,20] (Fig. 8A). The results presented herein demonstrated that *subolesin/akirin* are evolutionarily conserved at several levels (primary sequence, gene organization and function), thus showing that these molecules have a

Table 2. Primers and PCR conditions used for *subolesin* intron amplification.

Intron	Upstream/downstream primer sequence (5'–3')	PCR conditions (annealing, extension)
INTRON 1	56: TCAGCCATTTCAAAGTTGCG 3PR: GCTCCGGATGATGAACCTTT	56°C, 30 s 72°C, 1 min
INTRON 2	INT23: CAAATCAACCCTCACCTTC INT231: TATCTGTAGGGTGCAACGCA INT252: ACAGTGAATGCATCGGCTT INT25: ACTCCAGGGGAGACGAGAA	52°C, 30 s 72°C, 4 min 56°C, 30 s 72°C, 4 min
INTRON 3	INT33: ATGATGAAGGAGCGCGAGA INT331: TTCGGTGCAGTCCCTATTA INT351: AAAAGTGGAAGAGGAGGGA INT35: CCTCAAACCGCTTCTGAATT	56°C, 30 s 72°C, 5 min 30 s 56°C, 30 s 72°C, 3 min
INTRON 4	INT45: CCAAATTCAGAAGCGGTTG INT43: CCACCATGGGTTCTCTTT	56°C, 30 s 72°C, 3 min

doi:10.1371/journal.pone.0065915.t002

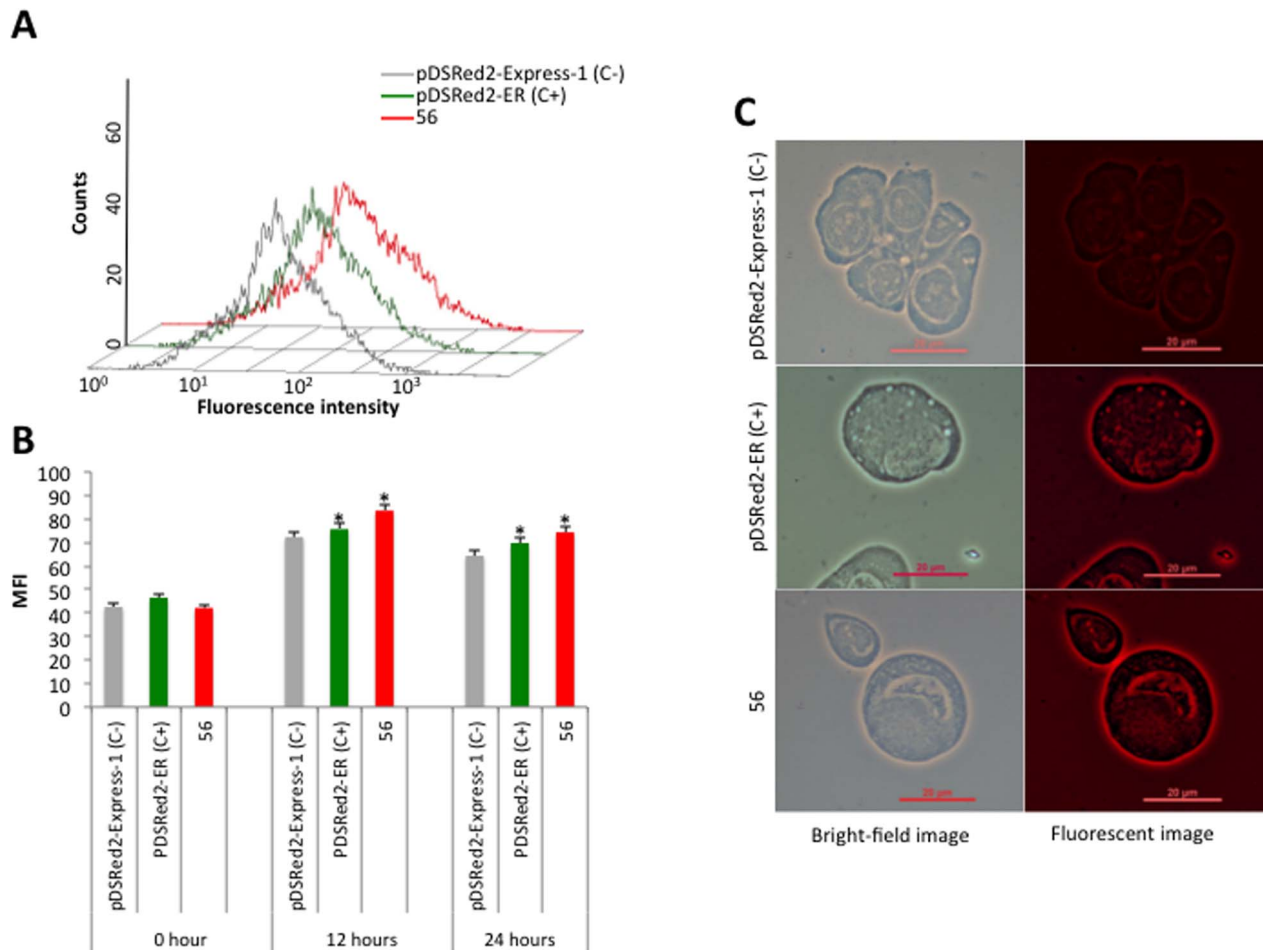


Figure 4. The *subolesin* core promoter is active in mosquito cells. (A) Flow cytometry profile histogram of mosquito Aag2 cells 12 h after transient transfection with pDsRed2-ER expressing the DsRed protein (positive control; C+), promoterless pDsRed-Express-1 (negative control; C-) and the construct containing the identified *subolesin* core promoter (fragment 56). (B) Geometric median fluorescence intensity (MFI) in transiently transfected Aag2 cells 0, 12 and 24 h after transfection. MFI values (Average \pm S.D.; N = 2) were compared between C+ and fragment 56 transfected cells and C- by Student's T-test (* $P \leq 0.05$). (C) Representative images of immunofluorescence analysis of transiently transfected Aag2 cells 12 h after transfection. Bars, 20 μ m. Two independent experiments were conducted with similar results. doi:10.1371/journal.pone.0065915.g004

crucial biological function in all metazoans. NF- κ B regulation of *subolesin* gene expression in ticks suggests that as in other organisms [19], different NF- κ B integral subunits and/or unknown interacting proteins regulate the specificity of the NF- κ B-mediated gene expression in ticks (Fig. 8A). Additionally, the possible implication of Subolesin in NF- κ B regulation in ticks adds to the complexity of this regulatory network with important implications in tick immune response (Fig. 8A). Taken together, these results suggest a regulatory network that includes cross-regulation between NF- κ B and Subolesin and Subolesin auto-regulation (Fig. 8B). These studies highlight the importance of characterizing gene regulation in ticks, with particular emphasis on immune response pathways and the role of NF- κ B and Subolesin in gene regulation to design new strategies for the control of tick infestations and pathogen transmission.

