

# NIH Public Access

**Author Manuscript** 

Med Hypotheses. Author manuscript; available in PMC 2014 August 01.

# Published in final edited form as:

Med Hypotheses. 2013 August ; 81(2): 274–278. doi:10.1016/j.mehy.2013.04.031.

# URINE miRNAs: POTENTIAL BIOMARKERS FOR MONITORING PROGRESION OF EARLY STAGES OF DIABETIC NEPHROPATHY

Yeyi Yang<sup>1</sup>, Li Xiao<sup>1</sup>, Jun Li<sup>1</sup>, Yashpal S. Kanwar<sup>2</sup>, Fuyou Liu<sup>1</sup>, and Lin Sun<sup>1</sup>

<sup>1</sup>Department of Nephropathy, The Second Xiangya Hospital, Kidney Institute of Central South University, Changsha, Hunan Province, China 410011

<sup>2</sup>Departments of Pathology & Medicine, Northwestern University, Chicago, Illinois, USA

## Abstract

With a steep increase in the incidence of type 1 and 2 diabetes globally, diabetic nephropathy (DN) has now become the leading cause of renal failure in the world. There are no suitable biomarkers for the diagnosis of early stages of DN. In recent years, tremendous efforts are being made worldwide to delineate the role of micro RNAs in the pathogenesis of DN. Circulating miRNAs in serum, plasma, urine and other body fluids, which reflect a response to various pathophysiological stresses, are being investigated in the context of diabetic nephropathy. Delineation of the changes in miRNA levels in patients with DN may lead to a better understanding of the progression of the disease. We present here an exhaustive survey of the miRNA literature, highlighting various studies performed over the last decade. The aim is to assess if changes in various miRNAs could correlate with the progression of diabetic nephropathy. Based on the survey, we found that miRNA-377, miRNA-192, miRNA-216/217 and miRNA-144 are increased in body fluids of patients with DN, while miRNA-21 and miRNA-375 are decreased. Overall, there are a very few miRNAs that are kidney specific, and although significant differences were observed in the urinary excretion of certain miRNAs, they were not correlative to their levels in the blood or plasma. Thus, it is completely plausible that urine-specific miRNAs could serve as novel biomarkers for the diagnosis of early stages of diabetic nephropathy.

#### Keywords

miRNA; diabetic nephropathy; biomarkers

# INTRODUCTION

Early stages of diabetic nephropathy (DN) are characterized by hyperfiltration, nephron enlargement and mesangial cell hypertrophy, which later on progress to glomerulosclerosis [1]. Initial stages of DN are associated with mild proteinuria that is traditionally described as

<sup>© 2013</sup> Elsevier Ltd. All rights reserved.

Correspondence Address: Lin Sun, M.D., Ph.D. Department of Nephrology, The Second Xiangya Hospital, Kidney Institute of Nephrology, Central South University, Changsha, Hunan Province, China 410011, sunlinnwu11@163.com.

*Conflict of interest:* No conflict of interest.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

microalbuminuria (albumin excretion: 30 - 300 mg/day), but with progression to overt nephropathy there is an associated increased risk of vascular disease and patient mortality [2]. Major pathophysiological mechanisms associated with DN complicate the outcome of DN, including activation of the renin-angiotensin-aldosterone system (RAAS), the protein kinase C (PKC) pathway, and upregulation of pro-inflammatory cytokines and various growth factors [3]. Angiotensin II and transforming growth factor- $\beta$  (TGF- $\beta$ ) are significant molecular mediators that influence DN pathogenesis, and have been suggested as potential targets for the development of therapeutics [4]. The generation of advanced glycation end products (AGEs) and increased oxidative stress in high glucose ambience are additional pathobiological processes that further exacerbate hyperglycemia-induced renal injury.

