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# Impact of Aging Brain Circuits on Cognition

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## **Abstract**

Brain networks that engage the hippocampus and prefrontal cortex are central for enabling effective interactions with our environment. Some of the cognitive processes that these structures mediate, such as encoding and retrieving episodic experience, wayfinding, working memory and attention are known to be altered across the lifespan. As illustrated by examples given below, there is remarkable consistency across species in the pattern of age-related neural and cognitive change observed in healthy humans and other animals. These include changes in cognitive operations that are known to be dependent on the hippocampus, as well as those requiring intact prefrontal cortical circuits. Certain cognitive constructs that reflect the function of these areas lend themselves to investigation across species allowing brain mechanisms at different levels of analysis to be studied in greater depth.

#### **Keywords**

prefrontal cortex; hippocampus; synapse; spatial learning; working memory

### **Overview**

Over the past several decades it has become clear that multiple cognitive trajectories can be experienced during the aging process, both in humans and in other animals. A fundamental dichotomy in the human case is whether individuals are on a path towards dementia, or on a path towards reasonably intact cognitive function over their lifespan. Epidemiological studies have resulted in varied estimates of what proportion of us will fall into one or the other category. While some of the apparently contradictory findings are attributable to issues of sampling bias, at least one group has used a design that has overcome this limitation. Plassman et al. (2007) examined the prevalence of dementia in a representative sample taken from all regions of the United States in people over 70 years of age. The proportion of those 71 and older who could be categorized as being demented was 14%, while 86% were not. This suggests to some (e.g., Wagster et al., 2012; Small et al., 2011; Roberson et al., 2012) that it is critical to understand normal cognitive aging processes in their own right, not only as a backdrop to understanding diseases that can co-occur in aging. The data reviewed here will be taken from studies that examine this non-dementia aging trajectory, focusing on the more moderate cognitive changes that occur across the 86% of us over 71 years. Within this category, there are clear individual differences, as the impact of the aging process is far from uniform.

Two primary brain circuits will be reviewed here – the hippocampus and frontal cortex. Both are known to be important for cognitive operations in humans and other animals and both show functional changes with age. Because no brain region operates independently, when the data are available, interactions among structures with age will also be discussed. This overview is not intended to be comprehensive. Rather, selected experiments in human subjects and animal models are highlighted that illustrate the types of neurobiological change that alter these neural circuits and contribute to cognitive aging across species.

## Age-related cognitive changes that depend on hippocampal circuits

The hippocampus is critically involved in the formation and utilization of 'cognitive maps'. Tolman's classic paper entitled "Cognitive maps in rats and man" (1948) outlined the kind of choices animals make in navigating mazes or finding ones way home. He described two learning strategies used to navigate: one that involves learning the configuration of landmarks in the environment (place), and the other involves learning a particular route (response). O'Keefe and Nadel (1978) argued that in addition to place and response strategies, cue strategies consisting of the approach or avoidance of some salient cue can be used. They also proposed that the hippocampus is critically involved in place learning and the formation and flexible utilization of cognitive maps that are independent of habitual routes or salient cues. Although spatial cognition is a broad psychological construct that can engage multiple brain circuits, the hippocampus appears to be necessary for wayfinding (place learning), while striatal systems are critical for route learning. Moreover, this general concept of regional specialization appears to hold across mammalian species. An example from the human literature is the finding that when humans navigate a virtual environment using a place strategy, the hippocampus is activated as assessed by neuroimaging, whereas when the participants use a response strategy to navigate, the caudate nucleus is activated (Iaria et al., 2003).

What happens to hippocampus-dependent behavior during aging? If rats are given the opportunity to learn a T-maze problem that can be solved equally effectively by using a place, response or cue strategy, each animal adopts a favored strategy to solve the problem. Probe trials can be used to test for spontaneous strategy use. When young and old rats are compared, there are no differences between age groups in number of trials to learn the task, but the predominant strategy chosen by young rats was 'place', whereas old rats chose 'response' (Barnes *et al.*,1980). These data indicate a shift away from hippocampus-dependent behaviors by old rats, if other solutions are equally effective. While this observation is consistent with hippocampal dysfunction, the experiment did not test spatial learning directly. When old rats are forced to use a place strategy for optimal task performance, direct evidence is found for spatial learning and memory deficits. Examples include deficits on the Barnes maze (e.g., Barnes, 1979) and Morris watermaze (e.g., Gage *et al.*, 1984) spatial learning and memory tasks (for review, Foster *et al.*, 2012). Rapp *et al.* (1997) has also shown spatial strategy changes in aged rhesus macaques.

Advanced age also impacts navigational abilities in humans (e.g., Uttl & Graf, 1993; Burns, 1999; Driscoll *et al.*, 2005; Moffat *et al.*, 2006; Iaria *et al.*, 2009; Jansen *et al.*, 2010). For example, Head and Isom (2010) examined young and older adult performance on two different types of navigational tasks – one that required wayfinding, and the other that required route learning. The virtual maze environment was identical in the two tasks. For the wayfinding task the participants were allowed to freely explore the entire environment, and then at test, were asked to find their way to a particular landmark using the shortest route. For the route learning condition, the participants learned a specific route through the virtual environment marked by arrows, and then at test, the arrows were removed. For the behavior, older participants ranging in age from 56-88, as compared to a young group (18-22 years),

showed significantly impaired performance in acquiring a specific route through the environment, although the difference was relatively small. There were large age differences, however, observed between age groups in wayfinding. Additionally, structural MRI scans were performed on the older subjects, and volumes of the hippocampus, caudate and prefrontal cortex were obtained. There were no significant associations between prefrontal cortex volume and navigation performance, but there were associations with the other two structures examined. The volume of the hippocampus (but not caudate, Figure 1 part C) was associated with wayfinding accuracy - those older individuals with the largest hippocampi showed the shortest distances to find the landmark (Figure 1 part A). The volume of the caudate (not hippocampus, Figure 1 part B), on the other hand was associated with accuracy in the route learning task – the older individuals with the largest caudate volume also exhibited the most accurate routes (Figure 1, part D). While this study did not explicitly examine whether the difficulty that older adults have in the use of cognitive maps is in their formation or their use, data from Iaria *et al.* (2009) suggest that older adults take longer to form effective maps and also use them less accurately once acquired.

While there are many more demonstrations that behaviors dependent on the hippocampus are altered in aging, those described above illustrate one consistent cognitive change that is observed across species boundaries – namely impaired wayfinding. This consistent observation provides an opportunity to examine these behaviors in relation to the neurobiological changes that may be responsible for this cognitive outcome.

# Age-related anatomical changes in the hippocampus

One possible contribution to age-related declines on hippocampus-dependent tests was mentioned in the previous section - change in volume. While noninvasive imaging methods have great power to assess brains in the absence of potential histological artifacts, the reasons for the volume changes cannot be specified at the resolution of these methods, and additional cell, synapse counts and morphological analyses are required. Nonetheless, various MRI techniques can be used across species to help dissect changes due to aging versus those of prodromal disease. Because the full pathological syndrome known as Alzheimer's disease (AD) only occurs spontaneously in humans, animal models that age, but do not exhibit AD, are helpful guides for understanding and separating what is normal from what is pathological. Not surprisingly, in the human cognitive aging literature, there are reports of hippocampal atrophy across age (e.g., O'Brien et al., 1997; Tisserand et al., 2000; Raz et al., 2004; Raz et al., 2010), along with reports of stability of overall hippocampal volume during aging (e.g., Van Petten, 2004; Sullivan et al., 2005). Studies examining aging in the nonhuman primate and rodent using structural MRI methods suggest that hippocampal volume in these animals is preserved across age (Shamy et al., 2006; Alexander et al., 2011); while frontal cortical gray matter volumes do show changes with age in both species (Alexander et al., 2008; Shamy et al., 2011; Alexander et al., 2011).

Using a high resolution variant of fMRI developed to evaluate resting state metabolism within hippocampal substructures, Small and his colleagues (Small *et al.*, 2002; Small *et al.*, 2004; Moreno *et al.*, 2007) have shown reduced metabolism in the dentate gyrus of aged mice, monkeys and humans. In animal models, this correlated with memory impairment. Thus, examining activity within hippocampal subregions provides a sensitive method to detect functional changes, even if volume does not differ. Furthermore, it has also been shown that taking individual health into account helps to explain subregion volume differences across age. Shing *et al.* (2011) report that reduced CA3 and dentate gyrus volume in older adults correlates with memory decline, while reduced volume of the CA1 region correlates with hypertension. Additionally, there is evidence in human samples for age-related signal degradation of white matter in the region of the perforant pathway (Yassa

et al., 2010), the main input to the hippocampus from the entorhinal cortex, reduced white matter volume in this region (Stoub et al., 2012), and dendritic diffusion defects in the dentate gyrus-CA3 region (Yassa et al., 2011). Interestingly, data obtained from electrically-evoked field potential recordings in the dentate gyrus of aged rats (Barnes & McNaughton, 1980; Foster et al., 1991) predicted entorhinal axon collateral pruning. The observation that led to this suggestion was the fact that there was no change in the stimulus current necessary to elicit responses from these axons (i.e., no threshold change), but the maximum amplitude of the compound action potential response was reduced in old compared to young rats. Assuming no layer II entorhinal cortical cell loss with aging (confirmed in rats, Merrill et al., 2001: Rapp et al., 2002; and in monkeys, Gazzaley et al., 1997), the reduced maximal amplitude in old rats suggested that there were fewer entorhinal axon collateral fibers running in the perforant pathway. This hypothesis fits rather well with the MRI observation of age-related reductions in perforant path white matter volume in normal aged humans reported by Stoub et al. (2012), but direct counts of entorhinal axon collaterals has yet to be made in aged rats.

With the advent of stereological methods, one feature of the aging hippocampus that can be ruled out as significantly contributing to volume or metabolic changes is cell number. That is, cell numbers are preserved in normal aging in the principal cell types of the hippocampus (granule cells, CA1 and CA3 pyramidal cells) in humans (e.g., West et al., 1993), nonhuman primates (e.g., Keuker et al., 2003) and rodents (Rapp & Gallagher, 1996; Rassmussen et al., 1996). A lack of dendritic deterioration has also been reported for hippocampal cells in rodents and humans (e.g., Flood et al., 1987; Turner & Deupree, 1991; Flood, 1993), and alterations in dendritic spines are region-specific, and will be discussed in terms of synapse number below. In rodents, there is loss of axospinous synapses from the layer II medial entorhinal cortex projection to granule cells (Geinisman et al., 1992) and reduced synaptophysin staining in the dendritic region of CA3 pyramidal cells (Smith et al., 2000) during aging. The synaptic input to CA1 pyramidal cells from CA3, however, does not show synapse reduction (Geinisman et al., 2004). However, a subset of the synaptic contacts in this region exhibit reduced postsynaptic density size (Nicholson et al., 2004), and electrophysiological evidence suggests that this group of synapses may reflect nonfunctional, 'silent' synapses (Barnes et al., 1997; Burke & Barnes, 2010).

Clearly, anatomical changes do occur within the hippocampus in normal aging, although they are rather subtle compared with those known to occur in pathological conditions that arise during aging, such as Alzheimer's disease (e.g., Ballard *et al.*, 2011). The impact that these neurological changes have on plasticity and circuit function is discussed below.

