

# Pharmacokinetics of First-Line Tuberculosis Drugs in Tanzanian Patients

## Alma Tostmann,<sup>a,b</sup>\* Charles M. Mtabho,<sup>c</sup> Hadija H. Semvua,<sup>c</sup> Jossy van den Boogaard,<sup>b,c</sup>\* Gibson S. Kibiki,<sup>c</sup> Martin J. Boeree,<sup>a,b</sup> Rob E. Aarnoutse<sup>d</sup>

Department of Respiratory Diseases, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands<sup>a</sup>; University Centre for Chronic Diseases Dekkerswald, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands<sup>b</sup>; Kilimanjaro Clinical Research Institute, Kilimanjaro Christian Medical Centre, Moshi, Tanzania<sup>c</sup>; Department of Pharmacy, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands<sup>d</sup>

East Africa has a high tuberculosis (TB) incidence and mortality, yet there are very limited data on exposure to TB drugs in patients from this region. We therefore determined the pharmacokinetic characteristics of first-line TB drugs in Tanzanian patients using intensive pharmacokinetic sampling. In 20 adult TB patients, plasma concentrations were determined just before and at 1, 2, 3, 4, 6, 8, 10, and 24 h after observed drug intake with food to estimate the areas under the curve from 0 to 24 h (AUC<sub>0-24</sub>) and peak plasma concentrations ( $C_{max}$ ) of isoniazid, rifampin, pyrazinamide, and ethambutol. Acetylator status for isoniazid was assessed phenotypically using the isoniazid elimination half-life and the acetylisoniazid/isoniazid metabolic ratio at 3 h postdose. The geometric mean AUC<sub>0-24</sub>s were as follows: isoniazid, 11.0 h  $\cdot$  mg/liter; rifampin, 39.9 h  $\cdot$  mg/liter; pyrazinamide, 344 h  $\cdot$  mg/liter; and ethambutol, 20.2 h  $\cdot$  mg/liter. The  $C_{max}$  was below the reference range for isoniazid in 10/19 patients and for rifampin in 7/20 patients. In none of the patients were the  $C_{max}$  for pyrazinamide and ethambutol below the reference range. Elimination half-life and metabolic ratio of isoniazid gave discordant phenotyping results in only 2/19 patients. A substantial proportion of patients had an isoniazid and/or rifampin  $C_{max}$  below the reference range. The drugs with food may partly explain these low drug levels, but such a drug intake reflects common practice. The finding of low TB drug concentrations is concerning because low concentrations have been associated with worse treatment outcome in several other studies.

he pharmacokinetic properties of tuberculosis (TB) drugs are well described, especially in Caucasian populations (1, 2). Pharmacokinetic data from East Africa are limited, especially data that are based on intensive pharmacokinetic sampling during the full dosing interval. Such intensive pharmacokinetic sampling enables an accurate assessment of the total exposure to TB drugs (area under the time-versus-plasma concentration curve from 0 to 24 h [AUC<sub>0-24</sub>]) and the peak plasma concentration ( $C_{max}$ ) in individual patients. Large interpatient variability in these key pharmacokinetic parameters of TB drugs generally exists (1). As a result, a proportion of patients achieve drug concentrations that are below the reference range for TB drugs. Several studies have shown that such lower exposures to TB drugs are associated with a suboptimal response (3-8) A recent study showed that pharmacokinetic variability to just a single drug in the multidrug TB drug regimen is associated with treatment failure and acquired drug resistance (9, 10).

On the other hand, unduly high exposures may cause toxicity and interruption of TB treatment. It is therefore important to have more knowledge on the pharmacokinetic properties of TB treatment in populations from East Africa, as a high TB incidence and a high TB mortality are found in this region (11). HIV infection (12–14) and malnutrition (15) have been associated with decreased plasma concentrations of TB drugs. Both are highly prevalent among East African TB patients, and therefore, low TB drug levels are to be expected in this population (11).

The objective of this study was to describe the pharmacokinetic parameters of isoniazid, rifampin, pyrazinamide, and ethambutol using intensive sampling during the full dosing interval in Tanzanian TB patients.

## MATERIALS AND METHODS

**Study design.** We conducted an observational pharmacokinetic study at the Kilimanjaro Clinical Research Institute at the Kilimanjaro Christian Medical Centre (KCMC) in Moshi, Tanzania, using the standard two-stage approach. With this approach, individual pharmacokinetic parameters are estimated in the first stage. In the second stage, population characteristics of each parameter are derived by obtaining measures of central tendency and spread of all the subjects' individual parameters.