Materials and Methods

ISE6 Tick Cells

The ISE6 tick cell line, derived originally from *I. scapularis* embryos (provided by U.G. Munderloh, University of Minnesota,

USA) was cultured in L15B medium as described previously [46,47]. The ISE6 tick cells were inoculated with the NY18 isolate of *A. phagocytophilum* or the Oklahoma isolate of *Anaplasma marginale* and maintained as described previously [31,40,41]. Uninfected and infected cultures with approximately 10^7 cells each were sampled at 13 days post-infection (dpi) with approximately 30% infected cells. Collected cells were centrifuged at 10,000 \times g for 3 min and used in various experiments.

RNA Interference in ISE6 Tick Cells

The dsRNA for *subolesin*, NF- κ B (*Relish*; XP_002434504) and *Rs86* control were generated with oligonucleotide primers containing T7 promoter sequences for *in vitro* transcription and synthesis of dsRNA using the Access RT-PCR system (Promega, Madison, WI, USA) and the Megascript RNAi kit (Ambion, Austin, TX, USA) as reported previously and using oligonucleotide primers RelRNAi5: 5'-TAATACGACTCACTATAGGG-TACTATGTTCCCTGTAATGTCCG-3' and RelRNAi3: 5'-TAATACGACTCACTATAGGGTACTGCTCGAGCAAAC-TCCCGTC-3' for *Relish* [2,3,48]. RNAi experiments were conducted in cell cultures by incubating ISE6 tick cells with

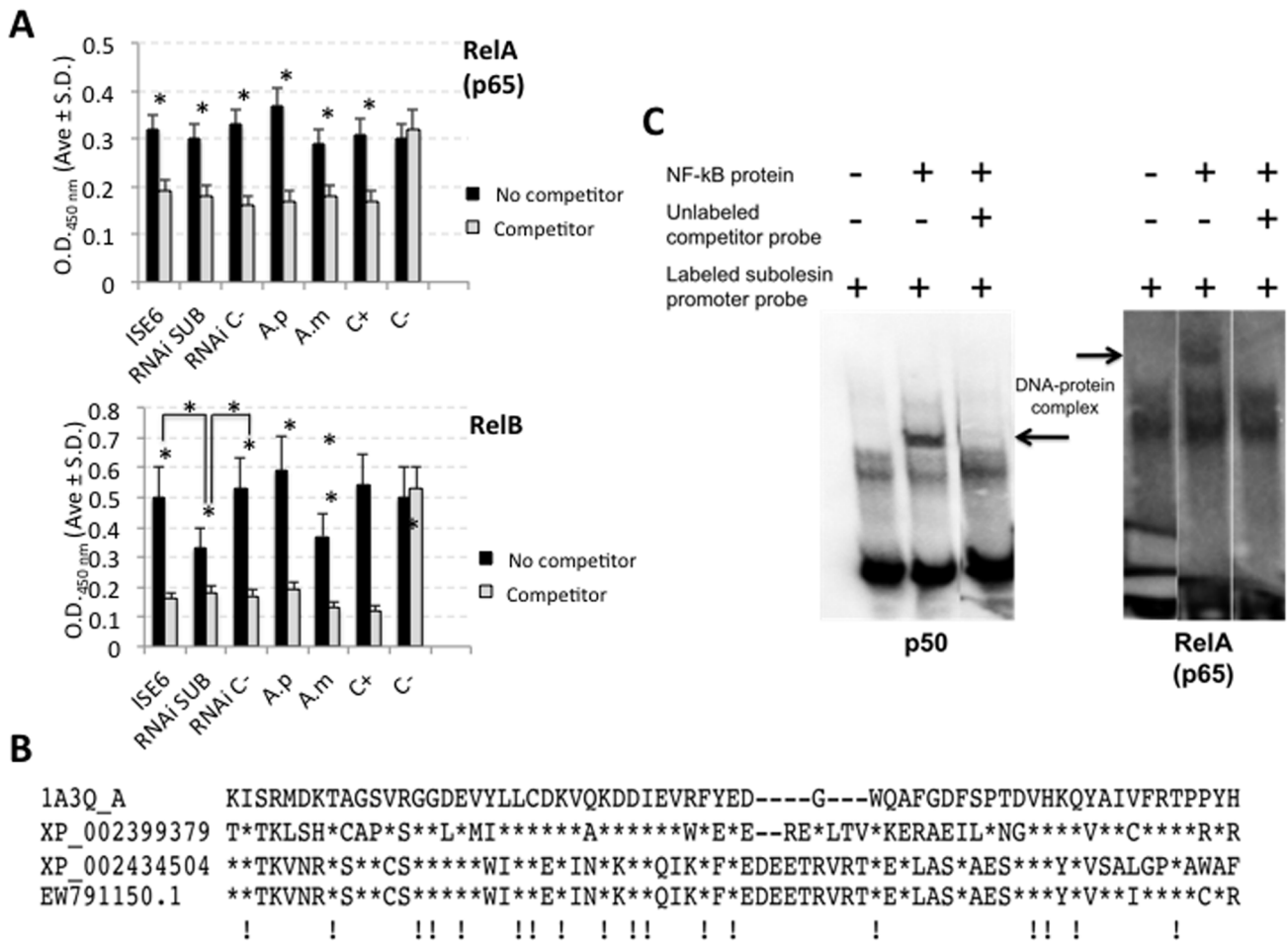


Figure 5. NF-kB factors are present in *I. scapularis* tick cells and bind to *subolesin* promoter. (A) NF-kB-binding activity in ISE6 tick cells nuclear extracts. A DNA-binding ELISA was used with RelA (p65) and RelB antibodies to characterize NF-kB-binding activity in ISE6 tick cells. Nuclear extracts were prepared and assayed from untreated ISE6 tick cells (ISE6), tick cells after incubation with *subolesin* dsRNA (RNAi SUB) or the unrelated *Rs86* dsRNA (RNAi C-), and *A. phagocytophilum* (A.p) and *A. marginale* (A.m) infected tick cells. The wild-type consensus NF-kB-binding oligonucleotide was used as a competitor for NF-kB binding in order to monitor the specificity of the assay when compared to non-competing conditions. The mutated consensus oligonucleotide which should have no effect on NF-kB binding was used as negative control (C-) while Raji cells nuclear extracts were used as positive control for NF-kB binding (C+). Results (Ave±S.D.; N=4) were compared between groups by Student's T-test (*P≤0.05). (B) Protein sequence alignment of the IPT-NF-kB domain (cd01177) in putative *I. scapularis* NF-kB factors. Asterisks denote identity to human reference sequence (1A3Q_A). Amino acids conserved in all sequences are shown with (!) at the bottom of the alignment. (C) NF-kB transcription factor binds to *subolesin* promoter. Representative electrophoretic mobility shift assay showing *subolesin* promoter-NF-kB interactions. Labeled dsDNA probe corresponding to the *subolesin* promoter was incubated with p50 and RelA (p65) NF-kB transcription factors. The unlabeled probe used to compete DNA-protein interactions corresponded to the predicted NF-kB-binding site in the *subolesin* promoter (GTGCTTTCC). Fast migrating unbound probes are found at the bottom of the gel whereas protein-DNA complexes have slower mobility (arrows). doi:10.1371/journal.pone.0065915.g005

10 µl dsRNA (5×10¹⁰–5×10¹¹ molecules/µl) and 90 µl L15B medium in 24-well plates using 4 wells per treatment [40]. Control cells were incubated with the unrelated *Rs86* dsRNA. After 48 hours of dsRNA exposure, tick cells were infected with *A. phagocytophilum* (NY18 isolate) or mock infected by adding culture medium alone as described previously [40,41]. Cells were incubated for an additional 72 hours, collected and used for RNA extraction. RNA was used to analyze *subolesin* and *NF-kB* (*Relish*) mRNA levels by real-time RT-PCR as described previously and using oligonucleotides RelPCR5:5'-GAGGACTGCTGC-TACGTCAC-3' and RelPCR3:5'-ACCAGGTTTCAGGTT-CAGCTC-3' for *Relish* [45]. Gene knockdown was analyzed by real-time RT-PCR with respect to *Rs86* control by comparing the normalized Ct values against tick *16S rRNA* for each treatment against the average control normalized Ct value (N=4) [3,41].

