Epithelial neoplasia in Drosophila entails switch to primitive cell states

Sumbul J. Khan^{1,2}, Anjali Bajpai¹, Mohammad Atif Alam^{1,3}, Ram P. Gupta, Sneh Harsh, Ravi K. Pandey, Surbhi Goel-Bhattacharya⁴, Aditi Nigam, Arati Mishra, and Pradip Sinha⁵

Department of Biological Sciences and Bioengineering, Indian Institute of Technology Kanpur, Kanpur 208 016, India

Edited by Matthew P. Scott, Stanford University, Howard Hughes Medical Institute, Stanford, CA, and approved April 25, 2013 (received for review July 25, 2012)

Only select cell types in an organ display neoplasia when targeted oncogenically. How developmental lineage hierarchies of these cells prefigure their neoplastic propensities is not yet well-understood. Here we show that neoplastic Drosophila epithelial cells reverse their developmental commitments and switch to primitive cell states. In a context of alleviated tissue surveillance, for example, loss of Lethal giant larvae (Lgl) tumor suppressor in the wing primordium induced epithelial neoplasia in its Homothorax (Hth)-expressing proximal domain. Transcriptional profile of proximally transformed mosaic wing epithelium and functional tests revealed tumor cooperation by multiple signaling pathways. In contrast, lg^r clones in the Vestigial (Vg)expressing distal wing epithelium were eliminated by cell death. Distal *lgl*[−] clones, however, could transform when both tissue surveillance and cell death were compromised genetically and, alternatively, when the transcription cofactor of Hippo signaling pathway, Yorkie (Yki), was activated, or when Ras/EGFR signaling was up-regulated. Furthermore, transforming distal lgl[−] clones displayed loss of Vg, suggesting reversal of their terminal cell fate commitment. In contrast, reinforcing a distal (wing) cell fate commitment in lgl⊤ clones by gaining Vg arrested their neoplasia and induced cell death. We also show that neoplasia in both distal and proximal *lgl*[−] clones could progress in the absence of Hth, revealing Hth-independent wing epithelial neoplasia. Likewise, neoplasia in the eye primordium resulted in loss of Elav, a retinal cell marker; these, however, switched to an Hth-dependent primitive cell state. These results suggest a general characteristic of "cells-of-origin" in epithelial cancers, namely their propensity for switch to primitive cell states.

It is now well-recognized that only specific cell types in a given
developmental hierarchy transform neoplastically when targeted t is now well-recognized that only specific cell types in a given by oncogenic lesions. For example, in mice, upon loss of the APC tumor suppressor, only intestinal crypt stem cells display neoplastic transformation while the transit amplifying cells, which are derived from these crypt stem cells, do not (1). Identification of cancer cells of origin is immensely important because biology of an oncogenically targeted cell is likely to provide clues to cancer pathogenesis (reviewed in ref. 2). It is conceivable that ontogeny, meaning developmental history, plays a crucial role in determining neoplastic propensity of oncogenically targeted cells. However, why different cell types of an organ display distinct neoplastic propensities have not yet been answered. Links between ontogeny and oncogeny (neoplasia) could be interrogated in genetically tractable organisms like the fruit fly, *Drosophila*, which carries a set of highly conserved tumor suppressor genes and display neoplastic growth when mutated (3).

Choice of a model organ to uncover the links between ontogeny and oncogeny is guided by a clarity in our understanding of lineage hierarchy among its different cell types. Epithelial primordia of Drosophila adult organ, the so-called "imaginal discs," are particularly suitable in this respect. The wing imaginal disc, for example, is a composite organ primordium that gives rise to notum (dorsal thorax) and hinge of the adult wing from its proximal domain whereas the wing blade proper is derived from its distal (pouch) domain (Fig. $1 A-C$). Distal (wing) cell fate is specified by the Vestigial (Vg) field selector (4) whereas EGFR signaling (5) and activities of the Teashirt (Tsh) field selector and a homeo-domain transcription factor, Homothorax (Hth) (6, 7), specify proximal cell fate. At early second-larval instar stage—that is, before its proximal-distal lineage separation—the wing primordium displays ubiquitous Hth expression (6, 7). During late-second and earlythird instar larval stages, a signaling axis set up by expression of Notch (N) and its target Wingless (Wg) specify wing cell fate by activating $Vg(4)$ and repressing Hth at the presumptive distal tip of the wing primordium (7). Subsequently, combined activities of Wg and Vg in the distal domain create short-range feed-forward signals in collaboration with Yorkie (Yki) (8), the transcription cofactor of the Hippo signaling pathway (reviewed in ref. 9), thereby recruiting neighboring proximal cells into an expanding distal (wing) field; the distal wing is thus carved out of a Hth-expressing ground state. It has also been seen that when wing cell fate is not specified, for example in the absence of distal Wg signaling, presumptive distal domain assumes a proximal cell state and displays a duplicated proximal element: the notum (10). Furthermore, it has also been seen that proximal domain of the wing primordium can regenerate surgically ablated distal elements; however, a vice versa-type event does not take place (reviewed in ref. 11). These findings mirror progressive restriction of developmental

Significance

Not all cell types in a lineage hierarchy succumb to cancer in the face of an oncogenic lesion. What turns only select cells within an organ into "cells-of-origin" in cancer is an enduring riddle of cancer biology. Here we show that in epithelial primordia of adult wings and eyes of Drosophila, oncogenically mutated cells lose their developmental commitments and, instead, switch to a primitive or progenitor-like cell state. Switch-to-a-primitive-cell-state could thus be a prevalent mechanism of epithelial carcinogenesis and, possibly, an essential characteristic of cancer cells-of-origin.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

Freely available online through the PNAS open access option.

Data deposition: The microarray data have been deposited in Array Express ([http://www.](http://www.ebi.ac.uk/arrayexpress/experiments/E-MEXP-2753/) [ebi.ac.uk/arrayexpress/experiments/E-MEXP-2753/\)](http://www.ebi.ac.uk/arrayexpress/experiments/E-MEXP-2753/) (accession no. [E-MEXP-2753\)](http://www.ebi.ac.uk/arrayexpress/experiments/E-MEXP-2753).

¹S.J.K., A.B., and M.A.A. contributed equally to this work.

²Present address: Department of Cell and Developmental Biology, University of Illinois at Urbana–Champaign, Urbana IL 61801.

³Present address: Molecular Biology Lab, Department of Pharmacology, Piramal Life Sciences, Goregoan (East), Mumbai 400 063, India.

4 Present address: Department of Genetics, Dartmouth Medical School, Hanover, NH 03755.

⁵To whom correspondence should be addressed. E-mail: pradips@iitk.ac.in.

This article contains supporting information online at [www.pnas.org/lookup/suppl/doi:10.](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental) [1073/pnas.1212513110/-/DCSupplemental](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental).

Author contributions: S.J.K. investigated tissue surveillance and tumor cooperation; A.B. investigated links between cell fate regulation and neoplasia; M.A.A. investigated tumor cooperation; R.P.G. carried out transcriptional profiling of mosaic discs; S.H., R.K.P., and S.G.-B. contributed to early stages of work on tissue surveillance and tumor invasion; A.N. and A.M. investigated scrib induced neoplasia; and P.S. designed the investigation and wrote the paper.

Fig. 1. Distinct neoplastic propensities of lg^F clones in proximal and distal wing imaginal disc epithelium. Cartoons of (A) an adult wing and (B) third-instar larval wing primordium. Notum and hinge (green) are parts of the proximal wing; wing pouch (gray) represents the distal wing blade. P→D designates proximal-distal axis. (C) Cartoon of lateral view of the wing imaginal disc epithelium, as seen in an optical section, displaying its proximal (green) and distal (gray) domains. (D–G) Images and data from mosaic discs where *lgl[−]M*⁺ clones (absence of β-gal) are surrounded by cell competition compromised M⁻/M⁺ heterozygous cells. (D) Note that lgl⁻M⁺ clones in the distal domain (blue stars) do not transform neoplastically as revealed by their intact cytoarchitecture (actin, gray). Proximally (yellow stars), however, their altered cytoarchitecture reveals their neoplastic transformation. (D') An $x-z$ optical section along the yellow dotted line in D to reveal intact and altered cytoarchitecture of distal (blue star) and proximally neoplastic clones (yellow star), respectively. (E) Proximal clones (yellow star) shows expression of matrix metalloproteinase-1 (MMP1, red) and breakdown of the basement membrane, marked by collagen-IV-GFP (Coll-IV, green) while the distal clone (blue star) does not; absence of β-gal (gray) marks clone boundary. Higher magnifications of boxed area of E is shown at the far right column to reveal correspondence between disrupted basement membrane (Coll-IV, arrowhead) with MMP1 expression (red) within the clonal area (absence of β-gal, gray). (F–F′′) In older (day 6) mosaic discs, distal (blue star) *lgl[−]M⁺* clones (absence of β-gal, green) were not seen in the apical plane (F), while in the basal plane (F′) these displayed extensive cell death (caspase, red). Optical section along the x–z plane (F′′) through the yellow dotted line shown in F and F' reveals basal extrusion of these distal (blue star), caspase-expressing (red) clones. Note the abundant neoplastic transformation of proximal clones (yellow stars) in all of the three optical planes. A←B indicates the direction of of apical-basal polarity. (G) Plot of total and transformed clonal area of proximal and

distal *Igl[−]M*+ clones in mosaic wing imaginal discs through successive days of clonal growth. (H) *Igl*[−] clones marked by GFP (green) and generated in a cell death compromised H99/+ genetic background. Distal clones (blue star) display intact cytoarchitecture (actin, gray) but proximally (yellow stars) these are neoplastically transformed. (I) Cartoon representation of Igl⊤ clones in mosaic wing disc epithelium in different genetic contexts. Clonal age is shown as days after clone induction. (Scale bars, 100 μm.)

capacities in the proximal-distal lineage hierarchy of the wing primordia.

