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## EMERGING FUNGAL INFECTIONS IN SOLID ORGAN TRANSPLANT RECIPIENTS

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### Abstract

The most important emerging and rare fungal pathogens in solid organ transplant recipients are the Zygomycetes, *Scedosporium*, *Fusarium* and the dark molds. Factors impacting the emergence of these fungi include the combination of intensive immunosuppressive regimens with increasingly widespread use of long-term azole antifungal therapy; employment of aggressive diagnostic approaches (e.g. sampling of bronchoalveolar lavage fluid) and changes in patients' interactions with the environment. Early diagnosis, differentiation between colonization and infection and institution of appropriate therapy is vital when contending with these fungi. Moreover, effective treatment often requires a multi-disciplinary approach. This article reviews the epidemiology, microbiology and clinical impact of emerging fungal infections in solid organ transplant recipients and provides up to date recommendations on their treatment.

### Introduction

Development of invasive fungal infections in solid organ transplant (SOT) recipients is dependent upon the interplay between host and fungal factors. Changes in either of these variables can favor emergence of infections in new populations and/or by previously nonpathogenic fungi.

Managing SOT recipients with emerging fungal infections can be a daunting task. Most clinicians have very limited personal experience in treating these infections. Therapy frequently requires a multidisciplinary approach that includes toxic medications and invasive procedures, and these infections have the potential for devastating outcomes including graft loss or death. The medical literature describing such infections is difficult to interpret, as it is mostly comprised of anecdotal experiences and small case series. Adding to the complexity is the evolving nomenclature of many of these fungi. The goals of this review are to demystify these infections and to serve as a resource for clinicians contending with emerging fungal infections in SOT recipients.

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## Overview

Emerging fungi are increasingly recognized as potential pathogens in SOT recipients. They account for 7-10% of invasive fungal infections in this population (1-4).

## Microbiology

Clinically significant emerging fungi include (1-3, 5, 6)(7-11):

- Zygomycetes (e.g. *Rhizopus*, *Mucor*, *Absidia* and *Mycoladus* species)
- *Scedosporium* (e.g. *S. apiospermum*, *S. auranticum* and *S. prolificans*)
- *Fusarium* (e.g. *F. solani* and *F. oxysporum*)
- Dark molds- also called dematiaceous fungi (e.g. *Ochroconis*, *Cladophialophora*, *Rhinocladiella*, *Bipolaris* and *Fonsecaea* species)
- *Paecilomyces*, *Acremonium*, *Trichoderma*
- Yeast-like organisms (e.g. *Trichosporon*, *Cryptococcus gattii* and *Rhodotorula* species)

## Environment

The emerging fungi are typically found in diverse environmental sources such as soil, water, decaying vegetation and sewage. Patients come into contact with these fungi either via inhalation of airborne spores, or less commonly by touching a contaminated source. The environmental microbiology and hence patients' exposures, can vary by geographic locale. For example, risk for infection due to the Zygomycetes and to *Scedosporium* species are particularly high in the Middle East and Australia respectively (12, 13).

Certain occupations and living circumstances can put patients at higher risk. Exposures to construction sites, farming operations, sandblasting work, air conditioning filters and flooded sites are important in that regard. Risks for contact with potentially pathogenic fungi and ways to reduce patients' exposure should be discussed with transplant recipients. If an infection is suspected, inquiring about a patient's travel, occupation and activities may provide important epidemiological clues. Sometimes exposures can occur within the healthcare setting (14, 15). Outbreaks of mucormycosis have been associated with contaminated adhesive bandages, wooden tongue depressors, ostomy bags, water circuitry damage, and adjacent building construction. Fusariosis may be acquired from contaminated hospital drains and showerheads.

Patterns of fungal exposure may also be impacted by environmental disruptions such as natural disasters and by the development of new ecological niches. Infections due to the Zygomycetes and *Scedosporium* species may be seen after floods, tornados and tsunamis (16, 17). The risk of transmitting such organisms may be relevant when evaluating potential organ donors who suffered drowning accidents (18, 19). The role of new ecological niches has been demonstrated in the recent outbreak of *Cryptococcus gattii* in the Pacific Northwest region of North America(20). Starting in 2004, cases of *C. gattii* infections have been identified in that region(21). Approximately 1/5 of those affected in that outbreak have been SOT recipients. With regards to climate change some have hypothesized that global warming may increase the prevalence of fungal diseases by increasing the geographic range of currently pathogenic species and by selecting for adaptive thermotolerance in species that are currently unable to survive at human body temperature(22).

## Host factors

Multiple arms of the immune system are impaired in transplant recipients. The first lines of defense are intact anatomical barriers. If there is exposure to an emerging fungal pathogen when such barriers are disrupted and there is ongoing high-level immunosuppression, a “perfect storm” can develop resulting in invasive infection. This can occur in any SOT recipient, but lung transplant recipients are at highest risk (1, 3, 23-25). Factors favoring fungal infections in lung transplant recipients include ongoing exposure of the graft to environmental fungi, underlying chronic respiratory disease and concomitant sinus and airway abnormalities impeding fungal clearance. Patients with impaired cutaneous barriers due to traumatic injury or an invasive medical procedure are also at risk. Skin infections due to the *Zygomycetes*, *Scedosporium*, *Fusarium* and the dark molds have been described in such circumstances (14, 26-28).

