

NIH Public Access Author Manuscript

Eur Urol. Author manuscript; available in PMC 2014 February 01

Published in final edited form as:

Eur Urol. 2013 February ; 63(2): 244–251. doi:10.1016/j.eururo.2012.09.054.

Metabolism of Kidney Cancer: From the Lab to Clinical Practice

Sunil Sudarshan^{1,*}, Jose Karam², James Brugarolas³, Houston Thompson⁴, Robert Uzzo⁵, Brian Rini⁶, Vitaly Margulis⁷, Jean-Jacques Patard⁸, Bernard Escudier⁹, and W. Marston Linehan¹⁰

¹Department of Urology, University of Alabama at Birmingham, Birmingham, AL, USA

²Department of Urology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

³Department of Internal Medicine, University of Texas Southwestern Medical Center, Dallas, TX, USA

⁴Department of Urology, Mayo Clinic, Rochester, MN, USA

⁵Department of Surgery, Urologic Oncology, Fox Chase Cancer Center, Philadelphia, PA, USA

⁶Department of Solid Tumor Oncology, The Cleveland Clinic Taussig Cancer Center, Cleveland, OH, USA

⁷Department of Urology, University of Texas Southwestern Medical Center, Dallas, TX, USA

⁸Department of Urology, CHU Bicêtre, Le Kremlin Bicêtre, France

⁹Department of Medical Oncology, Institut Gustave Roussy, Villejuif, France

¹⁰Urologic Oncology Branch, US National Cancer Institute, Bethesda, MD, USA

Abstract

- Patard, Sudarshan, Linehan.
- Statistical analysis: None.
- Obtaining funding: None.

Administrative, technical, or material support: None.

^{© 2012} European Association of Urology. Published by Elsevier B.V. All rights reserved.

^{*}Corresponding author. Department of Urology, University of Alabama at Birmingham, Birmingham, AL, 35294, USA. Tel.: +1 205 603 9330. sudarshan@uab.edu (S. Sudarshan).

Author contributions: Sunil Sudarshan had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Study concept and design: Thompson, Margulis, Brugarolas, Karam, Uzzo, Rini, Escudier, Patard, Sudarshan, Linehan. Acquisition of data: Sudarshan.

Analysis and interpretation of data: Thompson, Margulis, Brugarolas, Karam, Uzzo, Rini, Escudier, Patard, Sudarshan, Linehan. Drafting of the manuscript: Thompson, Margulis, Brugarolas, Karam, Uzzo, Rini, Escudier, Patard, Sudarshan, Linehan. Critical revision of the manuscript for important intellectual content: Thompson, Margulis, Brugarolas, Karam, Uzzo, Rini, Escudier,

Supervision: Linehan.

Other (specify): None.

Financial disclosures: Sunil Sudarshan certifies that all conflicts of interest, including specific financial interests and relationships and affiliations relevant to the subject matter or materials discussed in the manuscript (eg, employment/affiliation, grants or funding, consultancies, honoraria, stock ownership or options, expert testimony, royalties, or patents filed, received, or pending), are the following: None.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Context—There is increasing evidence for the role of altered metabolism in the pathogenesis of renal cancer.

Objective—This review characterizes the metabolic effects of genes and signaling pathways commonly implicated in renal cancer.

Evidence acquisition—A systematic review of the literature was performed using PubMed. The search strategy included the following terms: *renal cancer, metabolism, HIF, VHL.*

Evidence synthesis—Significant progress has been made in the understanding of the metabolic derangements present in renal cancer. These findings have been derived through translational, in vitro, and in vivo studies. To date, the most well-characterized metabolic features of renal cancer are linked to von Hippel-Lindau (*VHL*) loss. *VHL* loss and the ensuing increase in the expression of hypoxia-inducible factor affect several metabolic pathways, including glycolysis and oxidative phosphorylation. Collectively, these changes promote a glycolytic metabolic phenotype in renal cancer. In addition, other histologic subtypes of renal cancer are also notable for metabolic derangements that are directly related to the causative genes.

Conclusions—Current knowledge of the genetics of renal cancer has led to significant understanding of the metabolism of this malignancy. Further studies of the metabolic basis of renal cell carcinoma should provide the foundation for the development of new treatment approaches and development of novel biomarkers.

1. Introduction

Renal cell carcinoma (RCC) is among the top 10 most common malignancies in both men and women. In 2012, it is estimated that approximately 64 000 individuals will be diagnosed, with an estimated 13 570 dying from this disease in the United States [1]. Although the incidence has been increasing in the United States, the rates have leveled off or even declined in other parts of the world [2].

Is it is now clear that altered metabolism plays a key role in the pathogenesis of many disease states. Moreover, there has been increasing recognition of the role of metabolism in the pathogenesis of cancer, including RCC. To date, most studies have examined the risk of RCC in disease with known metabolic links. For example, the rising incidence of RCC has paralleled the increase in body mass index, particularly in Western countries [3]. This finding has been validated by several recent studies that reported increased incidence of RCC in disease states with known effects on metabolism, including obesity and diabetes [4]. Additionally, links between atherosclerosis and the risk of RCC have been identified [5].

Collectively, these data point to the emerging link between metabolism and cancer. Correspondingly, treatment of these disease processes, such as statins for atherosclerosis, may be associated with a reduced risk of RCC [6]. With continued examination of this link, identification of metabolic factors may point to new directions for therapeutics as well as biomarkers for diagnosis and prognosis. This review will focus on the metabolic alterations in the more common variants of RCC and the clinical relevance of these alterations.

2. Evidence acquisition

We performed a systematic search of PubMed through May 2012. Search terms included *renal cancer, metabolism, HIF,* and *VHL*. Searches were limited to articles written in English. All authors screened the potential references retrieved by the search and a final consensus on all publications included in the present review was reached.

3. Evidence synthesis

Evidence related to the review and that examined general aspects of cancer metabolism was acquired. Next, metabolic aspects of RCC were analyzed based on the histologic subtype. Metabolic aspects of RCC are described in the context of genes commonly alerted for each subtype.