Currently, DN is the most common cause of end-stage renal disease (ESRD) worldwide, and approximately 40% of patients require renal replacement therapy. Early identification of patients who are prone to develop renal complications would be an important step for their better management during the clinical course of this disease process [5]. Microalbuminuria has been the standard method for diagnosis of early stages of DN, however, this method has some drawbacks. Microalbuminuria can develop when advanced changes have already set in, as assessed by renal biopsy examination [6]. Also, the immunoassay that measures microalbuminuria can only detect the immunoreactive form of albumin, and its nonimmunoreactive forms are undetectable by this method [7]. The quantitation of immunoreactive albumin by RIA (radioimmunoassay) fails to detect approximately 80% of the total albumin; less than 28% of the non-immunoreactive albumin in urine could be accounted for by the total combination of all other proteins measured by ELISA [8]. During the past decade, proteomics have become a powerful tool for the discovery of biomarkers for various disease processes. However, these procedures are less than ideal to monitor the progression of DN because they are very time-consuming and restricted to a select group of proteins, excluding those that are highly hydrophobic. Thus, novel improved biomarkers are necessary to monitor the progression of early stages of diabetic nephropathy [9]. MicroRNAs comprise 21 to 23 nucleotides, and bind to the 3'-untranslated regions (UTRs) of their target mRNAs in a stable manner [10]. MiRNAs modulate a wide range of biological functions, including oncogenesis, apoptosis, cardiac development and insulin secretion [11][12][13][14][15]. MiRNAs are present in wide variety of body fluids, and their levels in these fluids usually reflect a tissue specific injury or expression, which makes them ideal candidates for potential biomarkers [16]. For example, serum miRNA-21 is a novel biomarker for the diagnosis of esophageal squamous cell carcinoma [17]. In addition miRNAs are potential biomarkers for smoking-related interstitial fibrosis [18]. This suggests a strong relationship between the expression of tissue miRNAs and their levels in body fluids. MiRNAs in are found at high concentrations in body fluids [20]. Further, miRNAs are extremely stable and notably less susceptible to RNase. This protection from RNase degradation has been shown to be a result of packaging within cytoplasmic microvesicles, exosomes and apoptotic bodies, and also due to specific interactions with protective proteins [21].

#### URINE-SPECIFIC miRNAs

We hypothesized that the unique stability of miRNAs in various body fluids would reveal urine-specific miRNAs that would have some promise to serve as biomarkers. As urine is an easily available source for molecular markers such as RNA; novel, highly sensitive, and specific urine-based diagnostic tools are particularly attractive. An analysis of miRNAs in urine from bladder cancer patients showed higher ratios of miR-126:miR-152 and miR-182:miR-152 [22]. Urinary miRNAs can be derived from glomerular ultrafiltrate or excreted by the renal tubules. Oftentimes, their levels may be reflective of intrinsic tissue injury in kidney or urinary tract. Melkonyan *et al.* detected 22 different urinary miRNAs, but

none were kidney-specific, suggesting these miRNAs were transported in the plasma and were filtered across the glomerular capillary barrier; their isolation in an intact form from the urine indicated their potential to serve as biomarkers [23][24]. In recent years, many studies have shown that miRNAs play an important role in regulating glucose and lipid metabolism in diabetes, and their levels in blood or urine should be of some value in stratifying a given stage of the disease process and predicting its clinical course [25]. Argyropoulos *et al.* found micro-albuminuria is associated with decreased levels of miR-323b-5p and increased urine concentration of miR-429 in patients with long standing type 1 diabetes [26]. Interestingly, miR-323b-5p regulates Claudin-16, a key component of the tight junction in the thick ascending limb, [27], while miR-429 correlates with the level of proteinuria and renal function in immunologically-mediated renal diseases such as IgA nephropathy [28]. Finally, the five most significant miRNAs were found to be of immense value in classifying cases of DN with a high degree of significance [29].

#### MIRNAS AS POTENTIAL DISEASE BIOMARKERS

There are 2214 and 848 known miRNAs in humans and mice, respectively (Data from miRBase Release 19.0: 2013.02.at http://microRNA.sanger.ac.uk). Recent clinical studies have shown that human serum contains large amounts of stable miRNAs derived from various cells and tissues, and their altered expression profile in body fluids of type 2 diabetic patients is considered of high clinical value [30].

#### MiRNA-377 in diabetic nephropathy and pulmonary neoplasms

MiR-377 seems to be epigenetically regulated [31]. It is normally expressed in human lung tissues, and is upregulated in lung tumors, and thus could serve as a potential biomarker for the management of patients with pulmonary neoplasms [32]. Interestingly, in mouse and in vitro models for DN, elevated miR-377 levels were shown to mediate decreased expression of p21-activated kinase (PAK1) and superoxide dismutase (SOD), enhancing production of the fibronectin protein. PAK1 plays an important role in tumor formation [33] and can lead to fibronectin production via Smad activation in DN. Superoxide dismutase genes are related to oxidative stress, and may be associated with vascular damage in patients with diabetic mellitus [34]. Therefore, miR-377 may play a critical role in the pathobiology of mesangial cells since they are known to undergo oxidative stress under high glucose ambience.

