

# HLA Identical Leukemia Cells and T Cell Growth Factor Activate Cytotoxic T Cell Recognition of Minor Locus Histocompatibility Antigens In Vitro

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**ABSTRACT** Lymphocytes from a healthy HLA-identical bone marrow transplant donor were tested for their ability to destroy her brother's acute myelogenous leukemia blasts in vitro. Primary mixed lymphocyte culture (MLC) and cell-mediated lysis (CML) responses between the patient's remission (pretransplant) and donor's lymphocytes were negative. Stimulation of donor lymphocytes for 7 d in vitro with irradiated leukemia cells, leukemia cells plus allogeneic irradiated lymphocytes, or a pool of irradiated lymphocytes from 10 donors, did not activate any cytotoxic cells able to destroy the HLA identical leukemic blasts. Further culturing for 7 additional d in T cell growth factor (TCGF) generated lymphocytes that induced effective cytotoxicity against the leukemic blasts, but not against autologous lymphocytes. Effective killing against the leukemia was observed only in cultures initially stimulated with the irradiated leukemia cells. These cytotoxic cells were maintained in TCGF and mediated persistent killing against the leukemic target cells. They were also able to destroy lymphocytes from the patient's mother and father, but not from an unrelated cell donor. This suggested specific recognition of non-HLA antigens inherited by the patient, that were foreign to the HLA identical bone marrow donor. These lymphocytes were cloned by a limiting dilution technique and one clone maintained cytotoxicity to the AML blasts and the father's lymphocytes, but not lymphocytes from the mother or an HLA-identical donor. This cytotoxicity was inhibited by a monoclonal anti-

HLA antibody. Thus, in vitro sensitization of this sibling's lymphocytes with AML blasts followed by TCGF expansion, and cloning, enabled the detection of HLA-restricted cytotoxic cells that recognize minor locus histocompatibility antigens. This immune recognition may be relevant to the "graft vs. leukemia" effect that has been observed in leukemic animals and patients following histocompatible hematopoietic transplants.

## INTRODUCTION

Human lymphocytes are readily activated in vitro by histocompatibility antigens controlled by the HLA region (1, 2). Lymphocytes from healthy donors who have never been directly immunized with foreign tissue mount a rapid proliferative response to foreign HLA-D antigens in the mixed lymphocyte culture (MLC)<sup>1</sup> and generate highly reactive cytotoxic T lymphocytes (T<sub>c</sub>) that recognize antigens controlled by the HLA A, B, and C regions (3). In contrast, lymphocytes from healthy HLA-identical siblings do not induce either an MLC proliferative response nor the generation of cytotoxic cells that react to each others tissues in vitro (4). However, tissue grafts from one HLA identical sibling to another are rejected when immune suppression is omitted. Thus the "weak" minor histocompatibility antigens that can affect graft rejection in vivo are not recognized by human lymphocyte populations in a primary in vitro response (5).

In the mouse, primary responses to minor locus (non-H-2) antigens are observed between some strains (6). Priming in vivo increases the ability to detect a minor locus immune response in vitro (7). Recognition of

<sup>1</sup> *Abbreviations used in this paper:* AML, acute myelocytic leukemia; BM, bone marrow; CML, cell-mediated lysis; GVD, graft vs. host; MLC, mixed lymphocyte culture; T<sub>c</sub>, cytotoxic T lymphocytes; TCGF, T cell growth factor.

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these minor locus antigens is H-2 restricted, since the T<sub>c</sub> must simultaneously recognize H-2 antigens (8, 9). Furthermore, depletion of suppressor lymphocyte populations results in a population that gives an enhanced response to minor locus antigens in vitro (10). In man, in vivo immunization, either by multiple transfusions, or tissue grafting, has provided adequate sensitization to enable the detection of in vitro immune responses to minor locus transplantation antigens (11–13). These cytotoxic cells are also MHC restricted (14, 15).