3.1. Cancer metabolism: general considerations

One of the most basic physiologic functions of cells is to convert nutrients into energy equivalents such as adenosine triphosphate (ATP). The most basic starting nutrient is glucose, which is broken down by a series of enzymatic reactions, referred to as glycolysis, to the metabolite pyruvate [7,8]. Under conditions where oxygen is present (ie, aerobic conditions), pyruvate is transferred to the mitochondria where it is converted to acetyl coenzyme A (acetyl coA). In turn, acetyl coA is metabolized in the tricarboxylic acid (TCA) cycle (also referred to as the citric acid cycle). This process yields reducing equivalents that drive the process of oxidative phosphorylation that, in turn, leads to ATP generation. Oxidative phosphorylation requires oxygen. Therefore, under low oxygen (anaerobic) conditions, oxidative phosphorylation is suppressed and pyruvate is converted to lactate via lactate dehydrogenase (LDH). One of the most notable alterations in cancer metabolism was initially described by Otto Warburg [9]. This observation, now referred to as the Warburg effect, refers to the increased rate of glycolysis with reduced oxidative phosphorylation in tumor tissues despite the presence of normal levels of oxygen. An unresolved question is whether the Warburg effect is merely the result of metabolic adaptations that tumor cells make to survive or whether this phenotype contributes to tumorigenesis. The finding that the loss of several tumor suppressor genes may promote the Warburg effect suggests the latter [8].

3.2. Clear cell renal cancer: the role of VHL

RCC is actually composed of several tumor types linked by the site of origin. Current understanding of the molecular biology of RCC has benefited significantly from the study of hereditary forms of renal cancer. Most notable is von Hippel-Lindau (VHL) disease. Patients with VHL disease have a genetic predisposition to the development of clear cell RCC (ccRCC) among other tumor types affecting organ sites, including the brain, spinal cord, pancreas, and adrenal gland [10]. Through genetic linkage performed in VHL families, the *VHL* gene was identified on the short arm of chromosome 3 and was eventually cloned [11,12]. Subsequent studies led to the finding that *VHL* alterations are common genetic events in sporadic ccRCC, indicating that, from the standpoint of tumor-initiating events, ccRCCs are a relatively homogenous population relative to other tumor types [13]. It is estimated that 70% to 90% of ccRCCs demonstrate alterations of the *VHL* gene either via mutation or gene silencing (ie, hypermethylation) [14,15]. As such, current understanding of the basic biology of ccRCC has been largely derived through the study of the functions of VHL protein.

3.3. The VHL pathway

Through protein interactions studies, it was subsequently determined that VHL was a member of the E3 ubiquitin ligases. VHL associates with the proteins elongin B, elongin C, and cullin 2 [16,17]. Together, the VHL complex ubiquitinates proteins, which targets them for degradation by the proteosome. Among the most studied targets of the VHL complex are the hypoxia-inducible factor (HIF) proteins HIF-1 α and HIF-2 α [18,19]. The labile HIF α proteins form a complex with the constitutively expressed HIF-1 β subunit [20]. Together, this complex binds to DNA motifs referred to as hypoxia response elements to regulate the expression of genes involved in angiogenesis, cell proliferation, and metabolism. Under

normal oxygen-containing (aerobic) conditions, HIFa proteins are hydroxylated on conserved proline residues by a series of enzymes referred to as prolyl hydroxylases [18,21]. Hydroxylation of HIF permits recognition by the VHL complex, which therefore targets these transcription factors for proteosomal degradation. Under hypoxic conditions when oxygen is limiting, HIFs remain unhydroxylated, and therefore stable, due to lack of recognition by VHL. HIFs are stabilized in the absence of VHL despite the presence of oxygen (referred to as a *pseudohypoxic state*). Alternatively, HIF may be stabilized under conditions in which proline hydroxylation of HIF is inhibited. Interestingly, recent data demonstrate the in vitro and in vivo antitumor efficacy of an HIF inhibitor in RCC cells [22].

3.4. Hypoxia-inducible factor: a master regulator of renal cell carcinoma metabolism

As VHL loss is a common event in ccRCC, a significant proportion of the current understanding of this tumor type has been derived from the study of HIF biology. In addition to effects on angiogenesis and growth factor expression, HIF has now been shown to have profound effects on cellular metabolism (Fig. 1). HIF is known to regulate the expression of glucose transporters that regulate glucose uptake from the extracellular milieu. Increased expression of the glucose transporter 1, which promotes cellular glucose uptake, has been observed in VHL-deficient ccRCC [23]. Additionally, HIF has been shown to transcriptionally upregulate many of the genes that encode the enzymes of glycolysis, including hexokinase 1 and 2 (HK1 and HK2) and glyceraldehyde 3-phosphate dehydrogenase (GAPDH) [24,25]. HIF shunts cellular metabolism away from the TCA cycle/oxidative phosphorylation through multiple mechanisms. As noted previously, the end product of glycolysis is pyruvate, which has one of two fates: (1) It may be converted to lactate via lactate dehydrogenase (LDH), or (2) it may be converted to acetyl CoA by the enzyme pyruvate dehydrogenase (PDH), which then proceeds through the reaction of the TCA cycle. HIF has been shown to upregulate LDH expression and thereby promote pyruvate's conversion to lactate. In addition, HIF shunts cellular metabolism away from the TCA cycle through the regulation of PDH [26,27]. PDH is subject to regulation by the enzyme pyruvate dehydrogenase kinase 1 (PDK1). Phosphorylation of PDH by PDK1 inactivates PDH, thereby blocking the conversion of pyruvate to acetyl CoA. Multiple studies have now demonstrated that HIF upregulates the expression of PDK1 [26,27]. Moreover, PDK-1 expression is increased in RCC [28]. Recent investigation has focused on the role of microRNAs (miRNAs) in cancer. miRNAs are noncoding RNA molecules that can regulate gene expression. HIF has previously been shown to regulate the expression of several miRNAs, including mir-210 [29]. Several reports have established that miR-210 is overexpressed in ccRCC [30-32]. Among the targets of miR-210 are the iron-sulfur cluster assembly proteins ISCU1 and ISCU2 [33]. ISCU1 and ISCU2 facilitate the assembly of iron-sulfur cluster proteins, including components of the electron transport chain that drives mitochondrial oxidative phosphorylation. Accordingly, miR-210 has been shown to block mitochondrial respiration. In agreement with these findings, ICSU1/ISCU2 expression is decreased in ccRCC as well as in animal models with genetic disruption of VHL [33,34]. Furthermore, constitutive HIF activation has been shown to inhibit mitochondrial respiration in vivo [35]. Interestingly, several recent studies have examined the potential for miRNAs as biomarkers for RCC [36-38].