#### MiRNA-192 in renal disease, hepatotoxicity and colon tumors

The value of miR-192, a liver-enriched miRNA, as a potential blood-based biomarker has been elucidated recently in a mouse model of acetaminophen (APAP)-induced hepatotoxicity [35]. The expression level of miR-192 is significantly decreased in colon tumors compared with normal tissues, suggesting that it could be used as a diagnostic tool to assess hepatotoxicity and neoplasms of the gastro-intestinal tract [36]. Urinary levels of miR-192 are downregulated in patients with IgA nephropathy [37]. Also, correlation of miRNA-192 levels with derangements in renal functional parameters and incremental increase in blood pressure has been recently reported [38]. More importantly, increased glomerular expression of miR-192 was found to be associated with heightened activity of renal TGF- $\beta$  in mouse models of DN. Furthermore, miR-192 could target ZEB1/2 in PTCs and enforced expression of E-cadherin while TGF-β led to a reduction in miR-192. ZEB1 (zinc finger E-box binding homeobox 1) and ZEB2 are E-Box-binding proteins and an important early stage of EMT [39]. E-cadherin expression was increased and remained after 96 hours of incubation with TGF- $\beta$  in PTC (proximal tubular epithelial cells) clones overexpressing miR-192 [40]. Interestingly, specific inhibition of renal miR-192 decreases renal fibrosis and dampens the proteinuric response [41]. Further support of miRNA-192 in the pathobiology of the kidney is derived from studies in patients having early stages of

diabetic nephropathy who had a higher expression of miR-192 compared to the late stages of clinical course of this disease process [42].

#### MiRNA-216 and miRNA-217 in DN and pancreatic ductal adenocarcinoma

MiR-216 and miR-217 are characteristically expressed in the pancreatic tissue. A uniquely controlled expression of miR-217 in the pancreatic tissues may be an additional mechanism for acinar cells to balance their  $\beta$ -catenin expression levels and secretory functions. miR-216 and miR-217 play important roles in early detection of pancreatic ductal adenocarcinoma, a condition known for its difficulty to diagnose [43]. The relevance of miR-216 and miR-217 in chronic kidney diseases has also been described (reviewed in [44]). Interestingly, miR-216 and miR-217 also play a role in the activation of Akt kinase, a key mediator of diabetic nephropathy. Specifically, TGF- $\beta$  was shown to activate Akt kinase by inducing miR-216 and miR-217, in turn downregulating phosphatase and tensin homologue (PTEN). Overexpression of PTEN has been observed in early stages of DN, adding significance to the role of miR-216 and miR-217. Intriguingly, these miRNAs are also upregulated by miRNA-192, a finding that is also relevant in the pathogenesis of diabetic nephropathy [45].

#### miRNA-144 in type 2 diabetes and pancreatic cancer

miRNA-144 has been shown to play a role in regulation of cell growth [46] and apoptosis [47]. MiR-144 is a regulator of genes that modulate embryonic-hemoglobin, and is downregulated in pancreatic cancer [48]. RT-qPCR analyses showed that miR-144 is overexpressed in colorectal cancerous tissues, thus yielding the possibility to serve as a clinical diagnostic marker [49]. Increased circulating levels of miR-144 also correlate with the downregulation of its predicted target: insulin receptor substrate 1 (IRS1), at both mRNA and protein levels; thus suggesting that miR-144 may also be a potential biomarker for type 2 diabetes (T2D) [50].

#### MiRNA-21 in DN and solid tumors

MiRNA-21 is commonly upregulated in solid tumors of the lung, breast, stomach, prostate, colon, brain, head and neck, esophagus, pancreas, and kidney [51]. Its overexpression inhibits proliferation of mesangial cells, and decreases urinary excretion of albumin in diabetic db/db mice. This suggests that miR-21 plays a protective role in glomerular hypertrophy and the onset of early DN. What is more interesting is that like miR-216a and miR-217, PTEN is also a potential target of miR-21 [52]. In addition, miR-21 prevented mesangial hypertrophy by targeting the PTEN/PI3K/Akt pathway. The main biological function of PTEN is to block PI3K signaling, a crucial pathway involved in metabolic responses to insulin [53]. Binding of insulin to the insulin receptor activates the PI3K pathway, while defects in PI3K signaling have been demonstrated in type 2 diabetes mellitus [54]. Altered PTEN expression is associated with diabetic nephropathy (DN), and miR-21 expression is downregulated in early stages of DN. Thus, it is conceivable that overexpression of miR-21 could prevent renal mesangial cell hypertrophy via downregulating PTEN in diabetic nephropathy [55].