**Study participants.** Study participants were recruited from an outpatient tuberculosis treatment clinic at Mawenzi Hospital in Moshi. Adult TB patients who were in the intensive phase of treatment and who were not using medication that could interfere with TB drug plasma concentrations were eligible for participation. All participants gave written informed consent. The study was approved by the local Institutional Research Board at KCMC and by the Tanzanian National Institute of Medical Research.

Tuberculosis was treated with fixed-dose combination tablets (FDC tablets) that were manufactured by Sandoz, Mumbai, India, and are on the WHO list of prequalified medicinal products. The drugs were donated by Novartis through the WHO Global Drug Facility (GDF), which provides only high-quality drugs that meet stringent WHO standards. Pa-

Received 24 December 2012 Returned for modification 3 February 2013 Accepted 21 April 2013

Published ahead of print 29 April 2013

Address correspondence to Alma Tostmann, A.Tostmann@elg.umcn.nl. \* Present address: Alma Tostmann, Dept of Primary and Community Care Medicine, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands; Jossy van den Boogaard, BELACD de Pala, Pala, Chad.

Copyright © 2013, American Society for Microbiology. All Rights Reserved. doi:10.1128/AAC.02599-12

tients with a body weight of less than 50 kg use three FDC tablets per day (i.e., 225 mg isoniazid, 450 mg rifampin, 1,200 mg pyrazinamide, and 875 mg ethambutol), and patients with a body weight of >50 kg use four FDC tablets per day (i.e., 300 mg isoniazid, 600 mg rifampin, 1,600 mg pyrazinamide, and 1,100 mg ethambutol) (16).

**Data collection.** Basic demographic and clinical information was collected from all participants, including age, sex, body weight and height (to calculate body mass index [BMI]), comorbidities (including HIV infection and hepatitis B and C), and concomitant drug use. Malnutrition was defined as a BMI of <18.5 kg/m<sup>2</sup> (17).

**Sample collection.** Pharmacokinetic sampling took place at KCMC hospital. Patients had to be on TB treatment for at least 10 days because of the expected steady state in the pharmacokinetics of the TB drugs at that point. Patients had refrained from food consumption for at least 8 h before drug intake. On the sampling day, patients took their drugs under our supervision at 8 a.m. They then had a standardized breakfast within 30 min after drug intake, which reflected the usual drug intake procedures in this population. The standardized breakfast consisted of a cup (125 ml) of tea with whole milk and sugar and either a small bowl of porridge or mandaazi, a typical East African fried pastry (analogous to a doughnut) that is rather high in fat.

Serial venous blood samples were collected just before, and at 1, 2, 3, 4, 6, 8, 10, and 24 h after observed TB drug intake. Plasma was separated and stored at  $-80^{\circ}$ C immediately until transport on dry ice to The Netherlands for bioanalysis.

**Bioanalysis.** The plasma concentrations of isoniazid, rifampin, pyrazinamide, and ethambutol were assessed by validated high-performance liquid chromatography (HPLC) as described before (18). Isoniazid and acetylisoniazid (acetyl-INH) were measured with liquid-liquid extraction followed by ultra-performance liquid chromatography (UPLC) with UV detection. Accuracy was between 97.8% and 106.7% for isoniazid and between 98.0% and 108.9% for acetylisoniazid, depending on the concentration level. The intra- and interassay coefficients of variation were less than 13.4% and less than 3.2% (depending on the concentration), respectively, over the range of 0.05 to 15.1 mg/liter for isoniazid and less than 4.2% and less than 5.7%, respectively, over the range of 0.16 to 16.2 mg/ liter for acetylisoniazid. Lower limits of quantification were 0.05 mg/liter for isoniazid and 0.16 mg/liter for acetylisoniazid. Isoniazid- and acetylisoniazid-containing samples were stable (<5% loss) for at least 12 months at  $-80^{\circ}$ C.

Pharmacokinetic analysis. Pharmacokinetic evaluations were performed using noncompartmental methods in WinNonLin version 5.3 (Pharsight Corp., Mountain View, CA). The highest observed plasma concentration was defined as the  $C_{\max}$  and the corresponding sampling time as the time to  $C_{\max}(T_{\max})$ . The log-linear period (log C versus t) was based on the last data points (at least three). The absolute value of the slope ( $\beta/2.303$ , in which  $\beta$  is the first-order elimination rate constant) was calculated using linear regression analysis. The elimination half-life  $(T_{1/2})$ was calculated as 0.693/ $\beta$ . If the concentration at 24 h ( $C_{24}$ ) was below the limit of quantification of the assay (which was often the case for isoniazid and rifampin), it was estimated based on the last measurable concentra-tion ( $C_{\text{last}}$ ) and  $\beta$  using the formula  $C_{24} = C_{\text{last}} \cdot e^{-[\beta \cdot (24 - T \text{last})]}$ . The area under the plasma concentration-time curve from 0 to 24 h (AUC<sub>0-24</sub>) was calculated using the linear/log-trapezoidal rule from zero up to the last concentration at 24 h. The apparent clearance of the drug (CL/F, where F is bioavailability) was calculated as dose/AUC<sub>0-24</sub>, and the apparent volume of distribution (V/F) was calculated as  $(CL/F)/\beta$ . The reference ranges for  $C_{\text{max}}$  were 3 to 5 mg/liter for isoniazid, 8 to 24 mg/liter for rifampin, 20 to 50 mg/liter for pyrazinamide, and 2 to 6 mg/liter for ethambutol (19).