The normalized Ct values were compared between treated and control cells by Student's t-test with unequal variance (P = 0.05).

Characterization of Subolesin Gene Organization and Sequence

To characterize *subolesin* gene organization and sequence, a cDNA encoding for *I. scapularis subolesin* (IDE8 cells; Genbank accession number AY652654) was aligned with an *I. scapularis* genomic sequence (Wikel tick colony; Genbank accession number DS936446). Total DNA was isolated from *I. scapularis* ISE6 tick cells using TRI Reagent (Sigma, St. Louis, Mo, USA) following manufacturer's recommendations. Primers were designed to amplify the predicted introns (Table 2) by PCR using the DyNAzyme EXT PCR kit (Finnzymes Oy, Espoo, Finland) in a 50 µl reaction mixture. Total RNA was isolated from ISE6 tick

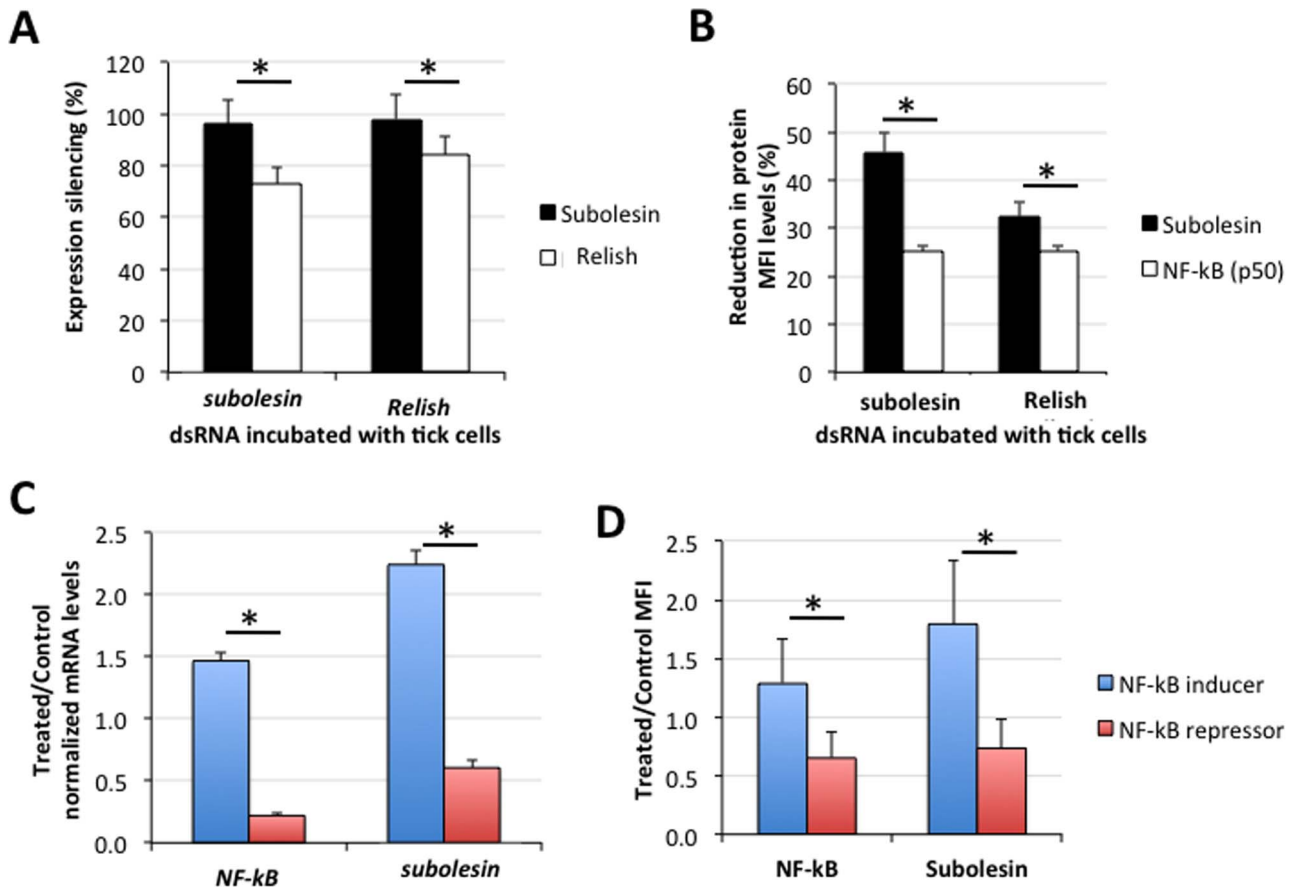


Figure 6. NF- κ B transcription factors are involved in the regulation of *subolesin* expression in ISE6 tick cells. (A) For gene knockdown by RNAi, ISE6 tick cells were incubated with *subolesin* or NF- κ B (*Relish*) dsRNAs and compared to control cells incubated with the unrelated *Rs86* dsRNA. The *subolesin* and *Relish* mRNA levels were determined by real-time RT-PCR and normalized against tick *16S rRNA*. The percent silencing of gene expression was calculated by comparing the normalized Ct values for each treated cell well against the average control normalized Ct value and plotted as average \pm S.D. (N=4). The normalized Ct values were compared between treated and control cells by Student's t-test with unequal variance and were statistically significant (*P \leq 0.05). (B) Subolesin and NF- κ B protein levels were determined after gene knockdown by flow cytometry. MFI for NF- κ B and Subolesin was calculated as the MFI of the test-labeled sample minus the MFI of the isotype control and used to calculate expression silencing by comparison to the MFI of the *Rs86* dsRNA-treated control. MFI values (Ave \pm S.D.; N=2) were compared between groups by Student's T-test with unequal variance and were statistically significant (*P \leq 0.05). (C) Subolesin and NF- κ B mRNA levels in ISE6 tick cells after induction and repression of NF- κ B. The mRNA levels were determined by RT-PCR and normalized Ct values were calculated for NF- κ B (*Relish*) and *subolesin*. (D) Subolesin and NF- κ B protein levels in ISE6 tick cells after induction and repression of NF- κ B. Protein levels were determined by flow cytometry and MFI for NF- κ B and Subolesin was calculated as the MFI of the test-labeled sample minus the MFI of the isotype control. For both mRNA and protein levels, Treated/Control ratios (Ave \pm S.D.; N=2) were compared between groups by Student's T-test (*P \leq 0.05). doi:10.1371/journal.pone.0065915.g006