#### MiRNA-375 in type 2 diabetes and colorectal cancer

While miRNA-375 expression is decreased in colorectal cancerous tissues, there is no significant correlation between the expression of miRNA-375 with tumor size, histological grade, or the tumor stage [56]. Like miRNA-144, miRNA-375 also inhibits tumor growth and metastasis of esophageal squamous cell carcinoma by repressing insulin-like growth factor 1 receptor (IGF-R1) [57]. Another study reported that pancreatic miR-375 expression was increased in type 2 diabetic (T2D) patients, and upregulated in pancreatic islet amyloid formation and A-cell deficit [24]. Of great interest here is that miR-375 seems to be an

important regulator of insulin secretion. It is a pancreatic islet cell specific miRNA in mice, and it upregulates insulin secretion by targeting myotropin [58]. Furthermore, a decrease of miR-375 levels has been seen in high glucose ambience. Such a regulation of miR-375 expression by glucose is also observed in rat islet cells[59]. In addition, miR-375 regulates PDK1 protein levels, by interacting directly with the 3' UTR, resulting in modulation of glucose stimulatory action on insulin gene expression and DNA synthesis. This is significant as PDK1 can regulate cell growth and organ development [60]. Finally, miR-375 expression is decreased in diabetic GK rat islets, thus emphasizing the significance of miR-375 in the pathophysiology of diabetes in rodents and humans [61] (Table 1).

This suggests that kidneys are not involved in the physiological urinary clearances of circulating miRNAs. The only exception to this is miR-638.. In general, it seems that in patients with severe chronic renal failure, the blood circulating levels of total and specific miRNAs are reduced in comparison to patients with mild renal impairment or normal renal functions. Intriguingly also the rates of *ex vivo* microRNA degradation of blood/plasma miR-210, miR-16 and miR-21 were higher in patients with end-stage renal disease (ESRD) compared to those individuals with normal renal functions [62].

# **HYPOTHESIS**

The above reviewed literature suggests that miRNAs play an important role in the pathogenesis of diabetic nephropathy and in the initiation of renal glomerular mesangial cell dysfunctions. Conceivably, miRNAs can modulate the pathogenesis of DN by affecting various different pathways. We hypothesize that urinary excretion of miR-377, miR-192, miR-216/217 and miR-144 increase in patients with diabetic nephropathy as they may exacerbate the perturbations in body homeostasis in a hyperglycemic milieu. On the other hand, the urinary excretion of miR-21 and miR-375 may decrease as they bind to their various tissue targets and henceforth serve to preserve body homoestasis in patients with diabetic nephropathy. In line with this contention is the study by Melkonyan et al., where 22 different urinary miRNAs were identified, but none of them specific for kidney tissues [19]. It is likely that miRNAs in urine versus in blood has no significant impact on a given disease characteristics, except miRNA-638 [56], which showed a significant increase in the urine of patients with Stage 4 CKD compared to normal and Stage 3 CKD patients. Neal et al. recently found a striking reduction in the overall levels of circulating miRNAs in patients with severe chronic kidney disease (CKD) having marked impairment in renal functions [62].

## CONCLUSIONS

Several miRNAs have now been identified, which may be upregulated or downregulated in the progression of DN, and their detection in very early stages may be of value in predicting the disease course. In addition, increasing the threshold of detection of miRNAs by various amplification methods and at the same time delineating miRNA tissue-restricted expression profiles will be very helpful in advancing this field by comprehensively determining their relevance in the pathogenesis of diabetic nephropathy.

Urine proteomic profiling studies have identified normoalbuminuric subjects with type 2 diabetes who subsequently develop diabetic nephropathy. As a further step, in this review we discuss a group of miRNAs that could serve as biomarkers in DN, including: miR-377, miR-21, miR-192, miR-216a, miR-217, miR-375 and miR-144. The blood levels of some of these miRNAs are increased in DN, suggesting they may be of some value in monitoring the progress of diabetic nephropathy. Whereas some of the other miRNAs in blood circulation are consumptively decreased as they bind to their targets in various tissues so as to maintain

body homeostasis in early stages of DN. Of course, there are also some changes in the urinary excretion of certain miRNAs in patients with diabetic nephropathy; for instance, there is an upregulation of miR-638 in the urine specimens. Further studies are needed to characterize miRNAs that are highly specific to DN in order to understand their role in the pathogenesis of diabetic nephropathy.

#### Acknowledgments

Supported by grants from the Creative Research Group Fund of the National Foundation Committee of Natural Sciences of China (30871169, 81100541), Doctoral Fund of Ministry of Education of China (20110162110012), Furong Scholars Fund from Hunan Province Education Department and Grants from the NIH DK 60635. We thank Dr. Elisabeth I. Wallner for proof reading the manuscript. We also thank Medjaden Bioscience Limited for assisting in the preparation of this manuscript.