# Age-related physiological changes in the hippocampus

Hippocampal cell function in aging animals is strikingly well preserved. In rats it is possible to study the detailed biophysics of individual hippocampal principal cells using *in vitro* recording methods. Most biophysical properties in these aging cells do not change (for reviews, Burke & Barnes, 2006; Hoang *et al.*, 2012), with a small number of exceptions, including a larger after-hyperpolarizing potential in CA1 pyramidal cells of old rats (e.g., Landfield & Pitler, 1984). This change may be due to an increased number of L-type calcium channels in old CA1 cells (e.g., Thibault & Landfield, 1996). This increase in channel numbers is hypothesized to lead to age-related disruption of neuronal calcium homeostasis, suggesting an interesting potential therapeutic target (for review, Kumar *et al.*, 2009). There are two additional electrophysiological changes that are observed in all 3 subregions of the hippocampus. These include reduced amplitude of the stimulation-induced cholinergic slow EPSP (Shen & Barnes, 1996), and an increase in gap junction-mediated electrotonic coupling between aged CA1 and CA3 pyramidal cells, as well as granule cells

(Barnes *et al.*, 1987). The former age-related change suggests reduced effectiveness of a modulatory input, and the latter increased electrical communication between cells. The alterations described above are consistent with both increased excitability (increased calcium conductance, increased electrotonic coupling) and decreased excitability (reduced cholinergic modulation) of old cells. Taken together the data suggest a complex set of mechanisms at play that may tend to keep overall cell function stable in the aged brain.

Two examples can be offered that may reflect cellular adaptation in aged hippocampal circuits. The first involves the fact that the synapses that arise from the medial entorhinal cortex and make contact within the middle third of the granule cell dendritic tree, are reduced in number by about one third in old rats (e.g., Geinisman et al., 1992). The remaining synapses in that dendritic region, however, are more powerful – the depolarization caused by activation of a single synapse is larger in the old rats (Barnes & McNaughton, 1980). Fewer but stronger synapses could be interpreted as an adaptive response, keeping overall depolarization levels of the granule cells within some optimal range. Another example involves the fact that there have been consistent reports of increased AHP amplitudes of old CA1 pyramidal cells measured in vitro (e.g., Landfield & Pitler, 1984; Disterhoft et al., 1996). The inference made from these intracellular recording studies is that this increased hyperpolarization after an action potential should slow the repolarization that enables another action potential to be generated, and thus predicts reduced behavior-induced firing rates for old CA1 pyramidal cells. A slowing of CA1 cell firing rates, however, is not observed in the intact, freely-behaving aged rat (e.g., Markus et al., 1994; Shen et al., 1997; Schimanski et al., 2013), suggesting that an adaptation has occurred that keeps output rates constant in these aged cells.

There are a number of examples of changes in the function of plasticity mechanisms that occur within the hippocampus. Because experimentally induced changes in synaptic communication are thought to underlie the acquisition, storage, consolidation and reconsolidation of memory (e.g., Bliss et al., 2007), the processes of long-term potentiation (LTP) and depression (LTD) are prime targets for studying the physiology of altered cognitive functions observed during aging. The first demonstration that LTP and behavioral performance may be related was provided by an experiment conducted in awake, freelybehaving young and old rats, in which LTP was induced at the perforant path-granule cell synapse. In this study, individual differences in the durability of LTP were significantly correlated with spatial memory accuracy, and this behavior/plasticity relationship was observed in each age group independently (Barnes, 1979). The same relationship between LTP durability and spatial behavior on the circular platform task was also observed at synapses in CA1 in young and old mice (Bach et al., 1999). Differences in induction of LTP have also been noted (e.g., Deupree et al., 1993; Moore et al., 1993; Barnes et al., 2000), and Foster and his colleagues have shown that LTD and LTP reversal are easier to induce in older, spatial memory-impaired rats (e.g., Norris et al., 1996). Additionally, a behaviorallyinduced form of plasticity dependent on NMDA receptor mechanisms (Ekstrom et al., 2001) is altered in aged rats (Shen et al., 1997). This behavior involves an expansion and backwards shift of place-specific firing of hippocampal cells that can be observed when rats engage in repeated route following behaviors. Mehta et al. (1997) have called this phenomenon place field expansion plasticity. Although the description of hippocampal cell firing characteristics is elaborated below, it is important to note here that along with agerelated deficits in plasticity measured in response to artificial electrical stimulation, behaviorally-driven LTP-like plasticity mechanisms are also observed to change with age. Moreover, this place field expansion plasticity is reminiscent of Hebb's (1949) theoretical idea of phase sequences in cell assemblies, which he postulated could provide a means to encode sequences or episodes of experience. Together, these data suggest clear changes in synaptic plasticity mechanisms in the normally aging brain, as well as potential mechanisms

through which therapeutic targets can be developed (e.g., Bach *et al.*, 1999; Burke *et al.*, 2005; Foster, 2006; Huang & Kandel, 2006; Rose *et al.*, 2007; Bodhinathan *et al.*, 2010).

There have been a number of experiments that have investigated the potential causes for these types of age-related plasticity deficits in aging. One approach has been to examine the role of immediate early genes (IEGs) in these processes. Arc (Lyford et al., 1995) has been useful in this regard because when Arc protein is knocked down in hippocampus of young rats, LTP decays significantly faster compared to the case when normal levels of Arc are present, and spatial memory consolidation is also disrupted (Guzowski et al., 2000; Plath et al., 2006). Penner et al. (2011) examined Arc mRNA activity in hippocampal cells of young and aged rats induced by spatial behaviors. The expression of Arc within cells provides an activity marker for those neurons that participate in a recent behavioral event (Guzowski et al., 1999). They used methods that allowed behavior-induced Arc-positive cells to be counted, and Arc mRNA to be quantified by rtPCR within the same animal and cell type. For example, in CA1, the same numbers of pyramidal cells across age groups express Arc following exploratory behavior, but old pyramidal cells transcribe less Arc (Penner et al., 2011). Epigenetic mechanisms such as DNA methylation are known to affect RNA expression, and can influence cell function by altering the amount of RNA transcribed from a gene. Interestingly, Penner et al. (2011) also observed a very distinct pattern of methylation change with age in the Arc gene in CA1 cells. Thus, it appears that aging is accompanied by significant changes in epigenetic regulation of at least this important plasticity gene. These data, taken together with more recent observations suggesting that there is reduced coordination of epigenetic regulation dynamics of plasticity genes in aging (Castellanos et al., 2012), strongly suggest that epigenetic mechanisms are fundamental to both age-related changes in circuit modifiability and cognition. Deeper understanding of how transcription is regulated by chromatin modification dynamics is going to be central in choosing targets for therapeutic optimization of cognition during aging.

When the activity of ensembles of hippocampal cells are examined in behaving young and old rats, interesting changes are observed in cell population dynamics between spatial experiences. O'Keefe and Dostrovksy (1971) first described the activity patterns of individual hippocampal cells as place-specific, and named these cells "place cells". As discussed earlier, this was a prime impetus behind the development of the idea that the hippocampus had an important role in cognitive mapping, and in spatial strategies that drive behavioral performance. All three principal hippocampal cell types show place-related firing, although only CA1 and CA3 pyramidal cells have been recorded and compared across age groups. While there is stability of CA1 and CA3 spatial firing patterns within a given recording session for young and old rats (i.e., the distribution of cell firing in the first half of the behavior session, is highly correlated with the distribution of place-specific firing in the second half of the session), between session dynamics are different between age groups, and across cell types. For CA1, the cell firing pattern, or "map", is stable in young rats across two daily sessions in an identical environment. For old rats, however, the cell firing pattern can completely change from one session to the next, and occurs on about a third of the recording days for any individual old rat. In other words, the hippocampus "remaps", as though the first and second session are recorded in different environments (Barnes et al., 1997). For CA3, on the other hand, when environments are changed between sessions, young rats remap appropriately between the first and second sessions. For old rats, however, the hippocampus appears to sometimes retrieve the same map for the two distinct environments. In this case, the hippocampus fails to remap (Wilson et al., 2005). It is likely that these altered dynamics of hippocampal representation of space contribute to the spatial behavioral differences noted between age groups.

In the context of changing circuit dynamics with age, it is important to highlight conditions under which the hippocampus is activated differentially in young and older adults in fMRI experiments. One, among a number of examples of this, is a study by Maguire and Frith (2003) who used fMRI imaging methods that assessed hippocampal and medial prefrontal cortical network activation in young and older adults. While in the scanner, the participants retrieved details of specific episodic memories for autobiographical events. These past experiences were obtained from each individual before being scanned, and the details of the autobiographic memories during test retrieval were matched for detail across age groups. Activation comparisons were between retrieval of autobiographical events and general semantic knowledge. There was no difference between age groups in prefrontal cortical activation during retrieval, but there were differences between groups in hippocampal activation. As in previous studies of autobiographical retrieval, there was significant activation of the left hippocampus in young participants. For the old participants, however, there was significant activation of both left and right hippocampi, suggesting that the older adults recruited additional circuits when recalling episodes from specific times and contexts. This result may suggest a neural compensatory process for recall of detailed episodes, or different strategies used for recall in the older adults. Regardless, it is likely that this difference in regional activation is initiated because of functional changes within the circuits responsible for these behaviors.

# Age-related cognitive changes that depend on frontal cortical circuits

One of the most replicated results in the cognitive aging literature is that cognitive processes that rely on frontal cortical areas are particularly vulnerable to the effects of aging. In particular, maintaining a representation through working memory is reliably affected (e.g., Alexander et al., 2012; Störmer et al., 2012). Older adults show a decline in performance on tasks that require updating items in working memory (e.g., Hartman et al., 2001), in accuracy during trials with larger memory loads (e.g., Cappell et al., 2010) and in responding after a delay (e.g., Lyons-Warren et al., 2004). Similarly, aged nonhuman primates and rats also show deficits in tasks that require working memory (for review Bizon et al., 2012). That is, when a delay is incorporated into the design of the task, aged animals are particularly disadvantaged (e.g., Bartus et al., 1978; Rapp & Amaral, 1989; Muir et al., 1999; Grottick & Higgins, 2002; Ramos et al., 2003; Smith et al., 2004; Bizon et al., 2009). Two widely used working memory tasks implemented in monkey experiments include the delayed response task (DR), which relies on the dorsolateral prefrontal cortex (PFC; Goldman & Rosvold, 1970; Passingham, 1985; Funahashi et al., 1993) and the delayed nonmatching-to-sample task (DNMS), which relies on the ventromedial PFC (Arnsten & Goldman-Rakic, 1990, Figure 2C). In the DR task, a monkey is required to remember a spatial location on a screen over a brief delay period, after which it must make a saccade towards that location in order to receive a juice reward. Aged monkeys are slower to acquire the task and are impaired when longer delays are imposed (e.g., Bartus et al., 1978; Rapp & Amaral, 1989; Bachevalier et al., 1991). In the DNMS task, a monkey is first exposed to one object that it displaces to receive a reward. After a delay period, the monkey is exposed to two objects and the task requires that the novel object is displaced for the 'nonmatch' requirement of the task. Reward is obtained when the displaced object was the one that was not previously presented. As with the DR task, aged monkeys are slower to learn the task and perform more poorly as delay intervals are increased (Shamy et al., 2011).

Behavioral flexibility is another frontal-dependent cognitive process that is compromised with aging. This has been studied using a variety of tasks, notably extradimentional set shifting and reversal tasks in humans (Ridderinkhof *et al.*, 2002; Marschner *et al.*, 2005; Weiler *et al.*, 2008), monkeys (Bartus *et al.*, 1979; Lai *et al.*, 1995; Voytko, 1999; Moore *et al.*, 2003) and rats (Stephens *et al.*, 1985; Barense *et al.*, 2002; Schoenbaum *et al.*, 2002;

Nicolle & Baxter, 2003; Mizoguchi *et al.*, 2010). Interestingly, lesions of area 9 in marmoset monkeys affected extradimentional set-shifting performance, whereas lesions of the orbital PFC affected reversal performance (Dias *et al.*, 1996). These data suggest that these tasks rely on different brain structures within the PFC. Extradimentional set-shifting (EDS) refers to the problem of switching attention between cues that are in different perceptual dimensions in order to perform the task correctly. An example of this is to train a rat to use light cues to determine which arm to select in a maze, and then shift the relevant cue to an auditory stimulus. When the EDS occurs, the rat must shift its strategy and follow the sound cue in order to select the correct baited arm (e.g., Insel *et al.*, 2012). In contrast, reversal learning refers to adapting a behavior to the changing contingencies required to reach a goal. For example, a rat can initially learn to press a lever in a 'light-on' condition to receive reward. After a reversal, the rat must adapt its behavior to press during the 'light-off' condition (e.g., Nomura *et al.*, 2004).

