



Published in final edited form as:

J Natl Compr Canc Netw. 2011 January ; 9(1): 13–25.

Personalized Medicine and Oncology Practice Guidelines: A Case Study of Contemporary Biomarkers in Colorectal Cancer

Robin K. Kelley, MD^a, Stephanie L. Van Bebber, MSc^{b,c}, Kathryn A. Phillips, PhD^{a,b,c,d}, and Alan P. Venook, MD^a

^aDepartment of Medicine, UCSF Helen Diller Family Comprehensive Cancer Center, University of California San Francisco, San Francisco, California

^bUCSF Center for Translational and Policy Research on Personalized Medicine (TRANSPERS), University of California San Francisco, San Francisco, California

^cDepartment of Clinical Pharmacy, School of Pharmacy, University of California San Francisco, San Francisco, California

^dPhilip R. Lee Institute for Health Policy Studies, San Francisco, California

Abstract

Predictive and prognostic biomarkers offer a potential means to personalize cancer medicine, although many reach the marketplace before they have been validated, and their adoption is often hindered by variable clinical evidence. Because of this variability in supporting evidence, clinical practice guidelines formulated by panels of subspecialty experts may be particularly important in guiding stakeholders' acceptance and use of new personalized medicine biomarker tests and other nascent technologies. This article provides a structured review of the clinical evidence supporting 4 contemporary biomarker tests in colorectal cancer: *K-ras* and *B-raf* mutation analyses, mismatch repair protein testing, and the *Oncotype DX* Colon Cancer Assay. All 4 tests have been evaluated for guideline inclusion by the NCCN Guidelines Panel for Colon Cancer. This case study shows significant variability in the level of clinical evidence associated with these tests. In the cases of *B-raf* and mismatch repair protein testing, the available evidence is also inconsistent as it pertains to the specific NCCN guideline recommendation. Based on this uncertainty in the evidence base, the authors conclude that expert clinical judgment, experience, and consensus may be more heavily weighted than published clinical trial data in the evaluation of new personalized medicine biomarker tests. Potential implications of this conclusion and future directions for research are discussed.

Keywords

Biomarker; personalized medicine; guidelines; KRAS; BRAF; microsatellite instability; *Oncotype DX* Colon Cancer Assay

Predictive and prognostic biomarkers offer the potential for personalized therapy in oncology. Despite a multitude of publications in the oncology literature identifying potential biomarkers, however, only a select few are recommended for use in clinical practice.^{1–5} A challenge to the adoption of personalized medicine biomarkers in oncology is the lack of a standardized validation process because of the heterogeneity of tumor types, treatments, and

tests themselves.^{1,3,6-8} Validation studies also may be limited by small data sets, long time intervals required to achieve end points, statistical complexity, cost, and the bias inherent in retrospective analysis.^{1,4,5,9-11} Randomized, controlled data are required to clearly define which markers are predictive and which are prognostic, because single-arm studies can be misleading.^{1,4} New biomarker tests may become available before the arduous validation process is complete, requiring practitioners to navigate the competing pressures of the existing data, the “blogosphere” recommendations, cost, and the patients’ best interests. This paradigm exists in stark contrast to the process of new drug development, in which the FDA requires rigorous demonstration of safety and efficacy before granting approval for commercialization.¹¹⁻¹³

Perhaps more so than in other medical specialties, oncology-focused consensus guidelines developed by multidisciplinary expert panels influence the decisions of clinicians, payers, and policy makers.¹⁴⁻²² Guideline adherence has been associated with improved outcomes in some studies.^{14,15,18,23} Worldwide, multiple cancer organizations produce practice guidelines. In the United States, ASCO, NCI, and NCCN publish clinical practice guidelines widely used in clinical practice. Among these, NCCN offers the most frequently updated practice guidelines specific to most common tumor types, and routinely include comprehensive diagnostic, risk stratification, treatment, and surveillance algorithms.²⁴⁻²⁶ The recent decision of the Centers for Medicare & Medicaid Services (CMS) to recognize the NCCN Drugs & Biologics Compendium (NCCN Compendium) underscores the integral role of these guidelines in the oncology community.²⁷

The published methodologies of the ASCO, NCI, and NCCN guidelines do not specifically describe the methods for evaluating nascent technologies such as new biomarkers, but they do acknowledge that expert clinical experience and judgment may be required when data pertaining to a specific intervention are incomplete.^{28,29} Currently, the NCCN guidelines are the only United States guidelines with recent updates reflecting new biomarker data across multiple malignancies. It stands to reason that timely, interim synthesis of the available evidence by expert guidelines panels may play a particularly important role in guiding practitioners’ use of new personalized medicine biomarkers with varying levels of supporting data at commercial release.

This article presents a case study of biomarker integration into the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines) using examples from colorectal cancer, a disease in which molecular markers of prognosis and treatment efficacy have been studied extensively.^{30,31} The objective for this case study is to describe and compare the available clinical evidence for 4 heterogeneous but contemporary biomarker tests in colorectal cancer: *K-ras* and *B-raf* mutation analyses, mismatch repair protein (MMR) testing, and the Oncotype DX Colon Cancer Assay (Genomic Health, Inc., Redwood City, California).²⁴ Each of these biomarker tests has been considered for guideline inclusion by the NCCN Guidelines Panel for Colon Cancer. The authors hypothesize that varying levels of and inconsistency in the clinical evidence supporting these biomarkers could impact the process of biomarker adoption by consensus guidelines, which rely on a combination of both clinical evidence and expert opinion.

Methods

Selection of Colorectal Cancer Biomarker Examples for Case Study

K-ras and *B-raf* mutation analyses and MMR testing were selected as examples of contemporary bio-marker tests that recently have been incorporated into the NCCN Guidelines for Colon Cancers.²⁴ The NCCN Categories of Evidence and Consensus are provided in the guidelines, available online, at www.NCCN.org. The Oncotype DX Colon

Cancer Assay was selected for this case study because of its recent entry into the marketplace for use in stage II colon cancer risk assessment. This test is currently not explicitly referenced in the NCCN Guidelines, but the data are in the public domain. The NCCN Guidelines Panel for Colon Cancer has evaluated the body of evidence for multi-gene assays in stage II colon cancer with the conclusion that the data are insufficient to support the use of assays such as the *Oncotype* DX Colon Cancer Assay for treatment decision-making.^{32,33} This case study is not intended to be a comprehensive review of all biomarkers with potential relevance to colorectal cancer; the 4 selected examples represent a purposive convenience sample.

Literature Search Methods and Retrieved References

This case study uses a structured review of the 4 selected biomarkers. Searches were targeted to capture data pertaining to the specific NCCN guideline recommendation for each biomarker test.

Structured Review of Clinical Evidence

A descriptive approach to characterizing the evidence was used because the body of evidence supporting the selected biomarkers is relatively small and heterogeneous and because both the NCCN and ASCO guidelines methodologies use similar qualitative analyses.^{28,29} For the published clinical studies identified, the study design, end points, and numbers of patients or specimens are summarized in table format. A review of the assay methods, analytic validity, and technology for each biomarker test is beyond the scope of this case study.

Results

The NCCN Guidelines recommendation, date of inclusion if applicable, and category of evidence and consensus for each of the selected biomarker tests in colorectal cancer are presented in Table 1. For each of these biomarkers, the clinical evidence is summarized descriptively later. Literature search results are also presented for each biomarker (Tables 2–5) to provide specific references according to study type, along with end points and numbers of patients. Table 6 compares the evidence across the 4 biomarkers and summarizes the conclusions regarding the consistency of the cumulative evidence for prognostic value, predictive value, or both for each test.

