# Detection of *Rickettsia parkeri* from within Piura, Peru, and the First Reported Presence of Candidatus Rickettsia andeanae in the Tick Rhipicephalus sanguineus

Carmen Flores-Mendoza,<sup>1</sup> David Florin,<sup>1,2</sup> Vidal Felices,<sup>1</sup> Edwar J. Pozo,<sup>3</sup> Paul C.F. Graf,<sup>1,4</sup> Roxanne G. Burrus,<sup>1</sup> and Allen L. Richards<sup>2,5</sup>

# Abstract

Domestic farm animals (n = 145) were sampled for the presence of ectoparasites in northwestern Peru during March, 2008. Ninety domestic animals (62%) were positive for the presence of an ectoparasite(s) and produced a total collection of the following: 728 ticks [Amblyomma maculatum, Anocentor nitens, Rhipicephalus (Boophilus) microplus, Rhipicephalus sanguineus, and Otobius megnini], 12 lice (Haematopinus suis), and 3 fleas (Ctenocephalides felis). A Rickettsia genus-specific qPCR assay was performed on nucleic acid preparations of the collected ectoparasites that resulted in 5% (37/743, 35 ticks and 2 fleas) of the ectoparasites positive for the presence of Rickettsia. DNA from the positive individual ticks was tested with 2 other qPCR assays for the presence of the ompB gene in Candidatus Rickettsia andeanae or Rickettsia parkeri. Candidatus R. andeanae was found in 25 A. maculatum ticks and in two Rh. sanguineus ticks, whereas R. parkeri was detected in 6 A. maculatum ticks. Two A. maculatum were co-infected with both Candidatus R. andeanae and R. parkeri. Rickettsia felis was detected in 2 fleas, Ctenocephalides felis, by multilocus sequence typing of the 17-kD antigen and ompA genes. These findings expand the geographic range of R. parkeri to include Peru as well as expand the natural arthropod vector of Candidatus R. andeanae to include Rhipicephalus sanguineus.

Key Words: Candidatus Rickettsia andeanae—Rickettsia parkeri—Ticks—Peru.

# Introduction

 $\mathbf{R}$  ICKETTSIAL DISEASES HAVE RE-EMERGED as a significant public health threat in South America (Labruna 2009). Evidence of the spotted fever group (SFG) rickettsial infections have been documented from diverse areas of Peru (Schoeler et al. 2005, Forshey et al. 2010), including a 2002 outbreak in Sapillica, Piura, Peru. Serological evidence from villagers during that outbreak showed that the prevalence of immunoglobulin M (IgM) antibody to SFG rickettsiae ranged from 10% to 19% among the human population (Blair et al. 2004a). In addition, a novel species of SFG rickettsiae, Candidatus Rickettsia andeanae, was detected from the ticks Amblyomma maculatum (Koch) and Ixodes boliviensis (Neumann) (Blair et al. 2004b). The purpose of this study was to detect and identify rickettsial agents from within the same area of Peru affected by the 2002 outbreak by collecting ectoparasites from domestic animals and testing for the presence of Rickettsia.

## **Materials and Methods**

Sample collections occurred in March, 2008, at 17 sites within 3 districts in the Department of Piura, Peru: Paimas (04°37'49"S and 79°56'85"W), Suyo (04°51'42"S and 80°00'42"), and Tambogrande (04°44'14"L and 80°17'69"W). A variety of domestic animals common to the typical rural farm were examined for external parasites. All visible ectoparasites to the unaided eye were removed with fine forceps, placed in appropriate containers, and later identified to species using morphological keys of Aragão and Fonseca (1961)

<sup>&</sup>lt;sup>1</sup>US Naval Medical Research Unit No. Six, Lima, Peru, Department of Entomology, Washington, DC.

<sup>&</sup>lt;sup>2</sup>Department of Preventive Medicine and Biometrics, Uniformed Services University of the Health Services, Bethesda, Maryland.

<sup>&</sup>lt;sup>3</sup>Ministry of Health, Sullana, Peru. <sup>4</sup>University of Rochester Medical Center, Clinical Microbiology Laboratory, Rochester, New York.

<sup>&</sup>lt;sup>5</sup>Naval Medical Research Center, Silver Spring, Maryland.

			Species of ecto	parasites	encountered				
Species	Surveyed	Rh. sanguineus	A. maculatum	A. nitens	Rh. (B.) micropilus	O. megnini	Ct. felis	H. suis	Total ectoparasites
Cow, Bos taurus	13	0	1	0	9	0	0	0	10
Dog, Canis familiaris	50	218	37	0	0	0	3	0	258
Goat, Capra hircus	9	0	0	0	0	0	0	0	0
Guinea pig, Cavia porcellus	2	0	0	0	0	0	0	0	0
Donkey, Equus asinus	26	0	11	156	0	0	0	0	167
Horse, Equus caballus	25	0	43	117	0	129	0	0	289
Cat, Felis catus	2	0	0	0	0	0	0	0	0
Chicken, Gallus gallus	1	0	0	0	0	0	0	0	0
Sheep, Ovis aries	5	0	0	0	0	0	0	0	0
Pig, Sus domesticus	12	3	3	1	0	0	0	12	19
Total	145	221	95	274	9	129	3	12	743

