

CROSSTALK

CrossTalk opposing view: There is not added benefit to providing permissive hypercapnia in the treatment of ARDSJeremy R. Beitler¹, Rolf D. Hubmayr² and Atul Malhotra^{1,3}¹Division of Pulmonary and Critical Care Medicine, Brigham and Women's Hospital, Boston, MA, USA²Division of Pulmonary and Critical Care Medicine, Mayo Clinic, Rochester, MN, USA³Division of Sleep Medicine, Brigham and Women's Hospital, Boston, MA, USA

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Current ventilator strategies for acute respiratory distress syndrome (ARDS) aim to impact lung function and clinical outcomes in several ways: low tidal volumes to minimize overdistension, titrated positive end-expiratory pressure (PEEP) to prevent alveolar derecruitment, and recruitment manoeuvres to promote parenchymal homogeneity. Among these interventions, only low tidal volume has been shown definitively to improve mortality from ARDS (Malhotra, 2007). During low tidal volume ventilation, practice varies substantially on whether to allow some degree of alveolar hypoventilation with incidental hypercapnic acidosis (Amato *et al.* 1998), or to increase respiratory rate to maintain alveolar ventilation, often requiring respiratory rates >30 breaths min^{-1} (Brower *et al.* 2000). As the independent effect of hypercapnia has

never been tested in humans, it remains unclear whether such permissive hypercapnia has protective or deleterious effects in ARDS.

Two major questions surrounding permissive hypercapnia include (1) whether the mechanical effects of limiting respiratory rate *per se* yield clinical benefit, and (2) what the effects are of hypercapnia, independent of minute ventilation, on end-organ injury.

Low tidal volumes, which may contribute to hypercapnia, minimize mechanical lung injury. Ventilation at high tidal volumes causes alveolar overdistension, leading to alveolar epithelial and capillary endothelial cell junction breaks, cell detachment from the basement membrane, intracapillary blebs, and alveolar and interstitial oedema (Fu *et al.* 1992). Further contributing to regional wall stress are shear forces created by inflation of normal alveoli adjacent to atelectatic or flooded alveoli (Perlman *et al.* 2011). Additionally, atelectrauma occurs from cyclic recruitment and collapse of atelectatic lung units during respirations. Resultant formation and destruction of foam bubbles contributes interfacial stress that disrupts intracellular plasma membrane–cytoskeletal adhesions, leading to bleb formation and plasma membrane disruption (Oeckler *et al.* 2010). Low tidal volumes may reduce atelectrauma if lung units with high opening pressures remain closed throughout the respiratory cycle.

In response to deformation-induced cell strain, alveolar epithelial cells rapidly translocate lipid molecules to the plasma

membrane to increase cell surface area and reduce strain, thereby minimizing risk of membrane rupture and facilitating repair when injury does occur (Vlahakis *et al.* 2002). Alveolar overdistension and cyclic atelectasis also induce extensive cytokine release that contributes further to end-organ injury (Imai *et al.* 2003). Low tidal volume ventilation directly reduces cyclic overdistension and thus risk of lung injury.

Additionally, limiting end-inspiratory plateau pressure is used commonly to minimize barotrauma. However, airway pressures do not delineate the often substantial contribution of the chest wall to respiratory system mechanics (Talmor *et al.* 2006). Plateau pressures may underestimate alveolar distension in patients with negative pleural pressures, such as during spontaneous respirations. In a recent study demonstrating reduced mortality in severe ARDS with neuromuscular blockade (Papazian *et al.* 2010), the incidence of barotrauma halved with paralysis compared to placebo despite similar initial airway plateau pressures. This may suggest that high transpulmonary pressures (defined as the pressure difference between the airway opening and pleural space) during spontaneous respirations worsened lung injury. Similarly, in a preclinical model of ARDS, lung injury worsened despite unchanged plateau pressures when spontaneous breathing generated high transpulmonary pressures (Yoshida *et al.* 2012). To account for patient differences in pleural pressures and chest wall mechanics, some advocate tailoring mechanical ventilation in ARDS to

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transpulmonary pressure by estimating pleural pressure with oesophageal manometry (Talmor *et al.* 2008).