## Primary site of infection (by route of exposure)

- Inhalation of airborne spores: Most likely sites are the sinuses, airways and/or lung parenchyma. Disease may then extend to involve adjacent sites or disseminate to distant organs.
- Direct inoculation: Most likely site is the skin and adjacent soft tissue structures
- Donor derived (e.g. Zygomycosis via contamination of the preservation fluid or from the organ itself): infection can be localized to the transplanted organ and the graft anastomosis or disseminate widely. Such cases are associated with high rates of graft loss and mortality and may be particularly problematic in commercial transplantation (19, 29, 30).

An additional factor in the evolution of infection due to emerging fungi may be the impact of antifungals used for prophylaxis and therapy. Decreased susceptibility to one or more commonly used antifungals is common in emerging fungi (Table 1). For example, the *Zygomycetes* are not susceptible to voriconazole and to the echinocandins. Use of these agents has been associated with increased risk for development of mucormycosis in some, but not all studies (31-33). *Trichosporon* and *Fusarium* species are frequently resistant to amphotericin B (AmB), and *Scedosporium prolificans* may be resistant to all commonly used antifungal agents. It is likely that development of infections with these fungi is related to antifungal selection pressure.

## Specific fungi

### Mucormycosis

Infections due to the *Zygomycetes* are the best characterized of the emerging fungal infections in SOT recipients(1-3). Exposure is generally via inhalation, but may also occur at the skin or GI tract. Direct contact with soil or water following natural disasters or near drowning episodes, and exposure to contaminated medical devices (as described above) are additional routes to infection (14).

Clinically important *Zygomycetes* include species of *Rhizopus*, *Mucor*, *Rhizomucor*, *Cunninghamella*, *Absidia*, *Apophysomyces* and *Mycocladius*. The predominant infecting organism can vary by geographic site of exposure (34). Additionally, pathogenic potential may differ by organism. For example *Mycocladius corymbifer* pulmonary infection has been associated with higher rates of disseminated disease (35).

## Epidemiology (6)

- Zygomycosis accounts for approximately 2% of fungal infections in SOT recipients.
- Lung and liver transplant recipients are the most affected
- Infections tend to occur at a median of 6 months after transplant except in liver transplant recipients where infection can occur in 1<sup>st</sup> month.
- Risk factors in SOT recipients include: Renal failure, diabetes mellitus, exposure to high doses of corticosteroids, and prior use of voriconazole.
- “Traditional” risk factors for mucormycosis (e.g. ketoacidosis, prolonged and profound neutropenia and deferoximine) are infrequently seen in SOT recipients.

## Clinical Presentations (6, 35)(36)(37)

- Lungs are the most common sites of infection. Disease presents as consolidation/ mass lesions nodules and cavities
- Infection at the sinuses and nose may remain localized or extend to orbits, brain and other intracranial structures.
- Primary cutaneous infection occurs at sites of surgical wounds or drains, intravenous catheter sites and after skin trauma. The typical appearance is of black necrotic lesions surrounded by cellulitis, thrombophlebitis or extension to deeper structures.
- Disseminated disease can involve virtually any organ including the lungs, pericardium, myocardium, endocardium, brain, liver, esophagus, stomach, small and large intestine, kidney, retroperitoneum, thyroid gland and skin.

Diagnosis usually requires an invasive procedure such as biopsy, fine needle aspiration, bronchoscopy, or surgical exploration, but occasionally the organism can be grown from expectorated sputum (37). Upon direct staining the Zygomycetes tend to appear as broad, ribbon-like and non-septate hyphae. Not infrequently, the organisms are identified on either histology or culture, but not both. Increasingly, PCR is being used for detection of Zygomycetes (34).

Outcomes depend upon timely initiation of appropriate antifungal therapy, the host immune status and the site and extent of infection. The cornerstones of management are effective antifungal therapy, improvement in host defenses and surgical resection of necrotic tissue generated by this angioinvasive fungus. Medical therapy alone can be attempted in some patients with pulmonary infection. However, surgery is typically required if there is extensive necrosis, infection at lung transplant anastomosis, or a threat to major vascular structures. Debridement of airway or sinus disease may be performed endoscopically.

## Antifungal treatment options

- Drugs of choice: Lipid formulation amphotericin B (5.0-7.5 mg/kg/day)
- Combination of an echinocandin + Lipid formulation amphotericin B may be considered based on data from animal studies and retrospective reports.
- Posaconazole may be considered for maintenance once clinical stability has been achieved and for salvage therapy in patients intolerant to or failing AmB (38-40).

## Dark (Dematiaceous) Molds

The dark molds are a diverse group of pigmented fungi that are associated with a variety of infections (41). Their nomenclature can be confusing and many organisms have undergone name changes in recent years. Invasive infection is called phaeohyphomycosis. Colonization of the respiratory tract and sinuses is common and does not necessarily indicate infection. Such colonization may be particularly common in lung transplant candidates and recipients. Infections tend to occur late, often several years after transplantation. The site of infection depends upon the mode of exposure and the fungal species (see below). Certain species tend to cause cutaneous infections at sites of inoculation, presenting as papules, nodules or pustules, while others cause pulmonary or disseminated (including CNS) disease. (7, 8, 42-45)(46, 47)(48).