In parallel to the age-related cognitive deficits discussed above, aging is also associated with changes in attentional processes (Gazzaley & D'Esposito, 2007; Prakash *et al.*, 2009; Hedden *et al.*, 2012). This is accompanied by a greater susceptibility to distraction or interference during the delay period of a working memory task in humans (Bowles & Salthouse, 2003; Campbell *et al.*, 2012) and monkeys (Bartus & Dean, 1979; Prendergast *et al.*, 1998). Additionally, fMRI studies in older adults have reported that there is increased activity in brain regions mediating distraction (Milham *et al.*, 2002; Stevens *et al.*, 2008), and in cases where task-irrelevant stimuli are presented (Gazzaley *et al.*, 2005).

## Age-related anatomical changes in frontal cortical circuits

One of the most consistent finding in the literature on aging brain is a decline in the volume of the prefrontal cortex (PFC) of humans, monkeys and rodents. This decline is one of the earliest changes detected, and for almost fifty years, it was thought that the decrease in frontal lobe volume was the result of cell loss (Haug, 1986; Peters, 2002). The early reports of cell loss, however, turned out to be an error resulting from differential shrinkage of young and aged tissue during processing (Haug et al., 1981; Terry et al., 1987; Haug & Eggers, 1991). It is now believed that cell numbers in the frontal cortex are preserved through aging in humans (Haug et al., 1981, 1984; Freeman et al., 2008). Similar conclusions have been drawn for frontal areas in non-human primates (Peters et al., 1996; Peters et al., 1998a; Smith et al., 2004), with the exception of prefrontal area 8A, a region of the dorsolateral PFC, which was shown to have a significant decline of Nissl-stained neurons (Smith et al., 2004). In rodents the cell counting results are conflicting. One group reports decreases in neuron numbers in the dorsal PFC areas but preservation in the ventral PFC areas (Stranahan et al., 2012), and another found the opposite, with cell loss in the ventral PFC and preservation in dorsal PFC (Yates et al., 2008). Because the same rat strain was utilized, Stranahan and colleagues (2012) suggest that different delineation of brain structures during counting could explain the disparate findings. Nonetheless, the current view is that the cell numbers in the PFC are reasonably well preserved during aging, although there may be focal points of cell loss in non-human primates and rodents.

In line with the overall reduction in frontal lobe volume mentioned above, age-related decreases in gray matter volumes and cortical thickness have been reported in humans (Haug & Eggers, 1991; Raz et al., 1997, 2005; Good et al., 2001; Tisserand et al., 2002; Salat et al., 2009; Bergfield et al., 2010; Giorgio et al., 2010; Thambisetty et al., 2010; Burzynska et al., 2012; Kalpouzos et al., 2012), nonhuman primates (Alexander et al., 2008; Shamy et al., 2011; Figure 2B) and rats (Alexander et al., 2011). However, an earlier stereological study performed using Nissl stained slices of monkeys reported a general preservation of area 46 (O'Donnell et al., 1999), which is in contrast with the findings from

MRI studies presented above. These differences may be the result of the research method employed or maybe caused by inter-individual variability of age effects on this part of the brain. Nonetheless, the changes in volume of the dorsolateral PFC in nonhuman primates was also shown to correlate with accuracy on a recognition memory task (Shamy *et al.*, 2011). Specifically, aged monkeys with larger PFC volumes identified more correct nonmatch objects on the DNMS task than did monkeys with smaller PFC volumes (Shamy *et al.*, 2011; Figure 2D). This correlation held even when the analysis was restricted to PFC gray matter or white matter volumes separately.

Rather than cell loss, the gray matter volume decrease in the PFC is in part caused by agerelated changes in neuron morphology, particularly the loss of synapses and the regression of apical dendrites (reviewed in Peters et al., 1996; Markham & Juraska, 2002; Dickstein et al., 2007; Luebke et al., 2010; Pannese, 2011; Morrison & Baxter, 2012). Decreases in spine numbers, density and changes in spine morphology have been reported in humans (Jacobs et al., 1997), non-human primates (Page et al., 2002; Duan et al., 2003; Peters et al., 2008; Dumitriu et al., 2010) and rats (Bloss et al., 2011, 2013). This change in spines represents the most consistent age-related alteration of cellular morphology reported in the frontal cortical literature, and is illustrated in Figure 3. With respect to the dendritic arbor, significant regression only occurs at the level of the apical dendrites in the PFC of aged humans (de Brabander et al., 1998), monkeys (Cupp & Uemura, 1980; Duan et al., 2003; Kabaso et al., 2009) and male rodents (Grill & Riddle, 2002; Markham & Juraska, 2002). The regression of terminal dendrites and synaptic loss that occur during aging likely affects dendritic excitability and plasticity processes in the PFC, thus contributing to the age-related decline in learning and working memory. In support of this, there is a decline in spine numbers and reduced thin spine volumes in area 46 in monkeys. This reduction was shown to correlate with acquisition and performance on a DNMS task (Peters et al., 1998b; Dumitriu et al., 2010). Additionally, a recent study was able to show that there is a correlation between the age-related overactivation of PKC, the length of basal dendrites and working memory performance in aged rats (Brennan et al., 2009), suggesting that altered PKC activity may be at the basis of some of the anatomical and functional deficits found in aged animals.

Despite cortical volume and cellular changes reported in the frontal cortex of older adults, many fMRI studies report areas of overactivation, greater bilateralization or recruitment of additional structures in PFC areas of older adults during performance certain cognitive tasks (e.g., Spreng *et al.*, 2010; Morcom & Friston, 2012; Spaniol & Grady, 2012). This is a phenomenon thought to reflect compensatory mechanisms and in support this hypothesis, greater activation of frontal areas was shown to be associated with better performance (Grady *et al.*, 2005). Thus, it is plausible that plastic mechanisms in the PFC compensate for changes occurring in the PFC and other parts of the brain in older adults, thereby contributing to preservation of cognitive function. In support of this idea, under some circumstances accurate retrieval of autobiographical events in older adults also show a similar pattern (as outlined previously). That is, during retrieval, the hippocampi of older adults show bilateral activation, whereas young adults show hippocampal activation lateralized to the left hemisphere (Maguire & Frith 2003).

In contrast to gray matter volumes that decrease linearly with age, white matter volume change across the lifespan follows a parabolic shape, with the largest volumes in the midfifties and an accelerated decline after 65 years of age (Allen *et al.*, 2005; Gunning-Dixon *et al.*, 2009; Bennett *et al.*, 2010; Giorgio *et al.*, 2010; Malykhin *et al.*, 2011). Decreases in white matter volume have also been reported in aged nonhuman primates (Peters & Sethares, 2002; Luebke *et al.*, 2010; Shamy *et al.*, 2011). A number of studies using diffusion tensor imaging have also revealed that the integrity of white matter is altered

during aging in humans and nonhuman primates, particularly in the frontal lobe (Gunning-Dixon et al., 2009; Madden et al., 2009; Bennett et al., 2010; Giorgio et al., 2010; Luebke et al., 2010; Samanez-Larkin et al., 2012). In addition, aging is associated with an increased incidence of white matter hyperintensities (WMH) around the ventricles and in the deep white matter (Gunning-Dixon et al., 2009). Greater numbers of WMH and reduced white matter integrity were both found to correlate with poorer cognitive performance in older adults, particularly processing speed and attention (Gunning-Dixon & Raz, 2000; Madden et al., 2009; Penke et al., 2010; Hedden et al., 2012). Reductions in white matter integrity could affect the connectivity between distributed brain networks, and contribute to some of the age-related changes observed in cognition (see Madden et al., 2009). In support of this, a correlation between white matter integrity in the genu of the corpus callosum, intrinsic functional connectivity, and choice reaction time was reported for older but not younger adults (Chen et al., 2009).

## Age-related physiological changes in frontal cortical circuits

## Age-related changes in gamma oscillations in humans and rodents

Older adults are more prone to have deficits in attentional control than are younger adults (Prakash *et al.*, 2009; Hedden *et al.*, 2012). They show a selective impairment in visual attention tasks in which the goal is to determine whether a target object is present among distractor objects that share features with it - a task condition called conjunctive search (Plude & Doussard-Roosevelt, 1989). Solving such a task requires subjects to intentionally focus their attention toward the various objects - a form of attention referred to as top-down (Talsma *et al.*, 2010; Awh *et al.*, 2012). A recent aging study found that under conjunctive search conditions, there were differences between age groups in the power of gamma in the PFC-posterior parietal network. Older adults fail to show an increase in low-gamma power (22-34 Hz) in the easier task condition (Phillips & Takeda, 2010), while younger adults show increases in low-gamma power at all difficulty levels of this task (Phillips & Takeda, 2009). This result adds further support to the inferences made in the imaging literature (e.g., Madden *et al.*, 2007; Gazzaley, 2011) that altered PFC-posterior parietal network activation in older adults may be responsible for a less efficient top-down attentional control of visual search.

Gamma rhythms have also been reported to be altered in aged rats. In aged rodents, behavioral slowing during decisions made in an extra-dimensional set-shifting task was found to correlate with slower gamma oscillations (30-100 Hz) in the anterior dorsal cingulate cortex, an area within the medial PFC (Insel et al., 2012). Specifically, the mean peak frequency of gamma oscillations, while performing this set-shifting task, was 56.4 Hz in young rats and 53.5 Hz in aged rats (Insel et al., 2012), and this difference was statistically reliable. Because gamma frequencies are thought to be mediated by network interactions between glutamatergic and GABAergic cells (Tiesinga et al., 2001; Börgers et al., 2005; Wang, 2010), the changes in gamma frequency suggest that the interaction between these cell types may be compromised in aged animals. In support of this, Insel and colleagues found that, during the performance of the task, putative excitatory and inhibitory neurons of the medial PFC fired preferentially at different phases of the gamma cycle in young and aged rats. When cross correlation analysis was applied to simultaneously recorded excitatory-inhibitory cell pairs, the interval between the excitatory drive onto inhibitory cells was lengthened in the older rats (Insel et al., 2012). While arguments for direct causation cannot be made, these studies suggest that GABAergic transmission is altered in the PFC of aged rodents and that this may contribute to altered gamma synchrony among medial PFC networks.

#### Decreased persistent firing neurons during delayed response tasks, role for cAMP

Converging evidence links age-related working memory impairments to dysfunction of adrenergic systems in primates. Indeed, age-related disinhibition of cyclic adenosine monophosphate (cAMP) signaling was shown to lead to decreases in persistent firing of area 46 neurons that are active through a delay period during working memory tasks (Ramos et al., 2003; Arnsten et al., 2010; Wang et al., 2011). These delay-firing neurons show a sustained activation that lasts for the duration of the cue delay period of a delayed response task (Goldman-Rakic, 1995). This increased activation is modulated by spatial location on a screen, and is greatest for the neurons' preferred direction. In aged monkeys, there is an agerelated loss in response modulation of these neurons to their preferred spatial location during working memory tasks, to a point where delay neurons show very little increase in firing rate during the cue delay period (Wang et al., 2011). The decrease in activity of delay neurons in aged monkeys could be rescued using local drug administration that either inhibited cAMP or the downstream potassium channels that cAMP is known to activate (HCN, KCNQ) (Wang et al., 2011). The same results could be obtained using local infusion of guanfacine, an α<sub>2A</sub> adrenergic agonist that inhibit cAMP signaling (Wang et al., 2011). Guanfacine and clonidine are both  $\alpha_{2A}$  adrenergic agonist known to enhance working memory performance in aged rats (Arnsten et al., 1988; Arnsten & Goldman-Rakic, 1990; Ramos et al., 2003). Because  $\alpha_{2A}$  adrenergic agonists have no effects on a visual pattern discrimination task, (Arnsten & Goldman-Rakic, 1985), the effects of guanfacine on working memory performance is likely through its action on the activity of PFC neurons. These findings support the notion that age-related working memory deficits are mediated, at least in part, by physiological changes occurring in the dorsolateral PFC.