3.5. The potential role of alternate clear cell renal cell carcinoma genes in metabolism

The recent utilization of deep-sequencing technologies has led to the identification of new ccRCC genes, many of which are involved in chromatin biology. Among the more commonly mutated genes in ccRCC is polybromo 1 (*PBRM1*)[39]. PBRM1 is a member of the switch/sucrose nonfermentable (SWI/SNF) protein complex that remodels chromatin [39]. Interestingly, gene-expression profiling of ccRCCs with *PBRM1* mutations falls into a

hypoxic gene profiling. Given the connection between hypoxia and metabolism as demonstrated through the studies on the VHL/HIF axis, it would seem likely that PBRM1 may have effects on cellular metabolism. In addition, several other genes, such as SET domain containing 2 (*SETD2*), that regulate histone modifications are mutated in a subset of ccRCC [40,41]. Given the role of histones in the regulation of gene expression, the function of these genes in the regulation of ccRCC metabolism warrants further investigation.

3.6. The IGF/IGF-1R axis and mechanistic target of rapamycin signaling in clear cell renal cell carcinoma

The type 1 insulin-like growth factor receptor (*IGF1R*) is commonly overexpressed in several tumor types, including RCC. Recent studies have demonstrated a role for *VHL* in the regulation of *IGF1R* expression in a HIF-independent fashion [42]. Downstream effectors of *IGF1R* include phosphoinositide-3-kinase (PI3K), which in turn activates AKT signaling. Additionally, PI3K regulates signaling by mechanistic target of rapamycin (mTOR).

MTOR's relevance to RCC has received much attention lately due to the emergence of mTOR inhibitors as US Food and Drug Administration-approved agents for the treatment of advanced renal cancer. The mTOR inhibitors used currently are analogues of the molecule rapamycin, which is an allosteric inhibitor of mTOR. MTOR is a key mediator of cellular physiology through effects on cellular metabolism. It serves as a nutrient sensor. Many tumor suppressor genes such as phosphatase and tensin homolog (PTEN), tuberous sclerosis 1 (TSC1), tuberous sclerosis 2 (TSC2), neurofibromin 1 (NF1), and liver kinase B1 (LKB1) are negative regulators of mTOR signaling [43]. Hence, loss or mutation of these genes may lead to aberrant mTOR activation. Hyperactivation of mTOR signaling has been observed in several cancer types, including RCC [44]. An enhanced understanding of the mTOR pathway has been derived from the study of patients with tuberous sclerosis (TS), who are at risk for the development of renal angiomyolipomas (AMLs). Patients with TS have germline mutations at either the TSC1 or TSC2 locus that encode for the proteins hamartin and tuberin, respectively. Hamartin and tuberin form a protein complex that blocks mTOR activation [43]. As such, loss of either TSC1 or TSC2 leads to mTOR activation. Consistent with this pathway are recent studies demonstrating the efficacy of sirolimus, a rapamycin analog and mTOR inhibitor, in the treatment of AMLs in patients with TS [45]. Of note are recent reports demonstrating mTOR and TSC1 mutations in ccRCC [46,47].

MTOR is known to regulate cell size and proliferation. mTOR can drive several metabolic processes, including fatty acid synthesis and the pentose phosphate pathway [48]. In addition, mTOR has been shown to promote glucose uptake and glycolysis, indicating that mTOR signaling promotes the Warburg phenotype in cancer. These latter effects are, in part, thought to be mediated by mTOR's effects on HIF-1a expression through promotion of HIF-1a mRNA translation [48,49]. Given mTOR's role in the regulation of metabolism, the efficacy of mTOR inhibition in renal cancer suggests the possibility of a metabolic *Achilles heel* of renal cancer, and that targeting the critical metabolic pathways may be of therapeutic benefit.

3.7. Metabolic aspects of papillary renal cancer

Papillary renal cancer accounts for approximately 15% of RCC and can be broadly classified as type 1 and type 2 papillary RCC (pRCC). As with ccRCC, our current understanding of pRCC has benefited from the study of families with hereditary papillary renal cancer (HPRC). Patients with HPRC harbor activating mutations of the met proto-oncogene (hepatocyte growth factor receptor) (*c-MET*) in the germline [50]. These individuals are predisposed to the development of type 1 pRCC. *c-MET* encodes for the receptor of hepatocyte growth factor [51]. MET is a receptor tyrosine kinase that may activate PI3K

signaling. Given the emerging role of PI3K signaling in cellular metabolism, pRCC type 1 cancers would be expected to demonstrate metabolic derangements. *c-MET* appears to have a role in sporadic cancer as well as mutations, and amplifications of this gene have been identified in a subset of tumors from patients with sporadic type 1 pRCC [52]. Of note is that c-MET inhibitors are currently being evaluated for efficacy in the treatment of pRCC.

Until recently, very little was known about metabolic aspects of type 2 pRCC. However, the report of genetic predisposition to type 2 pRCC of patients with germline mutations of the fumarate hydratase (*FH*) gene led to further understanding of the metabolic aspect of this disease [53]. *FH* encodes for the enzyme fumarate hydratase. This enzyme is part of the TCA cycle that contributes to oxidative phosphorylation and is responsible for converting fumarate to malate. As expected, tumors that lack *FH* have elevated levels of fumarate [54]. Consistent with the role of *FH* in the TCA cycle, cells lacking FH enzymatic activity are heavily reliant on glucose (and glycolysis) for cell survival relative to cells with intact *FH* and they demonstrate reduced oxidative phosphorylation [55,56]. *FH*-null tumor cells have recently been reported to demonstrate a Warburg phenotype [57]. Hence, targeting how these cells take up glucose and use it to generate energy equivalents required for cellular homeostasis may have therapeutic potential, as recently attempted [58].

