Danazol-induced Augmentation of Serum αl-Antitrypsin Levels in Individuals with Marked Deficiency of this Antiprotease

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A B S T R A C T Individuals with serum α 1-antitrypsin levels below 80 mg/dl are clearly at risk for the development of accelerated panacinar emphysema. One possible approach to the therapy of this disorder would be to raise serum levels of this major antiprotease to establish protease-antiprotease homeostasis within the lung parenchyma. Because danazol, an impeded androgen, elevates levels of C1 inhibitor in patients deficient of that serum antiprotease, we hypothesized that this agent might also increase α 1-antitrypsin levels in patients with α 1-antitrypsin deficiency. To evaluate this concept, seven patients with severe emphysema associated with α 1-antitrypsin deficiency (six PiZ and 1 M_{Duarte}Z) and one asymptomatic individual (PiSZ) received 600 mg of danazol daily for 30 d. Five of the six PiZ patients responded to danazol therapy with significant increases in serum α 1-antitrypsin levels (mean increase of 37%; P < 0.03). The two individuals who were heterozygous for the Z protein increased their serum levels by 85% (PiM_{Duarte}Z) and 87% (PiSZ), respectively. These increases in serum α 1-antitrypsin antigen were accompanied by commensurate increases in serum trypsin inhibition. Crossed immunoelectrophoresis showed no alterations of the microheterogeneity of the α 1-antitrypsin or the presence of protease-antiprotease complexes in serum during danazol therapy. These data demonstrate that serum α 1-antitrypsin levels can be augmented by danazol therapy in PiZ individuals as well as those heterozygotes with severe deficiency of α 1-antitrypsin. The clinical relevance of these increases in serum α 1-antitrypsin remains speculative, but these findings suggest that danazol may provide a means of improving the protease-antiprotease balance in these individuals and thus impede the progression of their lung disease.

INTRODUCTION

 α l-Antitrypsin is a major serum antiprotease with more than 20 known phenotypes (1–4). Individuals homozygous for the Z variant (phenotype PiZ) or heterozygous for the Z and M_{Duarte} variants (Pi M_{Duarte}Z) and Z and S variants (Pi SZ) are distinguished by marked deficiency of serum α l-antitrypsin levels and a predisposition to severe panacinar emphysema (5–7).

The normal α 1-antitrypsin protein (M) is synthesized by the hepatocyte and released into the circulation at a rate of 38 mg/kg-d (8). Current evidence suggests that the serum α 1-antitrypsin deficiency in individuals with phenotypes PiZ, M_{Duarte}Z, and SZ seems to result from an inability of the hepatocyte to transport the variant protein(s) to the extracellular space (3). It is likely that in each variant, this abnormality derives from a base substitution in the structural gene for α 1-antitrypsin, resulting in an amino acid substitution that alters the conformation of the protein (9–11) and ultimately its secretion from the hepatocyte.

 α l-Antitrypsin is the major serum antiprotease capable of inhibiting neutrophil elastase (12). Current concepts of the pathogenesis of the emphysema associated with marked α l-antitrypsin deficiency hold that the consequence of low levels of circulating α lantitrypsin is progressive, unimpeded neutrophil elastase-mediated destruction of lung parenchyma (2-4). According to this concept, individuals with α lantitrypsin phenotypes PiZ, M_{Duarte}Z, and SZ are at risk for the development of emphysema owing to this protease-antiprotease imbalance in lung. Because the

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variant α l-antitrypsin phenotypes are as effective as the M protein as antiproteases (3), it follows therefore, that if serum α l-antitrypsin levels could be augmented in α l-antitrypsin deficient individuals, protease-antiprotease homeostasis would be established with resultant protection of lung structures from protease-induced emphysema.