#### References

- Mason RM, Wahab NA. Extracellular matrix metabolism in diabetic nephropathy. J Am Soc Nephrol. 2003; 14:1358–1373. [PubMed: 12707406]
- 2. Satchell SC, Tooke JE. What is the mechanism of microalbuminuria in diabetes: a role for the glomerular endothelium? Diabetologia. 2008; 51:714–725. [PubMed: 18347777]
- Navarro-Gonzalez JF, Mora-Fernandez C. The role of inflammatory cytokines in diabetic nephropathy. J Am Soc Nephrol. 2008; 19:433–442. [PubMed: 18256353]
- Zhu Y, Usui HK, Sharma K. Regulation of transforming growth factor beta in diabetic nephropathy: implications for treatment. Semin Nephrol. 2007; 27:153–160. [PubMed: 17418684]
- Parving HH, Lehnert H, Brochner-Mortensen J, Gomis R, Andersen S, Arner P. The effect of irbesartan on the development of diabetic nephropathy in patients with type 2 diabetes. N Engl J Med. 2001; 345:870–878. [PubMed: 11565519]
- Jin J, Ku YH, Kim Y, Kim K, Lee JY, Cho YM, et al. Differential Proteome Profiling Using iTRAQ in Microalbuminuric and Normoalbuminuric Type 2 Diabetic Patients. Exp Diabetes Res. 2012:168602. [PubMed: 22536212]
- Liu R, Li G, Cui XF, Zhang DL, Yang QH, Mu XY, et al. Methodological evaluation and comparison of five urinary albumin measurements. J Clin Lab Anal. 2011; 25:324–329. [PubMed: 21919065]
- Clavant, Steven P.; Sastra, Steve A.; Osicka, Tanya M.; Comper, Wayne D. The analysis and characterisation of immuno-unreactive urinary albumin in healthy volunteers. Clinical Biochemistry. 2006; 39:143–151. [PubMed: 16325791]
- Thongboonkerd V. Study of diabetic nephropathy in the proteomic era. Contrib Nephrol. 2011; 170:172–183. [PubMed: 21659770]
- Kuokkanen S, Chen B, Ojalvo L, Benard L, Santoro N, Pollard JW. Genomic profiling of microRNAs and messenger RNAs reveals hormonal regulation in microRNA expression in human endometrium. Biol Reprod. 2010; 82:791–801. [PubMed: 19864316]
- Divakaran V, Mann DL. The emerging role of microRNAs in cardiac remodeling and heart failure. Circ Res. 2008; 103:1072–1083. [PubMed: 18988904]
- Youssef YM, White NM, Grigull J, Krizova A, Samy C, Mejia-Guerrero S, et al. Accurate molecular classification of kidney cancer subtypes using microRNA signature. Eur Urol. 2011; 59:721–30. [PubMed: 21272993]
- Chen YQ, Wang XX, Yao XM, Zhang DL, Yang XF, Tian SF, et al. Abated microRNA-195 expression protected mesangial cells from apoptosis in early diabetic renal injury in mice. J Nephrol. 2012; 25:566–76. [PubMed: 21983986]
- Feng Y, Yu X. Cardinal roles of miRNA in cardiac development and disease. Sci China Life Sci. 2011; 54:1113–20. [PubMed: 22227903]
- Roggli E, Gattesco S, Caille D, Briet C, Boitard C, Meda P. Changes in microRNA expression contribute to pancreatic β-cell dysfunction in prediabetic NOD mice. Diabetes. 2012; 61:1742–51. [PubMed: 22537941]