In addition to reducing tidal stretch, limiting respiratory rate may prevent biomechanical injury by reducing the frequency of cyclic recruitment and collapse of damaged alveoli and of repetitive shear stress in the heterogeneously diseased lung. The cumulative effects of such repetitive stress may lead to frequency-dependent membrane failure known as fatigue behaviour (Marini *et al.* 2003), analogous to bending a paperclip back and forth repeatedly to cause its eventual breakage. Indeed, lower respiratory rate reduces lung injury in preclinical models of ARDS (Hotchkiss *et al.* 2000). Moreover, dynamic hyperinflation may occur with high respiratory rate, increasing risk of regional overdistension (Marini, 2011).

However, it is entirely unclear whether resultant hypercapnia offsets or augments the advantageous biomechanical effects of low tidal volume and respiratory rate. Among the proposed additional benefits, hypercapnia attenuates oxygen free radical formation, NF- κ B activation, and TNF α , IL-1, IL-6 and IL-8 cytokine production (Taylor & Cummins, 2011) increases sympathetic tone with associated haemodynamic improvements (Wang *et al.* 2008), and theoretically reduces the oxygen delivery required to meet cellular energetic demand (Hillered *et al.* 1984).

Many of these theoretical benefits of hypercapnia may be deleterious when translated into the clinical realm. The immunomodulatory effects of hypercapnia delay bacterial clearance and worsen lung injury in pulmonary sepsis models (O’Croinin *et al.* 2008), of particular concern because pneumonia is a leading risk factor for ARDS (Gajic *et al.* 2011). Permissive hypercapnia’s adrenergic effects should be viewed with caution in light of recent data demonstrating increased morbidity and mortality in ARDS with β -agonist therapy from unclear mechanisms (Matthay *et al.* 2011; Gao Smith *et al.* 2012). Ongoing β_3 adrenergic receptor stimulation may also lead to progressive deterioration in cardiac function (Gauthier *et al.* 1996). Moreover, hypercapnic pulmonary vasoconstriction augments hypoxic pulmonary vasoconstriction and may worsen right heart strain (Mekontso Dessap *et al.* 2009). The reduced oxygen consumption from hypercapnia is at least partly due to mitochondrial dysfunction, shown to

impair alveolar epithelial cell proliferation (Vohwinkel *et al.* 2011).

Hypercapnia additionally may have deleterious effects directly on the injured lung. While most alveolar cells survive deformation-induced injury by translocating lipids to the injured plasma membrane (Vlahakis *et al.* 2002), hypercapnic acidosis impairs plasma membrane wound healing in ventilator-injured lungs in a pH-dependent fashion (Caples *et al.* 2009). Hypercapnia, independent of pH, also impairs alveolar fluid reabsorption by inhibiting epithelial Na⁺-K⁺-ATPase (Briva *et al.* 2007).

Finally, the cardiopulmonary response to hypercapnia may lead further to harm. Heightened ventilatory drive during hypercapnic acidosis may require deeper sedation or paralysis to prevent patient-ventilator dyssynchrony, excess tidal excursions and double-triggering of the ventilator. If not blunted, this heightened ventilatory drive may generate more negative pleural pressures (Yan *et al.* 1993), thereby increasing transpulmonary pressure and alveolar stress. Enhanced pulmonary blood flow via hypercapnia-induced catecholamine release may worsen alveolar damage, capillary leak and oedema formation (Broccard *et al.* 1998). Even during moderate alveolar overdistension insufficient to cause lung injury alone, increasing pulmonary blood flow leads to lung injury that otherwise would not occur (Guery *et al.* 1998). The combined effects of occult increased transpulmonary pressure and increased pulmonary blood flow from permissive hypercapnia may create precisely this environment wherein occult lung injury is likely to occur.

As investigation of permissive hypercapnia transitions to humans, focus must be placed on distinguishing the effect attributable to minimizing biomechanical injury from that attributable to hypercapnia itself. Whether the independent effect of hypercapnia *per se* is harmful or protective is unknown. At the very least, there is sufficient concern to give pause. Most data suggesting improved outcomes associated with permissive hypercapnia are probably attributable primarily to the mechanical benefits of limiting tidal volume and respiratory rate.