Another interesting aspect of *FH*-null tumors is that they also demonstrate elevated levels of HIF protein expression (particularly HIF-1 α) [59]. As noted previously, HIF is usually very labile as it undergoes proline hydroxylation, which therefore targets it for proteosomal degradation. Fumarate has been shown to inhibit the proline hydroxylation of HIF and therefore lead to aberrant stabilization of HIF proteins [59]. Additionally, cellular models of *FH* loss demonstrated elevated levels of reactive oxygen species (ROS), which may also contribute to HIF stabilization [56]. The increase in ROS may be due to the highly glycolytic phenotype of these cells. Interestingly, two recent studies suggest that antioxidant pathways are activated in *FH*-null HPRC tumors as well as sporadic type 2 pRCC [60,61]. One possibility is that activation of antioxidant pathways may perhaps *cope* with the elevated levels of oxidative stress. From a therapeutic standpoint, ROS levels could be further raised above a certain threshold that would induce cell death in such tumor cells, as recently proposed by Sourbier et al[62].

3.8. Metabolic aspects of oncocytomas and chromophobe renal cell carcinoma

Recent evidence suggests that oncocytomas and chromophobe RCCs also have alterations in cellular metabolism relative to normal kidney. Gene-expression studies of these tumors demonstrate elevated levels of mitochondrial and oxidative phosphorylation gene expression in these tumors. These data are consistent with the observation that these tumor types have an abundance of mitochondria. In a similar fashion, tumors from patients with Birt-Hogg-Dubé (BHD) syndrome also demonstrate upregulation of mitochondrial/oxidative phosphorylation genes [63]. Patients with BHD syndrome are genetically predisposed to the development of chromophobe RCC, oncocytomas, and hybrid oncocytic tumors (tumors with a histologic mixture of both oncocytoma and chromophobe RCC) [64]. The implications of this finding as it pertains to tumor biology remains unknown at this time. However, it does underscore the finding that RCC is a spectrum of disease that varies at many levels, including metabolism.

Cellular studies of BHD syndrome demonstrate a potential link with 5' adenosine monophosphate-activated protein kinase (AMPK) [65]. APMK is a major cellular regulator of energy homeostasis and therefore a key metabolic regulator. AMPK is a negative regulator of mTOR. Interestingly, animal models of BHD syndrome demonstrate activation of mTOR signaling [66]. As such, mTOR inhibitors may have therapeutic potential in these

tumor types. Animal models of BHD syndrome demonstrate a renal cystic phenotype that can be abrogated by treatment with the mTOR inhibitor rapamycin.

3.9. Metabolomics in renal cancer

With an improved understanding of genetic alterations in RCC, particularly ccRCC, investigators are now in a position to exploit the ensuing metabolic alterations for clinical applications. Underlying gene defects such as VHL loss have profound impacts on the gene expression of many enzymes involved in key metabolic processes. As these enzymes interconvert small molecules, changes in the expression of these enzymes can affect levels of small molecules, that is, metabolites. The use of separation and identification technologies has now led to the development of metabolomics. Genomics, transcriptomics, and proteomics characterize a data set at the DNA, RNA, and protein levels, respectively. Metabolomics can characterize a biological sample at the level of small molecules (metabolites). Metabolomic analyses have generally applied two analytic approaches: nuclear magnetic resonance and mass spectrometry [67]. More recent approaches have used chromatography, either gas (GC) or liquid (LC), to separate metabolites in a biological sample from each other combined with mass spectrometry (MS) to allow for identification as well as quantification of molecules of interest. Given the relatively small number of molecules in the human metabolome (approximately 2200) as compared with the human genome and transcriptome, the potential for biomarker discovery exists for a variety of clinical applications [67].

The use of metabolomics for genitourinary malignancies has received considerable attention after the identification of the small molecule sarcosine as a potential mediator of prostate cancer progression. Using normal prostate tissue as well prostate cancer specimens (both localized and metastatic), Sreekumar et al were able to identify increasing levels of sarcosine as a function of cancer progression [68]. Moreover, they were able to identify sarcosine in urine samples of patients with prostate cancer. Metabolomics has begun to be applied to studies on renal cancer. Using GC-MS, Catchpole et al performed a metabolic profile of renal cancer tissues relative to normal tissue [69]. The primary histology analyzed in this data set was ccRCC. Consistent with a glycolytic phenotype representative of the Warburg effect, the levels of glucose were higher in RCC relative to normal tissue. In contrast, metabolites such as fumarate, malate, and succinate that participate in the mitochondrial TCA cycle/oxidative phosphorylation were reduced in RCC. These findings are consistent with the aforementioned metabolic biology of ccRCC. As most ccRCCs demonstrate loss of VHL, the enhanced HIF expression would be expected to promote glycolytic metabolism while downregulating mitochondrial metabolism. In addition, this study found markedly elevated levels (>5-fold) of α -tocopherol, a form of vitamin E, in RCC compared with normal tissue. As RCC cells are noted to have elevated levels of oxidative stress [70], the increased levels of α -tocopherol, which has antioxidant properties, may allow renal cancer cells to *cope* with increased levels of ROS. The potential for biomarker development was also investigated in this study. Using just α -tocopherol levels in a receiver operator curve analysis, this group was able to correctly classify almost 85% of tumors and >90% of normal tissues.