The eye imaginal disc of third-instar larva presents yet another model organ primordium, where cells of a developmental hierarchy representing distinct stages of retinal development are seen in spatially discrete domains (12). These organ primordia (wing and eye) therefore offer the advantage of spatial resolution of cells with distinct developmental capacities. Furthermore, in these epithelial primordia, cross-talks between a primary tumor (mutant cell) and its tissue microenvironment, namely the neighboring nontransformed cells, could be studied as well. A number of contact-dependent intercellular tissue microenvironmental surveillance (reviewed in refs. 13 and 14) mechanisms (subsequently referred to as tissue surveillance) has been suggested to restrain run away tumor growth in mammalian models. In Drosophila imaginal epithelia as well, tissue surveillance mechanisms, such as cell competition (15–18) and intrinsic tumor suppression (19), have been implicated in elimination of neoplastic mutant clones.

Here we have examined mutant somatic clones of a highly conserved tumor suppressor gene, lethal giant larvae (lgl), in the primordia of adult wings and eyes. Our results reveal that cell survival upon escape from tissue surveillance and gain of cooperative signaling pathways (or transcription factors) propel lgl[−] clones in both these organ primordia to lose their terminal cell fate commitments resulting in their switch to primitive cell states during the course of their tumor progression. In contrast, imposing a terminal cell fate commitment by gain of a field (wing) selector like Vg, for example, arrests neoplastic transformation of *lgl*[−] clones in the wing primordium and induces their large-scale cell death. Given the conservation of fundamental developmental and disease mechanisms, a ground rule of "switch-to-a-primitivecell-state" seen during epithelial neoplasia in Drosophila is therefore likely to be of wider relevance in our understanding of cancer cells of origin and their pathogenesis.

Results

lgl[−] Somatic Clones of Proximal and Distal Wing Primordia Display Disparate Neoplastic Propensities. Cell competition is a tissue-surveillance mechanism for organ homeostasis (reviewed in ref. 20). Somatic clones mutant for the *lgl* tumor suppressor gene [\(Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)B), and also those of other Drosophila neoplastic tumor suppressors, are eliminated from mosaic imaginal disc epithelium by cell competition. Therefore, alleviation of cell competition in lgl[−] clones (and other neoplastic mutants) in a *Minute* (M^{-}/M^{+}) surrounding is permissive for their survival and eventual neoplasia (15–18) through successive days of their clonal growth [\(Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)C). However,

despite growth and survival upon escape from cell competitionmediated elimination, these lgl^- (henceforth depicted as lg^+M^+) clones displayed neoplastic transformation in only the proximal domain (Fig. 1D). Neoplastic transformation of these lgl[−] clones is accompanied by: (i) loss of apico-basal polarity as assayed by mislocalization of either the actin cytoskeleton (18, 21) (Fig. 1D) or apical/lateral membrane markers, such as Discs Lost, Dlt ([Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)D) and, subsequently, (ii) by their invasive transformation, marked by breakdown of their basement membrane (Fig. 1E). Proximal-distal identity of these *lgl*[−]*M*⁺ clones could be readily ascertained by their characteristic epithelial folds (Fig. 1D) or by marking the proximal domain with Hth (Fig. SLE) (6, 7). However, despite a permissive tissue microenvironmental context, distal $lg[−]M⁺$ clones were eliminated from the mosaic disc epithelium, after an initial period of survival, because of cell death (blue star in Fig. 1F), and were basally extruded from the wing epithelium (Fig. 1 $F-F'$). In contrast, neoplasia in their proximal counterparts continued unabated (yellow stars in Fig. 1 $\overline{F}-F'$ ′); bulk of the neoplastically transformed $lg\overline{F}$ clones in mosaic wing imaginal discs therefore is of proximal origin (Fig. 1G). We also noted that basal extrusion of lgl[−] clones and their cell death could not be arrested by expressing a baculovirus inhibitor of apoptosis, namely p35 (22) (lgl[−] UAS-p35 clones in [Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)H). Furthermore, distally extruded lgl[−] clones also did not delaminate as revealed by their intact basement membrane ([Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)G). In these respects, extrusion of distal lgl[−] clones seen here is distinct from mutant clones displaying genomic instability, which often delaminate and turn neoplastic when rescued from cell death by a gain of p35 (23).

Finally, because tissue surveillance-mediated elimination of disadvantaged cells invokes apoptosis, we further argued that a genetic compromise of cell death could rescue lgl[−] clones from tissue surveillance as well. Heterozygosity for a deficiency, Df(3L) H99 (H99/+, in brief), which deletes three closely linked proapoptotic genes—namely, hid, grim, and reaper (24)—is known to compromise cell death. We found that lgl[−] clones generated in a H99/+ genetic background displayed distal survival and proximal transformation (day 5, Fig. 1H); however, in this genetic background as well, such as those in \dot{M} ⁺/M⁻, distal *lgl*⁻ clones displayed basal extrusion after day 5 of their induction ([Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)F). Taken together, these results reveal that neoplastic propensity of lgl[−] clones is linked to their proximal-distal lineage hierarchy, distal being more resistant (Fig. 1I).

Multiple Signaling Pathways Can Cooperate for Proximal Neoplasia.

Neoplastic transformation of somatic clones mutant for a given Drosophila tumor suppressor gene could be triggered by gain of cooperating signaling pathways (25, 26). Selective neoplastic transformations of *lgl*[−] clones in the proximal wing primordium imply activation of such cooperating signaling pathways in this spatial domain. To obtain an unbiased view of these misregulated cellular signaling pathways, we performed genome-wide transcriptional profiling of mosaic wing primordia carrying proximally transformed *lgl*[−] M ⁺ clones in an M [−] $/M$ ⁺ surrounding (for details, see [Fig. S1\)](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1) and compared them with that of control mosaic wing discs that carried M^+ clones, also generated in an $M⁻/M⁺$ sur-rounding ([Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)*I*). Thus, control and test mosaic discs differed primarily in terms of their clonal genotypes, surrounding cells being identical. We noticed twofold up- or down-regulation of 356 genes in test samples compared with control or baseline samples ($P \leq 0.05$) [\(Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)J), besides tight correspondence among the biological replicates [\(Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)K). Heat-maps of genesets of different signaling pathways also displayed broad agreements among their respective replicates (Fig. $S1 L-O$ $S1 L-O$). However, to assess the status of different signaling pathways in these mosaic wing imaginal discs, we went beyond the fold-changes of individual genes and, instead, took a pathway centric approach using gene-set enrichment analysis (GSEA) (27), a statistical tool that uses an a priori-defined set of genes, to ascertain if these

gene-sets are enriched in one biological state as compared with another. We noted enrichment $(P<0.05)$ of several signaling pathways, suggesting their misregulation in proximally transformed lgl[−] clones; these include Hippo, Wnt/Wg, TGF-β, and EGFR pathways (Fig. 2 A–D). Quantitative PCR of representative members of individual signaling pathways correlated with their transcriptional status in the microarrays (Fig. 2A, \ddot{u} , B, \ddot{u} , C, \ddot{u} , and D, \ddot{u}). Finally, we noted that only proximally transformed lgl[−] clones displayed cellular up-regulation of Hippo signaling targets (reviewed in ref. 9), such as, Expanded (Ex) (Fig. 2E) (see also ref. 28) and Wg (Fig. 2F) (see also ref. 29), also a ligand for Wg signaling (Fig. 2F).

