

Developmental Expression of Rat Transforming Growth Factor- α mRNA

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Expression of the gene encoding transforming growth factor- α (TGF α) was examined in developing rat embryos by using a cloned TGF α cDNA as a hybridization probe. Northern blot analysis of RNA isolated from whole fetuses revealed that TGF α mRNA was present at relatively high levels in 8- through 10-day-old embryos and then declined to the low or undetectable level, which is characteristic of adult tissues before birth. The level of TGF α mRNA present during early gestation was similar to that present in retrovirus-transformed cells in culture, suggesting that TGF α expression is not highly localized in the embryo. These observations are consistent with the hypothesis that TGF α plays a role in development, possibly as a fetal growth factor.

Transforming growth factors (TGFs) are potent mitogenic polypeptides produced by a variety of retrovirus-transformed cells and certain tumors (17). In addition, TGFs confer on normal cells in culture some of the phenotypic properties associated with transformation, including loss of contact inhibition and growth in soft agar (3). Two distinct types of TGFs have been identified. TGF α is structurally related to epidermal growth factor (EGF) (6, 11, 12, 15) and competes for binding to the EGF receptor (13). TGF β , on the other hand, potentiates the growth-stimulating activity of TGF α and demonstrates no affinity for the EGF receptor (1).

Recently, cDNA clones encoding human (4) and rat (9) TGF α 's have been isolated; the nucleotide sequence of these cDNAs suggests that human and rat TGF α 's are initially synthesized as polypeptides of 160 and 159 amino acids, respectively, and that these larger forms exist as transmembrane proteins. Release of the smaller TGF α from the larger form apparently does not occur through the action of a serine-like protease but, rather, through an unusual proteolytic cleavage between alanine and valine residues at both the amino and carboxy termini of the rat and human molecules.

Using the cloned rat TGF α cDNA as a hybridization probe, we have previously shown that retrovirus-transformed cells express a 4.5-kilobase TGF α mRNA (9). A similar-sized transcript is also detected at considerably lower levels in some adult rat tissues but not in the corresponding untransformed cell line. Here we report that Northern analysis of developing rat embryos indicates that TGF α transcripts are expressed at relatively high levels during early embryogenesis but then decline to low levels before birth. These results are consistent with the hypothesis that TGF α functions as a developmental growth factor.

Pools of rat embryos were collected at various time points from time-mated Sprague-Dawley rats. RNA was isolated by homogenization of embryos or cells in 4 M guanidine

thiocyanate and sedimentation through 5.7 M CsCl (2, 5). Poly(A)⁺ RNA was selected by chromatography on oligo(dT)-cellulose, and samples (5 μ g) were electrophoresed through 1% agarose gels containing formaldehyde (10). RNA was transferred to nitrocellulose and probed with the nick-translated TGF α cDNA insert from prTGF0,2 (9). Hybridization was carried out at 45°C in 50% formamide-1 \times Denhardt reagent-5 \times SSC (1 \times SSC is 0.15 M NaCl plus 0.015 M sodium citrate)-25 mM NaH₂PO₄-herring sperm DNA (200 μ g/ml) and washes were performed in 2 \times SSC-0.1% sodium dodecyl sulfate at 68°C. At each time point, the quality of the poly(A)⁺ RNA was independently assessed by electrophoresis through 1% agarose-8 M urea gels, followed by staining with ethidium bromide.

Figure 1 shows a Northern analysis of poly(A)⁺ RNA isolated from pools of whole rat embryos at various stages of development. It is apparent that rat embryos contain, during the early stages of embryogenesis, a TGF α transcript identical in size to the TGF α mRNA contained in retrovirus-transformed rat cells (9; see Fig. 2). The levels of this transcript are highest in 8- through 10-day-old embryos (Fig. 1, lanes A to C) and then decline, beginning at day 11, (lane

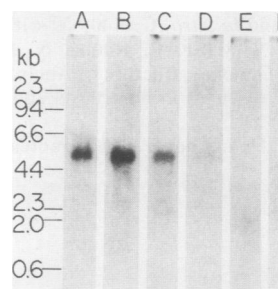


FIG. 1. Northern blot analysis of embryonic rat poly(A)⁺ RNAs probed with a cloned TGF α cDNA. Poly(A)⁺ RNAs are from pools of whole 8-day-old (lane A), 9-day-old (lane B), 10-day-old (lane C), 11-day-old (lane D), 13-day-old (lane E), and 18-day-old (lane F) rat embryos. The markers (in kilobases) are *Hind*III-digested λ DNA.