These reference ranges represent the normal  $C_{\text{max}}$ s that can be expected in adults after the standard doses of anti-TB drugs. They are based on data that were compiled from all available sources (for both healthy volunteers and TB patients) by, among others, Holdiness (1) and Peloquin (2). Subsequently, the ranges were validated in a range of phase I

studies in healthy volunteers (C. A. Peloquin, presented at the 51st Intersci. Conf. Antimicrob. Agents Chemother., Chicago, IL, 17 to 20 September 2011).

**Determination of acetylator status.** We determined the acetylator status phenotypically by assessing the  $t_{1/2}$  of isoniazid. Participants with a  $t_{1/2}$  of greater than 130 min were classified as slow metabolizers, and those with a  $t_{1/2}$  of less than 130 min were classified as fast/intermediate metabolizers (21–23). As another means to assess acetylator phenotype, the metabolic ratio of acetylisoniazid concentration to isoniazid concentration at 3 h postdose was calculated. Patients with a ratio above 1.5 were considered fast/intermediate metabolizers (21). In addition, we also explored a metabolic ratio of 0.55 at 3 h postdose as a cutoff to distinguish fast/intermediate from slow metabolizers, as this evolved from a study in African patients (23).

**Statistical analysis.** Most pharmacokinetic parameters were presented as geometric means. The sample size was considered too small to test for associations between patient characteristics and pharmacokinetic parameters. We tested the effect of acetylator status only on the pharmacokinetics of isoniazid. The correlations between the  $C_{\rm max}$  and AUC<sub>0-24</sub> of each of the drugs were explored using Spearman's rho correlation.

To explore the potential of limited sampling for estimating the  $AUC_{0-24}$  based on sampling early in the pharmacokinetic curve, univariate linear regression was used to determine the association between the plasma concentration at 2, 3, 4, and 6 h postdose and the  $AUC_{0-24}$  for each of the drugs. The  $r^2$  value was presented as a measure of variance in the  $AUC_{0-24}$  that is explained by the variance in the concentration at that time point. Statistical analyses were performed in STATA version 10.1 (Stata Corp. LP, College Station, TX).

#### RESULTS

Twenty tuberculosis patients were enrolled for this study. All were under community-based directly observed treatment. Their median age was 38 years (interquartile range [IQR], 30 to 42 years), 15 patients (75%) were male, seven (35%) were HIV positive, and seven (35%) were considered malnourished based on their BMI (Table 1).

**Descriptive pharmacokinetics.** Intensive pharmacokinetic sampling took place after a median of 19 days after the start of TB treatment (range, 11 to 49 days). The pharmacokinetic parameters for isoniazid, rifampin, pyrazinamide, and ethambutol are presented in Table 2. In 10 patients (53%), the isoniazid peak plasma concentration was below the reference range (3 to 5 mg/liter). In seven patients (35%), the rifampin  $C_{\text{max}}$  was below the reference range (8 to 24 mg/liter); in five of them, the isoniazid  $C_{\text{max}}$  was also below the reference range. In none of the patients was the  $C_{\text{max}}$  of pyrazinamide or ethambutol below the reference range.

There was a positive correlation between the AUC<sub>0-24</sub> and  $C_{\text{max}}$  of isoniazid (correlation coefficient, 0.68; P = 0.001), pyrazinamide (correlation coefficient, 0.69; P = 0.001), and ethambutol (correlation coefficient 0.72; P = 0.003) but not between those of rifampin (correlation coefficient, 0.35; P = 0.14).

Assessment of acetylator status. Based on the elimination half-life of isoniazid, 7 (37%) patients were fast or intermediate metabolizers and 12 (63%) were slow metabolizers. Based on the 3-hour acetyl-INH/INH ratio with a cutoff of 1.5, nine patients (47%) were fast/intermediate metabolizers and 10 patients (53%) were slow metabolizers (Fig. 1). Two patients had a discrepancy in acetylator phenotype determined by the two methods. With a cutoff of 0.55, 11 patients (58%) were fast/intermediate metabolizers and eight (42%) were slow metabolizers.