To test if these signaling pathways enriched in transcriptional profiles of mosaic discs are functionally relevant for lgl neoplasia, we misregulated these signaling pathways, one at at a time, in lgl[−] clones induced in a wild-type genetic background and asked if these would now override their elimination by tissue surveillance (see [Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)B). We misregulated Hippo signaling in lgl[−] clones by inducing loss of Fat (Ft), a receptor of Hippo signaling; loss of Ft triggers nuclear translocation of Yki, resulting in activation of its target genes (reviewed in ref. 9). We noticed overproliferation and neoplastic transformation of lgl[−] ft[−] double-mutant clones in the proximal domain but distally these were eliminated (Fig. 2A, iii). Similarly, gain of Wg signaling in lgl[−] clones induced by overexpression of Disheveled (Dsh), a down-stream transducer of Wg signaling (30), resulted in proximal neoplasia in lgl[−] UAS-dsh clones, but distally these were eliminated by cell death (Fig. 2B, iii). Proximal neoplasia of lgl[−] clones were also noted following gain of TGF-β or EGFR signaling pathways by expressing their respective activated receptors, namely Thick-veins (Tkv^{OD}) (Fig. 2C, iii) or EGFR (EGFR^{λ top}) (Fig. 2*D*, *iii*) but distally these were eliminated. Of note, individual misregulations of these signaling pathways, however, do not transform wing epithelial cells [\(Fig. S1](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1) *P–[S](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF1)*).

Together, these results thus reveal that lgl-induced neoplasia in proximal domain of wing imaginal disc could be driven by misregulation of signaling pathways that are seen to be perturbed in transcriptional profiles of mosaic discs.

Rescue of Distal lgl[−] Clones from Cell Competition and Cell Death Leads to Their Neoplastic Transformation. Distal elimination of lgl[−] clones prevailed despite cooperation by signaling pathways (Fig. 2) or when cell death ($H99/+)$ or cell competition (M^{-}/M^{+}) was individually compromised (Fig. 1). We thus asked if compromising both cell competition and cell death simultaneously would alleviate distal elimination of $lg[−]M⁺$ clones generated in an $M[−]/$ M^+ ; $H99/+$ genetic context. Indeed, in such a context, where both cell competition and cell death are compromised, distal $lg[−]M⁺$ clones were transformed (day 7, Fig. 3A) although onset of their neoplasia was substantially delayed [\(Fig. S2](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF2)A).

In light of the above results, it thus appeared surprising that misregulation of most signaling pathways, particularly Hippo, which promotes growth and cell competition $(9, 31)$, failed to override distal elimination of lgl[−] clones (Fig. 2A, iii). It is, however, likely that the consequences of loss of Ft in distal and proximal domains of the wing disc were not similar, presumably because of its cross-talks with locally active morphogen signalings (32, 33). To overcome this limitation of dissimilar outcomes of loss of Ft in these two developmental domains, we overexpressed Yki in lgl[−] clones under the ubiquitously active promoter of the tubulin gene. Indeed, unlike *lgl[−] ft[−]* clones (Fig. 2*A iii*), Yki overexpressing *lgl[−]* clones (lgl[−] UAS-yki) displayed massive overgrowth over the entire wing imaginal disc because of fusion of these fast-growing clones that obscured precise resolution of their proximal-distal origins [\(Fig. S2](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF2)B). We thus reduced the duration of heat shock from 30 min to 4 min so as to induce only few dispersed clones to help resolve their proximal-distal spatial distribution. These discrete lgl[−] UAS-yki clones, induced by a shorter heat pulse, were seen in both distal and proximal domains (Fig. $3 B$ and C). A closer examination revealed early signs of neoplasia in only some of these Fig. 2. Recruitment of multiple signaling pathways during proximal wing neoplasia. (A–D) Transcriptional status of Hippo (A), Wg (B), TGF-β (C), and EGFR (D) signaling pathways and their functional cooperation for lgl[−] neoplasia. (A, i–D, i) GSEA plots display enrichment of these pathways at $P < 0.05$; y axis displays enrichment score (ES) and x axis displays ranked order of genes in descending order of their fold-changes in lgl[−] vs. Wild-type; (A, ii–D, ii) Quantitative real-time PCR of representative members of the pathways show their up-regulation in lgl[−] mosaics. Error bars represent SEM. (A, iii–D, iii) Functional cooperation of individual signaling pathways for lal[−] neoplasia; n represents the number of mosaic discs examined in each case. (A, iii) Functional cooperation of Hippo signaling: mosaic wing discs bearing ft[−] or lgl[−] ft[−] clones (loss of GFP). Distal (blue star) and proximal (yellow star) twin spots in boxed areas are shown at higher magnification in the Center; the far right panel shows their cartoon representation. Note that proximal and distal ft[−] clones outgrow their wild-type twins (dark green, red line). In contrast, distal lgl[−] ft[−] clones are conspicuous by their absence (question mark, blue star), but their wild-type twin-spot grew as expected. Proximal *lgl[−] ft*[−] clones display neoplastic transformation (actin, gray) and outgrow their wild-type twins (dark green). *lgl*[−] clones (GFP, green) overexpressing *UAS-dsh (B, iii), UAS-tkv^{QD} (C, iii),* and $UAS-EGFR^{\lambda top}$ (D, iii). Note that proximally all these clones (yellow stars) are transformed (actin) but distally (blue stars) these are rarely seen in the apical optical plane; however, basally these were invariably

seen to be extruded. Mosaic disc in B, iii is marked for cell death (arrows, caspase, red), which are seen in the basal plane. (E) Selective up-regulation of Expanded, Ex (red) and (F) Wg (red) in proximally transformed lg/⊤ clones (absence of green, yellow stars). Note that distally, where lg/⊤ clones are not neoplastically transformed, Wg expression too was not altered in both 4- and 6-d-old clones (blue arrowheads). [Scale bar (applies to all panels), 100 μm.]

distal lgl[−] UAS-yki clones on the day 3 of clonal growth (Fig. 3B) although by this time many of their proximal counterparts were neoplastically transformed (Fig. 3B). Distal lgl[−] UAS-yki clones thus show a delayed onset of neoplasia. By day 4 of clone induction, however, the majority of the lgl[−] UAS-yki clones in both proximal and distal wing primordia were neoplastically transformed (Fig. 3C), while those that did not transform continued to display characteristics of nontransformed epithelial cells, namely, densely packed cells and compact nuclear morphology, as revealed by the nuclear GFP marker, besides their intact actin cytoarchitecture (arrow in Fig. 3C).

In summary, distal lgl[−] clones display neoplastic transformation (Fig. 3D) when their elimination is arrested by simultaneously compromising cell competition and cell death or when provided with a substantial advantage in cell proliferation or cell competition by gain of a cooperating partner, like Yki.

Loss of Wing Cell Fate During Neoplasia of Distal IgI[−] Clones. Proximal-distal disparity in neoplastic propensity could be linked to their respective lineage commitments. We thus asked if distally transforming lgl[−] clones retain their cellular memory by examining several exemplars of wing cell fate specification: namely, Vg (4), Distalless (Dll) (34), and Nubbin (Nub) (35). We considered *lgl[−] UAS-yki* clones ideal to examine this question because distally these are not eliminated (Fig. 3) and gain of Yki alone does not alter distal expression of these markers (Fig. 4 A– C). We noticed that down-regulation of Vg expression in lgl[−] UAS-yki clones sets in before the onset of neoplastic transformation (actin): that is, on the day 3 of clone induction (Fig. 4D). Furthermore, Vg is extinguished by the day 4 of clone induction ([Fig. S2](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF2)C), coinciding with neoplastic transformation of lgl[−] UAS-yki clones. Likewise, expressions of Dll (Fig. 4E) and Nub (Fig. 4F) were extinguished in distally transformed *lgl*[−]

UAS-yki clones (day 4) as well. We also noticed loss of distal cellfate marker, Dll, in mutant clones of another tumor suppressor, Scribble (Scrib) (21) [\(Fig. S2](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF2)D); loss of cell fate could thus be prevalent during distal neoplasia.

The Yki transcription cofactor, in collaboration with one of its binding partner, Scalloped (Sd), a TEA-domain transcription factor, positively regulates vg transcription (36). Furthermore, feedforward recruitment of Vg expression during expansion of the presumptive wing blade region because of Wg signaling too is Ykimediated (8). Thus, loss of a terminal cell fate commitment in distal lgl[−] UAS-yki clones (Fig. 4D) is unlikely to be because of gain of a cooperative partner, like Yki; instead, it is likely to be a latent characteristic of transforming lgl[−] clone. To test this possibility, we examined expression of Vg in $lg[−]M⁺$ clones generated in an $M[−]/$ M^+ ; H99/+ genetic context where these display distal survival and neoplasia (Fig. 3A) without the advantage of a forced gain of a cooperating partner, like Yki (Fig. 3 B and C). Early stage $lg\sqrt{M}$ + clones (day 5 after clone induction, larval growth being slow in this genetic context) did not display a loss of Vg and were characterized by wiggly clonal borders (Fig. 4G), thereby revealing their normal cell-cell adhesion with the surrounding (also Vg-expressing) cells. Older lgl[−]M⁺ clones (day 7 after clone induction) (Fig. 4H) were of two types: (i) those that displayed early signs of down-regulation of Vg and showed smooth clone borders (yellow box, Fig. 4H), suggesting their cell fate changes (37) ; and (ii) those that were neoplastically transformed (blue box, Fig. 4H) and displayed complete loss of Vg, besides smooth clone borders. Loss of distal cell fate in lgl[−] clones therefore is a latent characteristic that manifests under conditions permissive for their survival.