One therapeutic approach to increasing α 1-antitrypsin levels in PiZ, M_{Duarte}Z, and SZ individuals is to induce the hepatocyte to increase its output of α 1antitrypsin. Theoretically, optimal therapy would use an agent that specifically stimulates the hepatocyte to increase its output of α 1-antitrypsin. Although such an agent is unknown, it is recognized that anabolic steroids such as diethylstilbestrol will stimulate hepatocytes to increase the production of various proteins, including α 1-antitrypsin (13). Unfortunately, the use of diethylstilbestrol is associated with estrogenic side effects, thus precluding its chronic use in a disorder such as α 1-antitrypsin deficiency. Recently, however, it has been shown that a synthetic androgen, danazol (2,3-isoxazol-17 α -ethinyl testosterone), will also induce the hepatocyte to increase production of various proteins without many of the dangers associated with other anabolic steroid agents (14). This, together with the fact that danazol increases serum C1 inhibitor levels in hereditary angioedema (another serum antiprotease deficiency disorder), suggests that danazol might be an effective therapy for severe α 1antitrypsin deficiency (14-16). For these reasons, the present study was designed to assess the capacity of danazol to augment serum α 1-antitrypsin levels in individuals with α 1-antitrypsin phenotypes PiZ, $M_{Duarte}Z$, and SZ.

METHODS

Patient population. The study population consisted of eight individuals referred to the Pulmonary Branch, National Institutes of Health, with known decreased serum levels of α 1-antitrypsin (Table I). Seven had bibasilar lower lobe bullous disease and one had a normal chest roentgenogram. Of the seven patients with bullous disease, six possessed the phenotype PiZ. These homozygous patients included three males and three females with average age of 47±5 yr. This group had experienced pulmonary symptoms for 5±3 yr; none were active cigarette smokers at the time of the study, although four were ex-smokers who had averaged 25±4 pack-yr of cigarette consumption. Pulmonary function testing (17) of the PiZ individuals revealed severe obstruction to airflow and reduced diffusing capacity consistent with the clinical diagnosis of emphysema. One of the seven patients with bullous lung disease was a heterozygote with the PiM_{Duarte}Z phenotype. This individual was a 43-yr-old female. She was an ex-smoker who had consumed 20 pack-yr of cigarettes, and whose pulmonary function tests closely resembled those of the six homozygous patients. The only asymptomatic subject included in the study was a 19-yr-old with phenotype PiSZ, a nonsmoking female whose pulmonary function studies were normal.

 α l-Antitrypsin serum levels and α l-antitrypsin phenotyping. Serum levels of α l-antitrypsin in the study patients were measured by a standard radial immunodiffusion method using antibody-containing agarose plates and reference serum standard (Behring Diagnostics, American Hoechst Corp., Somerville, N. J.). Serum levels of α 2-macroglobulin and antithrombin III were measured using plates obtained from the same source; serum Cl inhibitor was measured by the same technique. All determinations were performed in duplicate according to the methods described by Mancini (18).

αl-Anti- trypsin phenotype	Patient	Age	Sex	Duration of symptoms	Smoking history	Serum α1- antitrypsin	Vital‡ capacity	Total lung capacity	RV/TLC§	FEV ₁ ^s	MMEF¶	D _{LC0} **
·		yr		yr	pack-yr	mg/dl	% predicted		% observed		% predicted	
PiZ	R. J.	49	М	4	30	15	78	128	51	24	10	36
PiZ	L.E.	37	Μ	2	20	35	49	112	67	23	10	35
PiZ	M.G.	66	F	5	0	43	111	103	38	51	8	40
PiZ	H.P.	50	F	15	30	27	47	71	54	21	4	26
PiZ	T.W.	34	Μ	7	20	32	47	95	64	12	4	19
PiZ	E.M.	49	F	2	0	42	51	81	36	29	12	38
		47±5*		5±3		32 ± 4	64 ± 10	98±8	52 ± 5	27±5	8±1	32±3
PiM _{Duarte} Z	D.K.	43	F	11	20	20	38	71	48	16	7	36
PiSZ	A.J.	19	F	0	0	85	96	87	18	103	91	77

 TABLE I

 Clinical and Physiological Description of the Study Population*

* Mean±SE of the M are given for the PiZ group.

§ RV/TLC, residual volume/total lung capacity.

¶ MMEF, maximum midexpiratory flow rate (25-75% of vital capacity).

** D_{LCO}, diffusing capacity; single breath method.

[‡] Methods for the physiologic studies (17).

[#] FEV₁, forced vital capacity in 1 s.

All data given as mean \pm SEM; all statistical comparisons are by the two-tailed Student's t test.

The eight patients were phenotyped by acid starch gel electrophoresis of their pretreatment serum (19). Six of the eight subjects were homozygous for the Z protein (Pi Z). The seventh was heterozygous for the Z protein and M_{Duarte} protein (Pi M_{Duarte} Z; the Duarte protein displayed an electrophoretic mobility identical to the M protein (7), yet was associated with serum levels that approximated those of the Z protein). The eighth individual was heterozygous for the S and Z proteins.