Acetylator status based on the half-life method was associated with the  $AUC_{0-24}$  and  $C_{max}$  of isoniazid. In slow metabolizers, the

TABLE 1 Characteristics of	of 20 tuberculosis	patients in northern
Tangania		

Characteristic	Value
Male sex, <i>n</i> (%)	15 (75)
Age, median, yr (IQR)	38 (30-42)
Body wt, mean, kg (SD)	55.7 (6.4)
BMI, mean (SD)	19.5 (2.4)
Malnutrition, $n (\%)^a$	7 (35)
HIV positive, <i>n</i> (%)	7 (35)
Type of tuberculosis, $n$ (%)	
Pulmonary	19 (95)
Extrapulmonary	1 (5)
Concomitant drugs, <i>n</i> (%)	
Amoxicillin	1 (5)
Antiretroviral treatment	2 (10)
Co-trimoxazole	3 (15)
Diclofenac	3 (15)
Vitamin B complex	2 (10)
Dose, mean, mg per kg body wt (SD)	
Isoniazid	5.2 (0.46)
Rifampin	10.4 (0.91)
Pyrazinamide	27.8 (2.4)
Ethambutol	19.1 (1.7)
Acetylator status, <i>n</i> (%,) based on:	
INH $t_{1/2}$	
Slow	12 (63)
Fast/intermediate	7 (37)
Acetyl-INH/INH ratio with cutoff of:	
1.5	
Slow	10 (53)
Fast/intermediate	9 (47)
0.55	
Slow	8 (42)
Fast/intermediate	11 (58)

<sup>a</sup> Malnutrition defined as BMI of <18.5 kg/m<sup>2</sup>.

geometric mean AUC<sub>0-24</sub> was 17.2 h · mg/liter (range, 9.5 to 22.7) and the  $C_{\text{max}}$  was 3.4 mg/liter (range, 2.0 to 4.6), and in intermediate/fast metabolizers, the geometric mean AUC<sub>0-24</sub> was 5.2 h · mg/liter (range 3.7 to 8.1) and the  $C_{\text{max}}$  was 2.0 mg/liter (range, 1.0 to 3.3) (P < 0.001 and P = 0.01, respectively).

Association between the 2-, 3-, 4-, and 6-h concentrations and  $AUC_{0-24}$ . The isoniazid concentrations at 2, 3, 4, and 6 h postdose were associated with  $AUC_{0-24}$ , and the  $r^2$  value ranged from 0.91 to 0.96, meaning that 91 to 96% of the variance in the  $AUC_{0-24}$  is explained by the variance in the concentration at a particular sampling time point. For rifampin, the  $r^2$  value was highest for the concentration at 4 h postdose (0.82). For pyrazinamide, the  $r^2$  value was highest for the concentrations at 4 and 6 h postdose (0.84 and 0.86, respectively). For the different ethambutol concentrations, the  $r^2$  value was low, varying from 0.40 to 0.71. This indicates that limited sampling to estimate the  $AUC_{0-24}$  may be feasible for isoniazid, rifampin, and pyrazinamide but less for ethambutol.

## DISCUSSION

This report provides important data on the pharmacokinetic characteristics of isoniazid, rifampin, pyrazinamide, and ethambutol based on intensive 24-hour pharmacokinetic sampling in East African TB patients.

Half of the patients from our study had an isoniazid peak plasma concentration  $(C_{\text{max}})$  below the reference range, and a third of patients had a rifampin  $C_{\text{max}}$  below the reference range. This may partly be explained by the breakfast that patients took within half an hour after the intake of TB drugs. This approach was chosen to mimic the real-life situation for TB patients, as most TB patients take their pills just before or with their breakfast in order to prevent or alleviate gastrointestinal adverse effects. Previous studies have shown that intake of a meal with a high fat or carbohydrate content just before drug intake decreases the rate of absorption and significantly reduces the isoniazid  $C_{\max}$  by 20% to 51% (24–27). The rifampin  $C_{\text{max}}$  can be reduced up to 36% when it is taken directly after a meal (24, 28). There is no significant effect of food on the  $C_{\text{max}}$ s of pyrazinamide and ethambutol (24, 29, 30). Of note, in the current study the average  $T_{\text{max}}$  values for the TB drugs were short (about 1 h for isoniazid, rifampin, and pyrazinamide) (Table 2), indicating that the delay in absorption and a possible related decrease in  $C_{\max}$  were probably limited. Furthermore, it should be noted that pharmacokinetic studies from other African countries found even higher proportions of patients with  $C_{\text{max}}$ s below the reference range for isoniazid (30%) in Botswana [8, 31] and 89% in Kenya [32]) and rifampin (69% in South Africa [12], 78 to 84% in Botswana [27, 28], and 90% in Kenya [32]), despite administration of drugs on an empty stomach in the studies from Botswana and Kenya. The relatively high  $C_{\rm max}$ s of the TB drugs in our study among African patients may also relate to the average body weight of 55.7 kg (Table 1), which is