As with nonhuman primates, the activity of the PFC during the delay period of working memory tasks is altered in older adults. Indeed, a fMRI study revealed age differences in the pattern of activation of the lateral PFC that were dependent on the trial phase, with lower activation during task delays, and greater activation at the time of the probe in older adults (Paxton *et al.*, 2008). These results suggest that aging may also affect delay neurons, not only in monkeys, but perhaps in human as well.

### Decreased sensitivity of orbitofrontal cortex (OFC) neurons to delays

The activity of OFC neurons has been characterized in young and aged rats, while performing two different tasks, a delay discounting task and a reversal task (Schoenbaum et al., 2006; Roesch et al., 2012). In a delay discounting task, rats have the choice between a small immediate reward and a large reward delivered after a delay. In this task, aged rats were found to prefer the large reward, regardless of the length of the delay, whereas young rats were more prone to switch their behavior towards the small immediate reward as the delay increased (Simon et al., 2010). Using a delay discounting task, Roesch and colleagues (2012) addressed whether there are age-related differences in the activity of OFC neurons in response to varying the length of delays. They found a higher prevalence of neurons responsive to long delay rewards in aged rats. While about 50% of reward-responsive neurons were active during short delays in aged rats, about 75% of the neurons fired preferentially to short delays in young rats (Roesch et al., 2012). There was no age difference in the proportions of cells responding to large over small rewards (Roesch et al., 2012). Thus, aging appears to selectively affect OFC delay neurons. It is possible that agerelated changes in plastic processes in OFC biased the older neurons from adapting their activity in a manner similar to that of the younger animals. This lack of adaptation of OFC cells may be responsible for the lack of willingness of older animals to change their behavior towards receiving a large reward, in spite of the long delay associated with doing so.

Aged rats are known for their behavioral impairments on reversal tasks (Schoenbaum et al., 2002; Mizoguchi et al., 2010). Whereas older rats are able to acquire discrimination problems at high levels of performance, some are impaired when contingencies are reversed. Because the OFC is critical for reversal performance, Schoenbaum and colleagues (2006) recorded neurons from this brain region in young and aged rats while they performed a go, no-go task with reversals. In this task, rats learned to associate pairs of odors predicting either reward or punishment. Following presentation of a "go" odor, rats learned to go to the food port to receive a reward. Following a "no-go" odor, rats learned to avoid going to the food port where an aversive quinine solution was delivered. They found that age-impaired rats had fewer neurons that were cue selective, and most of these cells lost their cueselective firing after reversal (Schoenbaum et al., 2006). In contrast, young and agedunimpaired rats had a larger number of cells that were more sensitive to one of the odor cues, and a significant proportion of these cells reversed their activity in response to the new odor after reversal (Schoenbaum et al., 2006). These results suggest that a loss in flexible responding of OFC neurons to changing contingencies might underlie the behavioral deficits found in some aged rats during reversal performance.

### Increased excitability of cells in area 46

The electrical properties of pyramidal cells of area 46 of young and aged monkeys have been examined using *in vitro* preparations. The general findings suggest an increased excitability of pyramidal cells located in layer 2/3, but not in layer 5 (Luebke *et al.*, 2004; Chang *et al.*, 2005; Luebke & Chang, 2007; Dickstein *et al.*, 2012; Luebke & Amatrudo, 2012). Specifically, the authors report an age-related decrease in spontaneous excitatory post-synaptic currents and increases in spontaneous inhibitory post-synaptic currents (Luebke *et al.*, 2004). Additionally, the authors report an increased input resistance and firing frequency of layer 3 pyramidal neurons (Chang *et al.*, 2005). Layer 3 mainly contains pyramidal neurons that project to other cortical areas (Page *et al.*, 2002; Yeterian *et al.*, 2012), thus increased excitability suggests increased output from these cells. Because aged monkeys with the highest and lowest firing rates displayed the poorest performance levels in working memory tasks, a balance in the activity of area 46 might be necessary for optimal performance (Chang *et al.*, 2005). The exact impact that this age-related increase in excitability has on wider PFC networks in non-human primates remains to be explored.

# Summary

Overall, the patterns of age-related change in brain function and cognitive domains are remarkably conserved across mammals, as has been reviewed here. The depth of analytic approaches that can be used in animals other than humans has made it possible to understand the neurobiological processes that are vulnerable across the lifespan in greater detail. Equally striking in this comparison of temporal and frontal lobe systems is the apparent selectivity and differential vulnerability of these brain structures to the changes that do occur with age. While the reasons for these differences are the target of active investigation, there is no clear explanation for why frontal lobe systems appear to 'age at a different rate' (faster, earlier signs of change) than do temporal lobe systems. Clearly the brain region specificity of neural changes with aging needs to be taken into account in the development of strategies targeted at optimization of cognitive function across the lifespan.

Another important point to emphasize is that while it has been suggested that cognitive decline is not apparent until after 60 years of age (e.g., Hedden & Gabrieli, 2004), there are also data suggesting that changes in cognition can occur much earlier (e.g., Finch, 2009; Salthouse, 2009). In fact, one longitudinal study has reported that the decline in some domains can be detected across large populations of those in their forties (Singh-Manoux *et al.*, 2011). This suggests that it will be important to develop interventions that optimize

neural circuit function, in regions such as the hippocampus and prefrontal cortex, beginning at least in mid-age and probably earlier. As discussed here, progress in understanding the biology of lifespan development and how neural change drives cognitive change has led to a number of key insights that can now be directed towards the development of tools that can help maintain cognitive health across the lifespan.

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### References

- Alexander GE, Chen K, Aschenbrenner M, Merkley TL, Santerre-Lemmon LE, Shamy JL, Skaggs WE, Buonocore MH, Rapp PR, Barnes CA. Age-related regional network of MRI gray matter in the rhesus macaque. J. Neurosci. 2008; 28:2710–2718. [PubMed: 18337400]
- Alexander, GE.; Lin, L.; Yoshimaru, E.; Hoang, LT.; Bergfield, BL.; Lister, JP.; Chen, K.; Moeller, JR.; Barnes, CA.; Trouard, TP. Regional network pattern of MRI gray matter in the aged rat. Society for Neuroscience, Online; Washington, DC: 2011. Program No. 939.01. Abstract Viewer/ Itinerary Planner
- Alexander GE, Ryan L, Bowers D, Foster TC, Bizon JL, Geldmacher DS, Glisky EL. Characterizing cognitive aging in humans with links to animal models. Front Aging Neurosci. 2012; 4:21. [PubMed: 22988439]
- Allen JS, Bruss J, Brown CK, Damasio H. Normal neuroanatomical variation due to age: The major lobes and a parcellation of the temporal region. Neurobiology of Aging. 2005; 26:1245–1260. [PubMed: 16046030]
- Arnsten AF, Goldman-Rakic PS. Alpha 2-adrenergic mechanisms in prefrontal cortex associated with cognitive decline in aged nonhuman primates. Science. 1985; 230:1273–1276. [PubMed: 2999977]
- Arnsten AF, Cai JX, Goldman-Rakic PS. The alpha-2 adrenergic agonist guanfacine improves memory in aged monkeys without sedative or hypotensive side effects: evidence for alpha-2 receptor subtypes. J. Neurosci. 1988; 8:4287–4298. [PubMed: 2903226]
- Arnsten AF, Goldman-Rakic PS. Analysis of alpha-2 adrenergic agonist effects on the delayed nonmatch-to-sample performance of aged rhesus monkeys. Neurobiol. Aging. 1990; 11:583–590. [PubMed: 1980719]
- Arnsten AFT, Paspalas CD, Gamo NJ, Yang Y, Wang M. Dynamic network connectivity: A new form of neuroplasticity. Trends Cogn. Sci. 2010; 14:365–375. [PubMed: 20554470]
- Awh E, Belopolsky AV, Theeuwes J. Top-down versus bottom-up attentional control: a failed theoretical dichotomy. Trends Cogn. Sci. 2012; 16:437–443. [PubMed: 22795563]
- Bach ME, Barad M, Son H, Zhuo M, Lu YF, Shih R, Mansuy I, Hawkins RD, Kandel ER. Age-related defects in spatial memory are correlated with defects in the late phase of hippocampal long-term potentiation in vitro and are attenuated by drugs that enhance the cAMP signaling pathway. Proc. Natl. Acad. Sci. USA. 1999; 96:5280–5285. [PubMed: 10220457]
- Bachevalier J, Landis LS, Walker LC, Brickson M, Mishkin M, Price DL, Cork LC. Aged monkeys exhibit behavioral deficits indicative of widespread cerebral dysfunction. Neurobiol. Aging. 1991; 12:99–111. [PubMed: 2052134]
- Ballard C, Gauthier S, Corbett A, Brayne C, Aarsland D, Jones E. Alzheimer's disease. Lancet. 2011; 377:1019–1031. [PubMed: 21371747]
- Barense MD, Fox MT, Baxter MG. Aged rats are impaired on an attentional set-shifting task sensitive to medial frontal cortex damage in young rats. Learn. Mem. 2002; 9:191–201. [PubMed: 12177232]
- Barnes CA. Memory deficits associated with senescence: A neurophysiological and behavioral study in the rat. J. Comp. Physiol. Psychol. 1979; 93:74–104. [PubMed: 221551]

Barnes CA, McNaughton BL. Physiological compensation for loss of afferent synapses in rat hippocampal granule cells during senescence. J. Physiol. (Lond). 1980; 309:473–485. [PubMed: 7252877]