Imposition of Wing Cell Fate in *Igl*[−] Clones Arrests Neoplasia and **Induces Cell Death.** In the wing primordium (Fig. $5A$), Vg is regulated by inputs from two axial signaling cascades: N-Wg signaling

Fig. 3. Distal neoplasia of lgl[−] clones under conditions that override their tissue microenvironmental surveillance and cell death. (A) lgl[−] clones (absence of β-gal, green) in a genetic context compromising both cell competition (M⁻/ M⁺) and cell death (H99/+) display neoplastic transformation (actin, red) in both proximal (yellow star) and distal (blue star) domains. (B and C) lql[−] UASyki clones (GFP, green) induced by a short pulse of heat shock (4 min). On day 3, only proximal clones are transformed (yellow star #1), but distal clone (blue star #2) display only early signs of actin reorganization (actin, red) (B). On day 4, proximal clones displayed neoplasia (yellows star) but distally one of the two large clones is neoplastically transformed (blue star), while other is not (arrow head) (C); optical sections along the x–z plane through the dotted line across transformed (blue) and nontransformed (pink) clones are shown at the bottom right panels to reveal altered F-Actin cytoarchitecture in only the former. (D) Cartoon representation of transformed lqΓ clones in both distal and proximal territories under conditions of reduced tissue surveillance and cell death. $n =$ number of discs examined. Scores for distal (d) or proximal (p) clones showing neoplasia are displayed in the panel. A←B indicates apicalbasal polarity. [Scale bar (applies to all panels), 100 μm.]

inputs from the Dorsal-Ventral (D/V) axis and decapentaplegic (Dpp) signaling inputs from the Anterior-Posterior (A/P) axis. N signaling activates a D/V boundary enhancer of $vg(4)$, which is then maintained by Wg. Expression of Cut (Ct) (Fig. 5A), a neuronal cell fate marker on the D/V boundary (38), is also triggered by N signaling and is maintained by its cross-talks with Wg signaling (39). Furthermore, a combined N $(4, 40)$ and Wg $(35, 39)$ signaling and activities of Yki (8) positively regulate Vg expression in the wing pouch domain through its quadrant enhancer (Q-vg). Finally, secreted Dpp signaling from the A/P axis too positively regulates the Q -vg enhancer (4).

Loss of Vg in neoplastic lgl[−] clones (Fig. 4D) could thus be fallouts of down-regulation of these two A/P and D/V signaling inputs. To test these possibilities, we first examined expression of nuclear targets of these two axial signaling systems in distally transformed lgl[−] UAS-yki clones. Spalt is a short-range Dpp signaling target and its expression is compromised when Dpp signaling is down-regulated along the A/P axis (41). Expression of Spalt, however, was not affected in lgl[−] UAS-yki clones [\(Fig.](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF2) S_{2F}). Thus, apparently, Dpp signaling is not compromised in neoplastic lgl[−] clones. In contrast, the N-Wg signaling target, Ct, is down-regulated in lgl[−] UAS-yki clones ahead of their neoplastic transformation (boxed area, Fig. 5B) and is extinguished when these are neoplastically transformed (star, Fig. 5B). These results therefore suggest a down-regulation or perturbation in N-Wg signaling in distally transforming lgl[−] UAS-yki clones. We thus sought to test if a gain of N or Wg signaling would restore expression of Vg in distal $lg[−]$ clones. Gain of N, by expressing its activated receptor (N^{intra}) (4), in distal wing hindered clonal growth (Fig. $S2G$). This finding was not surprising because a high threshed of N signaling in developing wing disc at the D/V boundary corresponds to a zone of nonproliferating cells (4, 42). N activation in distal lgl^- clones ($lgl^- UAS-N^{intra}$) resulted in their elimination as well; however, proximally these were transformed [\(Fig.](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF2) $S2H$) and could ectopically turn on its targets, like Dll (Fig. [S2](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF2)I). Clones displaying gain of Wg signaling (UAS-dsh) grew well in both proximal and distal domains (Fig. 5C). Furthermore, distally these inevitably displayed up-regulation of Vg (blue arrowheads, Fig. 5C) but, in contrast, proximally only about 10% of these clones showed ectopic gain of Vg (yellow arrowhead, Fig. 5C) (see also ref. 35). Gain of Wg signaling in distal lgl[−] clones (lgl[−] UAS-dsh), however, resulted in their elimination by

Fig. 4. Loss of cell fate during distal neoplasia of lgl[−] clones. (A–C) Ykiexpressing (UAS-yki, green) clones do not alter distal expression of Vg (A), Dll (B), or Nub (C) but (D–F) their lgl[−] UAS-yki counterparts (GFP, green) do (blue stars). Distal *lgl[−]* UAS-yki clones shown in D display intact cytoarchitecture (actin), whereas those shown in E and F are neoplastically transformed (actin). (G and H) Expression of Vg in lgl[−]M⁺ clones (loss of GFP, green) induced in a cell competition (M[−]/M⁺) and cell death (H99/+) compromised context. Boxed area in a mosaic distal wing imaginal disc, dissected day 5 after clone induction highlights wiggly clonal borders (G); Vg (red) expression in these clones, however, is not affected (shown at higher magnification). At a later stage of their growth, such clones display both neoplastically transformed (blue star) and nontransformed (yellow arrows) territories (H). Yellow boxed area is shown at a higher magnification at far right column to reveal clonal territory (yellow dotted lines), which are not transformed yet (actin, gray) but show down-regulation of Vg (arrowhead) and smooth clone border. Blue boxed area shown at a higher magnification (shown at the far right panel) display neoplasia (Actin, gray), loss of Vg (blue star), and smooth clone border (dotted blue line). Scores of distal (d) clones are also shown. (Scale bars, $100 \mu m$.)

PNAS PLUS PNAS PLUS

Fig. 5. Distal cell fate is incompatible with neoplastic cell state. (A) Wild-type third-instar wing imaginal disc displaying expressions of a N target, Ct (red) along the dorsal-ventral wing margin, and Vg (green) in distal wing (pouch domain); expression of Hth (purple) marks the proximal wing. (B) In a neoplastically transformed (actin, gray, blue star) distal lgl[−] UAS-yki clones (green) expression of Ct (red) is extinguished. Boxed area marks a nontransformed distal clone (actin, gray), which is shown at a higher magnification on the right to reveal early signs of loss of Ct (between the arrowheads). (C and D) Clones displaying gain of Wg signaling (UAS-dsh, green) activate Vg in distal (blue arrowheads) and in select proximal (yellow arrowhead) clones (C). In contrast, lgl[−] UAS-dsh (green) clones are not seen in the distal wing (marked by Vg, red, D); proximally (outside the domain of Vg, red, D) these are neoplastically transformed, but fail to display gain of Vg, unlike their UAS-dsh counterparts (C). (E-H) Gain of Vg (UAS-vg, green) does not affect growth or survival in distal (blue star) or proximal (yellow star) domains of the wing primordium (E). However, Vg-expressing lgl[−] clones (lgl[−] UAS-vg, green), are distally eliminated because of cell death (caspase, red, F), but those that displayed proximal survival were not transformed (G). Higher magnification of boxed area in F and G is shown at the right and x-z sections over the dotted line are shown at the far right. Apical-Basal (A←B) orientation in the x–z sections is shown. Note the dispersed distribution of the Igl[−] UAS-vg clones (GFP, green) in the periphery of this proximal clone (marked by yellow arrowhead, G) and their basal extrusion as revealed in the x-z section. Nonextruded lgl[−] UAS-vg cells at the center of the clone are not transformed, as revealed by their intact cytoarchitecture (actin). (H) lgl UAS-vg; UAS-yki (green) clones display cell death (caspase, red) both distally (blue star) and proximally (yellow star). Higher magnifications of proximal clones (marked by blue box) are shown at the bottom panel to reveal densely packed cells and their intact cyto-architecture (actin). (I and J) Schematic presentation of the outcome of gain of Wg signaling (I) or the Vg transcription factor (J) in Igl[−] clones. Note that distally, where Wg signaling induces Vg expression (see C, above), Igl[−] UAS-dsh clones are eliminated while those that are proximally transformed do not activate Vg (see D, above). Gain of Vg in Igl[−] clones, however, inevitably induces distal elimination while proximally these remain growth-compromised and die extensively (see F-H). Scores for distal (d) or proximal (p) clones showing a given phenotype (loss or gain of a marker or neoplasia) are displayed in respective panels. [Scale bar (A, B, and E-J), 100 μm, (C and D), 200 μm.]