Pharmacokinetic parameter	Value, geometric mean (minimum-maximum) for:				
	Isoniazid <sup>a</sup>	Rifampin	Pyrazinamide	Ethambutol	
$\overline{AUC_{0-24}}$ , h · mg/liter	11 (3.7–22.7)	39.9 (27.4–68.3)	344 (209–610)	20.2 (13.4-32.0)	
$C_{\rm max}$ , mg/liter	2.8 (1.0-4.6)	8.9 (5.9–14.8)	38.2 (29.0-50.8)	3.3 (2.2-5.8)	
T <sub>max</sub> h	1.2 (0.7–2.9)	1.3 (0.9–3.0)	1.2 (0.7–3.0)	1.5 (0.9-2.2)	
CL/F, liters/h	25.8 (13.0-60.5)	14.4 (8.8–21.9)	4.5 (2.6-6.5)	52 (34.3–71.3)	
V/F, liters	99 (69.6–173.4)	37.3 (22.7–56.5)	40.3 (29.5–98.3)	719 (491–965)	
<i>t</i> <sub>1/2</sub> , h	2.9 (1.3–4.2)	1.8 (1.1–3.8)	5.5 (4.1–15.7)	9.5 (6.9–13.5)	
Reference range $C_{\text{max}}$ , mg/liter	3–5	8–24	20-50	2-6	
No. (%) with $C_{\text{max}}$ below reference range	10 (52)	7 (35)	0 (0)	0 (0)	

<sup>a</sup> Isoniazid plasma concentrations were determined for only 19 patients.

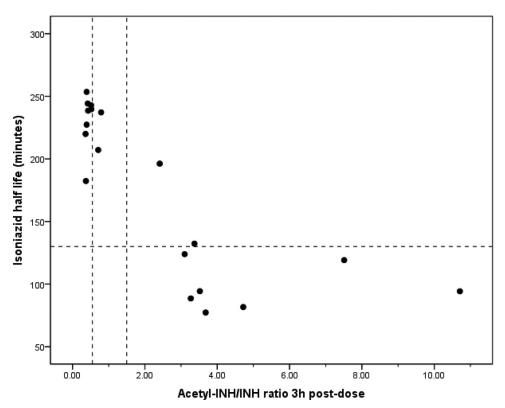


FIG 1 Relationship between the acetylator status (slow versus fast/intermediate) based on two methods: the  $t_{1/2}$  of isoniazid and the acetyl-INH/INH metabolic ratio in the 3-h-postdose plasma sample. Patients with a  $t_{1/2}$  below 130 min (below the horizontal dashed line) were considered fast/intermediate metabolizers (17–19). Patients with an acetyl-INH/INH ratio greater than 0.55 (19) or greater than 1.5 (17) (right of the first or second vertical dashed line, respectively) were considered fast/intermediate metabolizers based on metabolic ratio. Note that phenotyping by  $t_{1/2}$  and metabolic ratio resulted in discordant results (in the upper right quadrant) in 4 patients when a metabolic ratio of 0.55 was used as the cutoff, compared to 2 patients when a metabolic ratio of 1.5 was used as the cutoff.

just above 50 kg, above which 4 FDC tablets (rather than 3) are administered.

Values for total exposure  $(AUC_{0-24})$  may be even more relevant to the efficacy of first-line TB drugs than the  $C_{\text{max}}$  (33). AUC<sub>0-24</sub> values can best be compared to those recorded in Indonesian TB patients (18) and in a racially mixed population of patients in The Netherlands who used similar doses on a mg/kg basis (C. Magis-Escurra, H. M. J. Later-Nijland, J. W. C. Alffenaar, J. Broeders, D. M. Burger, R. van Crevel, M. J. Boeree, A. R. T. Donders, R. van Altena, T. S. van der Werf, and R. E. Aarnoutse, submitted for publication), as the pharmacokinetics in these studies were assessed with the same analytical methodology. Average exposures to rifampin were 39.9, 48.5, and 41.1 h · mg/liter in the Tanzanian patients, the Indonesian patients, and a mixed population, pyrazinamide AUC<sub>0-24</sub> values were 344, 473, and 380 h · mg/liter, respectively, and exposures to ethambutol were 20.2, 14.4, and 23.5 h · mg/liter, respectively. Isoniazid was not measured in the Indonesian TB patients, and we used another analytical method with the mixed population (the average AUC<sub>0-24</sub> was 15.2 h  $\cdot$  mg/liter, compared to 11.0 in the current study). These comparisons show no drastically lower total exposure to TB drugs between Tanzanian patients and the Indonesian patients and the mixed population from the Netherlands.