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References

- Amato MB, Barbas CS, Medeiros DM, Magaldi RB, Schettino GP, Lorenzi-Filho G, Kairalla RA, Deheinzelin D, Munoz C, Oliveira R, Takagaki TY & Carvalho CR (1998). Effect of a protective-ventilation strategy on mortality in the acute respiratory distress syndrome. *N Engl J Med* **338**, 347–354.
- Briva A, Vadász I, Lecuona E, Welch LC, Chen J, Dada LA, Trejo HE, Dumasius V, Azzam ZS, Myrianthefs PM, Batlle D, Gruenbaum Y & Sznajder JI (2007). High CO₂ levels impair alveolar epithelial function independently of pH. *PLoS One* **2**, e1238.
- Broccard AF, Hotchkiss JR, Kuwayama N, Olson DA, Jamal S, Wangenstein DO & Marini JJ (1998). Consequences of vascular flow on lung injury induced by mechanical ventilation. *Am J Respir Crit Care Med* **157**, 1935–1942.
- Brower RG, Matthay MA, Morris A, Schoenfeld D, Thompson BT & Wheeler A (2000). Ventilation with lower tidal volumes as compared with traditional tidal volumes for acute lung injury and the acute respiratory distress syndrome. *N Engl J Med* **342**, 1301–1308.
- Caples SM, Rasmussen DL, Lee WY, Wolfert MZ & Hubmayr RD (2009). Impact of buffering hypercapnic acidosis on cell wounding in ventilator-injured rat lungs. *Am J Physiol Lung Cell Mol Physiol* **296**, L140–L144.
- Fu Z, Costello ML, Tsukimoto K, Prediletto R, Elliott AR, Mathieu-Costello O & West JB (1992). High lung volume increases stress failure in pulmonary capillaries. *J Appl Physiol* **73**, 123–133.
- Gajic O, Dabbagh O, Park PK, Adesanya A, Chang SY, Hou P, Anderson H 3rd, Hoth JJ, Mikkelsen ME, Gentile NT, *et al.* (2011). Early identification of patients at risk of acute lung injury: evaluation of lung injury prediction score in a multicenter cohort study. *Am J Respir Crit Care Med* **183**, 462–470.
- Gao Smith F, Perkins GD, Gates S, Young D, McAuley DF, Tunnicliffe W, Khan Z & Lamb SE; BALTI-2 study investigators (2012). Effect of intravenous β -2 agonist treatment on clinical outcomes in acute respiratory distress syndrome. (BALTI-2): a multicentre, randomised controlled trial. *Lancet* **379**, 229–235.
- Gauthier C, Tavernier G, Charpentier F, Langin D & Le Marec H (1996). Functional β_3 -adrenoceptor in the human heart. *J Clin Invest* **98**, 556–562.