### Phaeohyphomycosis by site and organism

- Primary Skin: *Alternaria*, *Curvularia*, *Exophiala*
- Pulmonary: *Ochroconis gallopavum*, *Cladophialophora bantiana*, *Exophiala*, *Alternaria*, *Curvularia* and *Fonsecaea*
- CNS: *Cladophialophora bantiana*, *Ochroconis gallopavum*, *Rhinoctadiella mackenziei*, *Exophiala dermatitidis*, *Bipolaris* and *Fonsecaea*

The diagnosis can be straightforward as in the case of symptomatic disease and evidence of the fungus in histopathology and culture. However, simply growing these molds in culture (particularly from the nose or sinuses) does not necessarily imply infection. In tissue these fungi may be identified by the golden-brown coloration in the walls of the hyphae(44).The presence of melanin can be highlighted by Fontana-Masson staining.

Treatment depends upon the infecting organism and site of infection. In general, surgical excision or debridement is recommended whenever feasible. This may even be sufficient for isolated cutaneous disease. Conversely, when disease is limited to the respiratory tract, medical management alone may suffice. Voriconazole, posaconazole or itraconazole are typically first line agents, but there may also be a role for AmB and the echinocandins (48, 49). Susceptibility testing can be useful to guide therapy.

## Fusarium

Among SOT patients, fusariosis predominantly affects lung transplant recipients. Such infections accounts for <1% of fungal infections in SOT recipients (1, 3). Most of the infections are caused by *F. solani* and *F. oxysporum*, and to a lesser extent *F. proliferatum*, *F. moniliforme* and *F. sacchari* (15). Exposure is primarily via inhalation of airborne conidia or by contact with contaminated material (e.g. soil, plants and organic matter). *Fusarium* may also be found in water distribution systems, tap water, sinks and showerheads (15).

The clinical spectrum includes superficial, localized and disseminated infections (50). The specific presentation depends upon portal of entry, extent of immunosuppression and transplant type (51)(52)(53-55)(56).

### Clinical presentations

- Primary skin infection due to direct inoculation: Present as superficial or localized infection (e.g. nodules, ulcers, cellulitis, subcutaneous abscess) and usually very responsive to therapy.
- Respiratory tract and sinus infection: Due to inhalation and typically occurs in lung transplant recipients.

- Secondary dissemination to multiple organs including the GI tract, liver, heart valves, kidneys, lung, CNS and skin

### Diagnosis(53)(57, 58)

- Targetoid skin lesions with darkish discoloration are a clue to disseminated fusariosis. Skin biopsy can establish the diagnosis.
- Occasionally, the organism grows in blood cultures
- The role of non-culture based diagnostic tests (e.g. beta glucan and galactomannan) is currently evolving, but these may be useful as adjunctive tests.

Treatment options and outcomes depend on site and extent of infection and upon the species (59). Identification of the organism to the species level and antifungal susceptibility testing can help guide therapy. Surgical excision or debridement should be employed whenever possible.

### Treatment options(56)

- *F. solani* and *F. verticillioides*: AmB
- Other species: Voriconazole
- Limited skin disease: excision alone might suffice
- Combination therapy (AmB + voriconazole) for invasive infection while awaiting identification and susceptibility testing and in severe cases.

### Scedosporium/Pseudallescheria

Infections due to *Scedosporium* and *Pseudallescheria* species can be particularly difficult to treat. The organisms are frequently resistant to multiple antifungal agents and outcomes with invasive infections can be devastating. The nomenclature is complicated and still evolving. The teleomorph (sexual form) is referred to as *Pseudallescheria* and the asexual form (anamorph) as *Scedosporium*. The predominant species are *S. apiospermum* (teleomorph: *P. apiosperma*), *S. aurantiacum*, *P. boydii* (anamorph: *S. boydii*), *S. dehoogii*, and *S. prolificans* (60, 61).

These organisms are typically found in soil and contaminated water, including in urban environments (12). Exposure is generally via inhalation of airborne spores but may also occur after contact with contaminated water. Patients with cystic fibrosis are often colonized with *Scedosporium* even prior to transplant. Infections are predominantly seen in lung transplant recipients, but occur across the spectrum of organ transplants (3, 62). Primary sites of infection include the respiratory tract, sinuses and skin. Infection may progress or disseminate to involve additional organs including bone, joint, brain, eye, ear and vocal cords (62).

Treatment recommendations are listed below, however, antifungal susceptibility testing of all clinical isolates is essential for guiding therapy(63)(64)(65)(66, 67):

### Treatment options

- *S. apiospermum*: Voriconazole +/- echinocandin
- *S. aurantiacum*: Voriconazole
- *S. prolificans*: Resistant to multiple antifungal agents, but there may be a role for voriconazole + echinocandin, AmB + terbinafine or voriconazole + terbinafine

- Surgical debridement should be considered whenever feasible. Particularly in cases of multi-drug resistant fungal infection.

### Paecilomyces

Paecilomyces species, particularly *P. lilacinus*, have emerged as a cause of fungal infections in highly immunocompromised patients. These environmental fungi are generally found in the air and in soil, but have also been associated with an outbreak of infection related to contaminated skin lotion(68). The predominant clinical presentation in SOT recipients is subacute skin infection (69). Paecilomyces can sometimes cause such infections in association with other fungi or mycobacteria (70, 71). The antifungal agent with the best track record for *P. lilacinus* infections is voriconazole, but susceptibility testing may help guide therapy (72, 73).

### Trichosporon

Trichosporon species, particularly *T. asahii* and *T. mucoides* can cause systemic infection in SOT recipients (74, 75). These yeasts are found in diverse setting including soil, water, and vegetables and as commensals of the human skin and GI tract. Clinical presentations include fungemia and widely disseminated infection. The significance of *Trichosporon* funguria in renal transplant recipients is unclear and may not require antifungal therapy(76). When treatment is indicated, as in systemic disease, azoles are the mainstay of treatment (74, 75, 77). However, resistance to azoles and AmB is common and antifungal susceptibility testing is necessary to help guide therapy(10).