The potential for metabolomics in RCC biomarker development has also been applied to urine and serum analyses. Two recent analyses were able to distinguish patients with RCC from patients without based on the metabolic profile of urine specimens [71,72]. However, these studies did not identify the individual molecules that constituted this profile. Subsequent analysis by Kim et al was able to identify metabolites in the urine with differential levels in patients with RCC relative to unaffected controls [73]. Among the molecules identified with differential levels in patients with RCC relatives are involved in amino

acid and energetic metabolism. Zira et al recently reported on serum metabolomic analysis using nuclear magnetic resonance [74]. Using principal component analysis, they were able to identify a metabolic profile that could distinguish between patients with RCC and those with benign urologic disease. More recently, Lin et al were able to generate a metabolic profile using LC-MS of serum samples to distinguish with 100% sensitivity and 100% specificity patients with RCC from healthy volunteers [75]. In addition, they were able to identify candidate *metabolic* biomarkers with differential levels in patients with early and advanced stage disease. More recent studies have furthered the potential for metabolic biomarkers of RCC. Ganti et al implanted ccRCC cells (Caki-1 cell line) into nude mice and performed concurrent metabolic analysis of tumor tissue, serum, and urine [76]. Interestingly, they found that many metabolites that could be detected in tumor tissue could also be identified in serum and urine. Moreover, they found a general concordance of metabolite levels in tumor tissue and serum. These data suggest that serum metabolomics analysis may serve as a proxy for tumor metabolite alterations.

3.10. Altered metabolism in renal cancer: clinical implications

With recognition of the changes in metabolism characteristic of RCC histologic subtypes, and in particular ccRCC, opportunities are emerging for biomarker development that can assist with prognosis, diagnosis, and therapeutics. In some cases, the level of metabolic alteration may correlate with prognosis. Many of the studies have focused on HIF expression in RCC as a prognostic biomarker [77]. Interestingly, *VHL*-null ccRCC can be broadly characterized into two groups based on the expression of HIF-1 and HIF-2: (1) tumors that express both HIF-1 and HIF-2 and (2) tumors that express HIF-2 alone [78]. Recent data by Shen et al suggest that HIF-1 α may have a tumor suppressor function in ccRCC [79]. From a diagnostic standpoint, the gene expression of metabolic enzymes, or the metabolic profiles themselves, may assist in the diagnosis of renal cancer. A readily apparent application would be to assist in the interpretation of biopsy results of small renal masses. As up to 20% of biopsies in this setting may be nondiagnostic [80], the additional analysis of these metabolic parameters may improve the diagnostic yield of biopsy specimens.

Moreover, it may also help further stratify patients if additional metabolic prognostic factors could be assessed, once these are validated. The metabolic alterations found in ccRCC may also have therapeutic implications given recent evidence suggesting a correlation between tumor and serum metabolite levels in an animal model. Perhaps serum metabolite levels may serve as biomarker of tumor burden. This could be particularly helpful in the surveillance of patients who may be at high risk of recurrence. Alternatively, measurement of serum metabolite levels may be used to follow clinical response to therapy in patients with advanced/metastatic disease. For example, increases in serum cholesterol were recently reported to predict response to the mTOR inhibitor temsirolimus in patients with RCC [81]. At this time, serial imaging is the only reliable method of monitoring patients at high risk of recurrence and those on systemic therapy for advanced disease. Serum metabolite biomarkers theoretically could provide the advantage of assessing tumor burden in a more dynamic fashion. The technology currently exists to measure metabolites in serum. Hence, this could be easily incorporated into clinical practice once serum metabolites with differential levels in the presence or absence of RCC are identified and the measurements are standardized.

4. Conclusions

The last few decades have witnessed a marked advance in our understanding of the genetics of renal cancer, particularly ccRCC. ccRCC is remarkable for the fact that common genetic changes (ie, *VHL* loss) have dramatic effects on the metabolism that are biologically

relevant to the growth and survival of renal tumor cells. As such, further studies into ccRCC will provide a unique opportunity for researchers to apply understanding of tumor metabolism to clinically relevant applications. These characteristic changes in metabolism will provide avenues for the development of novel therapeutic strategies, perhaps in synergy with agents currently in use. Additionally, further understanding of RCC metabolism will likely open the door to biomarker development that will assist with diagnosis, prognosis, and monitoring therapeutic response.

Acknowledgments

Funding/Support and role of the sponsor: None.

References

- Siegel R, Naishadham D, Jemal A. Cancer statistics, 2012. CA Cancer J Clin. 2012; 62:10–29. [PubMed: 22237781]
- Chow WH, Dong LM, Devesa SS. Epidemiology and risk factors for kidney cancer. Nat Rev Urol. 2010; 7:245–57. [PubMed: 20448658]
- 3. McGuire BB, Fitzpatrick JM. BMI and the risk of renal cell carcinoma. Curr Opin Urol. 2011; 21:356–61. [PubMed: 21730854]
- Habib SL, Prihoda TJ, Luna M, Werner SA. Diabetes and risk of renal cell carcinoma. J Cancer. 2012; 3:42–8. [PubMed: 22232697]
- Hager M, Mikuz G, Kolbitsch C, Moser PL. Association between local atherosclerosis and renal cell carcinomas. Nutr Cancer. 2008; 60:364–7. [PubMed: 18444170]
- Liu W, Choueiri TK, Cho E. Statin use and the risk of renal cell carcinoma in 2 prospective US cohorts. Cancer. 2012; 118:797–803. [PubMed: 21751202]
- 7. Dang CV. Links between metabolism and cancer. Genes Dev. 2012; 26:877–90. [PubMed: 22549953]
- Levine AJ, Puzio-Kuter AM. The control of the metabolic switch in cancers by oncogenes and tumor suppressor genes. Science. 2010; 330:1340–4. [PubMed: 21127244]
- 9. Warburg O. On the origin of cancer cells. Science. 1956; 123:309-14. [PubMed: 13298683]
- Linehan WM, Lerman MI, Zbar B. Identification of the von Hippel-Lindau (VHL) gene. Its role in renal cancer. JAMA. 1995; 273:564–70. [PubMed: 7837390]
- Hosoe S, Brauch H, Latif F, et al. Localization of the von Hippel-Lindau disease gene to a small region of chromosome 3. Genomics. 1990; 8:634–40. [PubMed: 2276737]
- 12. Latif F, Tory K, Gnarra J, et al. Identification of the von Hippel-Lindau disease tumor suppressor gene. Science. 1993; 260:1317–20. [PubMed: 8493574]
- Gnarra JR, Tory K, Weng Y, et al. Mutations of the VHL tumour suppressor gene in renal carcinoma. Nat Genet. 1994; 7:85–90. [PubMed: 7915601]
- Kim WY, Kaelin WG. Role of VHL gene mutation in human cancer. J Clin Oncol. 2004; 22:4991– 5004. [PubMed: 15611513]
- Moore LE, Nickerson ML, Brennan P, et al. Von Hippel-Lindau (VHL) inactivation in sporadic clear cell renal cancer: associations with germline VHL polymorphisms and etiologic risk factors. PLoS Genet. 2011; 7:e1002312. [PubMed: 22022277]
- Kibel A, Iliopoulos O, DeCaprio JA, Kaelin WG Jr. Binding of the von Hippel-Lindau tumor suppressor protein to elongin B and C. Science. 1995; 269:1444–6. [PubMed: 7660130]
- Pause A, Lee S, Worrell RA, et al. The von Hippel-Lindau tumor-suppressor gene product forms a stable complex with human CUL-2, a member of the Cdc53 family of proteins. Proc Natl Acad Sci U S A. 1997; 94:2156–61. [PubMed: 9122164]
- Kaelin WG Jr. The von Hippel-Lindau protein, HIF hydroxylation, and oxygen sensing. Biochem Biophys Res Commun. 2005; 338:627–38. [PubMed: 16153592]
- Ivan M, Kondo K, Yang H, et al. HIFalpha targeted for VHL-mediated destruction by proline hydroxylation: implications for O2 sensing. Science. 2001; 292:464–8. [PubMed: 11292862]