Even though the exposure to TB drugs in Tanzanian patients may be high compared to exposure in other African studies and similar to those in other populations, a large number of patients had low plasma concentrations of isoniazid and rifampin., Several

July 2013 Volume 57 Number 7

clinical studies have pointed toward a possible association between low TB drug concentrations and poor treatment outcome (3–5). A preclinical model showed that interindividual variability in pharmacokinetics is relevant to the emergence of resistance (7), and a recent meta-analysis revealed that variability in exposure to isoniazid is associated with failure of therapy and acquired drug resistance (10). On the other hand, other studies have found no association between plasma concentrations and effect of first-line TB drugs (34, 35).

We could not readily explain the observed low isoniazid and rifampin AUC<sub>0-24</sub>s and C<sub>max</sub>s by patient characteristics, as the sample size was considered too small for analysis of association between patient characteristics and pharmacokinetic parameters. However, we evaluated the effect of genetic polymorphisms in N-acetyltransferase 2 (NAT2), a phase II metabolic enzyme, on the pharmacokinetics of isoniazid. Slow acetylation of isoniazid into acetylisoniazid is a homozygous recessive trait. Genotypically, homozygous fast, heterozygous fast (or intermediate), and slow acetylators are distinguished (22, 36). In our study, acetylator status was assessed phenotypically using the isoniazid elimination half-life and the acetylisoniazid/isoniazid ratio at 3 h postdose with a cutoff of 1.5. Based on these two methods, 63% or 53% of patients in our study, respectively, were slow metabolizers. This is consistent with available data showing that 50 to 60% of European (Caucasian), African, and Indian populations are slow metabolizers (37). Clearly, assessment of the acetylator status by the acetylisoniazid/isoniazid metabolic ratio at a single time point postdose is more convenient. There was good concordance between the two methods when 1.5 was used as the cutoff for the metabolic ratio (there were 2 discordant patients). In the future, limited sampling may allow for an assessment of acetylator status as well as estimation of AUC values of TB drugs, considering the high correlation between concentrations measured in the early part of the pharmacokinetic curve and exposures to isoniazid, rifampin, and pyrazinamide. The AUC<sub>0-24</sub>s and  $C_{\rm max}$ s of isoniazid, pyrazinamide, and ethambutol were also correlated, but for rifampin no such association was found. We have considered whether this may be explained by the nonlinear, saturable pharmacokinetics of rifampin, but disproportional changes in exposure above certain concentrations are expected to affect both  $C_{\rm max}$  and AUC<sub>0-24</sub>.

In summary, this study provides pharmacokinetic data for first-line TB drugs based on intensive 24-h sampling in East African TB patients. Half of the patients had isoniazid peak plasma levels and a third had rifampin peak plasma concentrations below the reference ranges. Although the pharmacodynamics of the currently used first-line TB drugs are not entirely clear, low isoniazid and rifampin concentrations have been associated with a suboptimal treatment response. Exploring the effect, tolerability, and pharmacokinetics of higher doses of isoniazid and rifampin in African tuberculosis patients is therefore recommended.

## ACKNOWLEDGMENTS

We thank the following people: all patients for participating in this study; C. Irongo (Regional TB and Leprosy Coordinator in the National TB and Leprosy Programme, Tanzania), the staff of the TB clinic at Mawenzi Hospital in Moshi and the research nurses at KCMC for their cooperation and effort; and the laboratory technicians at KCMC and at the Department of Pharmacy of the RUNMC, The Netherlands, for their technical support.

A. Tostmann received the UNESCO/L'Oreal for Young Women in Science Fellowship 2008 to coordinate this study. C. Mtabho, H. Semvua, J. van den Boogaard, and bioanalysis of drugs were sponsored by the African Poverty Related Infection Oriented Research Initiative (APRIORI), a research network funded by the Netherlands-African Partnership for Capacity Development and Clinical Interventions against Poverty-Related Diseases (NACCAP).