### Miscellaneous rare fungi

There are multiple rare fungi that have been described as causes of infection in SOT recipients at the case report level. These include *Acremonium*, *Scopulariopsis* and *Trichoderma* species (78-80). As general rules, surgical excision should be attempted when possible and therapy should be guided by susceptibility testing.

### Conclusion

Development of invasive fungal infections depends upon the confluence of host factors and environmental exposure. Changes in either of these parameters can favor emerging fungal infections. We are in the midst of such changes. Current trends in transplantation include increasingly diverse patient populations, use of novel and potent immunosuppressive regimens and expanded use of antifungal agents for prophylaxis and treatment. When coupled with environmental disruptions including natural disasters, new ecological niches and perhaps climate change we can expect an ongoing evolution of fungal epidemiology in SOT recipients and an increasingly important role for emerging fungi (tables 2-3).

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### REFERENCES

1. Neofytos D, Fishman JA, Horn D, Anaissie E, Chang CH, Olyaei A, et al. Epidemiology and outcome of invasive fungal infections in solid organ transplant recipients. *Transpl Infect Dis*. 2010; 12(3):220–9.
2. Pappas PG, Alexander BD, Andes DR, Hadley S, Kauffman CA, Freifeld A, et al. Invasive fungal infections among organ transplant recipients: results of the Transplant-Associated Infection Surveillance Network (TRANSNET). *Clin Infect Dis*. 2010; 50(8):1101–11. [PubMed: 20218876]

3. Park BJ, Pappas PG, Wannemuehler KA, Alexander BD, Anaissie EJ, Andes DR, et al. Invasive non-Aspergillus mold infections in transplant recipients, United States, 2001-2006. *Emerg Infect Dis.* 2011; 17(10):1855–64. [PubMed: 22000355]
4. Kubak BM, Huprikar SS. Emerging & rare fungal infections in solid organ transplant recipients. *Am J Transplant.* 2009; 9(Suppl 4):S208–26. [PubMed: 20070683]
5. Stelzmueller I, Lass-Floerl C, Geltner C, Graziadei I, Schneeberger S, Antretter H, et al. Zygomycosis and other rare filamentous fungal infections in solid organ transplant recipients. *Transpl Int.* 2008; 21(6):534–46. [PubMed: 18363572]
6. Singh N, Aguado JM, Bonatti H, Forrest G, Gupta KL, Safdar N, et al. Zygomycosis in solid organ transplant recipients: a prospective, matched case-control study to assess risks for disease and outcome. *J Infect Dis.* 2009; 200(6):1002–11. [PubMed: 19659439]
7. Shoham S, Pic-Aluas L, Taylor J, Cortez K, Rinaldi MG, Shea Y, et al. Transplant-associated *Ochroconis gallopava* infections. *Transpl Infect Dis.* 2008; 10(6):442–8. [PubMed: 18651872]
8. Rosow L, Jiang JX, Deuel T, Lechpammer M, Zamani AA, Milner DA, et al. Cerebral phaeohyphomycosis caused by *Bipolaris spicifera* after heart transplantation. *Transpl Infect Dis.* 2011; 13(4):419–23. [PubMed: 21323827]
9. Grossi P, Farina C, Fiocchi R, Dalla Gasperina D. Prevalence and outcome of invasive fungal infections in 1,963 thoracic organ transplant recipients: a multicenter retrospective study. Italian Study Group of Fungal Infections in Thoracic Organ Transplant Recipients. *Transplantation.* 2000; 70(1):112–6. [PubMed: 10919584]
10. Netsvyetayeva I, Swoboda-Kopec E, Paczek L, Fiedor P, Sikora M, Jaworska-Zaremba M, et al. *Trichosporon asahii* as a prospective pathogen in solid organ transplant recipients. *Mycoses.* 2009; 52(3):263–5. [PubMed: 18705664]
11. Riedel DJ, Johnson JK, Forrest GN. *Rhodotorula glutinis* fungemia in a liver-kidney transplant patient. *Transpl Infect Dis.* 2008; 10(3):197–200. [PubMed: 17605726]
12. Harun A, Gilgado F, Chen SC, Meyer W. Abundance of *Pseudallescheria/Scedosporium* species in the Australian urban environment suggests a possible source for scedosporiosis including the colonization of airways in cystic fibrosis. *Med Mycol.* 2010; 48(Suppl 1):S70–6. [PubMed: 21067333]
13. Einollahi B, Lessan-Pezeshki M, Pourfarziani V, Nemati E, Nafar M, Pour-Reza-Gholi F, et al. Invasive fungal infections following renal transplantation: a review of 2410 recipients. *Ann Transplant.* 2008; 13(4):55–8. [PubMed: 19034224]
14. Rammaert B, Lanternier F, Zahar JR, Dannaoui E, Bougnoux ME, Lecuit M, et al. Healthcare-associated mucormycosis. *Clin Infect Dis.* 2012; 54(Suppl 1):S44–54. [PubMed: 22247444]
15. Anaissie EJ, Kuchar RT, Rex JH, Francesconi A, Kasai M, Muller FM, et al. Fusariosis associated with pathogenic fusarium species colonization of a hospital water system: a new paradigm for the epidemiology of opportunistic mold infections. *Clin Infect Dis.* 2001; 33(11):1871–8. [PubMed: 11692299]
16. Weddle G, Gandy K, Bratcher D, Pahud B, Jackson MA. *Apophysomyces trapeziformis* infection associated with a tornado-related injury. *Pediatr Infect Dis J.* 2012; 31(6):640–2. [PubMed: 22301481]
17. Garzoni C, Emonet S, Legout L, Benedict R, Hoffmeyer P, Bernard L, et al. Atypical infections in tsunami survivors. *Emerg Infect Dis.* 2005; 11(10):1591–3. [PubMed: 16318701]
18. Katragkou A, Dotis J, Kotsiou M, Tamiolaki M, Roilides E. *Scedosporium apiospermum* infection after near-drowning. *Mycoses.* 2007; 50(5):412–21. [PubMed: 17714363]
19. Alexander BD, Schell WA, Siston AM, Rao CY, Bower WA, Balajee SA, et al. Fatal *Apophysomyces elegans* infection transmitted by deceased donor renal allografts. *Am J Transplant.* 2010; 10(9):2161–7. [PubMed: 20883549]
20. Mak S, Klinkenberg B, Bartlett K, Fyfe M. Ecological niche modeling of *Cryptococcus gattii* in British Columbia, Canada. *Environ Health Perspect.* 2010; 118(5):653–8. [PubMed: 20439176]
21. Emergence of *Cryptococcus gattii*-- Pacific Northwest, 2004-2010. *MMWR Morb Mortal Wkly Rep.* 2010; 59(28):865–8. [PubMed: 20651641]
22. Garcia-Solache MA, Casadevall A. Global warming will bring new fungal diseases for mammals. *MBio.* 2010; 1(1)