- Barnes CA, Nadel L, Honig WK. Spatial memory deficit in senescent rats. Can. J. Psychol. 1980; 34:29–39. [PubMed: 7388694]
- Barnes CA, Rao G, McNaughton BL. Increased electrotonic coupling in aged rat hippocampus: A possible mechanism for cellular excitability changes. J. Comp. Neurol. 1987; 259:547–558.
- Barnes CA, Suster MS, Shen J, McNaughton BL. Multistability of cognitive maps in the hippocampus of old rats. Nature. 1997; 388:272–275. [PubMed: 9230435]
- Barnes CA, Rao G, Houston FP. LTP induction threshold change in old rats at the perforant path granule cell synapse. Neurobiol. Aging. 2000; 21:613–620. [PubMed: 11016529]
- Bartus RT, Fleming D, Johnson HR. Aging in the rhesus monkey: Debilitating effects on short-term memory. J Gerontol. 1978; 33:858–871. [PubMed: 106081]
- Bartus RT, Dean RL. Recent memory in aged non-human primates: Hypersensitivity to visual interference during retention. Exp. Aging Res. 1979; 5:385–400. [PubMed: 118012]
- Bartus RT, Dean RL, Fleming DL. Aging in the rhesus monkey: Effects on visual discrimination learning and reversal learning. J Gerontol. 1979; 34:209–219. [PubMed: 108323]
- Bennett IJ, Madden DJ, Vaidya CJ, Howard DV, Howard JH Jr. Age-related differences in multiple measures of white matter integrity: A diffusion tensor imaging study of healthy aging. Hum. Brain Mapp. 2010; 31:378–390. [PubMed: 19662658]
- Bergfield KL, Hanson KD, Chen K, Teipel SJ, Hampel H, Rapoport SI, Moeller JR, Alexander GE. Age-related networks of regional covariance in MRI gray matter: reproducible multivariate patterns in healthy aging. Neuroimage. 2010; 49:1750–1759. [PubMed: 19796692]
- Bizon JL, LaSarge CL, Montgomery KS, McDermott AN, Setlow B, Griffith WH. Spatial reference and working memory across the lifespan of male Fischer 344 rats. Neurobiol. Aging. 2009; 30:646–655. [PubMed: 17889407]
- Bizon JL, Foster TC, Alexander GE, Glisky EL. Characterizing cognitive aging of working memory and executive function in animal models. Front. Ag. Neurosci. 2012; 4:19.
- Bliss, T.; Collingridge, G.; Morris, R. Synaptic plasticity in the hippocampus. In: Andersen, P.; Morris, R.; Amaral, D.; Bliss, T.; O'Keefe, J., editors. The Hippocampus Book. Oxford University Press; New York: 2007. p. 343-474.
- Bloss EB, Janssen WG, Ohm DT, Yuk FJ, Wadsworth S, Saardi KM, McEwen BS, Morrison JH. Evidence for reduced experience-dependent dendritic spine plasticity in the aging prefrontal cortex. J. Neurosci. 2011; 31:7831–7839. [PubMed: 21613496]
- Bloss EB, Puri R, Yuk F, Punsoni M, Hara Y, Janssen WG, McEwen BS, Morrison JH. Morphological and molecular changes in aging rat prelimbic prefrontal cortical synapses. Neurobiol. Aging. 2013; 34:200–210. [PubMed: 22727942]
- Bodhinathan K, Kumar A, Foster TC. Redox sensitive calcium stores underlie enhanced after hyperpolarization of aged neurons: Role for ryanodine receptor mediated calcium signaling. J. Neurophysiol. 2010; 104:2586–2593. [PubMed: 20884759]
- Börgers C, Epstein S, Kopell NJ. Background gamma rhythmicity and attention in cortical local circuits: A computational study. Proc. Natl. Acad. Sci. USA. 2005; 102:7002–7007. [PubMed: 15870189]
- Bowles RP, Salthouse TA. Assessing the age-related effects of proactive interference on working memory tasks using the Rasch model. Psychol Aging. 2003; 18:608–615. [PubMed: 14518820]
- Brennan AR, Yuan P, Dickstein DL, Rocher AB, Hof PR, Manji H, Arnsten AFT. Protein kinase C activity is associated with prefrontal cortical decline in aging. Neurobiol. Aging. 2009; 30:782–792. [PubMed: 17919783]
- Burke SN, Chawla MK, Penner MR, Crowell BE, Worley PF, Barnes CA, McNaughton BL. Differential encoding of behavior and spatial context in deep and superficial and layers of the neocortex. Neuron. 2005; 45:667–674. [PubMed: 15748843]
- Burke SN, Barnes CA. Neural plasticity in the ageing brain. Nat. Rev. Neurosci. 2006; 7:30–40. [PubMed: 16371948]

Burke SN, Barnes CA. Senescent synapses and hippocampal circuit dynamics. Trends Neurosci. 2010; 33:153–161. [PubMed: 20071039]

- Burns PC. Navigation and the mobility of older drivers. J. Gerontol. B Psychol. Sci. Soc. Sci. 1999; 54B:S49–S55. [PubMed: 9934402]
- Burzynska AZ, Nagel IE, Preuschhof C, Gluth S, Bäckman L, Li S-C, Lindenberger U, Heekeren HR. Cortical thickness is linked to executive functioning in adulthood and aging. Hum. Brain Mapp. 2012; 33:1607–1620. [PubMed: 21739526]
- Campbell KL, Grady CL, Ng C, Hasher L. Age differences in the frontoparietal cognitive control network: implications for distractibility. Neuropsychologia. 2012; 50:2212–2223. [PubMed: 22659108]
- Cappell KA, Gmeindl L, Reuter-Lorenz PA. Age differences in prefontal recruitment during verbal working memory maintenance depend on memory load. Cortex. 2010; 46:462–473. [PubMed: 20097332]
- Castellano JF, Fletcher BR, Kelley-Bell B, Kim DH, Gallagher M, Rapp PR. Age-related memory impairment is associated with disrupted multivariate epigentic coordination in the hippocamps. PLoS one. 2012; 7:e33249. [PubMed: 22438904]
- Chang Y-M, Rosene DL, Killiany RJ, Mangiamele LA, Luebke JI. Increased action potential firing rates of layer 2/3 pyramidal cells in the prefrontal cortex are significantly related to cognitive performance in aged monkeys. Cereb. Cortex. 2005; 15:409–418. [PubMed: 15749985]
- Chen N, Chou Y, Song AW, Madden DJ. Measurement of spontaneous signal fluctuations in fMRI: adult age differences in intrinsic functional connectivity. Brain Struct. Funct. 2009; 213:571–585. [PubMed: 19727810]
- Cupp CJ, Uemura E. Age-related changes in prefrontal cortex of Macaca mulatta: Quantitative analysis of dendritic branching patterns. Exp. Neurol. 1980; 69:143–163. [PubMed: 6771151]
- de Brabander JM, Kramers RJ, Uylings HB. Layer-specific dendritic regression of pyramidal cells with ageing in the human prefrontal cortex. Eur. J. Neurosci. 1998; 10:1261–1269. [PubMed: 9749780]
- Deupree DL, Bradley J, Turner DA. Age-related alterations in potentiation in the CA1 region in F344 rats. Neurobiol. Aging. 1993; 14:249–258. [PubMed: 8321393]
- Dias R, Robbins TW, Roberts AC. Dissociation in prefrontal cortex of affective and attentional shifts. Nature. 1996; 380:69–72. [PubMed: 8598908]
- Dickstein DL, Kabaso D, Rocher AB, Luebke JI, Wearne SL, Hof PR. Changes in the structural complexity of the aged brain. Aging Cell. 2007; 6:275–284. [PubMed: 17465981]
- Dickstein DL, Weaver CM, Luebke JI, Hof PR. Dendritic spine changes associated with normal aging. Neuroscience. 2012 in press.
- Disterhoft JF, Thompson LT, Moyer JR Jr. Mogul DJ. Calcium-dependent afterhyperpolarization and learning in young and aging hippocampus. Life Sci. 1996; 59:413–420. [PubMed: 8761329]
- Driscoll I, Hamilton DA, Yeo RA, Brooks WM, Sutherland RJ. Virtual navigation in humans: The impact of age, sex and hormones on place learning. Horm. Behav. 2005; 47:326–335. [PubMed: 15708762]
- Duan H, Wearne SL, Rocher AB, Macedo A, Morrison JH, Hof PR. Age-related dendritic and spine changes in corticocortically projecting neurons in macaque monkeys. Cereb. Cortex. 2003; 13:950–961. [PubMed: 12902394]
- Dumitriu D, Hao J, Hara Y, Kaufmann J, Janssen WGM, Lou W, Rapp PR, Morrison JH. Selective changes in thin spine density and morphology in monkey prefrontal cortex correlate with aging-related cognitive impairment. J. Neurosci. 2010; 30:7507–7515. [PubMed: 20519525]
- Ekstrom AD, Meltzer J, McNaughton BL, Barnes CA. NMDA receptor antagonism blocks experience-dependent expansion of hippocampal "place fields". Neuron. 2001; 31:631–638. [PubMed: 11545721]
- Finch CE. The neurobiology of middle-age has arrived. Neurobiol. Aging. 2009; 30:515–520. [PubMed: 19231030]
- Flood DG, Buell SJ, Horwitz GJ, Coleman PD. Dendritic extent in human dentate gyrus granule cells in normal aging and senile dementia. Brain Res. 1987; 402:205–216. [PubMed: 3828793]

Flood DG. Critical issues in the analysis of dendritic extent in aging humans, primates, and rodents. Neurobiol. Aging. 1993; 14:649–654. [PubMed: 8295674]

- Foster TC, Barnes CA, Rao G, McNaughton BL. Increase in perforant path quantal size in aged F-344 rats. Neurobiol. Aging. 1991; 12:441–448. [PubMed: 1770978]
- Foster TC. Biological markers of age-related memory deficits: Treatment of senescent physiology. CNS Drug. 2006; 20:153–166.
- Foster TC, DeFazio RA, Bizon JL. Characterizing cognitive aging of spatial and contextual memory in animal models. Front. Aging Neurosci. 2012; 4:12. [PubMed: 22988436]
- Freeman SH, Kandel R, Cruz L, Rozkalne A, Newell K, Frosch MP, Hedley-Whyte ET, Locascio JJ, Lipsitz L, Hyman BT. Preservation of neuronal number despite age-related cortical brain atrophy in elderly subjects without Alzheimer disease. J. Neuropathol. Exp. Neurol. 2008; 67:1205–1212. [PubMed: 19018241]
- Funahashi S, Bruce CJ, Goldman-Rakic PS. Dorsolateral prefrontal lesions and oculomotor delayed-response performance: evidence for mnemonic "scotomas". J. Neurosci. 1993; 13:1479–1497. [PubMed: 8463830]
- Gage FH, Dunnet SB, Björklund A. Spatial learning and motor deficits in aged rats. Neurobiol. Aging. 1984; 5:43–48. [PubMed: 6738785]
- Gazzaley AH, Thakker MM, Hof PR, Morrison JH. Preserved number of entorhinal cortex layer II neurons in aged macaque monkeys. Neurobiol. Aging. 1997; 18:549–553. [PubMed: 9390783]
- Gazzaley A, Cooney JW, Rissman J, D'Esposito M. Top-down suppression deficit underlies working memory impairment in normal aging. Nat. Neurosci. 2005; 8:1298–1300. [PubMed: 16158065]
- Gazzaley A, D'Esposito M. Top-down modulation and normal aging. Ann. N. Y. Acad. Sci. 2007; 1097:67–83. [PubMed: 17413013]
- Gazzaley A. Influence of early attentional modulation on working memory. Neuropsychologia. 2011; 49:1410–1424. [PubMed: 21184764]
- Geinisman Y, de Toledo-Morrell L, Morrell F, Persina IS, Rossi M. Age-related loss of axospinous synapses formed by two afferent systems in the rat dentate gyrus as revealed by the unbiased stereological disector technique. Hippocampus. 1992; 2:437–444. [PubMed: 1308200]
- Geinisman Y, Ganeshina O, Yoshida R, Berry RW, Disterhoft JF, Gallagher M. Aging, spatial learning, and total synapse number in the rat CA1 stratum radiatum. Neurobiol. Aging. 2004; 25:407–415. [PubMed: 15123345]
- Giorgio A, Santelli L, Tomassini V, Bosnell R, Smith S, De Stefano N, Johansen-Berg H. Age-related changes in grey and white matter structure throughout adulthood. Neuroimage. 2010; 51:943–951. [PubMed: 20211265]
- Goldman PS, Rosvold HE. Localization of function within the dorsolateral prefrontal cortex of the rhesus monkey. Exp. Neurol. 1970; 27:291–304. [PubMed: 4987453]
- Goldman-Rakic PS. Cellular basis of working memory. Neuron. 1995; 14:477–485. [PubMed: 7695894]
- Good CD, Johnsrude IS, Ashburner J, Henson RN, Friston KJ, Frackowiak RS. A voxel-based morphometric study of ageing in 465 normal adult human brains. Neuroimage. 2001; 14:21–36. [PubMed: 11525331]
- Grady CL, McIntosh AR, Craik FI. Task-related activity in prefrontal cortex and its relation to recognition memory performance in young and old adults. Neuropsychologia. 2005; 43:1466– 1481. [PubMed: 15989937]
- Grill JD, Riddle DR. Age-related and laminar-specific dendritic changes in the medial frontal cortex of the rat. Brain Res. 2002; 937:8–21. [PubMed: 12020857]
- Grottick AJ, Higgins GA. Assessing a vigilance decrement in aged rats: Effects of pre-feeding, task manipulation, and psychostimulants. Psychopharmacology (Berl.). 2002; 164:33–41. [PubMed: 12373417]
- Gunning-Dixon FM, Raz N. The cognitive correlates of white matter abnormalities in normal aging: A quantitative review. Neuropsychology. 2000; 14:224–232. [PubMed: 10791862]
- Gunning-Dixon FM, Brickman AM, Cheng JC, Alexopoulos GS. Aging of cerebral white matter: A review of MRI findings. Int J Geriatr Psychiatry. 2009; 24:109–117. [PubMed: 18637641]