the day 4 of clone induction (Figs. $2B$, *iii* and $5D$) following an initial and transient period of their survival when these showed gain of Vg [\(Fig. S2](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF2) J and K). In contrast, proximal lgV ⁻ UAS-dsh clones were extensively transformed (Fig. 2B, iii); however, none of these displayed gain of Vg (Fig. 5D), unlike their control UAS-dsh counterparts (Fig. 5C). It is therefore likely that proximal lgl[−] UAS-dsh clones displaying gain of Vg are preferentially eliminated (indeed, on day 2 after clone induction, proximal lgl[−] UAS-dsh clones displayed activation of Vg; see [Fig. S2](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF2)K). Cell survival and neoplasia in distal lgl[−] clones could thus be linked to loss of Vg. Conversely, signaling pathways which trigger expression of Vg may block neoplasia. To directly test this possibility we examined the consequences of an outright gain of Vg in lgl[−] clones. Control Vg-expressing clones (UASvg) grew well in both distal and proximal domains (Fig. 5E). In contrast, distal *lgl[−] UAS-vg* clones showed rapid elimination; thus, these were rarely recovered (6 of 24 mosaic discs) after day 4 of clone induction and appeared as small clusters of GFP-expressing cells displaying cell death and basal extrusion (Fig. 5F). Proximal survival of lgl[−] UAS-vg clones (Fig. 5G), on the other hand, appeared marginally better than their distal counterparts (Fig. 5F). However, these too displayed cell death [\(Fig. S2](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF2)L) and basal extrusion (Fig. 5G). Of note, in none of these $lgl^- UAS$ -vg clones (n = 20) examined in 14 mosaic discs were neoplastically transformed (Fig. 5G).

We further examined the consequences of gain of Yki in lgl[−] UAS-vg clones. However, despite gain of Yki, distal lgl[−] UAS-vg UAS-yki clones were poorly recovered (10 of 20 discs) and showed extensive cell death (Fig. 5H). Proximally, lgl[−] UAS-vg UAS-yki clones displayed a somewhat improved survival than their distal counterparts (Fig. 5H). However, these too displayed high caspase activity (Fig. 5H) that appeared to offset their growth advantage because of gain of Yki. Most significantly, none $(n = 30)$ of the proximal *lgl[−] UAS-vg UAS-yki* clones, of the 17 mosaic discs examined, were transformed till the day 4 of clone induction (Fig. 5H) and displayed dense cell packing and compact nuclear morphology, unlike their lgl[−] UAS-yki counterparts, which were inevitably transformed at this clonal age (see Figs. 3 and 4).

In summary, distal gain of Wg signaling or Vg transcription factor in lgl[−] clones produced identical outcome: both induced

Fig. 6. Hth-independent neoplasia in lg^r clones in wing primordium. (A–C) lgl[−] UAS-yki clones (GFP, green) at different clonal age; Dll (A and C) and Vg (B) mark the distal domain. Note the progressive loss of these markers in distal lgl[−] UAS-yki clones (blue star) and gain of Hth (gray) on day 4 (B) and day 5 (C), but not on day 3 (A) of clone induction. Yellow star in C marks a proximal clone. (D) A proximal lgl[−] UAS-yki; UAS-hth-RNAi (GFP, green) clone (yellow star) displaying loss of Hth (gray); it is, however, neoplastically transformed such as its distal (blue star) counterpart (actin, purple). $(E-G)$ Gain of Ras/EGFR signaling in lg/[−] clones represses distal cell fate and induces lgl neoplasia. lgl⁻ UAS-ras^{V12} clones (GFP, green, E) on day 3 of clone induction repress Vg (red) and display neoplasia (actin purple, F); boxed areas in these images are shown at a higher magnification on their respective right panels. (G) Schematic representation of proximal and distal neoplasia in *lgl* clone upon gain of Ras/EGFR signaling. Scores displayed in A-C, E, and F are for distal clones while scores in D are for for distal (d) or proximal (p) clones. (Scale bars, 100 μm.)

cell death (Fig. 5 I and J). Proximally, however, these displayed two distinct outcomes: only those lgl[−] UAS-dsh clones were transformed neoplastically, which did not gain Vg, while their *lgl*[−] UAS-vg counterparts failed to transform neoplastically and displayed cell death (Fig. $5 I$ and J). Imposition of wing cell fate (gain of Vg, for example) in *lgl*[−] clones therefore blocks their neoplastic transformation, induction of cell death apart.

Neoplastic Cell State in the Wing Imaginal Disc is Hth-Independent. Roles of field selectors in cell fate determination (43) can be likened to binary switches: an ON state of Vg, for example, either because of its ectopic activation (4) or developmental regulation (44), imposes a wing cell fate by repressing or overriding a nonwing cell state. Loss of Vg (OFF state) in distally transforming lgl[−] clones would thus entail their reversal to a nonwing cell state. In order to test if distal lgl[−] clones assume a proximal-like cell state during clonal growth, expression of Hth, which is required for maintenance of proximal cell fate (6, 7), was examined. On the day

was more pronounced in clones closer to the hinge region. Older (day 5) and distally neoplastic lgl[−] UAS-yki clones (see Fig. 3D) displayed uniform gain of Hth (blue star, Fig. 6C), matching their proximally transformed counterparts (yellow star, Fig. 6C). It is thus likely that distally transformed lgl[−] clones, upon loss of Vg, acquire an Hth-dependent proximal-like cell state. In other

words, Hth might cooperate for distal neoplasia of lgl[−] UAS-yki clones. We thus examined the consequences of misregulation of Hth in lgl[−] clones. lgl[−] clones expressing Hth (lgl[−] UAS-hth), however, are mostly eliminated from distal wing, although proximally these were neoplastically transformed ([Fig. S3](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3)B). Hth therefore does not cooperate for distal neoplasia; this appears consistent with the observation that distal gain of Hth does not repress Vg ([Fig. S3](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3)A) (7). Expectedly, down-regulation of Hth expression in lgl[−] clones (lgl[−]UAS-yki UAS-hth-RNAi clones) did not affect distal neoplasia (blue star, Fig. 6D). However, surprisingly, these clones displayed proximal transformation, despite loss of Hth (yellow star, Fig. 6D). These results reveal that distal gain of Hth in $lg-$ clones (Fig. 6 B and C), or its persistent expression in proximal lgl[−] clones (Fig. 6C), are functionally redundant for neoplasia in the wing primordium. By extension, both distally and proximally transformed lgl[−] clones switch to cell states, which are Hth-independent.

3 of clone induction—that is, prior to neoplastic transformation (Fig. 3A)—distal lgl[−] UAS-yki clones did not display a gain of Hth (Fig. 6A). On the day 4, only half of the distal $l\dot{gl}$ ⁻ UAS-yki clones displayed low-to-moderate gain in Hth expression (Fig. 6B), which