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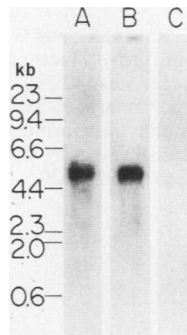


FIG. 2. Northern blot analysis of rat poly(A)⁺ RNAs probed with the TGF α cDNA clone. Samples are from a pool of whole 9-day-old rat embryos (lane A) and feline sarcoma virus-transformed (lane B) and untransformed (lane C) Fischer rat embryo cells. The markers (in kilobases) are *Hind*III-digested λ DNA.

D) to undetectable levels by day 13 (lane E) and day 18 (lane F). In other experiments, we have observed that TGF α mRNA sequences can be detected at day 18, but they are consistently present at much lower levels than at the earlier stages of development (data not shown). In addition, we have repeatedly observed that the levels of TGF α mRNA are higher at day 9 than at either day 8 or day 10, suggesting that it represents a peak of TGF α expression; we have not examined embryos before day 8 due to the difficulty in obtaining pure material.

The levels of TGF α transcripts present in 9-day-old embryos (Fig. 2, lane A) are similar to those observed in a clone of feline sarcoma virus-transformed rat fibroblasts (lane B) originally isolated because of its aggressive growth in soft agar and relatively high production of TGF α (19). Lane C contains, for comparison, an equivalent amount of poly(A)⁺ RNA isolated from the untransformed rat fibroblast cell line. The finding that poly(A)⁺ RNA from whole rat embryos contains levels of TGF α mRNA similar to those found in a relatively high-producing cell line suggests that TGF α expression in the fetus is not highly localized.

The observation that rat embryos contain relatively high levels of TGF α transcripts suggests that TGF α plays an important role in normal development. Viewed in this light, the abnormal expression of TGF α in the transformed state is analogous to the abnormal expression of developmentally regulated oncogenes (16). Although the appearance of TGF α transcripts in embryos does not necessarily imply that these transcripts are translated to produce TGF α polypeptide, our results are consistent with the earlier observation of Nexo et al. (14) that fetal mice at 11 days of gestation contained material that was capable of binding the EGF receptor but was not recognized in an EGF radioimmunoassay. Subsequently, Twardzik et al. (18) showed that growth factors with some of the characteristic properties of TGF α could be isolated from 12- to 13-day-old mouse embryos. The observation here that rat embryos of a similar gestational age contain TGF α transcripts supports the hypothesis that this growth factor activity derives from the embryo and is not of maternal origin. Finally, it is of interest to note that the growth factor activity isolated from mouse by Twardzik et al. (18) comprised two size classes with apparent molecular weights of 20,000 and 10,000. The relationship of the larger activity to the initial translation product predicted by the

TGF α cDNA sequence, as well as the structural form of TGF α expressed in the rat embryos, is presently unclear.

The expression of TGF α in embryos suggests that it might have a normal function as a fetal growth factor, possibly as a developmental analog of EGF. Interestingly Nexo et al. (14) found a significant rise in immunoreactive mouse EGF between days 15 and 17. This is approximately the time when in rat gestation the levels of TGF α mRNA have declined. An alternate explanation is suggested by studies which implicate the action of TGF α in the process of bone resorption (7, 8). Consistent with this model, the expression of TGF α in embryos may be instrumental in the embryonic remodeling which occurs during development.