We have studied the in vitro responses of human lymphocytes to HLA-identical leukemic myeloblasts. Prior reports have demonstrated that simultaneous in vitro sensitization with leukemic cells and unrelated histoincompatible cells can together act to induce the generation of cytotoxic lymphocytes able to destroy HLA-identical or autologous leukemic cells (16–18). We present here results of experiments investigating the in vitro response of a healthy lymphocyte donor to her brother's acute myelocytic leukemic (AML) cells. Initial in vitro sensitization with irradiated leukemia cells or allogeneic lymphocytes did not induce the generation of any antileukemic cytotoxicity. Subsequent expansion of these same in vitro primed lymphocytes in conditioned medium containing T cell growth factor yielded cytotoxic lymphocytes able to kill the leukemic blasts in vitro. Additional experiments were performed to determine the specificity of these cytotoxic lymphocytes. Assays using bulk or cloned cytotoxic T cells on leukemic and normal target cells demonstrated that reactivity was not against a "leukemia specific" antigen, but directed against minor locus histocompatibility antigens expressed on normal tissues.

## METHODS

**Lymphocytes and bone marrow cells.** Lymphocytes were obtained by venipuncture from patients, their relatives, and healthy volunteers. Peripheral blood lymphocytes (PBL) were obtained by Ficoll sedimentation of heparinized blood. Heparinized bone marrow aspirates were obtained from leukemic patients and purified by Ficoll density flotation (16). All donors signed informed consent forms approved by the University of Wisconsin Committee for the Protection of Human Subjects.

**Generation of conditioned medium containing T cell growth factor (TCGF).** Pooled irradiated PBL from eight separate donors were cultured at  $10^6$  cells/ml in the presence of 1% phytohemagglutinin (Difco Laboratories, Detroit, MI). Supernatant was harvested after 48 h, filtered, and tested for potency by previously described methods (19, 20). Such unpurified supernatants are known to contain many lymphokines, some of which influence T cells, including interleukin-2 (19–22). This conditioned medium was used here to expand and clone human T cell populations; it is designated "TCGF" (19–25) to functionally distinguish it from the multiple other forms of conditioned medium that have been described.

**In vitro cultures, MLC and cell-mediated lysis (CML) assays.** MLC. The MLC was performed using conditions

described previously (26). In brief,  $0.1 \times 10^6$  responding cells were cultured in 0.25-ml round-bottomed Linbro microwells (Flow Laboratories, McLean, VA) with  $0.1 \times 10^6$  irradiated stimulating lymphocytes (2,500 rad) or  $0.2 \times 10^6$  irradiated (4,000 rad) cryopreserved AML blasts in 0.2 ml of HS-RPMI (media RPMI 1640 supplemented with 25 mM Hepes buffer, L-glutamine, penicillin, streptomycin [Flow Laboratories] and 15% [by volume] heat-inactivated nontransfused human male serum). After 6 d of incubation at 37°C in a 5% CO<sub>2</sub> humidified incubator, the cultures were pulsed with 1  $\mu$ Ci of <sup>3</sup>H-labeled methyl thymidine (<sup>3</sup>H]TdR) (New England Nuclear, Boston, MA). Radiolabeled cultures were incubated for 8 h and harvested with a MASH device (Otto Hiller, Madison, WI). The <sup>3</sup>H]TdR incorporation was measured in a liquid scintillation counter. Each experimental group was assayed in quadruplicate and expressed as mean counts per minute  $\pm$  the SEM.