Ras/EGFR Signaling Represses Vg and Induces Distal Neoplasia. Given the obligatory requirement of loss of Vg for lgl neoplasia (Figs. 4 and 5), it is likely that signaling pathways that negatively regulate Vg too could cooperate for distal neoplasia of lgl[−] clones. The Ras/ EGFR signaling pathway, which regulates the choice of proximal cell fate in the developing wing primordium, might belong to this category of cooperative signaling partners for neoplasia in *lgl*[−] clones (5, 10); gain of EGFR signaling, for example, down-regulates Vg ([Fig. S3](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3)G). However, because a gain of EGFR signaling in lgl[−] UAS-EGFR^{λ top} clones did not induce distal neoplasia (Fig. 2D, iii), we sought to examine the consequences of gain of an activated Ras kinase (Ras^{V12}), which phenocopy homeotic functions of Ras/ EGFR signaling in the wing primordium (5, 10). We thus examined the consequences of distal gain of Ras^{V12} in discrete *lgl[−] UAS* ras^{V12} clones induced by a short pulse of heat shock (4 min) (Fig. $6E$). These clones displayed repression of Vg (Fig. $6E$), like their $UAS-ras^{V12}$ counterparts (5). Furthermore, these clones were neoplastically transformed at a pace (day 3 of clone induction) matching that of their proximal counterparts (Fig. 6F). Control Ras^{V12}-expressing clones (UAS-ras^{v12}) on the other hand tend to sort out from their neighbors [\(Fig. S3](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3) C–[E](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3)), presumably as a consequence of a switch in their cell fate (37), but otherwise remained monolayered (45) and displayed intact cytoarchitecture [\(Fig. S3](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3)C).
Thus, oncogenic cooperation of Ras^{V12} for distal neoplasia as seen here, and also those reported earlier (31), is likely to be aided by its ability to induce Ras/EGFR-mediated reversal of cell fate commitments (5) (summarized in Fig. 6G). Consistent with such a role of Ras/EGFR signaling for distal neoplasia, we further noticed that lgl[−] UAS-EGFR^{λtop} clones too displayed distal neoplasia at an elevated growth temperature (29 °C) ([Fig. S3](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3)H); a stronger Gal4 activity at 29 °C, may therefore elevate the level of $EGFR^{\lambda top}$, which is presumably required to for its distal cooperation during neoplasia of lgl[−] clones. In contrast, lgl[−] UAS-tkv^{QD} or lgl[−] UAS-dsh clones did not induce distal neoplasia at an elevated (29 °C) growth temperature [\(Fig. S3](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3) I and J), consistent with the fact that gain of TGF-β or Wg signaling in distal wing would reinforce Vg cell fate, which is incompatible with a neoplastic cell state (Fig. 5).

It may be further noted that although Ras^{V12}-expressing clones display distal gain of Hth [\(Fig. S3](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3)D), presumably because of a loss of Vg, proximally it repressed Hth in the hinge domain [\(Fig. S3](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3)E).

Fig. 7. Neoplastic cells of the eye primordium switch to an Hth-dependent progenitor-like cell state. (A) Cartoon of third-instar larval eye imaginal disc displaying three distinct stages of retinal development. The anterior-most progenitor cells express Hth (green) followed by a zone of Dac-expressing committed cells (peach) preceding the morphogenetic furrow (MF, arrowhead), which is succeeded by differentiated retinal cells (Elav, red). (B) An eye imaginal disc displaying domains of Elav and Hth expression. (C) scrib[−] retinal clone (loss of GFP, blue star). Boxed area in C is shown at a higher magnification on the right to reveal loss of Elav (red) in the clone and its neoplastic transformation (actin). (D and E) scrib[−]M⁺ clones (absence of GFP, blue star) generated in a M^{−/}M⁺ surrounding. Boxed area in D is magnified on its right panels to display gain of Hth (red) on day 3 of clone induction (D). Expansion of clonal area on day 5 of clone induction results in near ubiquitous expression of Hth in the mosaic eye primordium. (F-J) Switch to a progenitor-like cell state drives neoplasia in the eye primordium. An UAS-hth clone (F, green) in the eye primordium; boxed area is shown on the right panels to reveal loss of Elav (red, arrow) corresponding to the domain of gain of Hth (GFP, green, arrow). scrib[−] UAS-hth RNAi clones (G, green, yellow arrows) do not lose Elav (red) and display characteristic ommatidial cytoarchitecture (actin, gray) and, thus, are no longer neoplastically transformed. In lgl[−] M⁺ clones (loss of GFP, green), generated in M[−]/M⁺ surrounding, endogenous domains of Elav (red, H) or Hth (red, I) are not altered. In contrast, Iq Γ UAS-hth clones (green, blue star, J) display loss of Elav (red) and neoplastic transformation (actin, blue). The central large and neoplastically transformed clone (blue star) has popped out of the plane of the rest of the eye disc. The clone on the margin of the eye disc (yellow star) represents neoplastic transformation of margin cells. [Scale bar (for all panels), 100 μm.]

Furthermore, massively overgrown lgl[−] UAS-ras^{V12} clones spanning both distal and proximal domains display down-regulation of Hth ([Fig. S3](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=SF3)F). Thus, RasV12-mediated lgl[−] neoplasia in the wing primordium too, like those mediated by Yki (Fig. 6D), is Hthindependent.

Neoplastic Cell State in the Eye Primordium Is Hth-Dependent. Cell fate switch seen during neoplasia in the wing primordium could be a general feature of tumor progression in Drosophila epithelia. We examined this possibility in the eye imaginal disc where progenitors of the adult ommatidia (eye) and their derivatives are seen in discrete spatial domains. These include a domain of Hth-expressing eye progenitor cells, succeeded by a population of preproneural cells that are committed to retinal cell fate and marked by expression of transcription factors, like Dachshund (Dac) and Eyes absent (Eya), which is then followed by differentiated retinal cells marked by Elav (Fig. $7A$ and B) (12).

Previously, it has been reported that *lgl*[−] retinal clones neither extinguish Elav nor transform neoplastically (18). Thus, we sought to examine the consequences of loss of scrib (21). In the eye disc, unlike their lgl[−] counterparts, scrib[−] clones displayed loss of Elav (red, Fig. 7C), and were neoplastically transformed (actin) (Fig. 7C) (16, 25, 46). Importantly, scrib⁻ M^+ eye clones (in an M^-/M^+ surrounding) displayed an early gain of the retinal progenitor cell marker, Hth (day 3, Fig. 7D), which turned ubiquitous as the clonal territory expanded in the eye primordium (day 5; Fig. 7E). Previously, it has been shown that Hth expression arrests retinal cell differentiation by blocking Elav expression (47), as is evident in UAS-hth clones (Fig. 7F). Reversal in retinal cell differentiation and neoplasia in scrib[−] clones could thus be linked to their gain of Hth. If true, loss of Hth in *scrib*[−] eye clones would restore Elav expression. Indeed, scrib[−] UAS-hth-RNAi eye clones displayed expression of Elav (red, Fig. 7G) and restoration of their characteristic retinal cell architecture (actin, Fig. 7G). Given these findings in the *scrib*[−] clones (Fig. 7 D and E), we further argued that a persistent state of retinal differentiation in lgl[−] clones in the eye primordium (Elav, Fig. 6H) (18, 47) could be linked to their inability to recruit Hth activity (Fig. 6I), unlike their scrib[−] counterparts (Fig. $7 D$ and E). Conversely, gain of Hth should then render the retinal *lgl*[−] clones neoplastic. Indeed, gain of Hth in *lgl*[−] clones in the eye primordium (lgl[−] UAS-hth clones) resulted in their neoplastic transformation (Fig. 7J), reminiscent of their scrib[−] counterparts (Fig. 7C).

Taken together, these results reveal that neoplasia in an oncogenically targeted eye primordium sets in upon a cooperative gain of Hth that blocks or reverses their retinal cell differentiation, thereby switching to an Hth-dependent primitive/progenitor-like cell state.

Discussion

Switch-to-a-Primitive-Cell-State and Escape from Cell Death: A Framework for Drosophila Epithelial Neoplasia. Hallmarks of cancer, such as limitless replicative potential, escape from apoptosis or self-sufficiency in growth signals (reviewed in ref. 48) suggest subversion of normal developmental regulatory mechanisms during neoplastic growth. Developmental contexts that enable such neoplastic transformations, however, are not yet fully understood. By examining the consequences of loss of tumor suppressor genes in wing and eye primordia of Drosophila, we have herein asked if ontogeny of a mutated cell could foretell its neoplastic propensities. Although these two organ primordia display disparate developmental origins and their cell fates are specified by distinct sets of transcription factors, our results reveal a common paradigm of neoplasia: namely, their obligatory reversal to progenitor-like or primitive cell states, characterized by two hallmarks: first, reversal of their terminal cell fate commitments (loss of Vg and Elav in wing and eye, respectively), and second, gain/activation of transcription factors that are linked to primitive or progenitor-like

Fig. 8. Both lineage-committed and differentiated cells switch to primitive cell states during carcinogenesis. Cartoons of (A) wing and (B) eye discs displaying the cells which are committed to proximal (Hth), distal (Vg) wing, or retinal (Dac) cell fates or differentiated into retinal cells proper (red, Elav). Neoplastic transformations (irregular mass of cells) of these cells entail the following: first, loss of transcription factors that display their cell commitment (Vg) or differentiation (Elav) and, second, switch to a primitive/progenitor-like cell state (light green). Primitive cell state assumed by transformed lgl[−] clones in the wing is Hthindependent (A) while in the eye discs it is Hth-dependent (B).

developmental ground states (Fig. 8). Progression of an oncogenically targeted cell to a cancerous state therefore depends on its ability to recruit cellular partners, which promotes switch to a developmentally primitive cell state: for instance, by a gain of Ras/EGFR signaling in the distal wing or by a gain of a transcription factor, like Hth, in the eye primordium.