23. Carneiro HA, Coleman JJ, Restrepo A, Mylonakis E. Fusarium infection in lung transplant patients: report of 6 cases and review of the literature. *Medicine (Baltimore)*. 2011; 90(1):69–80. [PubMed: 21200188]
24. Cooley L, Spelman D, Thursky K, Slavin M. Infection with *Scedosporium apiospermum* and *S. prolificans*, Australia. *Emerg Infect Dis*. 2007; 13(8):1170–7. [PubMed: 17953087]
25. Castiglioni B, Sutton DA, Rinaldi MG, Fung J, Kusne S. *Pseudallescheria boydii* (Anamorph *Scedosporium apiospermum*). Infection in solid organ transplant recipients in a tertiary medical center and review of the literature. *Medicine (Baltimore)*. 2002; 81(5):333–48. [PubMed: 12352630]
26. Gallelli B, Viviani M, Nebuloni M, Marzano AV, Pozzi C, Messa P, et al. Skin infection due to *Alternaria* species in kidney allograft recipients: report of a new case and review of the literature. *J Nephrol*. 2006; 19(5):668–72. [PubMed: 17136699]
27. Talbot TR, Hatcher J, Davis SF, Pierson RN 3rd, Barton R, Dummer S. *Scedosporium apiospermum* pneumonia and sternal wound infection in a heart transplant recipient. *Transplantation*. 2002; 74(11):1645–7. [PubMed: 12490804]
28. Palmore TN, Shea YR, Childs RW, Sherry RM, Walsh TJ. *Fusarium proliferatum* soft tissue infection at the site of a puncture by a plant: recovery, isolation, and direct molecular identification. *J Clin Microbiol*. 2010; 48(1):338–42. [PubMed: 19923491]
29. Shoham S, Hinestrosa F, Moore J Jr, O'Donnell S, Ruiz M, Light J. Invasive filamentous fungal infections associated with renal transplant tourism. *Transpl Infect Dis*. 2010; 12(4):371–4. [PubMed: 20163566]
30. Tomazic J, Pirs M, Matos T, Ferluga D, Lindic J. Multiple infections after commercial renal transplantation in India. *Nephrol Dial Transplant*. 2007; 22(3):972–3. [PubMed: 17132708]
31. Chamilos G, Marom EM, Lewis RE, Lionakis MS, Kontoyiannis DP. Predictors of pulmonary zygomycosis versus invasive pulmonary aspergillosis in patients with cancer. *Clin Infect Dis*. 2005; 41(1):60–6. [PubMed: 15937764]
32. Llata E, Blossom DB, Khoury HJ, Rao CY, Wannemuehler KA, Noble-Wang J, et al. A cluster of mucormycosis infections in hematology patients: challenges in investigation and control of invasive mold infections in high-risk patient populations. *Diagn Microbiol Infect Dis*. 2011; 71(1):72–80. [PubMed: 21851872]
33. Saegeman V, Maertens J, Meersseman W, Spriet I, Verbeken E, Lagrou K. Increasing incidence of mucormycosis in University Hospital, Belgium. *Emerg Infect Dis*. 2010; 16(9):1456–8. [PubMed: 20735932]
34. Ruping MJ, Heinz WJ, Kindo AJ, Rickerts V, Lass-Flörl C, Beisel C, et al. Forty-one recent cases of invasive zygomycosis from a global clinical registry. *J Antimicrob Chemother*. 2010; 65(2):296–302. [PubMed: 20008047]
35. Sun HY, Aguado JM, Bonatti H, Forrest G, Gupta KL, Safdar N, et al. Pulmonary zygomycosis in solid organ transplant recipients in the current era. *Am J Transplant*. 2009; 9(9):2166–71. [PubMed: 19681829]
36. Sun HY, Forrest G, Gupta KL, Aguado JM, Lortholary O, Julia MB, et al. Rhino-orbital-cerebral zygomycosis in solid organ transplant recipients. *Transplantation*. 2010; 90(1):85–92.
37. Almyroudis NG, Sutton DA, Linden P, Rinaldi MG, Fung J, Kusne S. Zygomycosis in solid organ transplant recipients in a tertiary transplant center and review of the literature. *Am J Transplant*. 2006; 6(10):2365–74. [PubMed: 16925570]
38. Alexander BD, Perfect JR, Daly JS, Restrepo A, Tobon AM, Patino H, et al. Posaconazole as salvage therapy in patients with invasive fungal infections after solid organ transplant. *Transplantation*. 2008; 86(6):791–6. [PubMed: 18813103]
39. van Burik JA, Hare RS, Solomon HF, Corrado ML, Kontoyiannis DP. Posaconazole is effective as salvage therapy in zygomycosis: a retrospective summary of 91 cases. *Clin Infect Dis*. 2006; 42(7):e61–5. [PubMed: 16511748]
40. Kontoyiannis DP. Invasive mycoses: strategies for effective management. *Am J Med*. 2012; 125(1 Suppl):S25–38. [PubMed: 22196206]
41. Revankar SG, Sutton DA. Melanized fungi in human disease. *Clin Microbiol Rev*. 2010; 23(4):884–928. [PubMed: 20930077]