TABLE 1. ECTOPARASITES COLLECTED FROM DOMESTIC ANIMALS

and Guglielmone and Viñabal (1994). Specimens were transported alive to the laboratory, where they were flash-frozen and maintained at  $-80^{\circ}$ C until DNA extraction. Each specimen was surface-sterilized using sequential washes of sodium hypochlorite (0.05%), benzalkonium chloride (0.5%), ethanol (70%), and distilled sterile water. The specimens were then dried on filter paper and cut in half with a sterile scalpel. One half was used for DNA extraction and PCR analysis and the other half stored for future analysis. *Candidatus* R. andeanae and *R. parkeri* were detected by species-specific qPCR, whereas *R. felis* was found by multilocus sequence typing of the 17-kD antigen and *ompA* genes. DNA extraction, PCR, and sequencing methodologies were performed according to Jiang et al. (2005).

## Results

A total of 145 domestic farm animals were surveyed, of which 90 animals were found to have ectoparasites (Table 1). From these 90 animals, 743 ectoparasites were collected: 728 ticks, 3 fleas, and 12 lice. Percentages of ectoparasites collected in relation to the domestic animals were the following: 38.9% on horses, 34.6% on dogs, 22.5% on donkeys, 2.7% on pigs, and 1.3% on cows. Cats, chickens, goats, guinea pig, and sheep did not yield any ectoparasites. SFG rickettsiae were detected in ticks and fleas from 22 domestic animals (11/50 dogs, 5/25 horses, 4/26 donkeys, 1/12 pigs, and 1/13 cows), resulting in 5% (37/743) of the total ectoparasite specimens positive for the presence of a SFG rickettsiae. Positive ectoparasites were 2 Ctenocephalides felis (Bounché) fleas and 35 ticks: 34.7% (33/95) of the collected Amblyomma maculatum Koch and 0.9% (2/221) of the collected Rhipicephalus sanguineus (Latreille). No Rickettsia was found in the other 3 tick species collected, Anocentor nitens (Neumann), Rhipicephalus (Boophilus) microplus (Canestrini), Otobius megnini (Dugès), or in the 12 louse specimens, all identified as the hog louse Haematopinus suis (Linnaeus). Candidatus R. andeanae was found in 27 A. maculatum ticks (collected from 7 dogs, 5 horses, 4 donkeys, and 1 cow) and in 2 Rh. sanguineus ticks (from 2 dogs) (Table 2). Rickettsia parkeri was detected in 8 A. maculatum ticks (collected from 4 dogs, 3 horses, and 1 pig). Rickettsia felis was detected in the 2 positive fleas, Ct. felis (from 2 dogs). Two specimens of *A. maculatum* were found coinfected with *Candidatus* R. andeanae and *R. parkeri* (Table 2).

## Discussion

In this study, we identified the presence of 3 species of SFG rickettsiae in ectoparasites collected in northwestern Peru. Two species of the SFG rickettsia, Candidatus R. andeanae and R. felis, were documented previously (Blair et al. 2004b). We now report the presence of R. parkeri in Peru. R. parkeri is commonly found in A. maculatum ticks in the southeastern United States (Paddock et al. 2010, Varela-Stokes et al. 2011), but an infected A. maculatum had not been documented in Peru until now. Two specimens of A. maculatum were detected with co-infections of Candidatus R. andeanae and R. parkeri, a mixed infection of the same tick with 2 species of Rickettsia that is not frequently reported in the literature. Also, our findings expand the number of tick species known to be infected with Candidatus R. andeanae to include Rh. sanguineus. We believe that this is the first report documenting detection of R. parkeri from within Peru and the first report of Candidatus R. andeanae from Rh. sanguineus.

#### Acknowledgments

We thank Dr. Rosario Mendez (San Martin de Porres University), Dany Viera (SRS Luciano Castillo Colona-Ministry of Health, Sullana), Leonardo Mendoza (National Institute of Health), and Christopher Cruz (NAMRU-6) for their assistance and support of this study. The views expressed in this article are those of the author(s) and do not necessarily reflect the official policy or position of the Department of the Navy, Department of Defense, or the US Government. This work was supported by DOD-Global Emerging Infection Surveillance, project number 847705.82000.25GB.A0074. The study protocol was approved by the Naval Medical Research Center Institutional Review Board in compliance with all applicable Federal regulations governing the protection of human subjects. This work was prepared as part of official duties of the lead author. Title 17 U.S.C. §105 provides that "Copyright protection under this title is not available for any work of the United States Government."