**CML.** 10 million lymphocytes were cultured with various combinations of irradiated lymphocytes or AML blasts in 25-cm<sup>2</sup> flasks (Falcon Labware, Oxnard, CA) on end containing 15 ml of HS-RPMI. After 6 d in vitro, lymphocytes were harvested, resuspended, and serially diluted to give the appropriate effector to target (E:T) ratio as indicated in the text. Effectors were added in 0.1 ml to round-bottomed microwells. Target cells were fresh or cryopreserved lymphocytes or AML blasts cultured alone in HS-RPMI for 2–6 d immediately before use in the CML assay. Target cells were labeled with 250  $\mu$ Ci of <sup>51</sup>Cr (New England Nuclear) in 0.3 cm<sup>3</sup> for 3 h, then washed, diluted and  $5 \times 10^5$  cells were added to the microwells in 0.1 ml. After addition of the target cells to the effector cells in the microwells, the plates were centrifuged at 200 g for 5 min at room temperature and then incubated for 4 h at 37°C. The plates were then centrifuged at 500 g at 4°C, supernatants harvested using Flow harvesting frames, and <sup>51</sup>Cr counted in a gamma counter. Percent cytotoxicity was calculated using the formula, percent cytotoxicity = (experimental cpm – spontaneous cpm / maximum cpm – spontaneous cpm)  $\times$  100. Spontaneous and maximum release were determined by incubating target cells in HS-RPMI or cetrimide detergent (Sigma Chemical Co., St. Louis, MO), respectively. Spontaneous release values ranged from 8 to 25% of the maximum counts per minute. In all experiments two or three effector to target ratios were tested.

**Monoclonal antibody-CML inhibition.** Hybridoma PA 2.6 derived by Dr. Peter Parham was obtained as a gift from Dr. Steven Shaw (National Institutes of Health, Bethesda, MD) and Dr. Peter Parham (Stanford University, Palo Alto, CA). This monoclonal reagent reacts with a monomorphic antigenic determinant thought to be present on all HLA class I (45,000 mol wt) antigens (27). This hybridoma was grown in ascites by Dr. William Sugden (University of Wisconsin). This ascites fluid was added to <sup>51</sup>Cr-labeled target cells at a final dilution of 1:200 just before addition of cytotoxic cells in experiments testing HLA restriction (28, 29). This same concentration of ascites fluid has been shown in our laboratory to inhibit specific lysis of allogeneic targets by bulk cultures of alloactivated T<sub>c</sub> without inhibiting destruction of K562 target cells, and inhibit the HLA restricted recognition of some, but not all, T<sub>c</sub> clones that destroy autologous lymphoblastoid cell line cells. We have also shown that other monoclonal antibodies have no inhibitory effect on cytolysis by these same T<sub>c</sub>.<sup>2</sup>

**Cryopreservation.** Fresh lymphocytes, AML blasts, and in vitro activated lymphocytes were cryopreserved in 10%

<sup>2</sup> Hank, J. A., and P. M. Sondel. Submitted for publication.

dimethyl sulfoxide using controlled rate freezing (Cryo-Med, Mt. Clemens, MI) as described previously (16, 30).

## RESULTS

A previously healthy 26-yr-old man (P) was diagnosed with acute myelogenous leukemia (by standard histologic criteria) in June 1981. Before any antileukemic therapy, an aliquot of his leukemic bone marrow (L) was aspirated, purified, and cryopreserved. Immunologic marker testing on these myeloblasts revealed they were typical AML cells; negative for terminal deoxynucleotidyl transferase, the "common" acute lymphoblastic leukemia antigen, and T and B cell markers, but heterogeneous for the Ia antigen (31). 4 mo later, while in a stable remission, he was given 1,000 rad fractionated total body irradiation, 60 mg/kg per d for 2 d of cyclophosphamide followed by  $2.5 \times 10^8$ /kg nucleated bone marrow cells from his HLA identical sister (S). HLA and MLC data are presented in Table I. Donor S is a healthy primiparous 45-yr-old white female, never transfused with allogeneic blood. Her only potential allosensitization was a full term pregnancy 13 yr before bone marrow (BM) donation.

Lymphocytes from S (obtained at the time of BM transplant donation) were cultured with irradiated leukemic cells ( $SL_x$ ), with irradiated leukemic cells and irradiated lymphocytes from an unrelated donor ( $SL_xX_x$ ), or with an irradiated pool of lymphocytes from 10 unrelated donors (S Pool<sub>x</sub>). After 7 d of in vitro sensitization, these lymphocytes were tested for their cytotoxic capacity in the CML assay using leukemic blasts as target cells as well as other populations of target cells (Table II, experiment 1). Note that sensitization of the sister's lymphocytes with irradiated allogeneic cells ( $SX_x$  or S Pool<sub>x</sub>) generated effective  $T_c$  directed against alloantigens present on target cells