cells and *I. scapularis* tick strains from Oklahoma (OSU; Tick Rearing Facility, Oklahoma State University, Stillwater, OK, USA), New York (NY; D. Sonenshine, Old Dominion University, Norfolk, VA, USA) and Rhode Island (RI1; T.N. Mather, University of Rhode Island, Kingston, RI, USA and RI2; R.F. Massung, Centers for Disease Control and Prevention, Atlanta, GA, USA) using TRI Reagent (Sigma). The *subolesin* cDNAs were amplified by RT-PCR using oligonucleotide primers 4D8R5: (5'-GCTTGCGCAACATTAAGCGAAC-3') and 4D8R: (5'-TGCTTGTTCGAGATGCCCATCA-3') and the Access RT-PCR system (Promega) as previously described [2]. Control reactions were performed without DNA or RNA to monitor contamination of the PCR reaction. PCR products were electrophoresed in 1% agarose gels to check the size of amplified fragments by comparison to a DNA molecular weight marker (1 Kb Plus DNA Ladder; Promega, Madison, WI, USA). Amplified fragments were purified using a MinElute PCR purification kit (Qiagen, Valencia, CA, USA) and cloned into

the pGEM-T vector (Promega) for sequencing both strands by double-stranded dye-determination cycle sequencing and primer walking (Core Sequencing Facility, Department of Biochemistry and Molecular Biology, Noble Research Center, Oklahoma State University, Stillwater, OK, USA). At least three clones were sequenced for each fragment. Sequence alignments were performed between Wikel tick colony and ISE6 tick cells genomic sequences for introns and between Wikel tick colony genomic sequence and ISE6 tick cells and tick strains cDNAs for exons to characterize single-nucleotide polymorphisms (SNPs) and insertion/deletions (INDELs). Sequences were deposited in the Genbank under accession numbers JX898528 (for *subolesin* genomic sequence in ISE6 tick cells) and JX193813-JX193816 (for tick strains cDNAs).

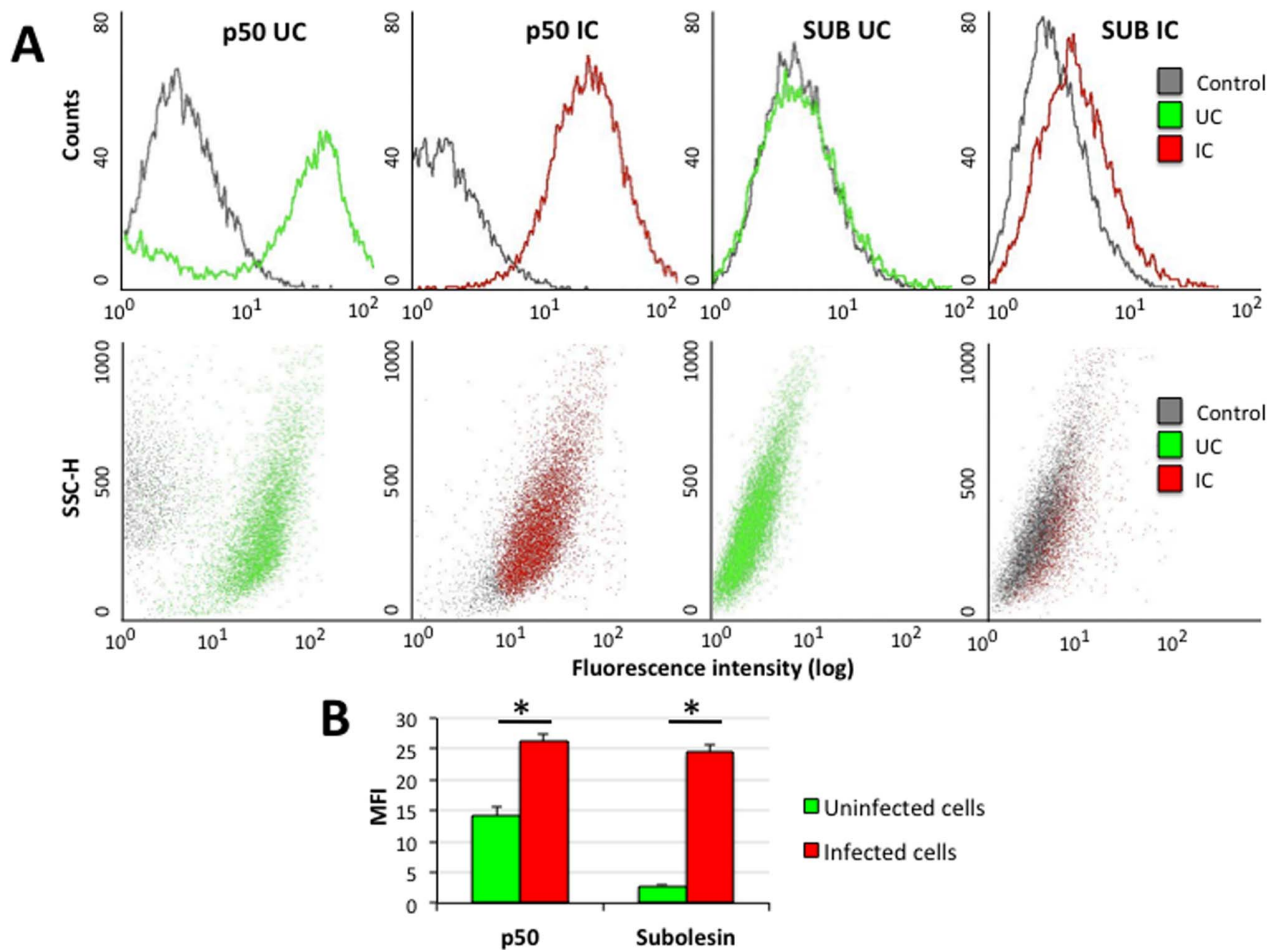


Figure 7. Subolesin and NF- κ B protein levels are higher in ISE6 tick cells infected with *A. phagocytophilum*. (A) Flow cytometric profile histogram and plot for the isotype control (grey), NF- κ B (p50) and Subolesin (SUB) in uninfected (UC; green) and *A. phagocytophilum*-infected (IC; red) tick cells. (B) MFI for NF- κ B and Subolesin in infected and uninfected tick cells. MFI was calculated as the MFI of the test-labeled sample minus the MFI of the isotype control. Results (Ave \pm S.D.; N = 2) were compared between groups by Student's T-test (*P \leq 0.05). doi:10.1371/journal.pone.0065915.g007

Mapping of the *subolesin* Transcription Start Site by Automated Capillary Electrophoresis