Guzowski JF, McNaughton BL, Barnes CA, Worley PF. Environment-specific expression of the immediate-early gene *Arc* in hippocampal neuronal ensembles. Nat. Neurosci. 1999; 2:1120–1124. [PubMed: 10570490]

- Guzowski JF, Lyford GL, Stevenson GD, Houston FP, McGaugh JL, Worley PF, Barnes CA. Inhibition of activity-dependent *Arc* protein expression in the rat hippocampus impairs the maintenance of long-term potentiation and the consolidation of long-term memory. J. Neurosci. 2000; 20:3993–4001. [PubMed: 10818134]
- Hartman M, Bolton E, Fehnel SE. Accounting for age differences on the Wisconsin Card Sorting Test: Decreased working memory, not inflexibility. Psychol Aging. 2001; 16:385–399. [PubMed: 11554518]
- Haug H, Knebel G, Mecke E, Orün C, Sass NL. The aging of cortical cytoarchitectonics in the light of stereological investigations. Prog. Clin. Biol. Res. 1981; 59B:193–197. [PubMed: 7279939]
- Haug H, Kühl S, Mecke E, Sass NL, Wasner K. The significance of morphometric procedures in the investigation of age changes in cytoarchitectonic structures of human brain. J. Hirnforsch. 1984; 25:353–374. [PubMed: 6481152]
- Haug H. History of neuromorphometry. J. Neurosci. Methods. 1986; 18:1–17. [PubMed: 3540464]
- Haug H, Eggers R. Morphometry of the human cortex cerebri and corpus striatum during aging. Neurobiol. Aging. 1991; 12:336–338. discussion 352–355. [PubMed: 1961364]
- Head D, Isom M. Age effects on wayfinding and route learning skills. Behav. Brain Res. 2010; 209:49–58. [PubMed: 20085784]
- Hebb, DO. The Organization of Behavior. John Wiley & Sons; New York: 1949.
- Hedden T, Gabrieli JDE. Insights into the ageing mind: A view from cognitive neuroscience. Nat. Rev. Neurosci. 2004; 5:87–96. [PubMed: 14735112]
- Hedden T, Van Dijk KRA, Shire EH, Sperling RA, Johnson KA, Buckner RL. Failure to modulate attentional control in advanced aging linked to white matter pathology. Cereb. Cortex. 2012; 22:1038–1051. [PubMed: 21765181]
- Hoang, LT.; Lister, JP.; Barnes, CA. The ageing hippocampus. In: Bartsch, T., editor. Clinical Neurobiology of the Hippocampus. Oxford University Press; New York: 2012. p. 152-173.
- Huang YY, Kandel ER. Age-related enhancement of a protein synthesis-dependent late phase of LTP induced by low frequency paired-pulse stimulation in hippocampus. Learn. Mem. 2006; 13:298–306. [PubMed: 16741282]
- Iaria G, Petrides M, Dagher A, Pike B, Bohbot VD. Cognitive strategies dependent on the hippocampus and caudate nucleus in human navigation: Variability and change with practice. J. Neurosci. 2003; 23:5945–5952. [PubMed: 12843299]
- Iaria G, Palermo L, Committeri G, Barton JJS. Age differences in the formation and use of cognitive maps. Behav. Brain Res. 2009; 196:187–191. [PubMed: 18817815]
- Insel N, Patron LA, Hoang LT, Nematollahi S, Schimanski LA, Lipa P, Barnes CA. Reduced gamma frequency in the medial frontal cortex of aged rats during behavior and rest: Implications for agerelated behavioral slowing. J. Neurosci. 2012; 32:16331–16344. [PubMed: 23152616]
- Jacobs B, Driscoll L, Schall M. Life-span dendritic and spine changes in areas 10 and 18 of human cortex: a quantitative Golgi study. J. Comp. Neurol. 1997; 386:661–680. [PubMed: 9378859]
- Jansen P, Schmelter A, Heil M. Spatial knowledge acquisition in younger and elder adults: A study in a virtual environment. Exp. Psychol. 2010; 57:54–60. [PubMed: 20178963]
- Kabaso D, Coskren PJ, Henry BI, Hof PR, Wearne SL. The electrotonic structure of pyramidal neurons contributing to prefrontal cortical circuits in macaque monkeys is significantly altered in aging. Cereb. Cortex. 2009; 19:2248–2268. [PubMed: 19150923]
- Kalpouzos G, Persson J, Nyberg L. Local brain atrophy accounts for functional activity differences in normal aging. Neurobiol. Aging. 2012; 33:623.e1–623.e13. [PubMed: 21524432]
- Keuker JI, Luiten PG, Fuchs E. Preservation of hippocampal neuron numbers in aged rhesus monkeys. Neurobiol. Aging. 2003; 24:157–165. [PubMed: 12493561]
- Kumar A, Bodhinathan K, Foster TC. Susceptibility of calcium dysregulation during brain aging. Front. Aging Neurosci. 2009; 1:2. [PubMed: 20552053]

Lai ZC, Moss MB, Killiany RJ, Rosene DL, Herndon JG. Executive system dysfunction in the aged monkey: spatial and object reversal learning. Neurobiol. Aging. 1995; 16:947–954. [PubMed: 8622786]

- Landfield PW, Pitler TA. Prolonged Ca<sup>2+</sup>-dependent after hyperpolarizations in hippocampal neurons of aged rats. Science. 1984; 226:1089–1092. [PubMed: 6494926]
- Luebke J, Barbas H, Peters A. Effects of normal aging on prefrontal area 46 in the rhesus monkey. Brain Res. Rev. 2010; 62:212–232. [PubMed: 20005254]
- Luebke JI, Chang Y-M, Moore TL, Rosene DL. Normal aging results in decreased synaptic excitation and increased synaptic inhibition of layer 2/3 pyramidal cells in the monkey prefrontal cortex. Neuroscience. 2004; 125:277–288. [PubMed: 15051166]
- Luebke JI, Chang Y-M. Effects of aging on the electrophysiological properties of layer 5 pyramidal cells in the monkey prefrontal cortex. Neuroscience. 2007; 150:556–562. [PubMed: 17981400]
- Luebke JI, Amatrudo JM. Age-related increase of sI(AHP) in prefrontal pyramidal cells of monkeys: Relationship to cognition. Neurobiol. Aging. 2012; 33:1085–1095. [PubMed: 20727620]
- Lyford GL, Yamagata K, Kaufmann WE, Barnes CA, Sanders LK, Copeland NG, Gilbert DJ, Jenkins NA, Lanahan AA, Worley PF. Arc, a growth factor and activity-regulated gene, encodes a novel cytoskeleton-associated protein that is enriched in neuronal dendrites. Neuron. 1995; 14:433–445. [PubMed: 7857651]
- Lyons-Warren A, Lillie R, Hershey T. Short- and long-term spatial delayed response performance across the lifespan. Dev Neuropsychol. 2004; 26:661–678. [PubMed: 15525563]
- Madden DJ, Spaniol J, Whiting WL, Bucur B, Provenzale JM, Cabeza R, White LE, Huettel SA. Adult age differences in the functional neuroanatomy of visual attention: A combined fMRI and DTI study. Neurobiol. Aging. 2007; 28:459–476. [PubMed: 16500004]
- Madden DJ, Bennett IJ, Song AW. Cerebral white matter integrity and cognitive aging: contributions from diffusion tensor imaging. Neuropsychol Rev. 2009; 19:415–435. [PubMed: 19705281]
- Maguire EA, Frith CD. Aging effects the engagement of the hippocampus during autobiographical memory retrieval. Brain. 2003; 126:1511–1523. [PubMed: 12805116]
- Malykhin N, Vahidy S, Michielse S, Coupland N, Camicioli R, Seres P, Carter R. Structural organization of the prefrontal white matter pathways in the adult and aging brain measured by diffusion tensor imaging. Brain Struct. Funct. 2011; 216:417–431. [PubMed: 21559982]
- Markham JA, Juraska JM. Aging and sex influence the anatomy of the rat anterior cingulate cortex. Neurobiol. Aging. 2002; 23:579–588. [PubMed: 12009507]
- Markus EJ, Barnes CA, McNaughton B.L, Gladden, V.L. Skaggs WE. Spatial information content and reliability of hippocampal CA1 neurons: Effects of visual input. Hippocampus. 1994; 4:410–421. [PubMed: 7874233]
- Marschner A, Mell T, Wartenburger I, Villringer A, Reischies FM, Heekeren HR. Reward-based decision-making and aging. Brain Res. Bull. 2005; 67:382–390. [PubMed: 16216684]
- Mehta MR, Barnes CA, McNaughton BL. Experience-dependent, asymmetric expansion of hippocampal place fields. Proc. Natl. Acad. Sci. USA. 1997; 94:8918–8921. [PubMed: 9238078]
- Merrill DA, Chiba AA, Tuszynski MH. Conservation of neuronal number and size in the entorhinal cortex of behaviorally characterized aged rats. J. Comp. Neurol. 2001; 438:445–456. [PubMed: 11559900]
- Milham MP, Erickson KI, Banich MT, Kramer AF, Webb A, Wszalek T, Cohen NJ. Attentional control in the aging brain: Insights from an fMRI study of the stroop task. Brain Cogn. 2002; 49:277–296. [PubMed: 12139955]
- Mizoguchi K, Shoji H, Tanaka Y, Tabira T. Orbitofrontal dopaminergic dysfunction causes age-related impairment of reversal learning in rats. Neuroscience. 2010; 170:1110–1119. [PubMed: 20736050]
- Moffat SD, Elkins W, Resnick SM. Age differences in the neural systems supporting human allocentric spatial navigation. Neurobiol. Aging. 2006; 27:965–972. [PubMed: 15982787]
- Moore CI, Browning MD, Rose GM. Hippocampal plasticity induced by primed burst, but not long-term potentiation, stimulation is impaired in area CA1 of aged Fischer 344 rats. Hippocampus. 1993; 3:57–66. [PubMed: 8364683]

Moore TL, Killiany RJ, Herndon JG, Rosene DL, Moss MB. Impairment in abstraction and set shifting in aged rhesus monkeys. Neurobiol. Aging. 2003; 24:125–134. [PubMed: 12493558]