from unrelated donor X. However, neither of these cultures generated effective killing against the leukemic blasts (32). Even sensitization in the "three cell" protocol ( $SL_xX_x$ ) generated no killing of the leukemic blasts (16, 17). However, this same culture was able to kill the allogeneic cells from donor X, demonstrating that immune activation had occurred in this culture. Lastly, lymphocytes from a second unrelated donor (Y) were sensitized with irradiated leukemic cells. This culture ( $YL_x$ ) generated effective killing against the leukemic blasts, as well as against remission bone marrow cells from the patient, and lymphocytes from the healthy sibling. Thus, the leukemic blasts were effective at presenting alloantigens in a sensitization culture. Furthermore, the leukemic blasts, remission bone marrow, and sibling's lymphocytes could be destroyed by appropriately alloactivated killer cells recognizing the identical HLA antigens on these three target cells.

Replicate flasks of these same cultures were maintained in vitro in fresh media containing 20% TCGF. These were retested on the same target populations 7 d later (Table II, experiment 2). As noted in the primary cultures, alloactivation of sibling's (S) lymphocytes with irradiated pooled lymphocytes or cells from donor X induced  $T_c$  that effectively destroyed target lymphocytes from donor X. However, activation with the HLA identical leukemic blasts, alone ( $SL_x$ ) or in combination with the irradiated allogeneic cells ( $SL_xX_x$ ), now induced 26.3 and 32.8% killing against the leukemic blasts. These same two effector cell populations also mediated weak killing against remission bone marrow (11.6% and 13.1% cytotoxicity), but were unable to destroy the sister's autologous lymphocytes.

Destruction of the leukemic cells and "remission" bone marrow by HLA identical sibling's lymphocytes could result from  $T_c$  recognition of minor locus (non-HLA) antigens present on both tissues, or from  $T_c$  de-

TABLE I  
MLC Identity of Patient and Sister

Responding cells	$^3\text{H}]\text{TdR cpm} \times 10^3$ induced by stimulating cells				
	Media	$P_x$	$S_x$	$X_x$	$L_x$
P	1.9±0.6	0.7±0.4	1.4±0.2	38.7±5.5	3.4±1.3
S	1.8±0.4	0.9±0.3	1.5±0.4	58.4±6.2	1.8±0.5
X	1.4±0.4	54.4±3.0	60.5±2.5	1.7±2.4	13.3±0.2

PBL ( $50 \times 10^3$  per well) from the patient (P), his HLA-identical sister (S), and an unrelated donor (X) were stimulated in MLC with irradiated (2,500 rad) lymphocytes ( $100 \times 10^3$ ) from donors P, S, or X, or the same number of irradiated (4,000 rad) cryopreserved AML blasts (L) from patient P. After 120 h, cultures were labeled with  $^3\text{H}]\text{TdR}$  for 12 h and harvested. Results are  $^3\text{H}]\text{TdR cpm} \times 10^3$  of quadruplicate cultures±SEM. Both patient and donor were HLA A-2, A-X, B-15, B-17, and Bw4, Bw6 by standard serologic microcytotoxicity testing with alloantisera.

TABLE II  
Generation of Cytotoxic Cells against HLA-identical AML Blasts

Effector	Percent cytotoxicity on target cells				
	L	BM	S	X	
Experiment 1					
Primary culture 7 d in vitro	SL <sub>x</sub>	-2.0±0.4	-0.8±0.3	-1.2±0.4	-5.7±0.6
	SX <sub>x</sub>	1.8±0.6	1.3±0.9	0.9±0.8	35.9±1.7
	S Pool <sub>x</sub>	3.8±0.7	3.3±1.1	-1.6±0.4	24.9±9.5
	SL <sub>x</sub> X <sub>x</sub>	2.6±0.4	2.0±0.6	2.0±0.2	30.0±1.3
	YL <sub>x</sub>	65.4±2.4	70.6±2.3	57.0±1.5	0.8±1.0
Experiment 2					
14 d in vitro (7 d in TCGF)	SL <sub>x</sub>	26.3±0.8	11.6±1.7	-1.7±1.0	-0.5±0.4
	SX <sub>x</sub>	0.3±0.5	0.9±1.5	0.7±0.4	55.7±1.5
	S Pool <sub>x</sub>	7.3±0.5	1.7±1.0	0.1±0.6	56.2±1.7
	SL <sub>x</sub> X <sub>x</sub>	32.8±1.8	13.1±0.7	1.9±0.6	61.2±1.7
	YL <sub>x</sub>	78.3±2.1	39.1±2.5	66.1±2.3	24.5±0.8