The analysis of *subolesin* transcription start site was performed using mRNA start site analysis by automated capillary electrophoresis (MSACE; Plant-Microbe Genomics Facility, Ohio State University, OH, USA). MSACE consists on a primer extension assay where the extension product is labeled with a fluorescent dye that is analyzed by automated capillary electrophoresis. RNA from ISE6 cells was isolated using RNeasy mini kit (Qiagen) following manufacturer's recommendations. The oligonucleotide primer PESUB (5'-ACTATGCAGCGGATCCCAATCG-3') was labeled with 6FAM (Applied Biosystems, Carlsbad, CA, USA) on the 5' end and used for the primer extension assay. PESUB is located between positions +27 to +48 after the translation start site of the *subolesin* gene (AY652654). Ten μ g RNA were mixed with 100 pmol 6FAM-PESUB primer in a 11 μ l total volume. The mix was heated at 90°C for 3 min and then slowly cooled to 30°C in a PCR thermocycler. Primer extension was completed by adding AMV reverse transcriptase (AMV RT) 5X reaction buffer (Promega), 10 mM dNTP mix, 40 U of RNasin ribonuclease inhibitor (Promega) and 25 U of AMV RT enzyme (Promega). The reaction was incubated for 1 h at 42°C. RNA was degraded using 10 ng RNase A (10 μ l at 1 ng/ μ l) for 30 min at 37°C [49].

The resulting cDNA was purified with the MinElute PCR purification kit (Qiagen) and eluted in 15 μ l water. The template necessary for the DNA sequencing reaction (520 bp) included 469 bases upstream of the translation start site. The DNA template was amplified with 56PE (5'-GACAGAATCCGACACCCAGA-3') and PESUB primers (5'-ACTATGCAGCGGATCCCAATCG-3') by PCR using GoTaq Flexi DNA polymerase (Promega). The specific PCR conditions were 95°C for 2 min followed of 35 cycles of denaturation at 95°C for 30 sec, annealing at 60°C for 30 sec and extension at 72°C for 1 min. The PCR product was purified with the MinElute PCR purification kit (Qiagen) and cloned into the pGEM-T vector (Promega). The extension product was analyzed on a 3730 DNA Analyzer (Applied Biosystems). Results from MSACE analysis were corroborated by 5' System for Rapid Amplification of cDNA Ends (RACE; Life Technologies S.A., Madrid, Spain) using gene-specific primers GSP1 (4D8R: 5'-TGCTTGTTTGCAGATGCCCATCA-3') and GSP2 (3PRO: 5'-CCGGATCCGCTCCGGATGATGAACCTTTT-3) and following manufacturer's recommendations [50]. The PCR product was purified, cloned into pGEM-T and 20 clones were sequenced as previously described.

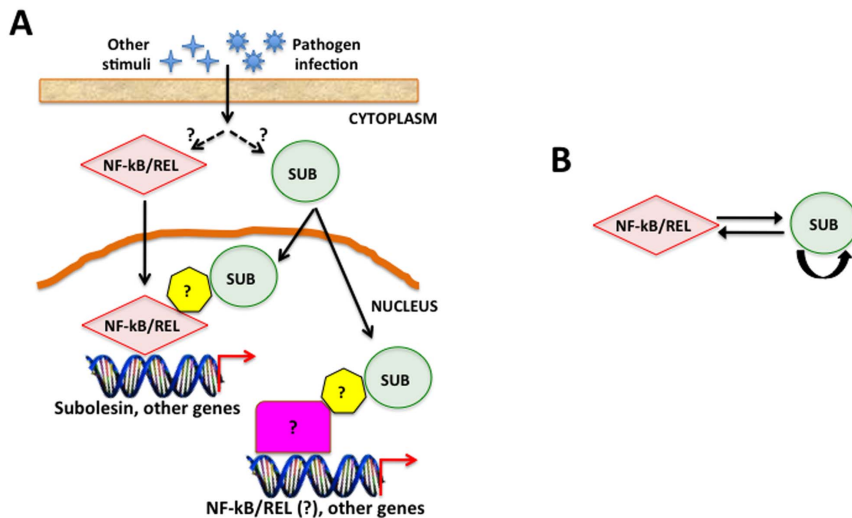


Figure 8. Model for Subolesin regulation in ticks. (A) Proposed model for NF- κ B/Relish (REL) and Subolesin (SUB) regulation of gene expression in ticks. The mechanisms resulting in NF- κ B/Subolesin activation and/or increased protein levels are still unknown. NF- κ B/REL directly regulates the expression of SUB and other genes. SUB interacts with NF- κ B/REL subunits through unknown auxiliary proteins and may be involved in the regulation of NF- κ B/REL expression and the regulation of NF- κ B/REL-independent gene expression in ticks. (B) Proposed regulatory network that includes cross-regulation between NF- κ B/REL and Subolesin and Subolesin auto-regulation. doi:10.1371/journal.pone.0065915.g008

Characterization of *subolesin* Core Promoter

A 5,503 pb region was used to characterize *subolesin* promoter sequence. DNA was isolated from ISE6 tick cells as previously described. Oligonucleotide primers were designed to amplify by PCR 7 different fragments with 5' or 3' deletions in the *subolesin* promoter region (Table 3). PCR reactions were performed using the Advantage Genomic LA Polymerase Mix (Clontech, Mountain View, CA, USA) in a 25 μ l reaction mixture. Control reactions were performed without DNA to monitor contamination of the PCR reaction. PCR products were electrophoresed, purified, cloned into pGEM-T and sequenced as previously described. Positives clones from the seven fragments were digested with *Bam*HI and *Bgl*II (Promega), purified using the Qiaquick Gel Extraction kit (Qiagen) and cloned into the pDsRed-Express-1 vector (Clontech), which is a promoterless vector containing the *Discosoma* sp. red fluorescent protein (DsRed) marker. Resulting constructs were purified and 4 μ g were used to transfect ISE6 tick cells using Effectene Transfection Reagent (Qiagen). Two independent experiments were conducted. The pDsRed2-ER expressing the DsRed protein and promoterless pDsRed-Express-1 were used as positive and negative controls, respectively. After transfection, cells were incubated in L15B medium for 24 h [51] and then analyzed for DsRed protein expression by flow cytometry FACS analysis using a FACSCalibur cytometer (Becton Dickinson, Franklin Lakes, NJ, USA).

To confirm these results, the construct containing the identified *subolesin* core promoter (fragment 56) was more efficiently transfected into *Aedes aegypti* mosquito Aag2 cells (kindly provided by Alain Kohl and Claudia Rueckert; [52]). As in the experiment with tick cells, pDsRed2-ER expressing the DsRed protein and promoterless pDsRed-Express-1 were used as positive and negative controls, respectively. Cells were grown at 31°C in L15 medium containing 10% tryptose phosphate broth (Sigma) and 10% fetal bovine serum. Cells were seeded for transfection 24 h prior to use in 24-well plates. Approximately 2×10^5 cells/well were transfected with 100 μ l/well of Lipofectamine 2000 (Invitrogen, Oregon, USA; 1 μ l/well) and plasmid DNA (4 μ g/well) in Optimem added to 400 μ l of medium/well and incubated for

5 h at 31°C following manufacturer's instructions. After transfection, cells were incubated in L15 medium for 12 and 24 h and collected for FACS as described before. Cells were also observed under a Nikon Eclipse Ti-U fluorescent microscope with a Nikon Digital Sight DS Vi1 camera. Two independent experiments were conducted with similar results.