- Li L, Kaelin WG Jr. New insights into the biology of renal cell carcinoma. Hematol Oncol Clin North Am. 2011; 25:667–86. [PubMed: 21763962]
- 21. Kaelin WG. Proline hydroxylation and gene expression. Ann Rev Biochem. 2005; 74:115–28. [PubMed: 15952883]
- 22. Carew JS, Esquivel JA 2nd, Espitia CM, et al. ELR510444 inhibits tumor growth and angiogenesis by abrogating HIF activity and disrupting microtubules in renal cell carcinoma. PLoS One. 2012; 7:e31120. [PubMed: 22295124]
- 23. Mandriota SJ, Turner KJ, Davies DR, et al. HIF activation identifies early lesions in VHL kidneys: evidence for site-specific tumor suppressor function in the nephron. Cancer Cell. 2002; 1:459–68. [PubMed: 12124175]
- Semenza GL. Regulation of cancer cell metabolism by hypoxia-inducible factor 1. Semin Cancer Biol. 2009; 19:12–6. [PubMed: 19114105]
- Semenza GL. HIF-1: upstream and downstream of cancer metabolism. Curr Opin Genet Devel. 2010; 20:51–6. [PubMed: 19942427]
- 26. Kim JW, Tchernyshyov I, Semenza GL, Dang CV. HIF-1-mediated expression of pyruvate dehydrogenase kinase: a metabolic switch required for cellular adaptation to hypoxia. Cell Metab. 2006; 3:177–85. [PubMed: 16517405]
- Papandreou I, Cairns RA, Fontana L, Lim AL, Denko NC. HIF-1 mediates adaptation to hypoxia by actively downregulating mitochondrial oxygen consumption. Cell Metab. 2006; 3:187–97. [PubMed: 16517406]
- 28. Baumunk D, Reichelt U, Hildebrandt J, et al. Expression parameters of the metabolic pathway genes pyruvate dehydrogenase kinase-1 (PDK-1) and DJ-1/PARK7 in renal cell carcinoma (RCC). World J Urol. In press. 10.1007/s00345-012-0874-5
- 29. Huang X, Ding L, Bennewith KL, et al. Hypoxia-inducible mir-210 regulates normoxic gene expression involved in tumor initiation. Mol Cell. 2009; 35:856–67. [PubMed: 19782034]
- Juan D, Alexe G, Antes T, et al. Identification of a microRNA panel for clear-cell kidney cancer. Urology. 2010; 75:835–41. [PubMed: 20035975]
- Valera VA, Walter BA, Linehan WM, Merino MJ. Regulatory effects of microRNA-92 (miR-92) on VHL gene expression and the hypoxic activation of miR-210 in clear cell renal cell carcinoma. J Cancer. 2011; 2:515–26. [PubMed: 22043236]
- White NM, Bao TT, Grigull J, et al. miRNA profiling for clear cell renal cell carcinoma: biomarker discovery and identification of potential controls and consequences of miRNA dysregulation. J Urol. 2011; 186:1077–83. [PubMed: 21784468]
- Chan SY, Zhang YY, Hemann C, Mahoney CE, Zweier JL, Loscalzo J. MicroRNA-210 controls mitochondrial metabolism during hypoxia by repressing the iron-sulfur cluster assembly proteins ISCU1/2. Cell Metab. 2009; 10:273–84. [PubMed: 19808020]
- 34. Neal CS, Michael MZ, Rawlings LH, Van der Hoek MB, Gleadle JM. The VHL-dependent regulation of microRNAs in renal cancer. BMC Med. 2010; 8:64. [PubMed: 20964835]
- Kucejova B, Sunny NE, Nguyen AD, et al. Uncoupling hypoxia signaling from oxygen sensing in the liver results in hypoketotic hypoglycemic death. Oncogene. 2011; 30:2147–60. [PubMed: 21217781]
- 36. Redova M, Poprach A, Nekvindova J, et al. Circulating miR-378 and miR-451 in serum are potential biomarkers for renal cell carcinoma. J Transl Med. 2012; 10:55. [PubMed: 22440013]
- 37. Von Brandenstein M, Pandarakalam JJ, Kroon L, et al. MicroRNA 15a, inversely correlated to PKCalpha, is a potential marker to differentiate between benign and malignant renal tumors in biopsy and urine samples. Am J Pathol. 2012; 180:1787–97. [PubMed: 22429968]
- 38. Zhai Q, Zhou L, Zhao C, et al. Identification of miR-508-3p and miR-509-3p that are associated with cell invasion and migration and involved in the apoptosis of renal cell carcinoma. Biochem Biophys Res Commun. 2012; 419:621–6. [PubMed: 22369946]
- 39. Varela I, Tarpey P, Raine K, et al. Exome sequencing identifies frequent mutation of the SWI/SNF complex gene PBRM1 in renal carcinoma. Nature. 2011; 469:539–42. [PubMed: 21248752]
- 40. Dalgliesh GL, Furge K, Greenman C, et al. Systematic sequencing of renal carcinoma reveals inactivation of histone modifying genes. Nature. 2010; 463:360–3. [PubMed: 20054297]