Carcinogenesis is also proposed to be an outcome of essentially two broad classes of cooperating events: first, oncogenic lesions that provide proliferative drive in mutated cells and, second, obligatory cellular cooperative events, such as genetic lesions or epigenetic changes in cooperating partners that ensure survival of the oncogenically targeted cells. Combination of these two cellular events culminate in escape of oncogenically targeted cells from cell death and, eventually, neoplasia (49, 50). We note that neoplasia of lgl[−] clones does not set in unless conditions permissive for their escape from cell death are available. Furthermore, loss of a terminal cell fate commitment (loss of Vg) is an essential requirement for escape from cell death as well, and thereby, neoplasia. Gain of Vg induces cell death in lgl[−] clones, both in the endogenous domain of its expression in distal wing (Fig. $5F$) and outside: that is, proximal wing (Fig. $5G$). These findings underscore primacy of cell fate reversal among the obligatory cellular events that cooperate for survival of the neoplastic cells. Loss of terminal cell fate and reversal to a primitive cell state are thus two intertwined cellular events that dictate cell survival and transformation of oncogenically targeted cells.

Primitive Cell States, Cancer Cells-of-Origin, and Context-Dependent

Tumor Cooperation. Cells targeted by a neoplastic lesion are referred to as "cells-of-mutation," and those that finally display neoplastic transformation are called "cells-of-origin." All cells-of-mutation do not turn into cells-of-origin in cancer (2), an enduring reason for cell type-specific carcinogenesis. Cells-of-origin in cancer often tend to display stem cell-like characteristics; for example, crypt stem cells in intestinal epithelium that give rise to colorectal cancer (reviewed in ref. 2). Conversely, cells-of-mutation in breast epithelial cells tend to de-differentiate into a stem cell state before turning into cellsof-origin in breast cancer (51). Furthermore, in breast carcinoma, luminal cells with BRCA-1 mutation undergo basal-like cell fate switch before their neoplastic transformation (52). These and other examples notwithstanding, it is not yet clearly established if cells of origin in cancer are inevitably progenitor cell-like in nature or if lineage-committed or differentiated (nonprogenitor) cells too could turn into cancer cells-of-origin through reversals in their cell states. While our observations supports the latter alternative, namely switch in cell fates during tumor progression (Fig. 8), we also note that strategies for reversal in cell fate commitments are distinct in different developmental contexts. In the distal wing, for example, cellular reprogramming, exemplified by loss of Vg (Fig. 4, also see ref. 53), is a latent characteristic of $lg\bar{t}$ clones that appears to be hastened by down-regulation of N-Wg signaling, or possibly by a concomitant gain of Ras/EGFR signaling. In the eye primordium, switch-to-a-primitive-cell-state is triggered by recruitment of Hth (Fig. 7), a regulator of progenitor cell state in the eye primordium (12, 47). Furthermore, our results reveal developmental contextdependent tumor cooperative roles of different signaling pathways (Figs. 2, 5, and 6) based on their ability to regulate local cell fates. For example, N, Wg, or Dpp (TGF-β) signalings, which promote a wing cell fate commitment (positive regulators of Vg), do not cooperate for neoplasia of lgl[−] clone in the distal wing (Figs. 2 and 5). Signaling pathways that promote neoplasia in one developmental context can thus suppress tumor progression in another.

Core Gene-Set for Primitive and Neoplastic Cell States. An intractable link between primitive and neoplastic cell states seen here is a pointer towards recruitment of a core gene-set in both these cell states. The transcription cofactor of the Hippo pathway, Yki, (reviewed in ref. 9), its target dMyc (54), and its partner Hth (12), are thus likely to be crucial members of this core gene-set. It has been previously shown that Yki is activated upon loss of Lgl (46, 55) or Scrib (16, 31) while its target dMyc is a driver of neoplasia in lgl[−] (17) and scrib[−] (16) mutant cells. Further, mammalian homolog of Yki, namely, transcriptional co-activator with PDZ-binding motif (TAZ), induces stem cell-like characteristic in breast epithelial cells transformed by loss of the Scrib (56). In Drosophila too, Yki is likely to be implicated in maintenance of stem cell-like characteristic: it is recruited in regenerating epithelial cells (55) and also plays a role in the maintenance of intestinal cell homeostasis (57). Furthermore, a Myc network has been shown to be an active component of both human embryonic stem cells and cancer cells (58). Hth, on the other hand, in collaboration with Yki, plays a critical role in the maintenance of retinal progenitor cells (12) which is likely to to be evolutionarily conserved (59). Further, recruitment of Hth during retinal cell neoplasia in Drosophila (Fig. 7) implicates this core gene-set in their switch to a progenitor-like cell state (Fig. 6). During normal course of development, Yki recruits different transcription factors, such as Hth (12), Sd (36), and Mad, the transcription factor of Dpp/TGF-β signaling pathway, in a developmental context-dependent manner (60). We note that neoplasia in the wing primordium is Hth-independent (Fig. 6), unlike that of the eye primordium (Fig. 7). In the wing primordium therefore Yki presumably partners with transcription factor(s) other than Hth to induce neoplasia. Rich genetics of the fly model could thus be explored to screen for these diverse sets of transcription factors with spatially distinct cooperative or tumorsuppressive roles in carcinogenesis.

Experimental Procedures

Genetic stocks of Drosophila were received from Bloomington Drosophila Stock Center, while antibodies were procured from Developmental Studies Hybridoma Bank or received as gifts from other investigators. Detailed material and methods are provided in [SI Experimental Procedures](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1212513110/-/DCSupplemental/pnas.201212513SI.pdf?targetid=nameddest=STXT).

ACKNOWLEDGMENTS. We thank L. Johnston, G. Morata, H. Bellen, G. Haldar, K. Basler, and the Bloomington Drosophila Stock Center for fly stocks; S. Cohen, H. Sun, S. Carroll, J. F. de Celis, and Developmental Studies Hybridoma Bank for antibodies; S. S. Pandey and S. Srivastava for technical help; A. Awadhiya for the illustrations; and P. Srivastava for critical comments on the manuscript. This work was supported by the financial assistance from a Department of