K-ras Mutation Analysis in Metastatic Colorectal Cancer

The *K-ras* gene is a downstream target of epidermal growth factor receptor (EGFR) signaling. Activating mutations in codon 12 or 13 of this gene are present in approximately 30% to 40% of colorectal cancers.³⁴ Retrospective subset analyses of tumor tissue samples from small clinical trials initially showed that tumor *K-ras* gene mutations are associated with lack of response to cetuximab and panitumumab, the 2 EGFR-targeted monoclonal antibodies approved for use in colorectal cancer.^{35–61} The strength of this association was later substantiated in retrospective analyses of patients treated in 6 large randomized studies.^{62–67} Search results are summarized in Table 2.

The evidence shows with great consistency that patients whose tumors harbor a mutation in the *K-ras* gene do not benefit from cetuximab or panitumumab, whether in monotherapy or in combination with chemotherapy, whereas those whose tumors are wild-type have significantly higher response rates and longer survival. These findings clearly establish tumor *K-ras* mutation as a predictive factor for non-response to EGFR-targeted therapy. Data are mixed regarding whether *K-ras* mutation is a negative prognostic factor independent of treatment with EGFR-targeted therapy.^{61,65–70}

K-ras mutation analysis was included in the NCCN Guidelines as a Category 2A recommendation in 2008 before the package label for either antibody was changed and before FDA acknowledgment of *K-ras* testing as a standard. The guideline inclusion was based on the publication of a retrospective subset analysis of the randomized, phase III study of panitumumab versus best supportive care, and after national and international presentation of results from similar unplanned retrospective analyses of subsets from the randomized, phase III CRYSTAL and CAIRO2 trials, the randomized phase II OPUS trial, and a multicenter, multinational randomized phase III trial of cetuximab versus best supportive care.^{62,71–74}

***B-raf* Mutation Analysis in Metastatic Colorectal Cancer**

The *B-raf* gene encodes a protein kinase downstream in the *K-ras* pathway. Activating mutations in *B-raf* at the V600E site are present in approximately 10% of patients with metastatic colorectal cancer and seem to be mutually exclusive with activating *K-ras* mutations.^{40,68,75,76} Soon after publication and presentation of the data establishing *K-ras* as a predictive factor, a small, retrospective series suggested that patients with *K-ras* wild-type with mutations in *B-raf* at the V600E site who were treated with cetuximab or panitumumab had significantly poorer outcomes than those without the V600E mutation.⁴⁰ These data were reinforced by 3 other small, retrospective, uncontrolled studies with similar findings.^{35,46,50}

However, subset analysis of the randomized, phase III CAIRO-2 study was not consistent with these findings.⁷⁵ In both treatment arms of CAIRO-2, a *B-raf* V600E mutation was associated with shorter progression-free survival without any difference in response rate compared with wild-type tumors, suggesting prognostic as opposed to predictive value. This result was consistent with a retrospective analysis of samples from the randomized PET-ACC-3 adjuvant study of patients with locoregional disease and with subset data from the CRYSTAL trial presented at the ASCO 2010 Gastrointestinal Cancers Symposium.^{68,76} Among the 625 evaluable patients in the CRYSTAL trial whose tumors were nonmutated for the *K-ras* gene, *B-raf* mutation (present in 59 patients) was shown to be associated with significantly worse survival outcomes regardless of treatment arm. These preliminary results are cited in the NCCN Guidelines and therefore are included in Table 3 along with published search results.

Therefore, the *B-raf* V600E mutation seems to be a negative prognostic factor in patients with metastatic colorectal cancer, independent of treatment with EGFR-targeted agents. The evidence is not consistent regarding its predictive value for non-response to cetuximab and panitumumab.

Based on the data as of 2009, the NCCN guidelines added the statement that patients with nonmutated *K-ras* tumors known to harbor a *B-raf* V600E mutation are unlikely to benefit from cetuximab or panitumumab.²⁴ This was listed as a Category 2A recommendation, but no specific recommendation was made regarding the performance of *B-raf* mutation analysis. After presentation of conflicting results from *B-raf* subset data of the CRYSTAL trial, this recommendation was amended to include the statement, “although the data are somewhat inconsistent.”

MMR Testing as a Predictive Factor in Stage II Colon Cancer

MMR deficiency is present in approximately 15% to 20% of colorectal cancers and may be from sporadic or inherited inactivation of a mismatch repair protein: MLH1, MSH2, MSH6, or PMS2.^{77–80} Tumor MMR testing historically has been reserved for patients meeting the Revised Bethesda Guidelines clinical criteria for genetic testing for hereditary nonpolyposis

colorectal cancer (HNPCC).^{78,80} This case study focuses on the possible role of MMR deficiency as a predictive factor for lack of benefit from 5-fluorouracil chemotherapy in patients with stage II colon cancer. Among the 16 nonrandomized studies identified by this search, several suggest the possibility of improved outcomes with 5-fluorouracil-based treatment in patients with MMR-deficient locoregional colorectal cancers compared with patients with proficient MMR.^{82–85}

However, some of these studies suggest that patients with MMR deficiency treated with 5-fluorouracil-based therapy experience no benefit or have a potentially worse outcome.^{85–93} Interpretation of all of these studies is confounded by the known strong positive prognostic value of MMR deficiency, and whether other prognostic factors such as *B-raf* mutation, which seems to be enriched in sporadic cases of MMR deficiency, were balanced across arms is unknown. Heterogeneity of stage, tumor location, and treatment (particularly whether combination therapies were used) further confounds interpretation of these data. In the 8 published, randomized studies identified, the results are also inconsistent.^{94–101} Among these, the most compelling published data for MMR deficiency as a predictive marker for lack of benefit from adjuvant 5-fluorouracil in locoregional colon cancer were shown by a retrospective, pooled analysis of MMR status in 570 tumor specimens of patients enrolled in randomized studies of adjuvant 5-fluorouracil with levamisole or leucovorin compared with no adjuvant therapy.⁹⁸ This study showed no improvement in overall survival in patients with stage II and III colon cancer with MMR-deficient tumors treated with adjuvant 5-fluorouracil compared with those with MMR-proficient tumors, who did experience benefit. A recently published large meta-analysis, which was first presented at the 2008 ASCO Annual Meeting, corroborated this impression and further suggested the possibility of worse outcomes in patients with MMR-deficient stage II colon cancers treated with 5-fluorouracil.^{102,103}

In contrast to these 2 studies, several other randomized studies have suggested that patients with MMR-deficient tumors may derive benefit from treatment with 5-fluorouracil-based therapies, although again interpretation is confounded by inclusion of patients with stage III disease and heterogeneous treatment, including 5-fluorouracil-based combination therapy arms and single-agent arms.^{94,97,101} Search results are summarized in Table 4.

Cumulatively, the data mentioned earlier provide somewhat equivocal evidence for MMR deficiency as a predictive marker for lack of benefit from 5-fluorouracil therapy in patients with locoregional colorectal cancer in general, and specifically in those with stage II colon cancer. The strong positive prognostic value of MMR deficiency is consistent across many large, randomized studies.^{30,31,68,77,79,104}

The latest version of the NCCN Guidelines added MMR testing to the risk stratification algorithm for stage II colon cancer as a Category 2A recommendation based on the data for this test as a predictive marker for lack of benefit from adjuvant 5-fluorouracil therapy, citing results of the recent meta-analysis presented at the 2008 ASCO Annual Meeting.^{24,102}

Oncotype DX Colon Cancer Assay

The *Oncotype DX* Colon Cancer Assay became available on the market in January of 2010.^{24,32} This assay characterizes gene expression in fixed, paraffin-embedded tumor specimens using reverse transcriptase-polymerase chain reaction (RT-PCR) to generate a 7-gene recurrence score for patients with stage II colon cancers.¹⁰⁴ The assay showed quantitative precision and reproducibility in an initial development set from the C-01/C-02 randomized adjuvant studies of the National Surgical Adjuvant Breast and Bowel Project (NSABP), and subsequent results from its 4 development sets and validation set have been presented at national oncology conferences but remain unpublished at the time of

writing.^{104–108} Validation of the *Oncotype DX* Colon Cancer Assay has been performed by retrospective, subset analysis of 1436 tissue blocks from patients with stage II colon cancer enrolled in the randomized QUASAR study comparing postsurgical adjuvant 5-fluorouracil therapy with observation alone.¹⁰⁹ In the validation set, the test showed discrimination of recurrence risk as a continuum between low, intermediate, and high recurrence score groups with estimated recurrence risk at 3 years of 12%, 18%, and 22%, respectively; the hazard ratio for recurrence between the low- and high-risk groups was 1.47 ($P = .046$) using the Cox model. Search results are summarized in Table 5.