#### REFERENCES

- 1. Holdiness MR. 1984. Clinical pharmacokinetics of the antituberculosis drugs. Clin. Pharmacokinet. 9:511–544.
- 2. **Peloquin CA.** 1991. Antituberculosis drugs: pharmacokinetics, p 59–88. *In* Heifets L (ed), Drug susceptibility in the chemotherapy of mycobacterial infections. CRC Press, Boca Raton, FL.
- 3. Kimerling ME, Phillips P, Patterson P, Hall M, Robinson CA, Dunlap NE. 1998. Low serum antimycobacterial drug levels in non-HIV-infected tuberculosis patients. Chest 113:1178–1183.
- 4. Mehta JB, Shantaveerapa H, Byrd RP, Jr, Morton SE, Fountain F, Roy TM. 2001. Utility of rifampin blood levels in the treatment and follow-up of active pulmonary tuberculosis in patients who were slow to respond to routine directly observed therapy. Chest 120:1520–1524.
- 5. Weiner M, Burman W, Vernon A, Benator D, Peloquin CA, Khan A, Weis S, King B, Shah N, Hodge T. 2003. Low isoniazid concentrations and outcome of tuberculosis treatment with once-weekly isoniazid and rifapentine. Am. J. Respir. Crit. Care Med. 167:1341–1347.
- Weiner M, Benator D, Burman W, Peloquin CA, Khan A, Vernon A, Jones B, Silva-Trigo C, Zhao Z, Hodge T. 2005. Association between acquired rifamycin resistance and the pharmacokinetics of rifabutin and isoniazid among patients with HIV and tuberculosis. Clin. Infect. Dis. 40:1481–1491.
- 7. Srivastava S, Pasipanodya JG, Meek C, Leff R, Gumbo T. 2011. Multidrug-resistant tuberculosis not due to noncompliance but to betweenpatient pharmacokinetic variability. J. Infect. Dis. 204:1951–1959.

- Chideya S, Winston CA, Peloquin CA, Bradford WZ, Hopewell PC, Wells CD, Reingold AL, Kenyon TA, Moeti TL, Tappero JW. 2009. Isoniazid, rifampin, ethambutol, and pyrazinamide pharmacokinetics and treatment outcomes among a predominantly HIV-infected cohort of adults with tuberculosis from Botswana. Clin. Infect. Dis. 48:1685–1694.
- 9. Egelund EF, Peloquin CA. 2012. Pharmacokinetic variability and tuberculosis treatment outcomes, including acquired drug resistance. Clin. Infect. Dis. 55:178–179.
- Pasipanodya JG, Srivastava S, Gumbo T. 2012. Meta-analysis of clinical studies supports the pharmacokinetic variability hypothesis for acquired drug resistance and failure of antituberculosis therapy. Clin. Infect. Dis. 55:169–177.
- 11. WHO. 2012. Global tuberculosis control: WHO report 2012. World Health Organization, Geneva, Switzerland.
- 12. McIlleron H, Wash P, Burger A, Norman J, Folb PI, Smith P. 2006. Determinants of rifampin, isoniazid, pyrazinamide, and ethambutol pharmacokinetics in a cohort of tuberculosis patients. Antimicrob. Agents Chemother. 50:1170–1177.
- 13. Gurumurthy P, Ramachandran G, Hemanth Kumar AK, Rajasekaran S, Padmapriyadarsini C, Swaminathan S, Venkatesan P, Sekar L, Kumar S, Krishnarajasekhar OR, Paramesh P. 2004. Malabsorption of rifampin and isoniazid in HIV-infected patients with and without tuberculosis. Clin. Infect. Dis. 38:280–283.
- Sahai J, Gallicano K, Swick L, Tailor S, Garber G, Seguin I, Oliveras L, Walker S, Rachlis A, Cameron DW. 1997. Reduced plasma concentrations of antituberculosis drugs in patients with HIV infection. Ann. Intern. Med. 127:289–293.
- 15. Polasa K, Murthy KJ, Krishnaswamy K. 1984. Rifampicin kinetics in undernutrition. Br. J. Clin. Pharmacol. 17:481–484.
- 16. **Ministry of Health and Social Welfare.** 2006. Manual of the National Tuberculosis and Leprosy Programme in Tanzania, 5th ed. Ministry of Health and Social Welfare, Dar es Salaam, Tanzania
- WHO. 2012. The international classification of adult underweight, overweight and obesity according to BMI. BMI classification; global database on body mass index. WHO, Geneva, Switzerland. http://apps.who.int /bmi/index.jsp?introPage=intro\_3.html. Accessed 5 February 2013.
- Ruslami R, Nijland HM, Alisjahbana B, Parwati I, van Crevel R, Aarnoutse RE. 2007. Pharmacokinetics and tolerability of a higher rifampin dose versus the standard dose in pulmonary tuberculosis patients. Antimicrob. Agents Chemother. 51:2546–2551.
- Peloquin CA. 2002. Therapeutic drug monitoring in the treatment of tuberculosis. Drugs 62:2169–2183.
- 20. Reference deleted.
- Hutchings A, Routledge PA. 1986. A simple method for determining acetylator phenotype using isoniazid. Br. J. Clin. Pharmacol. 22:343–345.
- 22. Parkin DP, Vandenplas S, Botha FJ, Vandenplas ML, Seifart HI, van Helden PD, van der Walt BJ, Donald PR, van Jaarsveld PP. 1997. Trimodality of isoniazid elimination: phenotype and genotype in patients with tuberculosis. Am. J. Respir. Crit. Care Med. 155:1717–1722.
- Seifart HI, Parkin DP, Botha FJ, Donald PR, van der Walt BJ. 2001. Population screening for isoniazid acetylator phenotype. Pharmacoepidemiol. Drug Saf. 10:127–134.
- Lin MY, Lin SJ, Chan LC, Lu YC. 2010. Impact of food and antacids on the pharmacokinetics of anti-tuberculosis drugs: systematic review and meta-analysis. Int. J. Tuberc. Lung Dis. 14:806–818.
- Peloquin CA, Namdar R, Dodge AA, Nix DE. 1999. Pharmacokinetics of isoniazid under fasting conditions, with food, and with antacids. Int. J. Tuberc. Lung Dis. 3:703–710.
- Zent C, Smith P. 1995. Study of the effect of concomitant food on the bioavailability of rifampicin, isoniazid and pyrazinamide. Tuber. Lung Dis. 76:109–113.
- Joshi MV, Saraf YS, Kshirsagar NA, Acharya VN. 1991. Food reduces isoniazid bioavailability in normal volunteers. J. Assoc. Physicians India 39:470-471.
- Peloquin CA, Namdar R, Singleton MD, Nix DE. 1999. Pharmacokinetics of rifampin under fasting conditions, with food, and with antacids. Chest 115:12–18.
- 29. Peloquin CA, Bulpitt AE, Jaresko GS, Jelliffe RW, Childs JM, Nix DE. 1999. Pharmacokinetics of ethambutol under fasting conditions, with food, and with antacids. Antimicrob. Agents Chemother. 43:568–572.
- Peloquin CA, Bulpitt AE, Jaresko GS, Jelliffe RW, James GT, Nix DE. 1998. Pharmacokinetics of pyrazinamide under fasting conditions, with food, and with antacids. Pharmacotherapy. 18:1205–1211.