- Chen, Xi; Ba, Yi; Ma, Lijia; Cai, Xing; Yin, Yuan; Wang, Kehui. Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. Cell Research. 2008; 18:997–1006. [PubMed: 18766170]
- 17. Kurashige J, Kamohara H, Watanabe M, Tanaka Y, Kinoshita K, Saito S, et al. Serum microRNA-21 is a novel biomarker in patients with esophageal squamous cell carcinoma. J Surg Oncol. 2012; 21 10.1002.
- 18. Huang Y, Dai Y, Zhang J, Wang C, Li D, Cheng J, et al. Circulating microRNAs as potential biomarkers for smoking-related interstitial fibrosis. Biomarkers. 2012 10.3109.
- Gantier MP, McCoy CE, Rusinova I, Saulep D, Wang D, Xu D, et al. Analysis of microRNA turnover in mammalian cells following Dicer1 ablation. Nucleic Acids Res. 2011; 39:5692–5703. [PubMed: 21447562]
- Cortez MA, Bueso-Ramos C, Ferdin J, Lopez-Berestein G, Sood AK, Calin GA. MicroRNAs in body fluids--the mix of hormones and biomarkers. Nat Rev Clin Oncol. 2011; 8:467–477. [PubMed: 21647195]
- 21. Wang K, Zhang S, Weber J, Baxter D, Galas DJ. Export of microRNAs and microRNA-protective protein by mammalian cells. Nucleic Acids Res. 2010; 38:7248–7259. [PubMed: 20615901]
- 22. Hanke M, Hoefig K, Merz H, Feller AC, Kausch I, Jocham D, et al. A robust methodology to study urine microRNA as tumor marker: microRNA-126 and microRNA-182 are related to urinary bladder cancer. Urologic Oncology-Seminars and Original Investigations. 2010; 28:655–661.
- Melkonyan HS, Feaver WJ, Meyer E, Scheinker V, Shekhtman EM, Xin Z, et al. Transrenal nucleic acids: from proof of principle to clinical tests. Ann N Y Acad Sci. 2008; 1137:73–81. [PubMed: 18837928]
- 24. Yamada Y, Enokida H, Kojima S, Kawakami K, Chiyomaru T, Tatarano S, et al. MiR-96 and miR-183 detection in urine serve as potential tumor markers of urothelial carcinoma: correlation with stage and grade, and comparison with urinary cytology. Cancer Science. 2011; 102:522–529. [PubMed: 21166959]
- Ciesla M, Skrzypek K, Kozakowska M, Loboda A, Jozkowicz A, Dulak J. MicroRNAs as biomarkers of disease onset. Analytical and Bioanalytical Chemistry. 2011; 401:2051–2061. [PubMed: 21544542]
- 26. Argyropoulos C, Wang K, McClarty S, Huang D, Bernardo J, Ellis D, et al. Urinary MicroRNA Profiling in the Nephropathy of Type 1 Diabetes. PLoS One. 2013; 8:e54662. 10.1371. [PubMed: 23358711]
- Hou J, Shan Q, Wang T, Gomes AS, Yan Q, et al. Transgenic RNAi Depletion of Claudin-16 and the Renal Handling of Magnesium. Journal of Biological Chemistry. 2007; 282:17114–17122. [PubMed: 17442678]
- Wang G, Kwan BC-H, Lai FM-M, Chow K-M, Kam-Tao Li P, et al. Expression of microRNAs in the urinary sediment of patients with IgA nephropathy. Dis Markers. 2010; 28:79–86. [PubMed: 20364043]
- Zampetaki A, Kiechl S, Drozdov I, Willeit P, Mayr U, Prokopi M, et al. <a>Plasma MicroRNA Profiling Reveals Loss of Endothelial MiR-126 and Other MicroRNAs in Type 2 Diabetes. Circulation Research. 2010; 107:810–U359. [PubMed: 20651284]
- Zhao HL, Guan J, Lee HM, Sui Y, He L, Siu JJ, et al. Up-Regulated Pancreatic Tissue MicroRNA-375 Associates With Human Type 2 Diabetes Through beta-Cell Deficit and Islet Amyloid Deposition. Pancreas. 2010; 39:843–846. [PubMed: 20467341]
- Zhang, Lin; Volinia, Stefano; Bonome, Tomas; Calin, George Adrian; Greshock, Joel; Yang, Nuo, et al. Genomic and epigenetic alterations deregulate microRNA expression in human epithelial ovarian cancer. PNAS. 2008; 105:7004–7009. [PubMed: 18458333]
- Melkamu T, Zhang XX, Tan JK, Zeng Y, Kassie F. Alteration of microRNA expression in vinyl carbamate-induced mouse lung tumors and modulation by the chemopreventive agent indole-3carbinol. Carcinogenesis. 2010; 31:252–258. [PubMed: 19748927]
- Chow HY, Jubb AM, Koch JN, Jaffer ZM, Stepanova D, Campbell DA, et al. p21-Activated kinase 1 is required for efficient tumor formation and progression in a Ras-mediated skin cancer model. Cancer Res. 2012; 72:5966–75. [PubMed: 22983922]