Based on these preliminary data, the *Oncotype DX* Colon Cancer Assay recurrence score seems to provide prognostic information independent of conventional risk factors, discriminating the absolute increase in recurrence risk at 3 years between low- and high-risk patients by approximately 10%.¹⁰⁴ The *Oncotype DX* Colon Cancer Assay is not predictive of 5-fluorouracil benefit, however, because the recurrence risk reduction from chemotherapy seemed proportional across all risk groups in the QUASAR dataset.¹⁰⁴

The *Oncotype DX* Colon Cancer Assay has not been included in NCCN Guidelines, which currently state that data are insufficient to recommend the use of multigene assay panels to determine adjuvant therapy.²⁴

Conclusions

This article presents a case study of 4 contemporary examples of personalized medicine biomarkers in colorectal cancer, describing the available clinical evidence for each example. All 4 of these biomarkers have been evaluated for inclusion in the wide-reaching NCCN Guidelines. Based on this structured review, the authors conclude that the level of published clinical evidence for these biomarkers is variable, and in some cases, discordant in content. This finding suggests that, by necessity, the domains of expert experience, clinical judgment, and consensus may play a greater role than published clinical trial data in guideline development for new personalized medicine biomarkers.

Reliance on expert opinion may be both a strength and a potential limitation when evaluating new technologies with rapidly evolving data, such as biomarkers. On the one hand, reliance on expert opinion enables timely review and incorporation of new information, resulting in the most up-to-date, accessible, and useful guidelines for stakeholders who must assimilate new technologies. The above-discussed qualification in *B-raf* recommendations soon after presentation of new data reflects the dynamic and adaptive nature of the NCCN Guidelines. Other guidelines that rely on formal systematic review of the evidence are more laborious, require mature data sets and studies, and are therefore slow to respond to new data, rendering them less useful to practitioners.²⁶ In a recent survey of 459 breast cancer surgeons, expert opinion followed by guidelines and consensus statements have been shown to have the strongest influence on decision-making in areas of scientific uncertainty.²²

Conversely, however, the strong reliance on expert opinion introduces potential for bias. The NCCN Guidelines development process uses rigorous safeguards, including strict conflict of interest disclosure requirements, an iterative process with review by and input from practitioners at member institutions, and inclusion of panel members representing diverse specialties and viewpoints.^{29,110} Among the 4 examples studied, the 3 tests that have been recommended by the NCCN Panel for Colon Cancer (*K-ras* and *B-raf* mutation analyses and MMR testing) are not proprietary to any single commercial entity, unlike the *Oncotype DX* Colon Cancer Assay, which has not been adopted by the guidelines.

These measures, however, are not protective of subtle factors that might influence the uptake of new personalized medicine technologies into oncology guidelines. For example, in the case of the new *B-raf* mutation analysis recommendation, it is possible that the momentum and enthusiasm generated by the *K-ras* biomarker discovery influenced panel members' impression of level of evidence and likelihood of improvement in patient outcomes from the related downstream biomarker, *B-raf*V600E mutation. The threshold for biomarker recommendation may also vary by the type of malignancy because of differences in research funding, patient advocacy, the risk inherent to the specific tumor type, treatment efficacy and toxicity, and subspecialty bias, factors that are not addressed by this case study.

This case study highlights several other intriguing aspects of biomarker evaluation and uptake by these influential guidelines. First, the examples of *K-ras* and *B-raf* mutation analysis, MMR testing, and the *Oncotype* DX Colon Cancer Assay highlight the challenge of applying a uniform methodology to categorize the variable level of data generally associated with biomarker studies.^{1,111} Across the 4 tests, the clinical evidence consists largely of retrospective subset analyses of patient subsets derived from case series, cohorts, and prospective, randomized trials. The current NCCN Categories of Evidence and Consensus ratings do not clearly discriminate differences in level of available evidence for *K-ras*, *B-raf*, and MMR testing. All 3 of these recommendations are assessed as Category 2A level of evidence and consensus, whereas this structured review suggests a significantly lower level of evidence for *B-raf* testing compared with the other 2 tests at the time of its inclusion in the guidelines.

These examples also show that it is very difficult to show improvements in patient outcome, one of the domains included in the NCCN Guideline evaluation methodology, for biomarkers. Biomarker studies may require a different set of standardized end points than studies of therapeutic modalities.¹ Inherent to the challenge of selecting appropriate end points for biomarker studies is the importance of determining whether a biomarker's association with a specific outcome is because it is a predictive marker for response to the treatment being studied, or because it is a strong prognostic marker independent of treatment. Many studies claiming predictive value do not uniformly include a control arm to exclude the contamination of a strong prognostic marker, as exemplified by studies of both *B-raf* mutation analysis and MMR testing. In the case of the *B-raf*V600E mutation, the lack of controlled studies impedes clear designation of this marker as prognostic, predictive, or both; because of this, the initial NCCN Guideline recommendation for *B-raf* interpretation has been amended to reflect the inconsistency suggested by new, controlled data sets.^{24,76}

Another observation from this case study is that NCCN Guideline inclusion often follows presentation of preliminary data at a national conference such as the ASCO Annual Meeting or a subspecialty symposium. This coincident timing likely follows naturally from the maturation of available evidence and consensus expert opinion from both organizations. However, given the previously discussed importance of guideline inclusion in the decisions of stakeholders, including policymakers, payors, and practitioners, this association in timing may merit further study.

Finally, although this case study focuses specifically on personalized medicine biomarker tests, the findings may apply to other types of nascent technologies and treatments in oncology that may be available to practitioners before the supporting evidence base is complete.

Future Directions

As long as medical technologies can reach the market before their optimal use has been comprehensively defined by validation studies, oncology practitioners are likely to continue

to rely on the recommendations of recognized bodies of experts, such as NCCN panelists, for guidance in their use. The methods of evaluation and decision by these guideline bodies, therefore, are likely to play a significant, ongoing role in the adoption of personalized medicine bio-markers in oncology. Further study is warranted to understand the complex balance between emerging clinical evidence and expert opinion in the integration of personalized medicine technology into oncology practice guidelines.

Acknowledgments

This study was supported by a grant from the National Cancer Institute (P01CA130818) through the Center for Translational and Policy Research on Personalized Medicine (TRANSPERS) at the University of California, San Francisco. This study was also supported in part by the Maisin Foundation.