42. Raparia K, Powell SZ, Cernoch P, Takei H. Cerebral mycosis: 7-year retrospective series in a tertiary center. *Neuropathology*. 2010; 30(3):218–23. [PubMed: 19845862]
43. Qureshi ZA, Kwak EJ, Nguyen MH, Silveira FP. *Ochroconis gallopava*: a dematiaceous mold causing infections in transplant recipients. *Clin Transplant*. 2012; 26(1):E17–23. [PubMed: 21955216]
44. Harrison DK, Moser S, Palmer CA. Central nervous system infections in transplant recipients by *Cladophialophora bantiana*. *South Med J*. 2008; 101(3):292–6. [PubMed: 18364660]
45. Taj-Aldeen SJ, Almaslamani M, Alkhalaf A, Al Bozom I, Romanelli AM, Wickes BL, et al. Cerebral phaeohyphomycosis due to *Rhinocladiella mackenziei* (formerly *Ramichloridium mackenziei*): a taxonomic update and review of the literature. *Med Mycol*. 2010; 48(3):546–56. [PubMed: 19886775]
46. Lief MH, Caplivski D, Bottone EJ, Lerner S, Vidal C, Huprikar S. *Exophiala jeanselmei* infection in solid organ transplant recipients: report of two cases and review of the literature. *Transpl Infect Dis*. 2011; 13(1):73–9. [PubMed: 20738833]
47. Vermeire SE, de Jonge H, Lagrou K, Kuypers DR. Cutaneous phaeohyphomycosis in renal allograft recipients: report of 2 cases and review of the literature. *Diagn Microbiol Infect Dis*. 2010; 68(2):177–80. [PubMed: 20846592]
48. Boyce RD, Deziel PJ, Otley CC, Wilhelm MP, Eid AJ, Wengenack NL, et al. Phaeohyphomycosis due to *Alternaria* species in transplant recipients. *Transpl Infect Dis*. 2010; 12(3):242–50. [PubMed: 20002611]
49. Nakai T, Uno J, Otomo K, Ikeda F, Tawara S, Goto T, et al. In vitro activity of FK463, a novel lipopeptide antifungal agent, against a variety of clinically important molds. *Chemotherapy*. 2002; 48(2):78–81. [PubMed: 12011539]
50. Cocuroccia B, Gaido J, Gubinelli E, Annessi G, Girolomoni G. Localized cutaneous hyalohyphomycosis caused by a *Fusarium* species infection in a renal transplant patient. *J Clin Microbiol*. 2003; 41(2):905–7. [PubMed: 12574313]
51. Sampathkumar P, Paya CV. *Fusarium* infection after solid-organ transplantation. *Clin Infect Dis*. 2001; 32(8):1237–40. [PubMed: 11283817]
52. Halpern M, Balbi E, Carius L, Roma J, Gonzalez AC, Agoglia L, et al. Cellulitis and nodular skin lesions due to *Fusarium* spp in liver transplant: case report. *Transplant Proc*. 2010; 42(2):599–600. [PubMed: 20304202]
53. Kleinschmidt-Demasters BK. Disseminated *Fusarium* infection with brain abscesses in a lung transplant recipient. *Clin Neuropathol*. 2009; 28(6):417–21. [PubMed: 19919815]
54. Lodato F, Tame MR, Montagnani M, Sambri V, Liguori G, Azzaroli F, et al. Systemic fungemia and hepatic localizations of *Fusarium solani* in a liver transplanted patient: an emerging fungal agent. *Liver Transpl*. 2006; 12(11):1711–4. [PubMed: 17058254]
55. Guinvarc'h A, Guilbert L, Marmorat-Khuong A, Lavarde V, Chevalier P, Amrein C, et al. Disseminated *Fusarium solani* infection with endocarditis in a lung transplant recipient. *Mycoses*. 1998; 41(1-2):59–61. [PubMed: 9610136]
56. Nucci M, Anaissie E. *Fusarium* infections in immunocompromised patients. *Clin Microbiol Rev*. 2007; 20(4):695–704. [PubMed: 17934079]
57. Ostrosky-Zeichner L, Alexander BD, Kett DH, Vazquez J, Pappas PG, Saeki F, et al. Multicenter clinical evaluation of the (1->3) beta-D-glucan assay as an aid to diagnosis of fungal infections in humans. *Clin Infect Dis*. 2005; 41(5):654–9. [PubMed: 16080087]
58. Cuetara MS, Alhambra A, Moragues MD, Gonzalez-Elorza E, Ponton J, del Palacio A. Detection of (1->3)-beta-D-glucan as an adjunct to diagnosis in a mixed population with uncommon proven invasive fungal diseases or with an unusual clinical presentation. *Clin Vaccine Immunol*. 2009; 16(3):423–6. [PubMed: 19158282]
59. Lortholary O, Obenga G, Biswas P, Caillot D, Chachaty E, Bienvenu AL, et al. International retrospective analysis of 73 cases of invasive fusariosis treated with voriconazole. *Antimicrob Agents Chemother*. 2010; 54(10):4446–50. [PubMed: 20625156]
60. Gilgado F, Cano J, Gene J, Guarro J. Molecular phylogeny of the *Pseudallescheria boydii* species complex: proposal of two new species. *J Clin Microbiol*. 2005; 43(10):4930–42. [PubMed: 16207945]