PBL ( $10 \times 10^6$ ) from the HLA-identical sibling (S) or an unrelated donor (Y) were stimulated in duplicate flasks with irradiated lymphocytes ( $10 \times 10^6$ ) from an unrelated donor (X<sub>x</sub>), a pool of PBL from 10 unrelated donors (Pool<sub>x</sub>) or AML cells (L<sub>x</sub>) from patient P. After 7 d (experiment 1), cells from each sensitization mixture were harvested and tested in the cytotoxicity assay on the following target cells: L, AML blasts from P; BM, remission (pretransplant) nucleated bone marrow cells from P; S, PBL from donor S; X, PBL from donor X. For experiment 2, the duplicate flask of each sensitization combination was recultured after the addition of 20% (vol/vol) TCGF. After an additional 7 d in TCGF (day 14 of culture), lymphocytes were harvested and tested on <sup>51</sup>Cr-labeled target cells from the same four cell populations used in experiment 1. All results represent percent cytotoxicity±SD in a 4-h assay of  $5 \times 10^3$  target cells and  $150 \times 10^3$  effectors in quadruplicate wells.

tection of a "leukemia-specific" antigen present on both the AML blasts and the morphologically remission bone marrow. To differentiate these possibilities it was essential to test for cytotoxicity on other nonleukemic tissues that may be bearing the same minor locus antigens. Because this patient had already received an allogeneic bone marrow transplant, we were unable to recover any lymphoid populations from this patient that could clearly be of host origin and bear exactly the same minor locus antigens as the leukemic cells or remission bone marrow cells. We thus planned to test these effector cells on target lymphocytes from the parents of this patient. To prevent the loss of cytotoxic function of these effector cells while arranging for subsequent in vitro testing, these activated cytotoxic lymphocytes were cryopreserved.

PBL were obtained from both mother (M) and father (F) of this patient and also cryopreserved. After 6 mo in liquid nitrogen, the lymphocytes and effector cells were thawed and placed in culture and assayed in the chromium release test (Table III). As before, the sister's effector population that had been alloactivated (SX<sub>x</sub>) was still highly efficient at destroying target cells from donor X. There was low level cross-reactive killing of

this effector population on the mother's target cells and minimal cytotoxicity against the father's target cells. Minimal detectable cytotoxicity was seen against autologous lymphocytes from the sister or against the leukemic blasts by these alloactivated cytotoxic cells. In contrast, the leukemia cells were effectively destroyed by the sister's cells that had been activated with leukemic blasts (SL<sub>x</sub>). This culture also mediated excellent killing against both mother's and father's target cells, but not against autologous lymphocytes (S) or cells from donor X.

To further examine the specificity of this T<sub>c</sub> response, T<sub>c</sub> clones were generated and maintained in vitro (23). Effector cells from the SL<sub>x</sub> culture maintained in TCGF and tested in Table III were seeded at 1 cell/well into 192 microwells with  $10^4$  irradiated leukemic cells from the patient as feeder cells and 20% TCGF. After 2 wk, 23 clones showed abundant growth by visual inspection. These were tested in two separate screening assays and on each screening only clones 11 and 17 showed significant destruction of HLA-identical AML blasts (note that these clones have not been subcloned; even though they were initially seeded at one cell per well and recovered with a "cloning efficiency" of <23 per