References

1. McShane LM, Altman DG, Sauerbrei W, et al. REporting recommendations for tumour MARKer prognostic studies (REMARK). *Eur J Cancer*. 2005; 41:1690–1696. [PubMed: 16043346]
2. Puzstai L. Perspectives and challenges of clinical pharmacogenomics in cancer. *Pharmacogenomics*. 2004; 5:451–454. [PubMed: 15212580]
3. Simon R. Clinical trial designs for evaluating the medical utility of prognostic and predictive biomarkers in oncology. *Per Med*. 2010; 7:33–47. [PubMed: 20383292]
4. Koopman M, Venderbosch S, Nagtegaal ID, et al. A review on the use of molecular markers of cytotoxic therapy for colorectal cancer, what have we learned? *Eur J Cancer*. 2009; 45:1935–1949. [PubMed: 19473832]
5. Taube SE. Biomarkers in oncology: trials and tribulations. *Ann N Y Acad Sci*. 2009; 1180:111–118. [PubMed: 19906265]
6. Chau CH, Rixe O, McLeod H, et al. Validation of analytic methods for biomarkers used in drug development. *Clin Cancer Res*. 2008; 14:5967–5976. [PubMed: 18829475]
7. Kelley R, Venook AP. Drug development in advanced colorectal cancer: challenges and opportunities. *Curr Oncol Rep*. 2009; 11:175–185. [PubMed: 19336009]
8. Simon R, Altman DG. Statistical aspects of prognostic factor studies in oncology. *Br J Cancer*. 1994; 69:979–985. [PubMed: 8198989]
9. Dupuy A, Simon RM. Critical review of published microarray studies for cancer outcome and guidelines on statistical analysis and reporting. *J Natl Cancer Inst*. 2007; 99:147–157. [PubMed: 17227998]
10. Fan X, Shi L, Fang H, et al. DNA microarrays are predictive of cancer prognosis: a re-evaluation. *Clin Cancer Res*. 2010; 16:629–636. [PubMed: 20068095]
11. Gutman S, Kessler LG. The US Food and Drug Administration perspective on cancer biomarker development. *Nat Rev Cancer*. 2006; 6:565–571. [PubMed: 16794639]
12. Feldman MD, Petersen AJ, Karliner LS, et al. Who is responsible for evaluating the safety and effectiveness of medical devices? The role of independent technology assessment. *J Gen Intern Med*. 2008; 23(Suppl 1):57–63. [PubMed: 18095046]
13. Phillips KA, Van Bebber SL. Regulatory perspectives on pharmacogenomics: a review of the literature on key issues faced by the United States Food and Drug Administration. *Med Care Res Rev*. 2006; 63:301–326. [PubMed: 16651395]
14. Fukuda H, Imanaka Y, Ishizaki T, et al. Change in clinical practice after publication of guidelines on breast cancer treatment. *Int J Qual Health Care*. 2009; 21:372–378. [PubMed: 19700780]
15. Heneghan HM, Prichard RS, Devaney A, et al. Evolution of breast cancer management in Ireland: a decade of change. *BMC Surg*. 2009; 9:15. [PubMed: 19765289]
16. O'Malley AS, Pham HH, Reschovsky JD. Predictors of the growing influence of clinical practice guidelines. *J Gen Intern Med*. 2007; 22:742–748. [PubMed: 17387556]
17. Foster JA, Abdolrasulnia M, Doroodchi H, et al. Practice patterns and guideline adherence of medical oncologists in managing patients with early breast cancer. *J Natl Compr Canc Netw*. 2009; 7:697–706. [PubMed: 19635225]

18. Smith TJ, Hillner BE. Ensuring quality cancer care by the use of clinical practice guidelines and critical pathways. *J Clin Oncol.* 2001; 19:2886–2897. [PubMed: 11387362]
19. Ray-Coquard I, Philip T, de Laroche G, et al. Persistence of medical change at implementation of clinical guidelines on medical practice: a controlled study in a cancer network. *J Clin Oncol.* 2005; 23:4414–4423. [PubMed: 15994151]
20. Graham ID, Evans WK, Logan D, et al. Canadian oncologists and clinical practice guidelines: a national survey of attitudes and reported use. *Provincial Lung Disease Site Group of Cancer Care. Ontario Oncology.* 2000; 59:283–290.
21. Pavlidis N. Towards a convenient way to practice medical oncology. *Ann Oncol.* 2007; 18(Suppl 2):iii3–4. [PubMed: 17491034]
22. Schroen AT, Brenin DR. Breast cancer treatment beliefs and influences among surgeons in areas of scientific uncertainty. *Am J Surg.* 2010; 199:491–499. [PubMed: 20359569]
23. Ray-Coquard I, Philip T, de Laroche G, et al. A controlled “before-after” study: impact of a clinical guidelines programme and regional cancer network organization on medical practice. *Br J Cancer.* 2002; 86:313–321. [PubMed: 11875690]
24. Engstrom, PF.; Arnoletti, JP.; Benson, AB., III, et al. [Accessed December 1, 2010] NCCN Clinical Practice Guidelines in Oncology: Colon Cancer. Version 2. 2010. Available at: http://www.nccn.org/professionals/physician_gls/PDF/colon.pdf
25. Shekelle PG, Ortiz E, Rhodes S, et al. Validity of the Agency for Healthcare Research and Quality clinical practice guidelines: how quickly do guidelines become outdated? *JAMA.* 2001; 286:1461–1467. [PubMed: 11572738]
26. Somerfield MR, Einhaus K, Hagerty KL, et al. American Society of Clinical Oncology clinical practice guidelines: opportunities and challenges. *J Clin Oncol.* 2008; 26:4022–4026. [PubMed: 18711193]
27. NCCN Drugs & Biologics Compendium. [Accessed December 1, 2010] Available at: http://www.nccn.org/professionals/drug_compendium/content/contents.asp
28. [Accessed December 1, 2010] American Society of Clinical Oncology Guidelines Procedures Manual Expert Panel Version 3.0. Available at: <http://www.doc-stoc.com/docs/40319747/American-Society-of-Clinical-Oncology-Guideline-Procedures-Manual>
29. National Comprehensive Cancer Network. [Accessed December 1, 2010] About the NCCN Clinical Practice Guidelines in Oncology. Available at: http://www.nccn.org/professionals/physician_gls/about.asp
30. Shankaran V, Wisinski KB, Mulcahy MF, et al. The role of molecular markers in predicting response to therapy in patients with colorectal cancer. *Mol Diagn Ther.* 2008; 12:87–98. [PubMed: 18422373]
31. Walther A, Johnstone E, Swanton C, et al. Genetic prognostic and predictive markers in colorectal cancer. *Nat Rev Cancer.* 2009; 9:489–499. [PubMed: 19536109]
32. Genomic Health. Announces Worldwide Availability of the On-cotype DX(R) Colon Cancer Test [press release]. Redwood City, CA: PRNewswire; Jan 21. 2010
33. Genomic Health Pipeline Colon Cancer. *Colon Cancer.* Genomic Health, Inc; 2010.
34. Andreyev HJ, Norman AR, Cunningham D, et al. Kirsten ras mutations in patients with colorectal cancer: the multicenter “RASCAL” study. *J Natl Cancer Inst.* 1998; 90:675–684. [PubMed: 9586664]
35. Benvenuti S, Sartore-Bianchi A, Di Nicolantonio F, et al. Oncogenic activation of the RAS/RAF signaling pathway impairs the response of metastatic colorectal cancers to anti-epidermal growth factor receptor antibody therapies. *Cancer Res.* 2007; 67:2643–2648. [PubMed: 17363584]
36. Bibeau F, Lopez-Crapez E, Di Fiore F, et al. Impact of Fc{gamma}RIIa-Fc{gamma}RIIIa polymorphisms and KRAS mutations on the clinical outcome of patients with metastatic colorectal cancer treated with cetuximab plus irinotecan. *J Clin Oncol.* 2009; 27:1122–1129. [PubMed: 19164213]
37. Cappuzzo F, Varella-Garcia M, Finocchiaro G, et al. Primary resistance to cetuximab therapy in EGFR FISH-positive colorectal cancer patients. *Br J Cancer.* 2008; 99:83–89. [PubMed: 18577988]