- Temneanu RO, Motoc A, Zugun FE, Folescu R, Lupu oru CE, Zamfir CL. The relevance of circadian rhythms disruption on pulmonary SOD expression in rat. Rom J Morphol Embryol. 2012; 53:789–93. [PubMed: 23188441]
- Wang K, Zhang S, Marzolf B, Troisch P, Brightman A, Hu Z, Hood LE, et al. Circulating microRNAs, potential biomarkers for drug-induced liver injury. Proc Natl Acad Sci USA. 2009:4402–4407. [PubMed: 19246379]
- 36. Karaayvaz M, Pal T, Song B, Zhang C, Georgakopoulos P, Mehmood S, et al. Prognostic significance of miR-215 in colon cancer. Clin Colorectal Cancer. 2011; 10:340–347. [PubMed: 21752725]
- Wang G, Kwan BCH, Lai FMM, Chow KM, Li PKT, Szeto CC. Expression of microRNAs in the urinary sediment of patients with IgA nephropathy. Disease Markers. 2010; 28:79–86. [PubMed: 20364043]
- Wang G, Kwan BC, Lai FM, Choi PC, Chow KM, Li PK, Szeto CC. Intrarenal expression of miRNAs in patients with hypertensive nephrosclerosis. Am J Hypertens. 2010; 23:78–84. [PubMed: 19910931]
- Wang B, Herman-Edelstein M, Koh P, Burns W, Jandeleit-Dahm K, Watson A, et al. E-cadherin expression is regulated by miR-192/215 by a mechanism that is independent of the profibrotic effects of transforming growth factor-beta. Diabetes. 2010; 59:1794–802. [PubMed: 20393144]
- 40. Krupa A, Jenkins R, Luo DD, Lewis A, Phillips A, Fraser D. Loss of MicroRNA-192 promotes fibrogenesis in diabetic nephropathy. J Am Soc Nephrol. 2010; 21:438–47. [PubMed: 20056746]
- Putta S, Lanting L, Sun G, Lawson G, Kato M, Natarajan R. Inhibiting microRNA-192 ameliorates renal fibrosis in diabetic nephropathy. J Am Soc Nephrol. 2012; 23:458–469. [PubMed: 22223877]
- Krupa A, Jenkins R, Luo DD, Lewis A, Phillips A, Fraser D. Loss of MicroRNA-192 Promotes Fibrogenesis in Diabetic Nephropathy. Journal of the American Society of Nephrology. 2010; 21:438–447. [PubMed: 20056746]
- Szafranska AE, Davison TS, John J, Cannon T, Sipos B, Maghnouj A, et al. MicroRNA expression alterations are linked to tumorigenesis & non-neoplastic processes in pancreatic ductal adenocarcinoma. Oncogene. 2007; 26:4442–4452. [PubMed: 17237814]
- 44. Li JY, Yong TY, Michael MZ, Gleadle JM. Review: The role of microRNAs in kidney disease. Nephrology (Carlton). 2010; 15:599–608. [PubMed: 20883280]
- 45. Kato M, Putta S, Wang M, Yuan H, Lanting L, Nair I, et al. TGF-beta activates Akt kinase through a microRNA-dependent amplifying circuit targeting PTEN. Nat Cell Biol. 2009; 11:881–889. [PubMed: 19543271]
- 46. Liu Y, Wang X, Jiang J, Cao Z, Yang B, Cheng X, et al. Modulation of T cell cytokine production by miR-144\* with elevated expression in patients with pulmonary tuberculosis. Mol Immunol. 2011; 48:1084–90. [PubMed: 21367459]
- 47. Huang F, Huang XY, Yan DS, Zhou X, Yang DY, et al. MicroRNA-144 over-expression induced myocytes apoptosis. Zhonghua Xin Xue Guan Bing Za Zhi. 2011; 39:353–7. [PubMed: 21624313]
- Sureban SM, May R, Lightfoot SA, Hoskins AB, Lerner M, Brackett DJ, et al. DCAMKL-1 Regulates Epithelial-Mesenchymal Transition in Human Pancreatic Cells through a miR-200a-Dependent Mechanism. Cancer Research. 2011; 71:2328–2338. [PubMed: 21285251]
- Kalimutho M, Del Vecchio Blanco G, Di Cecilia S, Sileri P, Cretella M, Pallone F, et al. Differential expression of miR-144\* as a novel fecal-based diagnostic marker for colorectal cancer. J Gastroenterol. 2011; 46:1391–1402. [PubMed: 21863218]
- Karolina DS, Armugam A, Tavintharan S, Wong MT, Lim SC, Sum CF, et al. MicroRNA 144 impairs insulin signaling by inhibiting the expression of insulin receptor substrate 1 in type 2 diabetes mellitus. PLoS One. 2011; 6:e22839. [PubMed: 21829658]
- 51. Chow TFF, Youssef YM, Lianidou E, Romaschin AD, Honey RJ, Stewart R, et al. Differential expression profiling of microRNAs and their potential involvement in renal cell carcinoma pathogenesis. Clinical Biochemistry. 2010; 43:150–158. [PubMed: 19646430]
- Zhang Z, Peng H, Chen J, Chen X, Han F, Xu X, et al. MicroRNA-21 protects from mesangial cell proliferation induced by diabetic nephropathy in db/db mice. FEBS Lett. 2009; 583:2009–2014. [PubMed: 19450585]