TABLE III  
Cytotoxic Cells Recognize Minor Histocompatibility Antigens

Cryopreserved effector	Stimulator	E:T	Percent cytotoxicity on target				
			L	S	M	F	X
SL <sub>x</sub>	L <sub>x</sub>	10	24.3±1.3	3.7±1.4	25.8±2.8	34.9±2.9	0.3±0.7
		3	13.0±1.3	1.0±3.1	11.9±0.9	16.9±2.9	-2.0±0.5
SL <sub>x</sub> X <sub>x</sub>	X <sub>x</sub>	10	6.2±0.5	-0.2±0.8	27.6±0.3	19.9±2.0	60.3±4.7
		3	3.0±0.8	-0.7±2.2	16.9±4.8	9.8±0.5	40.5±1.5
SX <sub>x</sub>	X <sub>x</sub>	10	3.5±0.5	3.5±2.2	16.8±1.5	5.7±0.6	47.5±1.3
		3	1.9±1.0	-0.1±1.2	7.9±1.3	6.7±0.8	32.3±0.7
YL <sub>x</sub>	L <sub>x</sub>	10	25.7±2.1	39.9±3.2	32.2±1.3	45.9±0.4	1.9±0.4
		3	12.0±1.3	22.5±0.8	9.3±0.8	23.1±1.7	-0.9±0.4

Excess effector cells harvested after 14 d in vitro (experiment 2, Table II) were cryopreserved in 10% dimethyl sulfoxide. 6 mo later these cells were thawed, cultured in TCGF, and restimulated with the original cryopreserved stimulating cells. Cultures were harvested after 7 d and tested on target cells: cryopreserved AML blasts from P (L), and chromated cryopreserved PBL from the patients HLA-identical sister (S), their mother (M), their father (F), and the unrelated donor (X). Cytotoxicity was measured in a 4-hr <sup>51</sup>Cr release assay with 5 × 10<sup>3</sup> target cells with both 15 and 50 × 10<sup>3</sup> effector cells in quadruplicate wells, making the effector to target (E:T) ratio, 10 and 3 to 1.

192 wells, it remains possible that they do not represent single T cell-derived clones). These clones were then tested (Table IV) for cytotoxicity against the same target cell populations tested previously with bulk generated T<sub>c</sub> in Table III. In this test (and two subsequent tests) cells from clone 11 had lost their ability to destroy the AML blasts, as well as all other targets. In contrast, lymphocytes from clone 17 mediated significant destruction of the AML targets and the father's lymphocytes, but not target cells from the mother or autologous S targets. Table V shows that cytotoxicity on L and F targets by cells from clone 17 is inhibited by the PA 2.6 monoclonal antibody directed against a class I HLA monomorphic determinant. Thus, lymphocytes from clone 17 recognize a minor locus antigen on L and F target cells, and that recognition appears to be HLA restricted.

TABLE IV  
Cloned Human T<sub>c</sub> Recognize Minor Locus Antigen

Effector clone	Percent cytotoxicity on target			
	L	S	M	F
Clone 11	5.6±2.5	-4.9±2.0	1.4±0.9	-1.9±2.5
Clone 17	26.3±3.3	6.2±1.1	2.2±2.2	15.5±3.6

Cells from T<sub>c</sub> clones 11 and 17 (see text) were assayed on the indicated target cells in the 4-h <sup>51</sup>Cr release assay: L, AML blasts; S, M, and F are PBL from the patient's sister, mother, and father, respectively. Because of low effector cell yields, clone 11 was tested at 0.1 effector per target cell and clone 17 was tested at 1.0 effector per target cell.