38. De Roock W, Piessevaux H, De Schutter J, et al. KRAS wild-type state predicts survival and is associated to early radiological response in metastatic colorectal cancer treated with cetuximab. *Ann Oncol.* 2008; 19:508–515. [PubMed: 17998284]
39. Di Fiore F, Blanchard F, Charbonnier F, et al. Clinical relevance of KRAS mutation detection in metastatic colorectal cancer treated by Cetuximab plus chemotherapy. *Br J Cancer.* 2007; 96:1166–1169. [PubMed: 17375050]
40. Di Nicolantonio F, Martini M, Molinari F, et al. Wild-type BRAF is required for response to panitumumab or cetuximab in meta-static colorectal cancer. *J Clin Oncol.* 2008; 26:5705–5712. [PubMed: 19001320]
41. Frattini M, Saletti P, Romagnani E, et al. PTEN loss of expression predicts cetuximab efficacy in metastatic colorectal cancer patients. *Br J Cancer.* 2007; 97:1139–1145. [PubMed: 17940504]
42. Freeman DJ, Juan T, Reiner M, et al. Association of K-ras mutational status and clinical outcomes in patients with metastatic colorectal cancer receiving panitumumab alone. *Clin Colorectal Cancer.* 2008; 7:184–190. [PubMed: 18621636]
43. Garm Spindler KL, Pallisgaard N, Rasmussen AA, et al. The importance of KRAS mutations and EGF61A>G polymorphism to the effect of cetuximab and irinotecan in metastatic colorectal cancer. *Ann Oncol.* 2009; 20:879–884. [PubMed: 19179548]
44. Goncalves A, Esteyries S, Taylor-Smedra B, et al. A polymorphism of EGFR extracellular domain is associated with progression free-survival in metastatic colorectal cancer patients receiving cetuximab-based treatment. *BMC Cancer.* 2008; 8:169. [PubMed: 18544172]
45. Khambata-Ford S, Garrett CR, Meropol NJ, et al. Expression of epiregulin and amphiregulin and K-ras mutation status predict disease control in metastatic colorectal cancer patients treated with cetuximab. *J Clin Oncol.* 2007; 25:3230–3237. [PubMed: 17664471]
46. Laurent-Puig P, Cayre A, Manceau G, et al. Analysis of PTEN, BRAF, and EGFR status in determining benefit from cetuximab therapy in wild-type KRAS metastatic colon cancer. *J Clin Oncol.* 2009; 27:5924–5930. [PubMed: 19884556]
47. Lievre A, Bachet JB, Boige V, et al. KRAS mutations as an independent prognostic factor in patients with advanced colorectal cancer treated with cetuximab. *J Clin Oncol.* 2008; 26:374–379. [PubMed: 18202412]
48. Lievre A, Bachet JB, Le Corre D, et al. KRAS mutation status is predictive of response to cetuximab therapy in colorectal cancer. *Cancer Res.* 2006; 66:3992–3995. [PubMed: 16618717]
49. Loupakis F, Pollina L, Stasi I, et al. PTEN expression and KRAS mutations on primary tumors and metastases in the prediction of benefit from cetuximab plus irinotecan for patients with meta-static colorectal cancer. *J Clin Oncol.* 2009; 27:2622–2629. [PubMed: 19398573]
50. Loupakis F, Ruzzo A, Cremolini C, et al. KRAS codon 61, 146 and BRAF mutations predict resistance to cetuximab plus irinotecan in KRAS codon 12 and 13 wild-type metastatic colorectal cancer. *Br J Cancer.* 2009; 101:715–721. [PubMed: 19603018]
51. Lurje G, Nagashima F, Zhang W, et al. Polymorphisms in cyclo-oxygenase-2 and epidermal growth factor receptor are associated with progression-free survival independent of K-ras in metastatic colorectal cancer patients treated with single-agent cetuximab. *Clin Cancer Res.* 2008; 14:7884–7895. [PubMed: 19047118]
52. Molinari F, Martin V, Saletti P, et al. Differing deregulation of EGFR and downstream proteins in primary colorectal cancer and related metastatic sites may be clinically relevant. *Br J Cancer.* 2009; 100:1087–1094. [PubMed: 19293803]
53. Moroni M, Veronese S, Benvenuti S, et al. Gene copy number for epidermal growth factor receptor (EGFR) and clinical response to antiEGFR treatment in colorectal cancer: a cohort study. *Lancet Oncol.* 2005; 6:279–286. [PubMed: 15863375]
54. Oden-Gangloff A, Di Fiore F, Bibeau F, et al. TP53 mutations predict disease control in metastatic colorectal cancer treated with cetuximab-based chemotherapy. *Br J Cancer.* 2009; 100:1330–1335. [PubMed: 19367287]
55. Perrone F, Lampis A, Orsenigo M, et al. PI3KCA/PTEN deregulation contributes to impaired responses to cetuximab in metastatic colorectal cancer patients. *Ann Oncol.* 2009; 20:84–90. [PubMed: 18669866]

56. Personeni N, Fieuws S, Piessevaux H, et al. Clinical usefulness of EGFR gene copy number as a predictive marker in colorectal cancer patients treated with cetuximab: a fluorescent in situ hybridization study. *Clin Cancer Res.* 2008; 14:5869–5876. [PubMed: 18794099]
57. Prenen H, De Schutter J, Jacobs B, et al. PIK3CA mutations are not a major determinant of resistance to the epidermal growth factor receptor inhibitor cetuximab in metastatic colorectal cancer. *Clin Cancer Res.* 2009; 15:3184–3188. [PubMed: 19366826]
58. Sartore-Bianchi A, Martini M, Molinari F, et al. PIK3CA mutations in colorectal cancer are associated with clinical resistance to EGFR-targeted monoclonal antibodies. *Cancer Res.* 2009; 69:1851–1857. [PubMed: 19223544]
59. Sohn BS, Kim TW, Lee JL, et al. The role of KRAS mutations in predicting the efficacy of cetuximab-plus-irinotecan therapy in irinotecan-refractory Korean metastatic colorectal cancer patients. *Oncology.* 2009; 77:224–230. [PubMed: 19738388]
60. Souglakos J, Philips J, Wang R, et al. Prognostic and predictive value of common mutations for treatment response and survival in patients with metastatic colorectal cancer. *Br J Cancer.* 2009; 101:465–472. [PubMed: 19603024]
61. Yen LC, Uen YH, Wu DC, et al. Activating KRAS mutations and overexpression of epidermal growth factor receptor as independent predictors in metastatic colorectal cancer patients treated with cetuximab. *Ann Surg.* 2010; 251:254–260. [PubMed: 20010090]
62. Amado RG, Wolf M, Peeters M, et al. Wild-type KRAS is required for panitumumab efficacy in patients with metastatic colorectal cancer. *J Clin Oncol.* 2008; 26:1626–1634. [PubMed: 18316791]
63. Bokemeyer C, Bondarenko I, Makhson A, et al. Fluorouracil, leucovorin, and oxaliplatin with and without cetuximab in the first-line treatment of metastatic colorectal cancer. *J Clin Oncol.* 2009; 27:663–671. [PubMed: 19114683]
64. Hecht JR, Mitchell E, Chidiac T, et al. A randomized phase IIIB trial of chemotherapy, bevacizumab, and panitumumab compared with chemotherapy and bevacizumab alone for metastatic colorectal cancer. *J Clin Oncol.* 2009; 27:672–680. [PubMed: 19114685]
65. Karapetis CS, Khambata-Ford S, Jonker DJ, et al. K-ras mutations and benefit from cetuximab in advanced colorectal cancer. *N Engl J Med.* 2008; 359:1757–1765. [PubMed: 18946061]
66. Tol J, Koopman M, Cats A, et al. Chemotherapy, bevacizumab, and cetuximab in metastatic colorectal cancer. *N Engl J Med.* 2009; 360:563–572. [PubMed: 19196673]
67. Van Cutsem E, Kohne CH, Hitre E, et al. Cetuximab and chemotherapy as initial treatment for metastatic colorectal cancer. *N Engl J Med.* 2009; 360:1408–1417. [PubMed: 19339720]
68. Roth AD, Tejpar S, Delorenzi M, et al. Prognostic role of KRAS and BRAF in stage II and III resected colon cancer: results of the translational study on the PETACC-3, EORTC 40993, SAKK 60-00 trial. *J Clin Oncol.* 2010; 28:466–474. [PubMed: 20008640]
69. Nash GM, Gimbel M, Cohen AM, et al. KRAS mutation and microsatellite instability: two genetic markers of early tumor development that influence the prognosis of colorectal cancer. *Ann Surg Oncol.* 2010; 17:416–424. [PubMed: 19813061]
70. Nash GM, Gimbel M, Shia J, et al. KRAS mutation correlates with accelerated metastatic progression in patients with colorectal liver metastases. *Ann Surg Oncol.* 2010; 17:572–578. [PubMed: 19727962]
71. Jonker, D. The influence of K-RAS exon 2 mutations on outcomes in a randomized phase III trial of cetuximab + best supportive care (BSC) versus BSC alone in patients with pre-treated metastatic EGFR-positive colorectal cancer (NCIC CTG CO.17). Presented at the 10th World Congress on Gastrointestinal Cancer; June 25–28, 2008; Barcelona, Spain.
72. van Cutsem E, Lang I, D’Haens G, et al. KRAS status and efficacy in the first-line treatment of patients with metastatic colorectal cancer treated with FOLFIRI with or without cetuximab: the CRYSTAL experience [abstract]. *J Clin Oncol.* 2008; 26(Suppl 1):Abstract 2.
73. Bokemeyer C, Bondarenko I, Hartmann JT, et al. KRAS status and efficacy of first-line treatment of patients with metastatic colorectal cancer (mCRC) with FOLFOX with or without cetuximab: the OPUS experience [abstract]. *J Clin Oncol.* 2008; 26(Suppl 1):Abstract 4000.
74. Punt C, Tol J, Rodenburg CJ, et al. Randomized phase III study of capecitabine, oxaliplatin, and bevacizumab with or without cetuximab in advanced colorectal cancer (ACC), the CAIRO2 study