61. Gilgado F, Cano J, Gene J, Sutton DA, Guarro J. Molecular and phenotypic data supporting distinct species statuses for *Scedosporium apiospermum* and *Pseudallescheria boydii* and the proposed new species *Scedosporium dehoogii*. *J Clin Microbiol*. 2008; 46(2):766–71. [PubMed: 18077629]
62. Troke P, Aguirrebengoa K, Arteaga C, Ellis D, Heath CH, Lutsar I, et al. Treatment of scedosporiosis with voriconazole: clinical experience with 107 patients. *Antimicrob Agents Chemother*. 2008; 52(5):1743–50. [PubMed: 18212110]
63. Figueiredo RT, Fernandez PL, Dutra FF, Gonzalez Y, Lopes LC, Bittencourt VC, et al. TLR4 recognizes *Pseudallescheria boydii* conidia and purified rhamnomannans. *J Biol Chem*. 2010; 285(52):40714–23. [PubMed: 20959459]
64. Lackner M, de Hoog GS, Verweij PE, Najafzadeh MJ, Curfs-Breuker I, Klaassen CH, et al. Species-specific antifungal susceptibility patterns of *Scedosporium* and *Pseudallescheria* species. *Antimicrob Agents Chemother*. 2012; 56(5):2635–42. [PubMed: 22290955]
65. Cuenca-Estrella M, Alastruey-Izquierdo A, Alcazar-Fuoli L, Bernal-Martinez L, Gomez-Lopez A, Buitrago MJ, et al. In vitro activities of 35 double combinations of antifungal agents against *Scedosporium apiospermum* and *Scedosporium prolificans*. *Antimicrob Agents Chemother*. 2008; 52(3):1136–9. [PubMed: 18195067]
66. Rodriguez MM, Calvo E, Serena C, Marine M, Pastor FJ, Guarro J. Effects of double and triple combinations of antifungal drugs in a murine model of disseminated infection by *Scedosporium prolificans*. *Antimicrob Agents Chemother*. 2009; 53(5):2153–5. [PubMed: 19223617]
67. Meletiadis J, Mouton JW, Meis JF, Verweij PE. In vitro drug interaction modeling of combinations of azoles with terbinafine against clinical *Scedosporium prolificans* isolates. *Antimicrob Agents Chemother*. 2003; 47(1):106–17. [PubMed: 12499177]
68. Orth B, Frei R, Itin PH, Rinaldi MG, Speck B, Gratwohl A, et al. Outbreak of invasive mycoses caused by *Paecilomyces lilacinus* from a contaminated skin lotion. *Ann Intern Med*. 1996; 125(10):799–806. [PubMed: 8928986]
69. Pastor FJ, Guarro J. Clinical manifestations, treatment and outcome of *Paecilomyces lilacinus* infections. *Clin Microbiol Infect*. 2006; 12(10):948–60. [PubMed: 16961630]
70. Lavergne RA, Cassaing S, Nocera T, Pauwels C, Cointault O, Basse G, et al. Simultaneous cutaneous infection due to *Paecilomyces lilacinus* and *Alternaria* in a heart transplant patient. *Transpl Infect Dis*. 2012
71. Kim JE, Sung H, Kim MN, Won CH, Chang SE, Lee MW, et al. Synchronous infection with *Mycobacterium chelonae* and *Paecilomyces* in a heart transplant patient. *Transpl Infect Dis*. 2011; 13(1):80–3. [PubMed: 20412536]
72. Ounissi M, Abderrahim E, Trabelsi S, Khaled S, Bezzine H, Ben Hamida F, et al. Hyalohyphomycosis caused by *Paecilomyces lilacinus* after kidney transplantation. *Transplant Proc*. 2009; 41(7):2917–9. [PubMed: 19765473]
73. Van Schooneveld T, Freifeld A, Lesiak B, Kalil A, Sutton DA, Iwen PC. *Paecilomyces lilacinus* infection in a liver transplant patient: case report and review of the literature. *Transpl Infect Dis*. 2008; 10(2):117–22. [PubMed: 17605741]
74. Lacasse A, Cleveland KO. *Trichosporon mucoides* fungemia in a liver transplant recipient: case report and review. *Transpl Infect Dis*. 2009; 11(2):155–9. [PubMed: 18983414]
75. Biasoli MS, Carlson D, Chiganer GJ, Parodi R, Greca A, Tosello ME, et al. Systemic infection caused by *Trichosporon asahii* in a patient with liver transplant. *Med Mycol*. 2008; 46(7):719–23. [PubMed: 18651307]
76. Lussier N, Laverdiere M, Delorme J, Weiss K, Dandavino R. *Trichosporon beigeli* funguria in renal transplant recipients. *Clin Infect Dis*. 2000; 31(5):1299–301. [PubMed: 11073770]
77. Nettles RE, Nichols LS, Bell-McGuinn K, Pipeling MR, Scheel PJ Jr, Merz WG. Successful treatment of *Trichosporon mucoides* infection with fluconazole in a heart and kidney transplant recipient. *Clin Infect Dis*. 2003; 36(4):E63–6. [PubMed: 12567323]
78. Wuyts WA, Molzahn H, Maertens J, Verbeken EK, Lagrou K, Dupont LJ, et al. Fatal *Scopulariopsis* infection in a lung transplant recipient: a case report. *J Heart Lung Transplant*. 2005; 24(12):2301–4. [PubMed: 16364887]