DNA-binding ELISA for the Characterization of NF- κ B Binding Activity in Tick Cells

To characterize NF- κ B binding activity in ISE6 tick cells, a DNA-binding ELISA was used with RelA (p65) and RelB antibodies (Trans AM NF- κ B Family Transcription Factor Assay Kit, Active Motif, Carlsbad, CA, USA). ISE6 tick cells were collected in PBS and lysed following manufacturer's instructions. After centrifugation, the pellet contained the nuclear fraction at a concentration of 4 mg/ml. The wild-type consensus NF- κ B oligonucleotide (5'-GGGACTTTCC-3') was used (20 pmol/well) as a competitor for NF- κ B binding in order to monitor the specificity of the assay. Conversely, the mutated consensus oligonucleotide (Active Motif) should have no effect on NF- κ B binding and served as negative control. The Raji cells nuclear extract (Active Motif) was used as positive control for NF- κ B binding. Four replicates were done for each assay and results were compared between groups by Student's T-test ($P = 0.05$).

Electrophoretic Mobility Shift Assay for the Characterization of NF- κ B Binding to *subolesin* Promoter

NF- κ B binding to *subolesin* promoter was assessed by incubating 5' end biotin-labeled double stranded (ds)DNA oligonucleotide corresponding to the surrounding region of the predicted NF- κ B binding site in the *subolesin* promoter (nucleotides -309 to -252; Fig. 1B) with p50 and Rel (p65) proteins (Active Motif, Carlsbad, CA USA) according to the manufacturer's protocol (Gel Shift Chemiluminiscent EMSA kit, Active Motif). NF- κ B proteins (0.1 μ g) were incubated independently with 20 fmol of labeled dsDNA probe in binding buffer (EMSA kit, Active Motif) in a final volume of 20 μ l at room temperature for 20 min. The resulting

Table 3. Primers and PCR conditions used for *subolesin* promoter analysis.

Promoter fragment	Length (bp)	Upstream/downstream primer sequence (5'-3')	PCR conditions (annealing, extension)
51	5503	51PRO:GGAGATCTAATGCCGTC AATAGTGCCTT 3PRO: CCGGATCCGCTCCGGATGATGAAC TTTT	57°C, 30 s 72°C, 5 min
52	4584	52PRO:CCAGATCTACAAAAGGTTCTCTGACCC 3PRO: CCGGATCCGCTCCGGATGATGAAC TTTT	56°C, 30 s 72°C, 3 min
53	3592	53PRO:CCAGATCTTTT GAGGGAAGTGCTAGAGCG 3PRO: CCGGATCCGCTCCGGATGATGAAC TTTT	61°C, 30 s 72°C, 4 min
54	2571	54PRO: CCAGATCTAAAGCGTAGGTTTCTGCCA 3PRO: CCGGATCCGCTCCGGATGATGAAC TTTT	56°C, 30 s 72°C, 3 min
55	1673	55PRO: GGAGATCTAGTCCGCTTTTATCCTTGAAA 3PRO: CCGGATCCGCTCCGGATGATGAAC TTTT	56°C, 30 s 72°C, 1 min
56	763	56PRO: CCAGATCTTCAGCCATTTCAAAGTTGCG 3PRO: CCGGATCCGCTCCGGATGATGAAC TTTT	56°C, 30 s 72°C, 1 min
SIN56	4734	51PRO: GAGATCTAATGCCGTC AATAGTGCCTT SIN56: CAAACTACGCGCTGTATAAATATCCCC	56°C, 30 s 72°C, 3 min

doi:10.1371/journal.pone.0065915.t003

complexes were separated in a 6% DNA retardation gel (Invitrogen, Madrid, Spain). Competition with the unlabeled probe containing the predicted subolesin NF- κ B binding site (GTGTCTTTCC) were done in a 200:1 (unlabeled:labeled) ratio.

Characterization of Subolesin and NF- κ B Expression in Tick Cells

Subolesin and NF- κ B expression was characterized in ISE6 tick cells in response to *A. phagocytophilum* infection, after gene knockdown by RNAi and after activation and repression of the NF- κ B pathway. Tick cells were infected with *A. phagocytophilum* or left uninfected as described before. For the activation and repression of the NF- κ B pathway, ISE6 tick cells were incubated at 31°C for 24 hrs with 7.5 mg/ml NF- κ B inducer (2-Deoxy-D-glucose; Sigma, St. Louis, MO, USA) [53], 5 mM NF- κ B repressor (Sodium Salicylate; Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA) [54], and PBS as control and analyzed by RT-PCR and flow cytometry for subolesin and NF- κ B (Relish or p50) expression.

The *subolesin* and *NF- κ B (Relish)* mRNA levels were characterized by real-time RT-PCR as described before for RNAi. For flow cytometry analysis, ISE6 tick cells were washed in phosphate buffered saline (10 mM PBS) fixed and permeabilized with Intracell[®] fixation and permeabilization kit (Immunostep, Salamanca, Spain) following manufacturer recommendations. After permeabilization, the cells were washed in PBS and incubated in 100 μ l with primary unlabeled rabbit polyclonal antibodies (preimmune IgG isotype control; 5 μ g/ml, anti-subolesin; 5 μ g/ml) and labelled monoclonal antibodies (PE-anti-NF- κ B p50; 40 μ g/ml), washed in PBS and incubated in 100 μ l of PBS with FITC or Cy5-goat anti-rabbit IgG for subolesin (Sigma, Madrid, Spain) labeled antibodies (diluted 1/500) for 15 min at 4°C. Finally, the cells were washed with PBS and resuspended in 500 μ l of PBS. All samples were analyzed on a FACScalibur[®] Flow Cytometer, equipped with the CellQuest Pro[®] software (BD-Biosciences, Madrid, Spain). The viable cell population was gated according to forward scatter and side scatter parameters. The level of Subolesin and NF- κ B in the viable cells was determined as the geometric median fluorescence intensity (MFI) of the test-labeled sample minus the MFI of the isotype control [55]. Two replicates were done for each assay and results were compared between groups by Student's T-test (P = 0.05).

References

- Almazán C, Kocan KM, Bergman DK, Garcia-Garcia JC, Blouin EF, et al. (2003) Identification of protective antigens for the control of *Ixodes scapularis* infestations using cDNA expression library immunization. *Vaccine* 21: 1492–501.
- de la Fuente J, Almazán C, Blas-Machado U, Naranjo V, Mangold AJ, et al. (2006) The tick protective antigen, 4D8, is a conserved protein involved in modulation of tick blood digestion and reproduction. *Vaccine* 24: 4082–4095.