- 41. Duns G, Hofstra RM, Sietzema JG, et al. Targeted exome sequencing in clear cell renal cell carcinoma tumors suggests aberrant chromatin regulation as a crucial step in ccRCC development. Hum Mutat. 2012; 33:1059–62. [PubMed: 22461374]
- 42. Yuen JS, Cockman ME, Sullivan M, et al. The VHL tumor suppressor inhibits expression of the IGF1R and its loss induces IGF1R upregulation in human clear cell renal carcinoma. Oncogene. 2007; 26:6499–508. [PubMed: 17486080]
- Laplante M, Sabatini DM. mTOR signaling in growth control and disease. Cell. 2012; 149:274–93. [PubMed: 22500797]
- 44. Lin F, Zhang PL, Yang XJ, Prichard JW, Lun M, Brown RE. Morphoproteomic and molecular concomitants of an overexpressed and activated mTOR pathway in renal cell carcinomas. Ann Clin Lab Sci. 2006; 36:283–93. [PubMed: 16951269]
- 45. Bissler JJ, McCormack FX, Young LR, et al. Sirolimus for angiomyolipoma in tuberous sclerosis complex or lymphangioleiomyomatosis. N Engl J Med. 2008; 358:140–51. [PubMed: 18184959]
- 46. Gerlinger M, Rowan AJ, Horswell S, et al. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. N Engl J Med. 2012; 366:883–92. [PubMed: 22397650]
- 47. Kucejova B, Pena-Llopis S, Yamasaki T, et al. Interplay between pVHL and mTORC1 pathways in clear-cell renal cell carcinoma. Mol Cancer Res. 2011; 9:1255–65. [PubMed: 21798997]
- Duvel K, Yecies JL, Menon S, et al. Activation of a metabolic gene regulatory network downstream of mTOR complex 1. Mol Cell. 2010; 39:171–83. [PubMed: 20670887]
- 49. Brugarolas JB, Vazquez F, Reddy A, Sellers WR, Kaelin WG Jr. TSC2 regulates VEGF through mTOR-dependent and -independent pathways. Cancer Cell. 2003; 4:147–58. [PubMed: 12957289]
- Schmidt L, Duh FM, Chen F, et al. Germline and somatic mutations in the tyrosine kinase domain of the MET proto-oncogene in papillary renal carcinomas. Nat Genet. 1997; 16:68–73. [PubMed: 9140397]
- 51. Bottaro DP, Rubin JS, Faletto DL, et al. Identification of the hepatocyte growth factor receptor as the c-met proto-oncogene product. Science. 1991; 251:802–4. [PubMed: 1846706]
- 52. Schmidt L, Junker K, Nakaigawa N, et al. Novel mutations of the MET proto-oncogene in papillary renal carcinomas. Oncogene. 1999; 18:2343–50. [PubMed: 10327054]
- Tomlinson IP, Alam NA, Rowan AJ, et al. Germline mutations in FH predispose to dominantly inherited uterine fibroids, skin leiomyomata and papillary renal cell cancer. Nat Genet. 2002; 30:406–10. [PubMed: 11865300]
- Pollard PJ, Briere JJ, Alam NA, et al. Accumulation of Krebs cycle intermediates and overexpression of HIF1alpha in tumours which result from germline FH and SDH mutations. Human Mol Genet. 2005; 14:2231–9. [PubMed: 15987702]
- 55. O'Flaherty L, Adam J, Heather LC, et al. Dysregulation of hypoxia pathways in fumarate hydratase-deficient cells is independent of defective mitochondrial metabolism. Human Mol Genet. 2010; 19:3844–51. [PubMed: 20660115]
- 56. Sudarshan S, Sourbier C, Kong HS, et al. Fumarate hydratase deficiency in renal cancer induces glycolytic addiction and hypoxia-inducible transcription factor 1alpha stabilization by glucosedependent generation of reactive oxygen species. Mol Cell Biol. 2009; 29:4080–90. [PubMed: 19470762]
- Yang Y, Valera V, Sourbier C, et al. A novel fumarate hydratase-deficient HLRCC kidney cancer cell line, UOK268: a model of the Warburg effect in cancer. Cancer Genet. 2012; 205:377–90. [PubMed: 22867999]
- Yamasaki T, Tran TA, Oz OK, et al. Exploring a glycolytic inhibitor for the treatment of an FHdeficient type-2 papillary RCC. Nat Rev Urol. 2011; 8:165–71. [PubMed: 21304509]
- Isaacs JS, Jung YJ, Mole DR, et al. HIF overexpression correlates with biallelic loss of fumarate hydratase in renal cancer: novel role of fumarate in regulation of HIF stability. Cancer Cell. 2005; 8:143–53. [PubMed: 16098467]
- Adam J, Hatipoglu E, O'Flaherty L, et al. Renal cyst formation in Fh1-deficient mice is independent of the Hif/Phd pathway: roles for fumarate in KEAP1 succination and Nrf2 signaling. Cancer Cell. 2011; 20:524–37. [PubMed: 22014577]