79. Beaudreuil S, Buchler M, Al Najjar A, Bastides F, Francois M, Duong TH, et al. Acute septic arthritis after kidney transplantation due to *Acremonium*. *Nephrol Dial Transplant*. 2003; 18(4): 850–1. [PubMed: 12637668]
80. Chouaki T, Lavarde V, Lachaud L, Raccurt CP, Hennequin C. Invasive infections due to *Trichoderma* species: report of 2 cases, findings of in vitro susceptibility testing, and review of the literature. *Clin Infect Dis*. 2002; 35(11):1360–7. [PubMed: 12439799]

### Key Points

- The leading emerging fungal pathogens in transplant recipients are the Zygomycetes, *Scedosporium/Pseudallescheria*, *Fusarium* and the dark molds.
- Amphotericin B (AmB) products are the treatment of choice for mucormycosis.
- Identifying emerging fungi to the species level and performing susceptibility testing can help guide therapy.
- When there is evidence for an active infection even low virulence fungi that are isolated from the respiratory tract or sinuses generally require treatment.
- Treatment frequently necessitates a combined approach that includes antifungal therapy, debridement of infected material and efforts to improve host defenses.

**Table 1**

Commonly used antimicrobials for MDR pathogens. (62, 112, 113)

Organism	1st line antimicrobials	Contraindication to transplant
MDR. <i>P. aeruginosa</i>	2 of the following: Carbapenem, piperacillin/tazobactam, cefepime, aminoglycosides, quinolones	Rare
Pan resistant <i>P. aeruginosa</i>	any of above +/- colistin	
<i>B. cenocepacia</i>	Ceftazidime, tetracyclines, trimethoprim-sulfamethoxazole, carbapenem	Probable
<i>B. gladioli</i>	Piperacillin, aminoglycosides, carbapenem, ciprofloxacin	Possible
<i>A. baumannii</i>	Carbapenem, colistin, tigecycline, ampicillin/sulbactam	Possible
<i>M. abscessus</i>	Clarithromycin + amikacin	Possible
	2 <sup>nd</sup> line: Clarithromycin + imipenem or ceftoxitin	
<i>M. avium complex</i>	Clarithromycin, ethambutol, rifampin	Rare
<i>S. apiospermum</i>	Voriconazole +/- echinocandin	Possible
<i>S. prolificans</i>	Voriconazole +/- echinocandin +/-terbinafine	Possible
<i>A. terreus</i>	Voriconazole +/- echinocandin	Rare

**Table 2**

## Emerging Fungal Pathogens in SOT

Category	Important species
Zygomycetes	Species of <i>Rhizopus</i> , <i>Mucor</i> , <i>Absidia</i> and <i>Mycoladus</i> .
Dematiaceous molds	Species of <i>Ochroconis</i> , <i>Cladophialophora</i> , <i>Rhinoctadiella</i> , <i>Bipolaris</i> and <i>Fonsecaea</i>
Scedosporium/Pseudallescheria	<i>S. apiospermum</i> , <i>S. auranticum</i> and <i>S. prolificans</i>
Fusarium	<i>F. solani</i> and <i>F. oxysporum</i>
Other filamentous fungi	Species of <i>Paecilomyces</i> , <i>Acremonium</i> and <i>Trichoderma</i>
Yeasts	Species of <i>Trichosporon</i> , <i>Rodotorula</i> and <i>Cryptococcus gattii</i>

**Table 3**

## Clinical Characteristics of Emerging Fungi

Infection	Characteristic transplant recipient	Median time to infection	Typical sites of infection
Mucormycosis	All, but esp. liver, lung, kidney	Liver transplant: 2-3 months, others ~18-24 months	Respiratory tract/sinuses/CNS
Dematiaceous molds	All	18-24 months	Respiratory, sinuses/CNS, skin
Scedosporiosis	Lung	18-24 months	Respiratory tract
Fusariosis	All, but especially liver and lung	Range from 1-3 to 9 months	Respiratory tract sinus, skin
<i>Paecilomyces</i>	All, especially heart and lung	12-18 months	Respiratory tract, sinus, skin
<i>Trichosporon</i>	All, but especially kidney, Liver	Within first few weeks or 18-24 months	Endovascular/bloodstream surgical wounds