- Guery BP, deBoisblanc BP, Fialdes P, Sarphy TG, Nelson S, Chidiac C, Beaucaire G, Summer WR & Mason CM (1998). Pulmonary stress injury within physiological ranges of airway and vascular pressures. *J Crit Care* **13**, 58–66.
- Hillered L, Ernster L & Siesjö BK (1984). Influence of *in vitro* lactic acidosis and hypercapnia on respiratory activity of isolated rat brain mitochondria. *J Cereb Blood Flow Metab* **4**, 430–437.
- Hotchkiss JR, Blanch L, Murias G, Adams AB, Olson DA, Wangenstein OD, Leo PH & Marini JJ (2000). Effects of decreased respiratory frequency on ventilator-induced lung injury. *Am J Respir Crit Care Med* **161**, 463–468.
- Imai Y, Parodo J, Kajikawa O, de Perrot M, Fischer S, Edwards V, Cutz E, Liu M, Keshavjee S, Martin TR, Marshall JC, Ranieri VM & Slutsky AS (2003). Injurious mechanical ventilation and end-organ epithelial cell apoptosis and organ dysfunction in an experimental model of acute respiratory distress syndrome. *JAMA* **289**, 2104–2112.
- Malhotra A (2007). Low-tidal-volume ventilation in the acute respiratory distress syndrome. *N Engl J Med* **357**, 1113–1120.
- Marini JJ (2011). Dynamic hyperinflation and auto-positive end-expiratory pressure: lessons learned over 30 years. *Am J Respir Crit Care Med* **184**, 756–762.
- Marini JJ, Hotchkiss JR & Broccard AF (2003). Bench-to-bedside review: microvascular and airspace linkage in ventilator-induced lung injury. *Crit Care* **7**, 435–444.
- Matthay MA, Brower RG, Carson S, Douglas IS, Eisner M, Hite D, Holets S, Kallet RH, Liu KD, MacIntyre N, Moss M, Schoenfeld D, Steingrub J & Thompson BT (2011). Randomized, placebo-controlled clinical trial of an aerosolized β_2 -agonist for treatment of acute lung injury. *Am J Respir Crit Care Med* **184**, 561–568.
- Mekontso Dessap A, Charron C, Devaquet J, Aboab J, Jardin F, Brochard L & Vieillard-Baron A (2009). Impact of acute hypercapnia and augmented positive end-expiratory pressure on right ventricle function in severe acute respiratory distress syndrome. *Intensive Care Med* **35**, 1850–1858.
- O’Croinin DF, Nichol AD, Hopkins N, Boylan J, O’Brien S, O’Connor C, Laffey JG & McLoughlin P (2008). Sustained hypercapnic acidosis during pulmonary infection increases bacterial load and worsens lung injury. *Crit Care Med* **36**, 2128–2135.
- Oeckler RA, Lee W-Y, Park M-G, Kofler O, Rasmussen DL, Lee H-B, Belete H, Walters BJ, Stroetz RW & Hubmayr RD (2010). Determinants of plasma membrane wounding by deforming stress. *Am J Physiol Lung Cell Mol Physiol* **299**, L826–L833.
- Papazian L, Forel JM, Gacouin A, Penot-Ragon C, Perrin G, Loundou A, Jaber S, Arnal JM, Perez D, Seghboyan JM, Constantin JM, Courant P, Lefrant JY, Guérin C, Prat G, Morange S & Roch A (2010). Neuromuscular blockers in early acute respiratory distress syndrome. *N Engl J Med* **363**, 1107–1116.
- Perlman CE, Lederer DJ & Bhattacharya J (2011). Micromechanics of alveolar edema. *Am J Respir Cell Mol Biol* **44**, 34–39.
- Talmor D, Sarge T, Malhotra A, O’Donnell CR, Ritz R, Lisbon A, Novack V & Loring SH (2008). Mechanical ventilation guided by esophageal pressure in acute lung injury. *N Engl J Med* **359**, 2095–2104.
- Talmor D, Sarge T, O’Donnell CR, Ritz R, Malhotra A, Lisbon A & Loring SH (2006). Esophageal and transpulmonary pressures in acute respiratory failure. *Crit Care Med* **34**, 1389–1394.
- Taylor CT & Cummins EP (2011). Regulation of gene expression by carbon dioxide. *J Physiol* **589**, 797–803.
- Vlahakis NE, Schroeder MA, Pagano RE & Hubmayr RD (2002). Role of deformation-induced lipid trafficking in the prevention of plasma membrane stress failure. *Am J Respir Crit Care Med* **166**, 1282–1289.
- Vohwinkel CU, Lecuona E, Sun H, Sommer N, Vadász I, Chandel NS & Sznajder JI (2011). Elevated CO₂ levels cause mitochondrial dysfunction and impair cell proliferation. *J Biol Chem* **286**, 37067–37076.
- Wang Z, Su F, Bruhn A, Yang X & Vincent JL (2008). Acute hypercapnia improves indices of tissue oxygenation more than dobutamine in septic shock. *Am J Respir Crit Care Med* **177**, 178–183.
- Yan S, Lichros I, Zakynthinos S & Macklem PT (1993). Effect of diaphragmatic fatigue on control of respiratory muscles and ventilation during CO₂ rebreathing. *J Appl Physiol* **75**, 1364–1370.
- Yoshida T, Uchiyama A, Matsuura N, Mashimo T & Fujino Y (2012). Spontaneous breathing during lung-protective ventilation in an experimental acute lung injury model. *Crit Care Med* **40**, 1578–1585.

Conflict of interest

J. R. Beitler and R. D. Hubmayr have no conflicts of interest to declare. A. Malhotra previously received consulting and/or research income from Philips, SGS, SHC, Apnex, Apnicure and Pfizer, but has relinquished all outside personal income since May 2012.