- of the Dutch Colorectal Cancer Group (DCCG) [abstract]. *J Clin Oncol*. 2008; 26(Suppl 1):Abstract LBA4011.
75. Tol J, Nagtegaal ID, Punt CJ. BRAF mutation in metastatic colorectal cancer. *N Engl J Med*. 2009; 361:98–99. [PubMed: 19571295]
 76. Van Cutsem, ELI.; Folprecht, G.; Nowacki, M., et al. Cetuximab plus FOLFIRI in the treatment of metastatic colorectal cancer (mCRC): the influence of KRAS and BRAF biomarkers on outcome: updated data from the CRYSTAL trial. Presented at the 2010 ASCO Gastrointestinal Cancers Symposium; January 22–24, 2010; Orlando, Florida.
 77. Boland CR, Thibodeau SN, Hamilton SR, et al. A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. *Cancer Res*. 1998; 58:5248–5257. [PubMed: 9823339]
 78. Hampel H, Frankel WL, Martin E, et al. Feasibility of screening for Lynch syndrome among patients with colorectal cancer. *J Clin Oncol*. 2008; 26:5783–5788. [PubMed: 18809606]
 79. Popat S, Hubner R, Houlston RS. Systematic review of microsatellite instability and colorectal cancer prognosis. *J Clin Oncol*. 2005; 23:609–618. [PubMed: 15659508]
 80. Umar A, Boland CR, Terdiman JP, et al. Revised Bethesda Guidelines for hereditary nonpolyposis colorectal cancer (Lynch syndrome) and microsatellite instability. *J Natl Cancer Inst*. 2004; 96:261–268. [PubMed: 14970275]
 81. Hemminki A, Mecklin JP, Jarvinen H, et al. Microsatellite instability is a favorable prognostic indicator in patients with colorectal cancer receiving chemotherapy. *Gastroenterology*. 2000; 119:921–928. [PubMed: 11040179]
 82. Kumar S, Chang EY, Frankhouse J, et al. Combination of microsatellite instability and lymphocytic infiltrate as a prognostic indicator for adjuvant therapy in colon cancer. *Arch Surg*. 2009; 144:835–840. [PubMed: 19797108]
 83. Elsaleh H, Iacopetta B. Microsatellite instability is a predictive marker for survival benefit from adjuvant chemotherapy in a population-based series of stage III colorectal carcinoma. *Clin Colorectal Cancer*. 2001; 1:104–109. [PubMed: 12445368]
 84. Charara M, Edmonston TB, Burkholder S, et al. Microsatellite status and cell cycle associated markers in rectal cancer patients undergoing a combined regimen of 5-FU and CPT-11 chemotherapy and radiotherapy. *Anticancer Res*. 2004; 24:3161–3167. [PubMed: 15510606]
 85. Benatti P, Gafa R, Barana D, et al. Microsatellite instability and colorectal cancer prognosis. *Clin Cancer Res*. 2005; 11:8332–8340. [PubMed: 16322293]
 86. Carethers JM, Smith EJ, Behling CA, et al. Use of 5-fluorouracil and survival in patients with microsatellite-unstable colorectal cancer. *Gastroenterology*. 2004; 126:394–401. [PubMed: 14762775]
 87. Colombino M, Cossu A, Manca A, et al. Prevalence and prognostic role of microsatellite instability in patients with rectal carcinoma. *Ann Oncol*. 2002; 13:1447–1453. [PubMed: 12196371]
 88. de Vos tot Nederveen Cappel WH, Meulenbeld HJ, Kleibeuker JH, et al. Survival after adjuvant 5-FU treatment for stage III colon cancer in hereditary nonpolyposis colorectal cancer. *Int J Cancer*. 2004; 109:468–471. [PubMed: 14961589]
 89. Jensen SA, Vainer B, Kruhoffer M, et al. Microsatellite instability in colorectal cancer and association with thymidylate synthase and dihydropyrimidine dehydrogenase expression. *BMC Cancer*. 2009; 9:25. [PubMed: 19154585]
 90. Jover R, Zapater P, Castells A, et al. The efficacy of adjuvant chemotherapy with 5-fluorouracil in colorectal cancer depends on the mismatch repair status. *Eur J Cancer*. 2009; 45:365–373. [PubMed: 18722765]
 91. Lanza G, Gafa R, Santini A, et al. Immunohistochemical test for MLH1 and MSH2 expression predicts clinical outcome in stage II and III colorectal cancer patients. *J Clin Oncol*. 2006; 24:2359–2367. [PubMed: 16710035]
 92. Liang JT, Huang KC, Cheng AL, et al. Clinicopathological and molecular biological features of colorectal cancer in patients less than 40 years of age. *Br J Surg*. 2003; 90:205–214. [PubMed: 12555297]