- 1. Barker N, et al. (2009) Crypt stem cells as the cells-of-origin of intestinal cancer. Nature 457(7229):608–611.
- 2. Visvader JE (2011) Cells of origin in cancer. Nature 469(7330):314–322.
- 3. Hariharan IK, Bilder D (2006) Regulation of imaginal disc growth by tumor-suppressor genes in Drosophila. Annu Rev Genet 40:335–361.
- 4. Kim J, et al. (1996) Integration of positional signals and regulation of wing formation and identity by Drosophila vestigial gene. Nature 382(6587):133–138.
- 5. Zecca M, Struhl G (2002) Control of growth and patterning of the Drosophila wing imaginal disc by EGFR-mediated signaling. Development 129(6):1369–1376.
- 6. Azpiazu N, Morata G (2000) Function and regulation of homothorax in the wing imaginal disc of Drosophila. Development 127(12):2685–2693.
- 7. Casares F, Mann RS (2000) A dual role for homothorax in inhibiting wing blade development and specifying proximal wing identities in Drosophila. Development 127(7):1499–1508.
- 8. Zecca M, Struhl G (2010) A feed-forward circuit linking wingless, fat-dachsous signaling, and the warts-hippo pathway to Drosophila wing growth. PLoS Biol 8(6):e1000386.
- 9. Pan D (2010) The hippo signaling pathway in development and cancer. Dev Cell 19(4): 491–505.
- 10. Wang S-H, Simcox A, Campbell G (2000) Dual role for Drosophila epidermal growth factor receptor signaling in early wing disc development. Genes Dev 14(18):2271–2276.
- 11. Bergantiños C, Vilana X, Corominas M, Serras F (2010) Imaginal discs: Renaissance of a model for regenerative biology. Bioessays 32(3):207–217.
- 12. Peng HW, Slattery M, Mann RS (2009) Transcription factor choice in the Hippo signaling pathway: Homothorax and Yorkie regulation of the microRNA bantam in the progenitor domain of the Drosophila eye imaginal disc. Genes Dev 23(19): 2307–2319.
- 13. Klein G (2009) Toward a genetics of cancer resistance. Proc Natl Acad Sci USA 106(3): 859–863.
- 14. Bissell MJ, Hines WC (2011) Why don't we get more cancer? A proposed role of the microenvironment in restraining cancer progression. Nat Med 17(3):320–329.
- 15. Agrawal N, Kango M, Mishra A, Sinha P (1995) Neoplastic transformation and aberrant cell-cell interactions in genetic mosaics of lethal(2)giant larvae (lgl), a tumor suppressor gene of Drosophila. Dev Biol 172(1):218–229.
- 16. Chen C-L, Schroeder MC, Kango-Singh M, Tao C, Halder G (2012) Tumor suppression by cell competition through regulation of the Hippo pathway. Proc Natl Acad Sci USA 109(2):484–489.
- 17. Froldi F, et al. (2010) The lethal giant larvae tumour suppressor mutation requires dMyc oncoprotein to promote clonal malignancy. BMC Biol 8:33.
- 18. Grzeschik NA, Amin N, Secombe J, Brumby AM, Richardson HE (2007) Abnormalities in cell proliferation and apico-basal cell polarity are separable in Drosophila lgl mutant clones in the developing eye. Dev Biol 311(1):106–123.
- 19. Igaki T, Pastor-Pareja JC, Aonuma H, Miura M, Xu T (2009) Intrinsic tumor suppression and epithelial maintenance by endocytic activation of Eiger/TNF signaling in Drosophila. Dev Cell 16(3):458–465.
- 20. Johnston LA (2009) Competitive interactions between cells: Death, growth, and geography. Science 324(5935):1679–1682.
- 21. Bilder D, Li M, Perrimon N (2000) Cooperative regulation of cell polarity and growth by Drosophila tumor suppressors. Science 289(5476):113–116.
- 22. Hay BA, Wolff T, Rubin GM (1994) Expression of baculovirus P35 prevents cell death in Drosophila. Development 120(8):2121–2129.
- 23. Dekanty A, Barrio L, Muzzopappa M, Auer H, Milán M (2012) Aneuploidy-induced delaminating cells drive tumorigenesis in Drosophila epithelia. Proc Natl Acad Sci USA 109(50):20549–20554.
- 24. White K, et al. (1994) Genetic control of programmed cell death in Drosophila. Science 264(5159):677–683.
- 25. Brumby AM, Richardson HE (2003) Scribble mutants cooperate with oncogenic Ras or Notch to cause neoplastic overgrowth in Drosophila. EMBO J 22(21):5769–5779.
- 26. Pagliarini RA, Xu T (2003) A genetic screen in Drosophila for metastatic behavior. Science 302(5648):1227–1231.
- 27. Subramanian A, et al. (2005) Gene set enrichment analysis: A knowledge-based approach for interpreting genome-wide expression profiles. Proc Natl Acad Sci USA 102(43):15545–15550.
- 28. Hamaratoglu F, et al. (2006) The tumour-suppressor genes NF2/Merlin and Expanded act through Hippo signalling to regulate cell proliferation and apoptosis. Nat Cell Biol 8(1):27–36.
- 29. Cho E, et al. (2006) Delineation of a Fat tumor suppressor pathway. Nat Genet 38(10): 1142–1150.
- 30. Neumann CJ, Cohen SM (1996) A hierarchy of cross-regulation involving Notch, wingless, vestigial and cut organizes the dorsal/ventral axis of the Drosophila wing. Development 122(11):3477–3485.

Biotechnology's National Facility Grant and Research Council United Kingdom and Department of Science and Technology's United Kingdom-India Science Bridge grant (to P.S.); a Science and Engineering Research Council grant (to A.M.); the Indian Institute of Technology Kanpur support (to A.M.); and re-search fellowships from the Council of Scientific and Industrial Research (to S.J.K., M.A.A., R.P.G., and R.K.P.).

- 31. Menéndez J, Pérez-Garijo A, Calleja M, Morata G (2010) A tumor-suppressing mechanism in Drosophila involving cell competition and the Hippo pathway. Proc Natl Acad Sci USA 107(33):14651–14656.
- 32. Rogulja D, Rauskolb C, Irvine KD (2008) Morphogen control of wing growth through the Fat signaling pathway. Dev Cell 15(2):309–321.
- 33. Jaiswal M, Agrawal N, Sinha P (2006) Fat and Wingless signaling oppositely regulate epithelial cell-cell adhesion and distal wing development in Drosophila. Development 133(5):925–935.
- 34. Campbell G, Tomlinson A (1998) The roles of the homeobox genes aristaless and Distal-less in patterning the legs and wings of Drosophila. Development 125(22): 4483–4493.
- 35. Ng M, Diaz-Benjumea FJ, Vincent JP, Wu J, Cohen SM (1996) Specification of the wing by localized expression of wingless protein. Nature 381(6580):316–318.
- 36. Wu S, Liu Y, Zheng Y, Dong J, Pan D (2008) The TEAD/TEF family protein Scalloped mediates transcriptional output of the Hippo growth-regulatory pathway. Dev Cell 14(3):388–398.
- 37. Dahmann C, Basler K (2000) Opposing transcriptional outputs of Hedgehog signaling and engrailed control compartmental cell sorting at the Drosophila A/P boundary. Cell 100(4):411–422.
- 38. Jack J, Dorsett D, Delotto Y, Liu S (1991) Expression of the cut locus in the Drosophila wing margin is required for cell type specification and is regulated by a distant enhancer. Development 113(3):735–747.
- 39. Neumann CJ, Cohen SM (1997) Long-range action of Wingless organizes the dorsalventral axis of the Drosophila wing. Development 124(4):871–880.
- 40. Couso JP, Knust E, Martinez Arias A (1995) Serrate and wingless cooperate to induce vestigial gene expression and wing formation in Drosophila. Curr Biol 5(12): 1437–1448.
- 41. Nellen D, Burke R, Struhl G, Basler K (1996) Direct and long-range action of a DPP morphogen gradient. Cell 85(3):357–368.
- 42. Johnston LA, Edgar BA (1998) Wingless and Notch regulate cell-cycle arrest in the developing Drosophila wing. Nature 394(6688):82–84.
- 43. Carroll SB, Grenier JK, Weatherbee SD (2005) From DNA to Diversity: Molecular Genetics and the Evolution of Animal Design (Blackwell Scientific, Malden, MA), 2nd Ed.
- 44. Wu J, Cohen SM (2002) Repression of Teashirt marks the initiation of wing development. Development 129(10):2411–2418.
- 45. Karim FD, Rubin GM (1998) Ectopic expression of activated Ras1 induces hyperplastic growth and increased cell death in Drosophila imaginal tissues. Development 125(1):1–9.
- 46. Grzeschik NA, Parsons LM, Allott ML, Harvey KF, Richardson HE (2010) Lgl, aPKC, and Crumbs regulate the Salvador/Warts/Hippo pathway through two distinct mechanisms. Curr Biol 20(7):573–581.
- 47. Lopes CS, Casares F (2010) Hth maintains the pool of eye progenitors and its downregulation by Dpp and Hh couples retinal fate acquisition with cell cycle exit. Dev Biol 339(1):78–88.
- 48. Hanahan D, Weinberg RA (2000) The hallmarks of cancer. Cell 100(1):57–70.
- 49. Green DR, Evan GI (2002) A matter of life and death. Cancer Cell 1(1):19–30.
- 50. Lowe SW, Cepero E, Evan G (2004) Intrinsic tumour suppression. Nature 432(7015): 307–315.
- 51. Chaffer CL, et al. (2011) Normal and neoplastic nonstem cells can spontaneously convert to a stem-like state. Proc Natl Acad Sci USA 108(19):7950–7955.
- 52. Molyneux G, et al. (2010) BRCA1 basal-like breast cancers originate from luminal epithelial progenitors and not from basal stem cells. Cell Stem Cell 7(3):403–417.
- 53. Smith-Bolton RK, Worley MI, Kanda H, Hariharan IK (2009) Regenerative growth in Drosophila imaginal discs is regulated by Wingless and Myc. Dev Cell 16(6):797–809.
- 54. Neto-Silva RM, de Beco S, Johnston LA (2010) Evidence for a growth-stabilizing regulatory feedback mechanism between Myc and Yorkie, the Drosophila homolog of Yap. Dev Cell 19(4):507–520.
- 55. Sun G, Irvine KD (2011) Regulation of Hippo signaling by Jun kinase signaling during compensatory cell proliferation and regeneration, and in neoplastic tumors. Dev Biol 350(1):139–151.
- 56. Cordenonsi M, et al. (2011) The Hippo transducer TAZ confers cancer stem cell-related traits on breast cancer cells. Cell 147(4):759–772.
- 57. Ren F, et al. (2010) Hippo signaling regulates Drosophila intestine stem cell proliferation through multiple pathways. Proc Natl Acad Sci USA 107(49):21064–21069.
- 58. Kim J, et al. (2010) A Myc network accounts for similarities between embryonic stem and cancer cell transcription programs. Cell 143(2):313–324.
- 59. Bessa J, et al. (2008) meis1 regulates cyclin D1 and c-myc expression, and controls the proliferation of the multipotent cells in the early developing zebrafish eye. Development 135(5):799–803.
- 60. Oh H, Irvine KD (2011) Cooperative regulation of growth by Yorkie and Mad through bantam. Dev Cell 20(1):109–122.