- Morcom AM, Friston KJ. Decoding episodic memory in ageing: a Bayesian analysis of activity patterns predicting memory. Neuroimage. 2012; 59:1772–1782. [PubMed: 21907810]
- Moreno H, Wu WE, Lee T, Brickman A, Mayeux R, Brown TR, Small SA. Imagining the  $A\beta$ -related neurotoxicity of Alzheimer disease. Arch. Neurol. 2007; 64:1467–1477. [PubMed: 17923630]
- Morrison JH, Baxter MG. The ageing cortical synapse: hallmarks and implications for cognitive decline. Nat. Rev. Neurosci. 2012; 13:240–250. [PubMed: 22395804]
- Muir JL, Fischer W, Björklund A. Decline in visual attention and spatial memory in aged rats. Neurobiol. Aging. 1999; 20:605–615. [PubMed: 10674426]
- Nicholson DA, Yoshida R, Berry RW, Gallagher M, Geinisman Y. Reduction in size of perforate postynaptic densities in hippocampal axosinous synapses and age-related spatial learning impairments. J. Neurosci. 2004; 24:7648–7653. [PubMed: 15342731]
- Nicolle MM, Baxter MG. Glutamate receptor binding in the frontal cortex and dorsal striatum of aged rats with impaired attentional set-shifting. Eur. J. Neurosci. 2003; 18:3335–3342. [PubMed: 14686906]
- Nomura M, Izaki Y, Takita M, Tanaka J, Hori K. Extracellular level of basolateral amygdalar dopamine responding to reversal of appetitive-conditioned discrimination in young and old rats. Brain Res. 2004; 1018:241–246. [PubMed: 15276884]
- Norris CM, Korol DL, Foster TC. Increased susceptibility to induction of long-term depression and long-term potentiation reversal during aging. J. Neurosci. 1996; 16:5382–5392. [PubMed: 8757251]
- O'Brien JT, Desmond P, Ames D, Schweitzer I, Tress B. Magnetic resonance imaging correlates of memory impairment in the healthy elderly: Association with medial temporal lobe atrophy but not white matter lesions. Int. J. Geriatr. Psychiatry. 1997; 12:369–374. [PubMed: 9152723]
- O'Donnell KA, Rapp PR, Hof PR. Preservation of Prefrontal Cortical Volume in Behaviorally Characterized Aged Macaque Monkeys. Exp Neurol. 1999; 160:300–310. [PubMed: 10630214]
- O'Keefe J, Dostrovsky J. The hippocampus as a spatial map. Preliminary evidence from unit activity in the freely-moving rat. Brain Res. 1971; 34:171–175. [PubMed: 5124915]
- O'Keefe, J.; Nadel, L. The Hippocampus as a Cognitive Map. Clarendon Press; Oxford: 1978.
- Page TL, Einstein M, Duan H, He Y, Flores T, Rolshud D, Erwin JM, Wearne SL, Morrison JH, Hof PR. Morphological alterations in neurons forming corticocortical projections in the neocortex of aged Patas monkeys. Neurosci. Lett. 2002; 317:37–41. [PubMed: 11750991]
- Pannese E. Morphological changes in nerve cells during normal aging. Brain Struct. Funct. 2011; 216:85–89. [PubMed: 21431333]
- Passingham RE. Memory of monkeys (Macaca mulatta) with lesions in prefrontal cortex. Behav. Neurosci. 1985; 99:3–21. [PubMed: 4041231]
- Paxton JL, Barch DM, Racine CA, Braver TS. Cognitive control, goal maintenance, and prefrontal function in healthy aging. Cereb. Cortex. 2008; 18:1010–1028. [PubMed: 17804479]
- Penke L, Maniega SM, Murray C, Gow AJ, Hernández MCV, Clayden JD, Starr JM, Wardlaw JM, Bastin ME, Deary IJ. A general factor of brain white matter integrity predicts information processing speed in healthy older people. J. Neurosci. 2010; 30:7569–7574. [PubMed: 20519531]
- Penner MR, Roth TL, Chawla MK, Hoang LT, Roth ED, Lubin FD, Sweatt DJ, Worley PF, Barnes CA. Age-related changes in *Arc* transcription and DNA methylation within the hippocampus. Neurobiol. Aging. 2011; 32:2198–2210. [PubMed: 20189687]
- Peters A, Rosene DL, Moss MB, Kemper TL, Abraham CR, Tigges J, Albert MS. Neurobiological bases of age-related cognitive decline in the rhesus monkey. J. Neuropathol. Exp. Neurol. 1996; 55:861–874. [PubMed: 8759775]
- Peters A, Morrison JH, Rosene DL, Hyman BT. Feature article: are neurons lost from the primate cerebral cortex during normal aging? Cereb. Cortex. 1998a; 8:295–300. [PubMed: 9651126]
- Peters A, Sethares C, Moss MB. The effects of aging on layer 1 in area 46 of prefrontal cortex in the rhesus monkey. Cereb. Cortex. 1998b; 8:671–684. [PubMed: 9863695]

Peters, A. Structural changes in the normally aging cerebral cortex of primates. In: Efrain, C.; Azmitia, JD., editors. Progress in Brain Research. Elsevier; New York: 2002. p. 455-465.

- Peters A, Sethares C. Aging and the myelinated fibers in prefrontal cortex and corpus callosum of the monkey. J. Comp. Neurol. 2002; 442:277–291. [PubMed: 11774342]
- Peters A, Sethares C, Luebke JI. Synapses are lost during aging in the primate prefrontal cortex. Neuroscience. 2008; 152:970–981. [PubMed: 18329176]
- Phillips S, Takeda Y. Greater frontal-parietal synchrony at low gamma-band frequencies for inefficient than efficient visual search in human EEG. Int. J. Psychophysiol. 2009; 73:350–354. [PubMed: 19481120]
- Phillips S, Takeda Y. Frontal-parietal synchrony in elderly EEG for visual search. Int. J. Psychophysiol. 2010; 75:39–43. [PubMed: 19903501]
- Plassman BL, Langa KM, Fisher GG, Heeringa SG, Weir DR, Ofstedal MB, Burke JR, Hurd MD, Potter GG, Rodgers WL, Steffens DC, Willis RJ, Wallace RB. Prevalence of dementia in the United States: The aging demographics, and memory study. Neuroepidemiology. 2007; 29:125–132. [PubMed: 17975326]
- Plath N, Ohana O, Dammermann B, Errington ML, Schmitz D, Gross C, Mao X, Engelsberg A,
  Mahlke C, Welzl H, Kobalz U, Stawrakakis A, Fernandez E, Waltereit R, Bick-Sander A,
  Therstappen E, Cooke SF, Blanquet V, Wurst W, Salmen B, Bosl MR, Lipp HP, Grant SG, Bliss TV, Wolfer DP, Kuhl D. Arc/Arg3.1 is essential for the consolidation of synaptic plasticity and memories. Neuron. 2006; 52:437–444. [PubMed: 17088210]
- Plude DJ, Doussard-Roosevelt JA. Aging, selective attention, and feature integration. Psychol. Aging. 1989; 4:98–105. [PubMed: 2803617]
- Prakash RS, Erickson KI, Colcombe SJ, Kim JS, Voss MW, Kramer AF. Age-related differences in the involvement of the prefrontal cortex in attentional control. Brain Cogn. 2009; 71:328–335. [PubMed: 19699019]
- Prendergast MA, Jackson WJ, Terry AV Jr, Kille NJ, Arneric SP, Decker MW, Buccafusco JJ. Agerelated differences in distractibility and response to methylphenidate in monkeys. Cereb. Cortex. 1998; 8:164–172. [PubMed: 9542895]
- Ramos BP, Birnbaum SG, Lindenmayer I, Newton SS, Duman RS, Arnsten AFT. Dysregulation of protein kinase a signaling in the aged prefrontal cortex: New strategy for treating age-related cognitive decline. Neuron. 2003; 40:835–845. [PubMed: 14622586]
- Rapp PR, Amaral DG. Evidence for task-dependent memory dysfunction in the aged monkey. J. Neurosci. 1989; 9:3568–3576. [PubMed: 2795141]
- Rapp PR, Gallagher M. Preserved neuron number in the hippocampus of aged rats with spatial learning deficits. Proc. Natl. Acad. Sci. USA. 1996; 93:9926–9930. [PubMed: 8790433]
- Rapp PR, Kansky MT, Roberts JA. Impaired spatial information processing in aged monkeys with preserved recognition memory. NeuroReport. 1997; 8:1923–1928. [PubMed: 9223078]
- Rapp PR, Deroche PS, Mao Y, Burwell RD. Neuron number in the parahippocampal region is preserved in aged rats with spatial learning deficits. Cereb. Cortex. 2002; 12:1171–1179. [PubMed: 12379605]
- Rasmussen T, Schliemann T, Sorensen JC, Zimmer J, West MJ. Memory impaired aged rats: No loss of principal hippocampal and subicular neurons. Neurobiol. Aging. 1996; 17:143–147. [PubMed: 8786797]
- Raz N, Gunning FM, Head D, Dupuis JH, McQuain J, Briggs SD, Loken WJ, Thornton AE, Acker JD. Selective aging of the human cerebral cortex observed in vivo: Differential vulnerability of the prefrontal gray matter. Cereb. Cortex. 1997; 7:268–282. [PubMed: 9143446]
- Raz N, Rodrigue KM, Head D, Kennedy KM, Acker JD. Differential aging of the medial temporal lobe: A study of a five-year change. Neurology. 2004; 62:433–438. [PubMed: 14872026]
- Raz N, Lindenberger U, Rodrigue KM, Kennedy KM, Head D, Williamson A, Dahle C, Gerstorf D, Acker JD. Regional brain changes in aging healthy adults: General trends, individual differences and modifiers. Cereb. Cortex. 2005; 15:1676–1689. [PubMed: 15703252]
- Raz N, Ghisletta P, Rodrigue KM, Kennedy KM, Lindenberger U. Trajectories of brain aging in middle-aged and older adults: Regional and individual differences. Neuroimage. 2010; 51:501– 511. [PubMed: 20298790]

Ridderinkhof KR, Span MM, Van der Molen MW. Perseverative behavior and adaptive control in older adults: performance monitoring, rule induction, and set shifting. Brain Cogn. 2002; 49:382–401. [PubMed: 12139960]

- Roberson ED, DeFazio RA, Barnes CA, Alexander GE, Bizon JL, Bowers D, Foster TC, Glisky EL, Levin BE, Ryan L, Wright CB, Geldmacher DS. Challenges and opportunities for characterizing cognitive aging across species. Front Aging Neurosci. 2012; 4:6. [PubMed: 22988434]
- Roesch MR, Bryden DW, Cerri DH, Haney ZR, Schoenbaum G. Willingness to wait and altered encoding of time-discounted reward in the orbitofrontal cortex with normal aging. J. Neurosci. 2012; 32:5525–5533. [PubMed: 22514314]
- Rose GM, Ong VS, Woodruff-Pak DS. Efficacy of MEM 1003, a novel calcium channel blocker, in delay and trace eyeblink conditioning in older rabbits. Neurobiol. Aging. 2007; 28:766–773. [PubMed: 16621170]
- Salat DH, Lee SY, Van der Kouwe AJ, Greve DN, Fischl B, Rosas HD. Age-associated alterations in cortical gray and white matter signal intensity and gray to white matter contrast. Neuroimage. 2009; 48:21–28. [PubMed: 19580876]
- Salthouse TA. When does age-related cognitive decline begin? Neurobiol. Aging. 2009; 30:507–514. [PubMed: 19231028]
- Samanez-Larkin GR, Levens SM, Perry LM, Dougherty RF, Knutson B. Frontostriatal white matter integrity mediates adult age differences in probabilistic reward learning. J. Neurosci. 2012; 32:5333–5337. [PubMed: 22496578]
- Schimanski LA, Lipa P, Barnes CA. Tracking the course of hippocampal representations during learning: When is the map required? J. Neurosci. 2013 in press.
- Schoenbaum G, Nugent S, Saddoris MP, Gallagher M. Teaching old rats new tricks: Age-related impairments in olfactory reversal learning. Neurobiol. Aging. 2002; 23:555–564. [PubMed: 12009505]
- Schoenbaum G, Setlow B, Saddoris MP, Gallagher M. Encoding changes in orbitofrontal cortex in reversal-impaired aged rats. J. Neurophysiol. 2006; 95:1509–1517. [PubMed: 16338994]
- Shamy JLT, Buonocore MH, Makaron LM, Amaral DG, Barnes CA, Rapp PR. Hippocampal volume is preserved and fails to predict recognition memory impairment in aged rhesus monkeys (mucaca mulatta). Neurobiol. Aging. 2006; 27:1405–1415. [PubMed: 16183171]
- Shamy JL, Habeck C, Hof PR, Amaral DG, Fong SG, Buonocore MH, Stern Y, Barnes CA, Rapp PR. Volumetric correlates of spatiotemporal working and recognition memory impairment in aged rhesus monkeys. Cereb. Cortex. 2011; 21:1559–1573. [PubMed: 21127015]
- Shen J, Barnes CA. Age-related decrease in cholinergic synaptic transmission in three hippocampal subfields. Neurobiol. Aging. 1996; 17:439–451. [PubMed: 8725906]
- Shen J, Barnes CA, McNaughton BL, Skaggs WE, Weaver KL. The effect of aging on experience-dependent plasticity of hippocampal place cells. J. Neurosci. 1997; 17:6769–6782. [PubMed: 9254688]
- Shing YL, Rodrigue KM, Kennedy KM, Fandakova Y, Bodammer N, Werkle-Bergner M, Lindenberger U, Raz N. Hippocampal subfield volumes: Age, vascular risk, and correlation with associative memory. Front. Aging Neurosci. 2011; 3:2. [PubMed: 21331174]
- Singh-Manoux A, Kivimaki M, Glymour MM, Elbaz A, Berr C, Ebmeier KP, Ferrie JE, Dugravot A. Timing of onset of cognitive decline: Results from Whitehall II prospective cohort study. BMJ. 2011; 344:d7622. [PubMed: 22223828]
- Simon NW, LaSarge CL, Montgomery KS, Williams MT, Mendez IA, Setlow B, Bizon JL. Good things come to those who wait: Attenuated discounting of delayed rewards in aged Fischer 344 rats. Neurobiol. Aging. 2010; 31:853–862. [PubMed: 18657883]
- Small SA, Tsai WY, DeLaPaz R, Mayeux R, Stern Y. Imaging hippocampal function across the human life span: Is memory decline normal or not? Ann. Neurol. 2002; 51:290–295. [PubMed: 11891823]
- Small SA, Chawla MK, Buonocore M, Rapp PR, Barnes CA. Imaging correlates of brain function in monkeys and rats isolates a hippocampal subregion differentially vulnerable to aging. Proc. Natl. Acad. Sci. USA. 2004; 101:7181–7186. [PubMed: 15118105]