93. Nehls O, Okech T, Hsieh CJ, et al. Studies on p53, BAX and Bcl-2 protein expression and microsatellite instability in stage III (UICC) colon cancer treated by adjuvant chemotherapy: major prognostic impact of proapoptotic BAX. *Br J Cancer*. 2007; 96:1409–1418. [PubMed: 17426704]
94. Bertagnolli MM, Niedzwiecki D, Compton CC, et al. Microsatellite instability predicts improved response to adjuvant therapy with irinotecan, fluorouracil, and leucovorin in stage III colon cancer: Cancer and Leukemia Group B Protocol 89803. *J Clin Oncol*. 2009; 27:1814–1821. [PubMed: 19273709]
95. French AJ, Sargent DJ, Burgart LJ, et al. Prognostic significance of defective mismatch repair and BRAF V600E in patients with colon cancer. *Clin Cancer Res*. 2008; 14:3408–3415. [PubMed: 18519771]
96. Halling KC, French AJ, McDonnell SK, et al. Microsatellite instability and 8p allelic imbalance in stage B2 and C colorectal cancers. *J Natl Cancer Inst*. 1999; 91:1295–1303. [PubMed: 10433618]
97. Kim GP, Colangelo LH, Wieand HS, et al. Prognostic and predictive roles of high-degree microsatellite instability in colon cancer: a National Cancer Institute-National Surgical Adjuvant Breast and Bowel Project Collaborative Study. *J Clin Oncol*. 2007; 25:767–772. [PubMed: 17228023]
98. Ribic CM, Sargent DJ, Moore MJ, et al. Tumor microsatellite-instability status as a predictor of benefit from fluorouracil-based adjuvant chemotherapy for colon cancer. *N Engl J Med*. 2003; 349:247–257. [PubMed: 12867608]
99. Sinicrope FA, Rego RL, Halling KC, et al. Prognostic impact of microsatellite instability and DNA ploidy in human colon carcinoma patients. *Gastroenterology*. 2006; 131:729–737. [PubMed: 16952542]
100. Watanabe T, Wu TT, Catalano PJ, et al. Molecular predictors of survival after adjuvant chemotherapy for colon cancer. *N Engl J Med*. 2001; 344:1196–1206. [PubMed: 11309634]
101. Westra JL, Schaapveld M, Hollema H, et al. Determination of TP53 mutation is more relevant than microsatellite instability status for the prediction of disease-free survival in adjuvant-treated stage III colon cancer patients. *J Clin Oncol*. 2005; 23:5635–5643. [PubMed: 16110022]
102. Sargent DJ, Marsoni S, Thibodeau SN, et al. Confirmation of deficient mismatch repair (dMMR) as a predictive marker for lack of benefit from 5-FU based chemotherapy in stage II and III colon cancer (CC): a pooled molecular reanalysis of randomized chemotherapy trials [abstract]. *J Clin Oncol*. 2008; 26(Suppl 1):Abstract 4008.
103. Sargent DJ, Marsoni S, Monges G, et al. Defective mismatch repair as a predictive marker for lack of efficacy of fluorouracil-based adjuvant therapy in colon cancer. *J Clin Oncol*. 2010; 28:3219–3226. [PubMed: 20498393]
104. Kerr D, Gray R, Quirke P, et al. A quantitative multigene RT-PCR assay for prediction of recurrence in stage II colon cancer: selection of the genes in four large studies and results of the independent, prospectively designed QUASAR validation study [abstract]. *J Clin Oncol*. 2009; 27(Suppl 1):Abstract 400.
105. Fazio VW, Tekkis PP, Remzi F, et al. Assessment of operative risk in colorectal cancer surgery: the Cleveland Clinic Foundation colorectal cancer model. *Dis Colon Rectum*. 2004; 47:2015–2024. [PubMed: 15657649]
106. Lavery, I.; Hammel, J.; Cowens, J., et al. Relationship between tumor gene expression and recurrence in an observational cohort of patients with stage II/III colon cancer treated with surgery only: quantitative RT-PCR assay of 375 genes in fixed paraffin-embedded (FPE) tissue. Presented at the 2008 ASCO Gastrointestinal Cancers Symposium; January 25–27, 2008; Orlando, Florida. p. Abstract 302
107. O’Connell MJ, Paik S, Yothers G, et al. Relationship between tumor gene expression and recurrence in stage II/III colon cancer: quantitative RT-PCR assay of 757 genes in fixed paraffin-embedded (FPE) tissue [abstract]. *J Clin Oncol*. 2006; 24(Suppl 1):Abstract 3518.
108. O’Connell, MJ.; Yothers, G.; Paik, S., et al. Relationship between tumor gene expression and recurrence in patients with stage II/III colon cancer treated with surgery + 5FU/LV in NSABP C-06: consistency of results with other independent studies. Presented at the 2008 ASCO Gastrointestinal Cancers Symposium; January 25–27, 2008; Orlando, Florida.

109. Gray R, Barnwell J, et al. Quasar Collaborative G. Adjuvant chemotherapy versus observation in patients with colorectal cancer: a randomised study. *Lancet*. 2007; 370:2020–2029. [PubMed: 18083404]
110. National Comprehensive Cancer Network. [Accessed December 1, 2010] NCCN Disclosure Policies and Potential Conflicts of Interest. Available at: <http://www.nccn.org/about/disclosure.asp>
111. Pentheroudakis G, Stahel R, Hansen H, et al. Heterogeneity in cancer guidelines: should we eradicate or tolerate? *Ann Oncol*. 2008; 19:2067–2078. [PubMed: 18662954]
112. Lamberti C, Lundin S, Bogdanow M, et al. Microsatellite instability did not predict individual survival of unselected patients with colorectal cancer. *Int J Colorectal Dis*. 2007; 22:145–152. [PubMed: 16724208]
113. Lim SB, Jeong SY, Lee MR, et al. Prognostic significance of microsatellite instability in sporadic colorectal cancer. *Int J Colorectal Dis*. 2004; 19:533–537. [PubMed: 15175889]
114. Zauber NP, Marotta SP, Berman E, et al. Molecular genetic changes associated with colorectal carcinogenesis are not prognostic for tumor regression following preoperative chemoradiation of rectal carcinoma. *Int J Radiat Oncol Biol Phys*. 2009; 74:472–476. [PubMed: 19304403]
115. Lanza G, Ferracin M, Gafa R, et al. mRNA/microRNA gene expression profile in microsatellite unstable colorectal cancer. *Mol Cancer*. 2007; 6:54. [PubMed: 17716371]
116. Lindor NM, Burgart LJ, Leontovich O, et al. Immunohistochemistry versus microsatellite instability testing in phenotyping colorectal tumors. *J Clin Oncol*. 2002; 20:1043–1048. [PubMed: 11844828]

Biography

Robin K. Kelley, MD is a member of the NCCN Hepatobiliary Cancers Panel and has received an honorarium from Genomic Health, Inc. for participating in a speaker training program. Alan P. Venook, MD is a member of the NCCN Panels for Colon/Rectal/Anal Cancers and Hepatobiliary Cancers and the NCCN Board of Directors. Dr. Venook has held an advisory role to and received honoraria from Genomic Health, Inc.

Table 1

NCCN Guidelines Recommendations for Selected PM Biomarker Testing

Biomarker Test	Recommended in NCCN Guidelines?	Date Included in Guidelines	Guideline Recommendation	Category of Evidence
<i>K-ras</i> codon 12 and 13 mutation analysis	Yes	Fall 2008	Predictive marker for nonresponse to EGFR-targeted therapy in metastatic disease	2A
<i>B-raf</i> V600E mutation analysis	Yes	January 2010	Predictive marker for nonresponse to EGFR-targeted therapy in metastatic disease	2A
MMR testing by IHC or PCR	Yes	January 2010	Predictive marker for lack of benefit from 5-FU in stage II colon cancer patients	2A
Oncotype DX Colon Cancer Test	No*	N/A	Not recommended	"Insufficient data"

Abbreviations: 5-FU, 5-fluorouracil; EGFR, epidermal growth factor receptor.

*The Oncotype DX Colon Cancer Assay is not named explicitly in the NCCN Clinical Practice Guidelines in Oncology for Colon Cancer. The guidelines state that, "There are insufficient data to recommend the use of multi-gene assay panels to determine adjuvant therapy."²⁴

Table 2
K-ras Codon 12 or 13 Mutation Analysis in Metastatic Colorectal Cancer Patients Treated With Cetuximab or Panitumumab