## DISCUSSION

These experiments demonstrate that under appropriate in vitro sensitization conditions, effector cells able to destroy HLA identical leukemic blasts can be generated and maintained in vitro, cryopreserved, thawed, recultured, cloned, and still retain their cytotoxic function and immunologic specificity. We feel that results presented in Table III demonstrate that non-HLA antigens are being recognized by the sister's lymphocytes that had been activated in vitro with irradiated leukemic cells and expanded with TCGF. The absence of killing by this population on autologous lymphocytes and target cells from donor X argues strongly against this being a totally "nonspecific" effector cell. In contrast, the activation of killing against the leukemic target might suggest the recognition of a leukemia specific antigen. But if this were the case, then this (SL<sub>x</sub>) population should not have destroyed nonleukemic target lymphocytes from the mother and father. This latter effect could occur only if leukemia-specific antigens on the leukemic blasts are immunologically cross-reactive with foreign histocompatibility antigens on the mother's and father's normal lymphoid cells (this phenomenon has been designated "alien" histocompatibility antigens on tumor cells) (33). However, if this were the case, one would expect even greater killing of the unrelated X target cells by the SL<sub>x</sub> culture since there is twice as great a chance of cross-reactivity on the target cells from unrelated donor X than on target cells from either the mother or father. This is due to donor X differing from the responding lym-

TABLE V  
Killing by Cloned T Cells Directed against Minor Locus Antigen Is Inhibited  
by Monoclonal Anti-HLA Class I Antibody

Effector	E:T	Anti-HLA added	Percent cytotoxicity on target			
			L	S	M	F
Clone 17	12	—	30.1±1.2	2.0±0.5	2.0±1.0	28.8±1.7
	4	—	21.1±0.7	2.0±0.5	-1.2±1.1	12.7±1.1
	1	—	11.3±1.3	0.5±0.8	0.7±0.5	13.2±1.5
Clone 17	12	+	8.2±0.7	ND*	-0.8±0.5	-0.6±0.5
	4	+	4.5±0.7	ND	-0.1±0.8	0.9±1.6
	1	+	2.1±1.1	ND	2.0±0.5	6.9±5.1

Cells from clone 17 were tested on the indicated target cells at E:T ratios of 12, 4, and 1:1 in a 4-h <sup>51</sup>Cr release assay. Just prior to the addition of effector cells, PA 2.6 monoclonal antibody (1:200 final concentration of ascites fluid) was added to the target cells as indicated. Separate control (spontaneous and maximum release) values were obtained for target cells treated with PA 2.6 and these were not significantly altered from those values for nontreated target cells.

\* ND, not determined.

phocyte donor by two HLA haplotypes rather than one haplotype, as is the case for both parents.

The activation of cytotoxic cells in the SL<sub>x</sub> culture that can destroy L, M, and F targets, but not S or X targets (Table III) is most consistent with recognition of at least one minor locus transplantation antigen. There are numerous such loci able to activate murine allograft reactions in vivo; some loci induce stronger reactions than others (34). The responses demonstrated in Table III by the SL<sub>x</sub> effector cells potentially reflect recognition of an allele controlled by a single non-HLA-locus. If so, the mother, father, and patient would all share an allele at that locus that was not inherited by the sister. Potential genotypes for this locus could be: father = *a/b*, mother = *a/b*, sister = *b/b*, and patient *a/b*. By this model the effectors from the SL<sub>x</sub> culture in Table III would recognize the *a* allele as foreign on F, M, and L targets, but not on the S or X target cells. However, the clonal analysis (Tables IV and V) shows this single locus hypothesis to be an oversimplification. Recognition by cells from clone 17 of a determinant shared by the patient's leukemia cells and the father's lymphocytes, but not shared by the mother's lymphocytes, proves that killers from the bulk culture SL<sub>x</sub> detected at least two separate minor locus determinants on L targets (Table III). While the determinant detected by clone 17 could potentially be a minor locus antigen controlled by the Y chromosome (14, 15), some other non-HLA antigen must be shared by M and L targets to account for killing of M targets (which contain no Y chromosome) by cells from the SL<sub>x</sub> bulk culture (Table III). Thus, at least two alleles at one or more non-HLA loci are the targets of the

killer cells stimulated by the leukemic blasts in these studies. In murine models, recognition of minor locus antigens by T<sub>c</sub> is MHC restricted (7-9). That clone 17 effectors are inhibited by antibody to the class I HLA molecules is consistent with HLA-restricted recognition of this minor locus antigen, as demonstrated for human bulk cultures and T cell clones from multiply transfused donors (13-15).