- Ooi A, Wong JC, Petillo D, et al. An antioxidant response phenotype shared between hereditary and sporadic type 2 papillary renal cell carcinoma. Cancer Cell. 2011; 20:511–23. [PubMed: 22014576]
- 62. Sourbier C, Valera-Romero V, Giubellino A, et al. Increasing reactive oxygen species as a therapeutic approach to treat hereditary leiomyomatosis and renal cell carcinoma. Cell Cycle. 2010; 9:4183–9. [PubMed: 20953139]
- 63. Klomp JA, Petillo D, Niemi NM, et al. Birt-Hogg-Dubé renal tumors are genetically distinct from other renal neoplasias and are associated with up-regulation of mitochondrial gene expression. BMC Med Genomics. 2010; 3:59. [PubMed: 21162720]
- Pavlovich CP, Walther MM, Eyler RA, et al. Renal tumors in the Birt-Hogg-Dubé syndrome. Am J Surg Pathol. 2002; 26:1542–52. [PubMed: 12459621]
- 65. Baba M, Hong SB, Sharma N, et al. Folliculin encoded by the BHD gene interacts with a binding protein, FNIP1, and AMPK, and is involved in AMPK and mTOR signaling. Proc Natl Acad Sci U S A. 2006; 103:15552–7. [PubMed: 17028174]
- 66. Hasumi Y, Baba M, Ajima R, et al. Homozygous loss of BHD causes early embryonic lethality and kidney tumor development with activation of mTORC1 and mTORC2. Proc Natl Acad Sci U S A. 2009; 106:18722–7. [PubMed: 19850877]
- Weiss RH, Kim K. Metabolomics in the study of kidney diseases. Nat Rev Nephrol. 2012; 8:22– 33. [PubMed: 22025087]
- Sreekumar A, Poisson LM, Rajendiran TM, et al. Metabolomic profiles delineate potential role for sarcosine in prostate cancer progression. Nature. 2009; 457:910–4. [PubMed: 19212411]
- 69. Catchpole G, Platzer A, Weikert C, et al. Metabolic profiling reveals key metabolic features of renal cell carcinoma. J Cell Mol Med. 2011; 15:109–18. [PubMed: 19845817]
- Block K, Gorin Y, Hoover P, et al. NAD(P)H oxidases regulate HIF-2alpha protein expression. J Biol Chem. 2007; 282:8019–26. [PubMed: 17200123]
- Kim K, Aronov P, Zakharkin SO, et al. Urine metabolomics analysis for kidney cancer detection and biomarker discovery. Mol Cell Proteomics. 2009; 8:558–70. [PubMed: 19008263]
- Kind T, Tolstikov V, Fiehn O, Weiss RH. A comprehensive urinary metabolomic approach for identifying kidney cancer. Anal Biochem. 2007; 363:185–95. [PubMed: 17316536]
- Kim K, Taylor SL, Ganti S, Guo L, Osier MV, Weiss RH. Urine metabolomic analysis identifies potential biomarkers and pathogenic pathways in kidney cancer. OMICS. 2011; 15:293–303. [PubMed: 21348635]
- 74. Zira AN, Theocharis SE, Mitropoulos D, Migdalis V, Mikros E. (1)H NMR metabonomic analysis in renal cell carcinoma: a possible diagnostic tool. J Proteome Res. 2010; 9:4038–44. [PubMed: 20527959]
- Lin L, Huang Z, Gao Y, Yan X, Xing J, Hang W. LC-MS based serum metabonomic analysis for renal cell carcinoma diagnosis, staging, and biomarker discovery. J Proteome Res. 2011; 10:1396– 405. [PubMed: 21186845]
- 76. Ganti S, Taylor SL, Aboud OA, et al. Kidney tumor biomarkers revealed by simultaneous multiple matrix metabolomics analysis. Cancer Res. 2012; 72:3471–9. [PubMed: 22628425]
- 77. Biswas S, Charlesworth PJ, Turner GD, et al. CD31 angiogenesis and combined expression of HIF-1alpha and HIF-2alpha are prognostic in primary clear-cell renal cell carcinoma (CC-RCC), but HIFalpha transcriptional products are not: implications for antiangiogenic trials and HIFalpha biomarker studies in primary CC-RCC. Carcinogenesis. 2012; 33:1717–25. [PubMed: 22777959]
- Gordan JD, Lal P, Dondeti VR, et al. HIF-alpha effects on c-Myc distinguish two subtypes of sporadic VHL-deficient clear cell renal carcinoma. Cancer Cell. 2008; 14:435–46. [PubMed: 19061835]
- 79. Shen C, Beroukhim R, Schumacher SE, et al. Genetic and functional studies implicate HIF1alpha as a 14q kidney cancer suppressor gene. Cancer Discov. 2011; 1:222–35. [PubMed: 22037472]
- Leveridge MJ, Finelli A, Kachura JR, et al. Outcomes of small renal mass needle core biopsy, nondiagnostic percutaneous biopsy, and the role of repeat biopsy. Eur Urol. 2011; 60:578–84. [PubMed: 21704449]

 Lee CK, Marschner IC, Simes RJ, et al. Increase in cholesterol predicts survival advantage in renal cell carcinoma patients treated with temsirolimus. Clin Cancer Res. 2012; 18:3188–96. [PubMed: 22472176]

Take-home message

Current knowledge of the metabolic alterations in renal cancer has been derived from both translational studies and genetic models (in vitro and in vivo). Further characterization of these alterations will provide opportunities for therapies and biomarkers for this malignancy. Sudarshan et al.



Fig. 1.

Hypoxia-inducible factor (HIF) accumulation resulting from von Hippel-Lindau (VHL) loss regulates several metabolic processes. HIF promotes the expression of many genes that encode glycolytic enzymes. HIF also prevents entry and metabolism through the tricarboxylic acid (TCA) cycle via several mechanisms including (1) promoting pyruvate conversion to lactate, (2) blocking pyruvate conversion to acetyl-coenzyme A, and (3) lowering the expression of enzymes involved in oxidative phosphorylation.