Randomized, Controlled Trial Subset Analyses	Mutant/N	End Point(s)	Single-Arm Studies	Mutant/N	End Point(s)
Amado et al. ⁶²	184/427	OS, PFS, R	Benvenuti et al. ³⁵	16/48	R, TTP
Bokemeyer et al. ⁶³	99/233	PFS, R	Bibeau et al. ³⁶	27/64	PFS, R
Hecht et al. ⁶⁴	346/865	OS, PFS, R	Capuzzo et al. ³⁷	42/80	R
Karapetis et al. ⁶⁵	167/394	OS, PFS, R	De Roock et al. ³⁸	42/108	OS, PFS, R
Tol et al. ⁶⁶	206/528	OS, PFS, R	Di Fiore et al. ³⁹	22/59	R, TTP
Van Cutsem et al. ⁶⁷	192/540	OS, PFS, R	Di Nicolantonio et al. ⁴⁰	34/113	OS, PFS, R
			Frattini et al. ⁴¹	10/27	R
			Freeman et al. ⁴²	24/62	OS, PFS, R
			Garm Spindler et al. ⁴³	22/64	OS, PFS, R
			Goncalves et al. ⁴⁴	14/32	R
			Khambata-Ford et al. ⁴⁵	30/80	DCR, PFS
			Laurent-Puig et al. ⁴⁶	53/169	OS, PFS, R
			Lievre et al. ⁴⁷	24/89	OS, PFS, R
			Lievre et al. ⁴⁸	13/30	OS, R
			Loupakis et al. ^{50,*}	8/76	OS, PFS, R [*]
			Loupakis et al. ⁴⁹	35/88	OS, PFS, R
			Lurje et al. ⁵¹	42/130	OS, PFS, R
			Molinari et al. ⁵²	16/37	R
			Moroni et al. ⁵³	10/31	R
			Oden-Gangloff et al. ⁵⁴	18/64	CD, TTP
			Perrone et al. ⁵⁵	7/29	R
			Personeni et al. ⁵⁶	29/87	OS, PFS
			Prenen et al. ⁵⁷	77/199	R
			Sartore-Bianchi et al. ⁵⁸	32/110	PFS, R
			Sohn et al. ⁵⁹	27/66	OS, PFS, R
			Souglakos et al. ⁶⁰	62/168	PFS, R
			Yen et al. ⁶¹	41/95	OS, PFS, R

All studies were retrospective.

Abbreviations: CD, controlled disease; DCR, disease control rate; N, number of patients/specimens in the subset that underwent *K-ras* biomarker testing; OS, overall survival; PFS, progression-free survival; R, any response-based end point (including best response, response rate, overall response rate, clinical response, and objective response); TTP, time to progression.

* Evaluated *K-ras* codon 61 and 146 mutations.

Table 3
B-raf V600E Mutation Analysis in Metastatic Colorectal Cancer Patients Treated With Cetuximab or Panitumumab

Randomized, Controlled Trial	Subset Analyses	Mutant/N	End Point(s)	Single-Arm Studies	Mutant/N	End Point(s)
Tol et al. ⁷⁵		45/516	OS, PFS, R	Benvenuti et al. ³⁵	6/48	R, TTP
Van Cutsem et al. ^{76, *} †		59/625	OS, PFS, R	Cappuzzo et al. ³⁷	4/79	OS, R, TTP
				Di Nicolantonio et al. ⁴⁰	11/113	OS, PFS, R
				Freeman et al. ⁴²	4/62	OS, PFS, R
				Laurent-Puig et al. ⁴⁶	5/171	OS, PFS, R
				Lievre et al. ⁴⁸	0/30	OS, R
				Loupakis et al. ^{50, *}	13/87	OS, PFS, R
				Molinari et al. ⁵²	2/36	R
				Moroni et al. ^{53, ‡}	1/31	R [‡]
				Perrone 2009 ⁵⁵	3/31	R
				Sohn et al. ⁵⁹	0/66	OS, PFS, R
				Souglakos et al. ⁶⁰	13/168	PFS, R

All studies were retrospective.

Abbreviations: DCR, disease control rate; N, number of patients/specimens in the subset that underwent *B-raf*/biomarker testing; OS, overall survival; PFS, progression free survival; R, any response-based endpoint (including best response, response rate, overall response rate, clinical response, and objective response); TTP, time to progression.

* Enriched for *K-ras* wild-type patients only.

† Preliminary results presented at national conference, included because referenced in NCCN Clinical Practice Guidelines in Oncology.

‡ Mutation identified was E599V in exon 15 of *B-raf* gene.

Table 4
MMR Deficiency Testing in Locoregional Colon Cancer Patients Treated With 5-Fluorouracil–Based Chemotherapy

Randomized, Controlled Trial Subset Analyses	Deficient [*] /N	End Point(s)	Non-Randomized and Single-Arm Studies	Deficient [*] /N	End Point(s)
Bertagnoli et al. ⁹⁴	96/702	DFS, OS	Benatti et al. ⁸⁵	256/1263	DFS
French et al. ⁹⁵	57/533	DFS, OS	Carethers et al. ⁸⁶	36/204	OS
Halling et al. ⁹⁶	76/508	DFS, TTR	Charara et al. ⁸⁴	5/57	R
Kim et al. ⁹⁷	98/542	DFS, OS	Colombino et al. ⁸⁷	17/91	DFS, OS
Ribic et al. ⁹⁸	95/570	OS	de Vos tot Nederveen Cappel et al. ^{88,†}	92/92 [‡]	OS [‡]
Sargent et al. ^{102,§}	47/341 [§]	DFS, OS [§]	Elsaleh et al. ⁸³	63/721	DFS, OS
Simicrope et al. ⁹⁹	95/528	DFS, OS	Hemminki et al. ⁸¹	11/95	DFS
Wantanabe et al. ¹⁰⁰	62/298	DFS, OS	Jensen et al. ⁸⁹	43/311	DFS, OS
Westra et al. ¹⁰¹	44/273	DFS	Jover et al. ⁹⁰	76/754	DFS, OS
			Kumar et al. ⁸²	30/149	DFS, OS
			Lamberti et al. ¹¹²	52/416	DFS, OS
			Lanza et al. ⁹¹	114/718	DFS
			Liang et al. ^{92,‡}	37/126 [‡]	OS [‡]
			Lim et al. ¹¹³	23/248	OS
			Nehls et al. ⁹³	16/174	DFS, OS
			Zauber et al. ¹¹⁴	2/51	R

All studies were retrospective.

Abbreviations: DFS, disease-free survival, disease-specific survival, or recurrence-free survival; N, number of patients/specimens in the subset that underwent MMR biomarker testing; OS, overall survival; R, any response-based end point (including best response, response rate, overall response rate, clinical response, and objective response); TTR, time to recurrence.

^{*} Deficient was defined by standard immunohistochemistry criteria or by polymerase chain reaction testing showing high microsatellite instability.^{80,115,116}

[†] Enriched for patients with hereditary nonpolyposis colorectal cancer.

[‡] Enriched for patients aged < 40 years.

[§] Preliminary results presented at national conference, included because referenced in NCCN Clinical Practice Guidelines in Oncology.

Table 5
Clinical Studies of *OncoType* DX Colon Cancer Assay in Stage II Colon Cancer

Randomized, Controlled Trial Subset Analyses	Recurrence Score (N) High/Intermediate/Low	End Point(s)	Single-Arm Studies	Recurrence score High/Intermediate/Low	End Point(s)
Kerr 2009 ^{104,*}	182/218/311	DFS, OS	None published to date		

This study was retrospective.

Abbreviations: DFS, disease-free survival; N, number of patients/specimens in the subset that underwent *OncoType* DX colon cancer testing; OS, overall survival.

* Preliminary results presented at national conference.

Table 6

Summary of Search Results

Biomarker	Approximate Incidence	Number of Randomized, Controlled Trial Subset Analyses	Number of Nonrandomized and Single-Arm Studies	Change in Outcome?
<i>K-ras</i> Codon 12 or 13 Mutation	30%–40%	6	27	Highly consistent evidence for predictive value; mixed evidence for prognostic value
<i>B-raf</i> V600E mutation	5%–10%	2	12	Limited, inconsistent evidence for predictive value; may be prognostic
MMR-deficient or MSI-high	15%–20%	9	16	Highly consistent evidence for prognostic value; inconsistent evidence for predictive value
Onco type DX Colon Cancer	26% high risk	1	0	Evidence for prognostic value in one RCT subset; not predictive
Assay Recurrence Score	31% intermediate risk 44% low risk			

Abbreviations: MMR, mismatch repair protein; MSI, microsatellite instability; RCT, randomized controlled trial.