		I ABLE Z. O	FELLES OF MICAELL	OFECIES OF MICKEI ISIA OF, FUUND BY FIUST AND JILE				
Sites	Meters above sea level	Geographic coordinates	Host code	Specie of ectoparasites	Candidatus R. andeanae	R. parkeri	R. felis	Candidatus R. <i>andeanae</i> +R. parkeri
				•		4		
Paimas	550	04°37'49''L,79°56'85''W	Dog-1	A. maculatum	1			
			þ	A. maculatum		<del>, -</del>		
				A. maculatum				1
			Pig-3	A. maculatum		1		
			Donkey-6	A. maculatum	1			
			$Dog-1\check{7}$	A. maculatum		1		
			þ	A. maculatum	1			
			Dog-19	A. maculatum		1		
			)	A. maculatum	1			
			Horse-35	A. maculatum	1			
				A. maculatum	1			
				A. maculatum	1			
				A. maculatum	1			
			Dog-38 Dog-39	Ct. felis Ct. felis				
Jambur	558	04°36′50″L, 79°56′66″W	Donkey-22	A. maculatum	1			
×			Horse-30	A. maculatum		1		
Guir-Guir	558	04°36′50′′L, 79°56′66′′W	Dog-46	A. maculatum	- 1			
				A. MUCHINIM	I			
Corrales	480	04°37′79′′L, 79°59′01′′W	Horse-50	A. maculatum	,	1		
				A. maculatum	1			
			Horse-53	A. maculatum	1			
				A. maculatum	← ,			
				A. maculatum	Ι			
La Tina	422	04°24′22′′L, 79°56′32′′W	Dog-71	A. maculatum	1			
				A. maculatum	1			
			Cow-80	$A.\ maculatum$	1			
Sarayuyo	526	04°33'05''L, 79°56'08''W	Dog-104	A. maculatum	1			
				A. maculatum	1			
			Donkey-108	A. maculatum	1			
				A. maculatum	1			
			Donkey-109	A. maculatum	1			
Panam GP-VI	206	04°75′06′′L, 80°29′12′′W	Dog-124	A. maculatum				
				A. macutatum	Ι			
Partidor	225	04°73′ 68″L, 80°28′02″W	Horse-128	A. maculatum				1
Bolognesi	236	04°68'07''L, 80°25'11''W	Dog-129	Rh. sanguineus	1			
Los Laureles	463	04°62'12″ L, 79°01'60″W	Dog-136	Rh. sanguineus	1			
Total					27	9	7	7

TABLE 2. SPECIES OF RICKETTSIA SP. FOUND BY HOST AND SITE

#### **Author Disclosure Statement**

No competing financial interests exist.

#### References

- Aragão H, Fonseca F. Notas de Ixodologia. VIII. Lista e chave para os representantes da fauna ixodologica brasileira. Mem Inst Oswaldo Cruz 1961; 59:115–129.
- Blair PJ, Schoeler GB, Morón C, Anaya E, et al. Evidence of rickettsial and leptospira infections in Andean Northern Peru. Am J Trop Med Hyg 2004a; 70:357–363.
- Blair PJ, Jiang J, Schoeler GB, Morón C, et al. Characterization of spotted fever group rickettsiae in flea and tick specimens from northern Peru. J Clin Microbiol 2004b; 42:4961–4967.
- Forshey BM, Stewart A, Morrison AC, Galvez H, et al. Epidemiology of spotted fever group and typhus group rickettsial infection in the Amazon Basin of Peru. Am J Trop Med Hyg 2010; 82:683–690.
- Guglielmone AA, Viñabal AE. Claves morfológicas dicotómicas e información ecológica para la identificación de garrapatas del género *Amblyomma* Koch, 1844 de la Argentina. Rev Inv Agropec 1994; 25:39–67.
- Jiang J, Blair PJ, Felices V, Morón C, et al. Phylogenetic analysis of a novel molecular isolate of spotted fever group rickettsiae

from northern Peru, *Candidatus* Rickettsia andeanae. Ann NY Acad Sci 2005; 1063:337–342.

- Labruna MB. Ecology of *Rickettsia* in South America. Ann NY Acad Sci 2009; 1166:156–166.
- Paddock CD, Fournier PE, Sumner JW, Goddard J, et al. Isolation of *Rickettsia parkeri* and identification of a novel spotted fever group *Rickettsia* sp. from Gulf Coast ticks (*Amblyomma maculatum*) in the United States. Appl Environ Microbiol 2010; 76:2689–2696.
- Schoeler GB, Morón C, Richards A, Blair PJ, et al. Evidence of human spotted fever rickettsial infections in Peru. Emerg Infect Dis 2005; 11:622–624.
- Varela-Stokes AS, Paddock CD, Engber B, Toliver M. Rickettsia parkeri in Amblyomma maculatum ticks, North Carolina, USA, 2009–2010. Emerg Infect Dis 2011; 17:2350–2353.

Address correspondence to: Roxanne G. Burrus U.S. Naval Medical Research Unit No. Six (NAMRU-6) Avenida Venezuela Cuadra 36 Callao 2 Peru E-mail: roxanne.burrus@med.navy.mil

**508**