- 31. Tappero JW, Bradford WZ, Agerton TB, Hopewell P, Reingold AL, Lockman S, Oyewo A, Talbot EA, Kenyon TA, Moeti TL, Moffat HJ, Peloquin CA. 2005. Serum concentrations of antimycobacterial drugs in patients with pulmonary tuberculosis in Botswana. Clin. Infect. Dis. 41:461–469.
- 32. Choudhri SH, Hawken M, Gathua S, Minyiri GO, Watkins W, Sahai J, Sitar DS, Aoki FY, Long R. 1997. Pharmacokinetics of antimycobacterial drugs in patients with tuberculosis, AIDS, and diarrhea. Clin. Infect. Dis. 25:104–111.
- 33. Hall RG, Leff RD, Gumbo T. 2009. Treatment of active pulmonary tuberculosis in adults: current standards and recent advances. Insights from the Society of Infectious Diseases Pharmacists. Pharmacotherapy 29:1468–1481.
- 34. Narita M, Hisada M, Thimmappa B, Stambaugh J, Ibrahim E, Hol-

lender E, Ashkin D. 2001. Tuberculosis recurrence: multivariate analysis of serum levels of tuberculosis drugs, human immunodeficiency virus status, and other risk factors. Clin. Infect. Dis. **32**:515–517.

- Chang KC, Leung CC, Yew WW, Kam KM, Yip CW, Ma CH, Tam CM, Leung EC, Law WS, Leung WM. 2008. Peak plasma rifampicin level in tuberculosis patients with slow culture conversion. Eur. J. Clin. Microbiol. Infect. Dis. 27:467–472.
- Aarnoutse RE. 2011. Pharmacogenetics of antituberculosis drugs, p 176– 190. *In* Donald PR, van Helden PD (ed), Antituberculosis chemotherapy. Progress in Respiratory Research, Basel, Switzerland.
- Ellard GA. 1984. The potential clinical significance of the isoniazid acetylator phenotype in the treatment of pulmonary tuberculosis. Tubercle 65:211–227.