Sequence Analysis

Multiple sequence alignments were performed using Geneious Pro 5.0.3 software (Biomatters Ltd., Auckland, New Zealand; <http://www.geneious.com>). Transcription factor binding sites to subolesin promoter were predicted using TRANSFAC at Biobase (<http://www.biobase-international.com/product/transcription-factor-binding-sites>). Blasting against nonredundant sequence database (nr) and databases of tick-specific (<http://www.ncbi.nlm.nih.gov> and <http://www.vectorbase.org/index.php>) and *D. melanogaster* release 5.30 genomic sequences was done using tblastx and blastn (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). Protein sequences were aligned using the CLUSTAL 2.1 multiple sequence alignment tool (EMBL-EBI; <http://www.ebi.ac.uk/Tools/>). Conserved protein domains (cd) were analyzed using Conserved Domains and Protein Classification at ncbi (<http://www.ncbi.nlm.nih.gov/Structure/cdd/cdd.shtml>) [56]. *D. melanogaster* akirin and NF- κ B-binding sites were mapped using the NCBI Map Viewer (<http://www.ncbi.nlm.nih.gov/mapview/>).

Supporting Information

Figure S1 NF- κ B-binding sites in *D. melanogaster* chromosome 3L (NT_037436.3) where akirin gene is located (3L: 7362900,7366958). Blasting was done against GTGTCTTTCC sequence in *D. melanogaster* release 5.30 genomic sequence using blastn (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) and mapped using the NCBI Map Viewer (<http://www.ncbi.nlm.nih.gov/mapview/>). (TIF)

Acknowledgments

We would like to thank Vishvanath Nene (International Livestock Research Institute, Nairobi, Kenya) for providing the initial genomic sequence of the *I. scapularis* (Wikel strain) *subolesin* locus. We thank Petr Kopacek and Ondra Hajdusek (Institute of Parasitology, Biology Centre, Academy of Sciences of the Czech Republic) for fruitful discussions.

Author Contributions

Conceived and designed the experiments: JF. Performed the experiments: VN NA JMPL RCG KMK EFB RM PA MV. Analyzed the data: VN NA JMPL RCG RM MV JF. Wrote the paper: JF VN KMK.