Alloactivation with pooled lymphocytes, or a single allogeneic donor, can occasionally (but not always), activate antileukemic cytotoxicity (16, 32). The primary responses demonstrated in Table II show that the sister's lymphocytes are not activated to kill leukemic blasts by alloactivation alone. This was the case even after prolonged culturing in TCGF with repeat alloantigenic stimulation (SX<sub>x</sub>, Table III) (35). In contrast, activation with leukemic cells and alloantigens together (SL<sub>x</sub>X<sub>x</sub>) did generate antileukemic killing in the 14-d culture. Thus in vitro activation of T<sub>c</sub> able to destroy these HLA identical leukemic blasts required the presence of irradiated stimulating cells that expressed the same foreign minor locus antigens that were recognized on the leukemic cells in the CML assay. That this same SL<sub>x</sub>X<sub>x</sub> culture generated much less killing against the leukemic cells in Table III may be due to the fact that this culture was reactivated with TCGF, L<sub>x</sub>, X<sub>x</sub> after thawing, while the SL<sub>x</sub> culture was reactivated only with TCGF and L<sub>x</sub>. In other words, the presence of the alloantigenic stimulus (X<sub>x</sub>) may have reactivated strong allorecognition in the bulk culture (SL<sub>x</sub>X<sub>x</sub>) that competitively inhibited the activation of "antileukemic" effectors recognizing minor locus antigens (36).

Previous studies demonstrating "primary" in vitro

cytotoxic responses to HLA-identical target cells bearing minor locus antigens all required extensive prior *in vivo* immunizations (4, 11–15). Thus, most responding cell donors were patients with aplastic anemia or renal failure receiving multiple transfusions or renal allografts. The healthy donor used here (S) was never transfused with any allogeneic blood product. It remains possible that her normal term pregnancy 13 yr before these studies caused *in vivo* immunization to fetal minor locus antigens. We were unable to obtain blood from her child or his father (unrelated to individuals S, P, M, or F) to screen for immune cross-reactivity between their minor locus antigens and those minor locus antigens of the AML patient (P) recognized as foreign by the donor. Nevertheless, no minor locus reactive T<sub>c</sub> were detected following the primary *in vitro* stimulation of the sister's lymphocytes with irradiated AML blasts. These were detected only after further culturing in TCGF. This suggests that the primary culture caused the differentiation of cytotoxic precursors, but their detection required the "helper" or "expansion" signal provided by the TCGF (19, 21, 22, 37).

Preliminary evidence now suggests that immune responses across minor locus histocompatibility barriers can have an antineoplastic effect. In human bone marrow transplants using HLA-identical sibling donors, stronger graft vs. host (GVH) reactions (presumably a minor locus-directed immune response) are associated with a decreased chance of recurrent leukemia post-transplant (38, 39). The patient reported here had biopsy proven clinical stage I acute GVH of the skin on day 52 that spontaneously disappeared, and developed extensive chronic GVH on day 258 that responded well to cyclophosphamide and prednisone. He is well and in remission 16 mo posttransplant. In mouse, transplantation of H-2 matched, but minor locus mismatched, bone marrow can cure a strain AKR host of leukemia if given under appropriate conditions (40). These conditions seem to include alloimmunization of the donor before transplant with tissues bearing the same minor locus incompatibilities as the leukemic host (Shih, C. Y., R. L. Truitt, A. V. Lefever, L. D. Tempelis, M. M. Bortin, 1983, *J. Cell. Biochem.*, 7A: 78, (Abstr.), 41). Thus, the "graft vs. leukemia" response may actually represent preferential destruction of leukemic blasts by minor locus reactive lymphocytes.

Current *in vitro* methods now make it possible to grow adequate numbers of human lymphocytes for potential adoptive immunotherapy (33). Preliminary testing shows no significant toxicity in primates or human patients of T cells grown in TCGF (42, 43). The results reported here of minor locus reactive T<sub>c</sub> stim-

ulated by HLA identical AML blasts and expanded in TCGF, coupled with data regarding minor locus-induced graft vs. leukemia in mouse and man, suggest that minor locus-reactive human lymphocytes be tested for toxicity and antileukemic efficacy in clinical trials of human adoptive immunotherapy.

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