- Zhao H, Yang J, Fan T, Li S, Ren X. RhoE functions as a tumor suppressor in esophageal squamous cell carcinoma and modulates the PTEN/PI3K/Akt signaling pathway. Tumour Biol. 2012; 33:1363–74. [PubMed: 22477709]
- 54. O'Connor JC, Sherry CL, Guest CB. Freund GG. Type 2 diabetes impairs insulin receptor substrate-2-mediated phosphatidylinositol 3-kinase activity in primary macrophages to induce a state of cytokine resistance to IL-4 in association with overexpression of suppressor of cytokine signaling-3. J Immunol. 2007; 178:6886–93. [PubMed: 17513737]
- Mahimainathan L, Das F, Venkatesan B, Choudhury GG. Mesangial cell hypertrophy by high glucose is mediated by downregulation of the tumor suppressor PTEN. Diabetes. 2006; 55:2115– 2125. [PubMed: 16804083]
- 56. Dai X, Chiang Y, Wang Z, Song Y, Lu C, Gao P, et al. Expression levels of microRNA-375 in colorectal carcinoma. Mol Med Report. 2012; 5:1299–1304.
- 57. Kong KL, Kwong DL, Chan TH, Law SY, Chen L, Li Y, et al. MicroRNA-375 inhibits tumour growth and metastasis in oesophageal squamous cell carcinoma through repressing insulin-like growth factor 1 receptor. Gut. 2012; 61:33–42. [PubMed: 21813472]
- Kaucsar T, Racz Z, Hamar P. Post-transcriptional gene-expression regulation by micro RNA (miRNA) network in renal disease. Adv Drug Deliv Rev. 2010; 62:1390–1401. [PubMed: 20940025]
- 59. El Ouaamari A, Baroukh N, Martens GA, Lebrun P, Pipeleers D, van Obberghen E, et al. miR-375 targets 3'-phosphoinositide-dependent protein kinase-1 and regulates glucose-induced biological responses in pancreatic beta-cells. Diabetes. 2008; 57:2708–17. [PubMed: 18591395]
- Fan R, Kim NG, Gumbiner BM. Regulation of Hippo pathway by mitogenic growth factors via phosphoinositide 3-kinase and phosphoinositide-dependent kinase-1. Proc Natl Acad Sci USA. 2013; 110:2569–74. [PubMed: 23359693]
- 61. El Ouaamari A, Baroukh N, Martens GA, Lebrun P, Pipeleers D, van Obberghen E. miR-375 targets 3'-phosphoinositide-dependent protein kinase-1 and regulates glucose-induced biological responses in pancreatic beta-cells. Diabetes. 2008; 57:2708–2717. [PubMed: 18591395]
- Neal CS, Michael MZ, Pimlott LK, Yong TY, Li JY, Gleadle JM. Circulating microRNA expression is reduced in chronic kidney disease. Nephrol Dial Transplant. 2011; 26:3794–3802. [PubMed: 21891774]

#### Table 1

#### Disease-associated miRNAs

miRNA	Location	Change	Disease Associations	Reference
miRNA377	Lung	Upregulation	Pulmonary neoplasm	Melkamu <i>et al.</i> [26]
	Kidney		Diabetic nephropathy	Zhao et al. [25]
miRNA192	Blood	Upregulation	Hepatotoxicity	Wang et al. [30]
	Colon		Colonic tumors	Karayvaz et al. [31]
	Urine		IgA nephropathy	Wang et al. [32]
	kidney		Diabetic nephropathy	Krupa et al. [37]
miRNA216/217	Pancreas	Upregulation	Pancreatic ductal adenocarcinoma	Szafranska <i>et al.</i> [38]
	Kidney		Diabetic nephropathy	Kato <i>et al.</i> [40]
miRNA144	Colorectal tissue	Upregulation	Colorectal cancer	Kalimutho et al. [42]
	Blood		Type 2 diabetes	Karolina <i>et al.</i> [43]
miRNA21	Lung, breast, stomach, prostate, colon, etc.	Upregulation	Lung, breast, stomach, prostate, colon tumor	Chow <i>et al.</i> [44]
	Kidney	Downregulation	Diabetic nephropathy	Mahimainathan et al. [48]
miRNA375	Colorectal tissue	Downregulation	Colorectal cancer	Dai <i>et al.</i> [49]
	Esophageal tissue		Esophageal squamous carcinoma	Kong et al. [50]
	islet cells		Type 2 diabetes	El Ouaamari et al. [53]
miRNA638	kidney	Upregulation	Chronic kidney disease	Neal et al. [54]