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LITERATURE CITED

1. Anzano, M. A., A. B. Roberts, C. A. Meyers, A. Komoriya, L. C. Lamb, J. M. Smith, and M. B. Sporn. 1982. Synergistic interaction of two classes of transforming growth factors from murine sarcoma cells. *Cancer Res.* **42**:4776-4778.
2. Chirgwin, J. M., A. E. Przybyla, R. J. MacDonald, and W. J. Rutter. 1979. Isolation of biologically active ribonucleic acid from sources enriched in ribonuclease. *Biochemistry* **18**:5294-5299.
3. DeLarco, J. E., and G. J. Todaro. 1978. Growth factors from murine sarcoma virus transformed cells. *Proc. Natl. Acad. Sci. USA* **75**:4001-4005.
4. Derynck, R., A. B. Roberts, M. E. Winkler, E. Y. Chen, and D. V. Goeddel. 1984. Human transforming growth factor- α : precursor structure and expression in *E. coli*. *Cell* **38**:287-297.
5. Glisin, V., R. Crkvenjakov, and C. Byus. 1974. Ribonucleic acid isolated by cesium chloride centrifugation. *Biochemistry* **13**:2633-2638.
6. Gregory, H. 1975. Isolation and structure of urogastrone and its relationship to epidermal growth factor. *Nature (London)* **257**:325-327.
7. Ibbotson, K. J., S. M. D'Sonza, K. W. Ng, C. K. Osborne, M. Niall, T. J. Martin, and G. R. Mundy. 1983. Tumor-derived growth factor increases bone resorption in a tumor associated with humoral hypercalcemia of malignancy. *Science* **221**:1292-1294.
8. Ibbotson, K. J., D. R. Twardzik, S. M. D'Sonza, W. R. Hargreaves, G. J. Todaro, and G. R. Mundy. 1985. Stimulation of bone resorption in vitro by synthetic transforming growth factor- α . *Science* **228**:1007-1009.
9. Lee, D. C., T. M. Rose, N. R. Webb, and G. J. Todaro. 1985. Cloning and sequence analysis of a cDNA for rat transforming growth factor- α . *Nature (London)* **313**:489-491.
10. Maniatis, T., E. F. Fritsch, and J. Sambrook. 1982. Molecular cloning. A laboratory manual. Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.
11. Marquardt, H., M. W. Hunkapiller, L. E. Hood, and G. J. Todaro. 1984. Rat transforming growth factor type 1: structure and relation to epidermal growth factor. *Science* **223**:1079-1082.
12. Marquardt, H., M. W. Hunkapiller, L. E. Hood, D. R. Twardzik, J. E. DeLarco, J. R. Stephenson, and G. J. Todaro. 1983. Transforming growth factors produced by retrovirus-transformed rodent fibroblasts and human melanoma cells: amino acid sequence homology with epidermal growth factor. *Proc. Natl. Acad. Sci. USA* **80**:4684-4688.
13. Marquardt, H., and G. J. Todaro. 1982. Human transforming growth factor. Production by a melanoma cell line, purification and initial characterization. *J. Biol. Chem.* **257**:5220-5225.
14. Nexo, E., M. D. Hollenberg, A. Figueora, and R. M. Pratt. 1980. Detection of epidermal growth factor-urogastrone and its receptor during fetal mouse development. *Proc. Natl. Acad. Sci. USA* **77**:2782-2785.
15. Savage, C. R., Jr., T. Inagami, and S. Cohen. 1972. The primary

- structure of epidermal growth factor. *J. Biol. Chem.* **247**:7612-7621.
16. **Slamon, D. J., and M. J. Cline.** 1984. Expression of cellular oncogenes during embryonic and fetal development of the mouse. *Proc. Natl. Acad. Sci USA* **81**:7141-7145.
 17. **Todaro, G. J., C. Fryling, and J. E. DeLarco.** 1980. Transforming growth factors produced by certain human tumors: polypeptides that interact with epidermal growth factor receptors. *Proc. Natl. Acad. Sci. USA* **77**:5258-5262.
 18. **Twardzik, D. R., J. E. Ranchalis, and G. J. Todaro.** 1982. Mouse embryonic transforming growth factors related to those isolated from tumor cells. *Cancer Res.* **42**:590-593.
 19. **Twardzik, D. R., G. J. Todaro, F. H. Reynolds, Jr., and J. R. Stephenson.** 1983. Similar transforming growth factors (TGFs) produced by cells transformed by different isolates of feline sarcoma virus. *Virology* **124**:201-207.