3. Galindo RC, Doncel-Pérez E, Zivkovic Z, Naranjo V, Gortazar C, et al. (2009) Tick subolesin is an ortholog of the akirins described in insects and vertebrates. *Dev Comp Immunol* 33: 612–617.
4. Mangold AJ, Galindo RC, de la Fuente J (2009) Response to the commentary of D. Macqueen on: Galindo RC, Doncel-Pérez E, Zivkovic Z, Naranjo V, Gortazar C, et al. Tick subolesin is an ortholog of the akirins described in insects and vertebrates [Dev. Comp. Immunol. 33 (2009) 612–617]. *Dev Comp Immunol* 33: 878–879.
5. Goto A, Matsushita K, Geselchen V, El Chamy L, Kutenkeuler D, et al. (2008) Akirins are highly conserved nuclear proteins required for NF-kappaB-dependent gene expression in drosophila and mice. *Nat Immunol* 9: 97–104.
6. Macqueen DJ, Johnston IA (2009) Evolution of the multifaceted eukaryotic akirin gene family. *BMC Evol Biol* 9: 34.
7. de la Fuente J, Almazán C, Naranjo V, Blouin EF, Meyer JM, et al. (2006) Autocidal control of ticks by silencing of a single gene by RNA interference. *Biochem Biophys Res Commun* 344: 332–338.
8. Nijhof AM, Taoufik A, de la Fuente J, Kocan KM, de Vries E, et al. (2007) Gene silencing of the tick protective antigens, *Bm86*, *Bm91* and *subolesin*, in the one-host tick *Boophilus microplus* by RNA interference. *Int J Parasitol* 37: 653–662.
9. Kocan KM, Manzano-Roman R, de la Fuente J (2007) Transovarial silencing of the subolesin gene in three-host ixodid tick species after injection of replete females with subolesin dsRNA. *Parasitol Res* 100: 1411–1415.
10. Almazán C, Blas-Machado U, Kocan KM, Yoshioka JH, Blouin EF, et al. (2005) Characterization of three *Ixodes scapularis* cDNAs protective against tick infestations. *Vaccine* 23: 4403–4416.
11. de la Fuente J, Almazán C, Blouin EF, Naranjo V, Kocan KM (2006) Reduction of tick infections with *Anaplasma marginale* and *A. phagocytophilum* by targeting the tick protective antigen subolesin. *Parasitol Res* 100: 85–91.
12. de la Fuente J, Blouin EF, Manzano-Roman R, Naranjo V, Almazán C, et al. (2007) Functional genomic studies of tick cells in response to infection with the cattle pathogen, *Anaplasma marginale*. *Genomics* 90: 712–722.
13. de la Fuente J, Maritz-Olivier C, Naranjo V, Ayoubi P, Nijhof AM, et al. (2008) Evidence of the role of tick subolesin in gene expression. *BMC Genomics* 9: 372.
14. Kocan KM, Zivkovic Z, Blouin EF, Naranjo V, Almazán C, et al. (2009) Silencing of genes involved in *Anaplasma marginale*-tick interactions affects the pathogen developmental cycle in *Dermacentor variabilis*. *BMC Dev Biol* 9: 42.
15. Zivkovic Z, Torina A, Mitra R, Alongi A, Scimeca S, et al. (2010) Subolesin expression in response to pathogen infection in ticks. *BMC Immunol* 11: 7.
16. Merino O, Almazán C, Canales M, Villar M, Moreno-Cid JA, et al. (2011) Targeting the tick protective antigen subolesin reduces vector infestations and pathogen infection by *Anaplasma marginale* and *Babesia bigemina*. *Vaccine* 29: 8575–8579.
17. de la Fuente J, Moreno-Cid JA, Canales M, Villar M, Pérez de la Lastra JM, et al. (2011) Targeting arthropod subolesin/akirin for the development of a universal vaccine for control of vector infestations and pathogen transmission. *Vet Parasitol* 181: 17–22.
18. Bensaci M, Bhattacharya D, Clark R, Hu LT (2012) Oral vaccination with vaccinia virus expressing the tick antigen subolesin inhibits tick feeding and transmission of *Borrelia burgdorferi*. *Vaccine* 30: 6040–6046.
19. Wan F, Lenardo MJ (2009) Specification of DNA binding activity of NF-kappaB proteins. *Cold Spring Harb Perspect Biol*. 1(4): a000067.
20. Nowak SJ, Aihara H, Gonzalez K, Nibu Y, Baylies MK (2012) Akirin links Twist-regulated transcription with the Brahma chromatin remodeling complex during embryogenesis. *PLoS Genet* 8: e1002547.
21. Lemaitre B (2004) The road to Toll. *Nature Rev Immunol* 4: 521–527.
22. Estrada-Peña A, Jongejan F (1999) Ticks feeding on humans: a review of records on human-biting Ixodoidea with special reference to pathogen transmission. *Exp Appl Acarol* 2: 583–593.
23. Barker SC, Murrell A (2004) Systematics and evolution of ticks with a list of valid genus and species names. *Parasitol* 129: S15–S36.
24. Jones KE, Patel NG, Levy MA, Storeygard A, Balk D, et al. (2008) Global trends in emerging infectious diseases. *Nature* 451: 990–994.
25. de la Fuente J, Estrada-Peña A, Venzal JM, Kocan KM, Sonenshine DE (2008) Overview: Ticks as vectors of pathogens that cause disease in humans and animals. *Frontiers Biosci* 13: 6938–6946.
26. Hill CA, Wikel SK (2005) The *Ixodes scapularis* Genome Project: an opportunity for advancing tick research. *Trends Parasitol* 21: 151–153.
27. Megy K, Hammond M, Lawson D, Bruggner RV, Birney E, et al. (2009) Genomic resources for invertebrate vectors of human pathogens, and the role of VectorBase. *Infect Genet Evol* 9: 308–313.
28. Jongejan F, Nene V, de la Fuente J, Pain A, Willadsen P (2007) Advances in the genomics of ticks and tick-borne pathogens. *Trends Parasitol* 23: 391–396.
29. Nene V (2009) Tick genomics—coming of age. *Front Biosci* 14: 2666–2673.
30. de la Fuente J, Kocan KM, Almazán C, Blouin EF (2007) RNA interference for the study and genetic manipulation of ticks. *Trends Parasitol* 23: 427–433.
31. de la Fuente J, Almazán C, Van Den Bussche RA, Bowman J, Yoshioka JH, et al. (2005) Characterization of genetic diversity in two populations of *Dermacentor andersoni* (Acari: Ixodidae). *Exp Parasitol* 109: 16–26.
32. Labruna MB, Naranjo V, Mangold AJ, Thompson C, Estrada-Peña A, et al. (2009) Allopatric speciation in ticks: genetic and reproductive divergence between geographic strains of *Rhipicephalus (Boophilus) microplus*. *BMC Evol Biol* 9: 46.
33. Krakowetz CN, Lindsay LR, Chilton NB (2011) Genetic diversity in *Ixodes scapularis* (Acari: Ixodidae) from six established populations in Canada. *Ticks Tick Borne Dis* 2: 143–150.
34. Sadusky T, Newman AJ, Dibb NJ (2004) Exon junction sequences as cryptic splice sites: Implications for intron origin. *Current Biol* 14: 505–509.
35. Butler JE, Kadonaga JT (2002) The RNA polymerase II core promoter: a key component in the regulation of gene expression. *Genes Dev* 16: 2583–2592.
36. Fujita T, Nolan GP, Ghosh S, Baltimore D (1992) Independent modes of transcriptional activation by the p50 and p65 subunits of NF-kB. *Genes Dev* 6: 775–787.
37. Peña-Rangel MT, Rodriguez I, Riesgo-Escovar JR (2002) A misexpression study examining dorsal thorax formation in *Drosophila melanogaster*. *Genetics* 160: 1035–1050.
38. DasGupta R, Kaykas A, Moon RT, Perrimon N (2005) Functional genomic analysis of the Wnt-wingless signaling pathway. *Science* 308: 826–833.
39. Sutterwala FS, Flavell RA (2008) Immunology: cascade into clarity. *Nature* 451: 254–255.
40. de la Fuente J, Blouin EF, Manzano-Roman R, Naranjo V, Almazán C, et al. (2008) Differential expression of the tick protective antigen subolesin in *Anaplasma marginale*- and *A. phagocytophilum*-infected host cells. *Ann N Y Acad Sci* 1149: 27–35.
41. Busby AT, Ayllón N, Kocan KM, Blouin EF, de la Fuente G, et al. (2012) Expression of heat-shock proteins and subolesin affects stress responses, *Anaplasma phagocytophilum* infection and questing behavior in the tick, *Ixodes scapularis*. *Med Vet Entomol* 26: 92–102.
42. Wan F, Lenardo MJ (2010) The nuclear signaling of NF-kappaB: current knowledge, new insights, and future perspectives. *Cell Res* 20: 24–33.
43. Beutler B, Moresco EM (2008) Akirins versus infection. *Nat Immunol* 9: 7–9.
44. Havliková S, Ličková M, Ayllón N, Roller L, Kazimirová M, et al. (2013) Immunization with recombinant subolesin does not reduce tick infection with tick-borne encephalitis virus nor protect mice against disease. *Vaccine* 31: 1582–1589.
45. Merino O, Almazán C, Canales M, Villar M, Moreno-Cid JA, et al. (2011) Control of *Rhipicephalus (Boophilus) microplus* infestations by the combination of subolesin vaccination and tick autocidal control after subolesin gene knockdown in ticks fed on cattle. *Vaccine* 29: 2248–2254.
46. Munderloh UG, Madigan J, Dumler JS, Goodman JL, Hayes SF, et al. (1996) Isolation of the equine granulocytic ehrlichiosis agent, *Ehrlichia equi*, in tick cell culture. *J Clin Microbiol* 34: 664–670.
47. Munderloh UG, Jaaron SD, Fingerle V, Leitritz L, Hayes SF, et al. (1999) Invasion and intracellular development of the human granulocytic ehrlichiosis agent in tick cell culture. *J Clin Microbiol* 37: 2518–2524.
48. de la Fuente J, Almazán C, Naranjo V, Blouin EF, Kocan KM (2006) Synergistic effect of silencing the expression of tick protective antigens 4D8 and Rs86 in *Rhipicephalus sanguineus* by RNA interference. *Parasitol Res* 99: 108–113.
49. Lloyd AL, Marshall BJ, Mee BJ (2005) Identifying cloned *Helicobacter pylori* promoters by primer extension using a FAM-labelled primer and GenScan analysis. *J Microbiol Methods* 60: 297–298.
50. Iwasaki H, Chiba K, Uchiyama T, Yoshikawa F, Suzuki F, et al. (2002) Molecular characterization of the starfish inositol 1,4,5-trisphosphate receptor and its role during oocyte maturation and fertilization. *J Biol Chem* 277: 2763–2772.
51. Blouin EF, Saliki JT, de la Fuente J, García-García JC, Kocan KM (2003) Antibodies to *Anaplasma marginale* major surface proteins 1a and 1b inhibit infectivity for cultured tick cells. *Vet Parasitol* 111: 247–260.
52. Lan Q, Fallon AM (1990) Small heat shock proteins distinguish between two mosquito species and confirm identity of their cells lines. *Am J Trop Med Hyg* 43: 669–676.
53. Pahl HL, Baeuerle PA (1995) A novel signal transduction pathway from the endoplasmic reticulum to the nucleus is mediated by transcription factor NF-kB. *EMBO J* 14: 2580–2588.
54. Yin M-J, Yamamoto Y, Gaynor RB (1998) The anti-inflammatory agents aspirin and salicylate inhibit the activity of Ikb kinase-β. *Nature* 396: 77–80.
55. Koutna I, Simara P, Ondračková P, Tesarova L (2012) Flow Cytometry Analysis of Intracellular Protein. In: *Flow Cytometry - Recent Perspectives*, Ingrid Schmid (Ed.), ISBN: 978-953-51-0626-5, InTech, 421–438. Available: <http://www.intechopen.com/books/flow-cytometry-recent-perspectives/flow-cytometry-analysis-and-determination-of-intracellular-protein>.
56. Marchler-Bauer A, Anderson JB, Chitsaz F, Derbyshire MK, DeWeese-Scott C, et al. (2009) CDD: specific functional annotation with the Conserved Domain Database. *Nucleic Acids Res* 37: D205–D210.