Small SA, Schobel SA, Buxton RB, Witter MP, Barnes CA. A pathophysiological framework of hippocampal dysfunction in ageing and disease. Nat. Rev. Neurosci. 2011; 12:585–601. [PubMed: 21897434]

- Smith TD, Adams MM, Gallagher M, Morrison JH, Rapp PR. Circuit-specific alterations in hippocampal synaptophysin immunoreactivity predict spatial learning impairment in aged rats. J. Neurosci. 2000; 20:6587–6593. [PubMed: 10964964]
- Smith DE, Rapp PR, McKay HM, Roberts JA, Tuszynski MH. Memory Impairment in Aged Primates Is Associated with Focal Death of Cortical Neurons and Atrophy of Subcortical Neurons. J. Neurosci. 2004; 24:4373–4381. [PubMed: 15128851]
- Spaniol J, Grady C. Aging and the neural correlates of source memory: over-recruitment and functional reorganization. Neurobiol Aging. 2012; 33:e3–18. [PubMed: 21111514]
- Spreng RN, Wojtowicz M, Grady CL. Reliable differences in brain activity between young and old adults: a quantitative meta-analysis across multiple cognitive domains. Neurosci Biobehav Rev. 2010; 34:1178–1194. [PubMed: 20109489]
- Stephens DN, Weidmann R, Quartermain D, Sarter M. Reversal learning in senescent rats. Behav. Brain Res. 1985; 17:193–202. [PubMed: 4084391]
- Stevens WD, Hasher L, Chiew KS, Grady CL. A neural mechanism underlying memory failure in older adults. J. Neurosci. 2008; 28:12820–12824. [PubMed: 19036975]
- Störmer VS, Passow S, Biesenack J, Li S-C. Dopaminergic and cholinergic modulations of visual-spatial attention and working memory: Insights from molecular genetic research and implications for adult cognitive development. Dev. Psychol. 2012; 48:875–889. [PubMed: 22103306]
- Stoub T, Barnes CA, Shah RC, Stebbins GT, Ferrari C, Detoledo-Morrell L. Age-related changes in the mesial temporal lobe: The parahippocampal white matter region. Neurobiol. Aging. 2012; 33:1168–1176. [PubMed: 21459484]
- Stranahan AM, Jiam NT, Spiegel AM, Gallagher M. Aging reduces total neuron number in the dorsal component of the rodent prefrontal cortex. J. Comp. Neurol. 2012; 520:1318–1326. [PubMed: 22020730]
- Sullivan EV, Marsh L, Pfefferbaum A. Preservation of hippocampal volume throughout adulthood in healthy men and women. Neurobiol. Aging. 2005; 26:1093–1098. [PubMed: 15748789]
- Talsma D, Senkowski D, Soto-Faraco S, Woldorff MG. The multifaceted interplay between attention and multisensory integration. Trends Cogn. Sci. 2010; 14:400–410. [PubMed: 20675182]
- Terry RD, DeTeresa R, Hansen LA. Neocortical cell counts in normal human adult aging. Ann. Neurol. 1987; 21:530–539. [PubMed: 3606042]
- Thambisetty M, Wan J, Carass A, An Y, Prince JL, Resnick SM. Longitudinal changes in cortical thickness associated with normal aging. Neuroimage. 2010; 52:1215–1223. [PubMed: 20441796]
- Thibault O, Landfield PW. Increase in single L-type calcium channels in hippocampal neurons during aging. Science. 1996; 272:1017–1020. [PubMed: 8638124]
- Tiesinga PHE, Fellous J-M, José JV, Sejnowski TJ. Computational model of carbachol-induced delta, theta, and gamma oscillations in the hippocampus. Hippocampus. 2001; 11:251–274. [PubMed: 11769308]
- Tisserand DJ, Visser PJ, van Boxtel MPJ, Jolles J. The relation between global and limbic brain volumes on MRI and cognitive performance in healthy individuals across the age range. Neurobiol. Aging. 2000; 21:569–576. [PubMed: 10924774]
- Tisserand DJ, Pruessner JC, Sanz Arigita EJ, Van Boxtel MPJ, Evans AC, Jolles J, Uylings HBM. Regional frontal cortical volumes decrease differentially in aging: An MRI study to compare volumetric approaches and voxel-based morphometry. NeuroImage. 2002; 17:657–669. [PubMed: 12377141]
- Tolman EC. Cognitive maps in rats and men. Psychol. Rev. 1948; 55:1–4.
- Turner DA, Deupree DL. Functional elongation of CA1 hippocampal neurons with aging in Fischer 344 rats. Neurobiol. Aging. 1991; 12:201–210. [PubMed: 1876226]
- Uttl B, Graf P. Episodic spatial memory in adulthood. Psychol. Aging. 1993; 8:257–273. [PubMed: 8323729]

Van Petten C. Relationship between hippocampal volume and memory ability in healthy individuals across the lifespan: Review and meta-analysis. Neuropsychologia. 2004; 42:1394–1413. [PubMed: 15193947]

- Voytko ML. Impairments in acquisition and reversals of two-choice discriminations by aged rhesus monkeys. Neurobiol. Aging. 1999; 20:617–627. [PubMed: 10674427]
- Wagster MV, King JW, Resnick SM, Rapp PR. The 87%. J. Gerontol. A Biol. Sci. Med. Sci. 2012; 67:739–740. [PubMed: 22773154]
- Wang X-J. Neurophysiological and computational principles of cortical rhythms in cognition. Physiol. Rev. 2010; 90:1195–1268. [PubMed: 20664082]
- Wang M, Gamo NJ, Yang Y, Jin LE, Wang X-J, Laubach M, Mazer JA, Lee D, Arnsten AFT. Neuronal basis of age-related working memory decline. Nature. 2011; 476:210–213. [PubMed: 21796118]
- Weiler JA, Bellebaum C, Daum I. Aging affects acquisition and reversal of reward-based associative learning. Learn. Mem. 2008; 15:190–197. [PubMed: 18353994]
- West MJ, Amaral DJ, Rapp PR. Preserved hippocampal cell number in aged monkeys with recognition memory deficits. Soc. Neurosci. Abst. 1993; 19:599.
- Wilson IA, Ikonen S, Gallagher M, Eichenbaum H, Tanila H. Age-associated alterations of hippocampal place cells are subregion specific. J. Neurosci. 2005; 25:6877–6886. [PubMed: 16033897]
- Yassa MA, Muftuler LT, Stark CEL. Ultrahigh-resolution microstructural diffusion tensor imaging reveals perforant path degradation in aged humans in vivo. Proc. Natl. Acad. Sci. USA. 2010; 107:12687–12691. [PubMed: 20616040]
- Yassa MA, Mattfeld AT, Stark SM, Stark CEL. Age-related memory deficits linked to circuit-specific disruptions in the hippocampus. Proc. Natl. Acad. Sci. USA. 2011; 108:8873–8878. [PubMed: 21555581]
- Yates MA, Markham JA, Anderson SE, Morris JR, Juraska JM. Regional variability in age-related loss of neurons from the primary visual cortex and medial prefrontal cortex of male and female rats. Brain Res. 2008; 1218:1–12. [PubMed: 18513705]
- Yeterian EH, Pandya DN, Tomaiuolo F, Petrides M. The cortical connectivity of the prefrontal cortex in the monkey brain. Cortex. 2012; 48:58–81. [PubMed: 21481342]

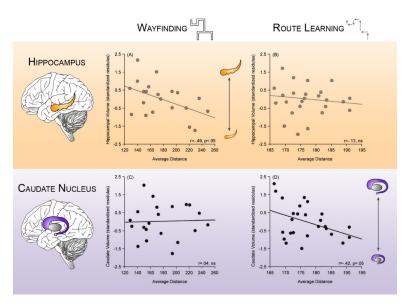


Figure 1.
Older participants (56-88 years) were trained to use either a wayfinding strategy or a route strategy to learn to navigate through a virtual environment. Correlations are shown between MRI-derived volumes of hippocampus and caudate nucleus in these individuals and performance levels on these two types of spatial navigation tasks. A) Participants with larger hippocampal volumes traveled shorter virtual distances to find a landmark when using a wayfinding strategy. B) There was no correlation between hippocampal volume and performance levels when using a route learning strategy. C) There was no significant relationship found between caudate nucleus volumes and wayfinding performance. D) Participants with larger caudate nucleus volumes, on the other hand, showed more accurate performance when they used a route learning strategy. Adapted with permissions from (Head and Isom, 2010).

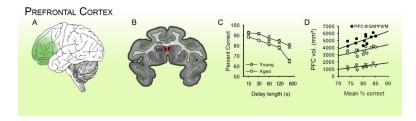


Figure 2.
Prefrontal cortical (PFC) volume correlates with performance on a DNMS task, in aged rhesus macaques. A. Illustration of a human brain with the frontal lobe highlighted in green.
B. Example coronal section of the PFC, illustrating the areas taken for ROIs for gray matter (dark grey) and white matter (light grey) volumes. The red marks the delineation of the ventricles. C. Aged monkeys (grey circles) were less accurate than were young monkeys (white circles) on a DNMS task, particularly at longer delays. D. The gray matter (grey circles), white matter (white triangles) and total PFC volumes (black circles) correlated with task accuracy in aged animals. Adapted with permissions from (Shamy et al., 2011).

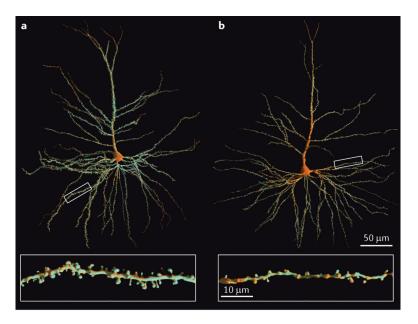


Figure 3. The spine density of prefrontal cortical (PFC) neurons is greatly reduced in aged monkeys. Illustration of a Lucifer yellow-filled PFC neurons from a young (left) and an aged (right) monkey. In the cells presented here, there are no age differences in the extent of dendritic arborization; however a marked decreased in spine density is observed. The rectangles show a higher magnification of a portion of a basal dendrite from each neuron. Reproduced with permissions from (Morrison & Baxter, 2012).