Drug protocol. Before initiation of the study, each patient was determined to be free of clinical or biochemical evidence of hepatic, renal, or hematologic disease. The participants each received 600 mg of danazol orally per d for 30 consecutive d. Serum was obtained on two occasions before administration of the test drug and thereafter throughout the 30-d study period at intervals that ranged from 3 to 7 d. These sera were kept in liquid N₂ vapor until assayed. Serum transaminase levels, alkaline phosphatase, and bilirubin, were measured at the conclusion of the study to assess any hepatic dysfunction secondary to the use of danazol. Given the short duration of this drug study, no attempt was made to ascertain the influence of this mode of therapy on the severe, chronic pulmonary disease manifested in the study patients.

Serum trypsin inhibitory capacity. To verify the functional integrity of the measured antigenic α l-antitrypsin in serum, base line and 30-d sera were assayed for the capacity to inhibit the enzymatic activity of bovine pancreatic trypsin (Sigma Chemical Co., St. Louis, Mo.) on the synthetic substrate benzoyl-arginine-p-nitroanilide as described by Eriksson (20).

Bidirectional antigen-antibody electrophoresis. To evaluate possible alterations in the microheterogeneity of α 1antitrypsin secondary to danazol therapy, pretreatment, and 30-d treatment sera were evaluated using crossed immunoelectrophoretic analysis (21). Isoelectric focusing was performed in polyacrylamide (pH 4-5); for the second dimension, the polyacrylamide strip containing the serum sample was electrophoresed into agarose (pH 8.2) containing monospecific goat antiserum to α 1-antitrypsin (Atlantic Antibodies, Westbrook, Maine). To evaluate sera for protease- α 1-antitrypsin complexes, crossed immunoelectrophoresis under alkaline conditions using agarose in the first dimension was performed as described by Laurell (22).

RESULTS

The group mean pretreatment α l-antitrypsin levels for the six homozygous PiZ patients was 32 mg/dl. After 30 d of danazol therapy, the group mean level was 44 mg/dl representing an average increase of 37% over pretreatment levels (P < 0.03) (Fig. 1). Evaluation of the sera of each of the homozygous PiZ patients demonstrated that, compared with their individual pretreatment levels, five of the six patients had significant increases in their serum α l-antitrypsin levels within 3 wk of therapy. The one PiZ patient who did not respond (M.G.) had a pretreatment level of 43 mg/dl and a posttreatment level of 42 mg/dl. In those patients that responded to danazol, the α l-antitrypsin levels returned to approximately pretreatment levels within 7 d after cessation of therapy (data not shown).

The two heterozygous patients (D.K., phenotype

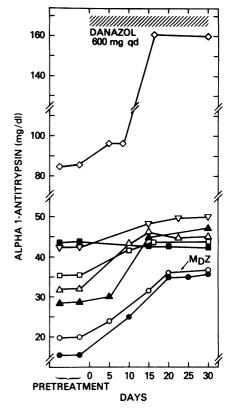


FIGURE 1 Effect of danazol administration on serum α lantitrypsin levels in the eight study patients (R.J., \oplus ; D.K. \bigcirc ; H.P., \blacktriangle ; T.W., \triangle ; M.G., \blacksquare ; L.E., \Box ; E.M., ∇ ; A.J., \diamondsuit). All individuals were type PiZ except D.K. (PiM_DZ) and A.J. (PiSZ).

Pi $M_{\text{Duarte}}Z$; A.J., phenotype Pi SZ) both responded to danazol therapy. The Pi $M_{\text{Duarte}}Z$ patient went from a serum α 1-antitrypsin pretreatment level of 20 mg/dl to a level of 37 mg/dl after 3 wk of therapy. The SZ patient had the largest absolute elevation of α 1-antitrypsin levels of all study patients (pretreatment, 85 mg/dl; after 3 wk of therapy, 160 mg/dl).

Crossed antigen-antibody electrophoresis demonstrated that the increment in serum α 1-antitrypsin in the patients who responded to danazol was not attributable to protease- α 1-antitrypsin complexes (data not shown). In addition, comparison of isoelectric focusing-crossed immunoelectrophoresis patterns in sera obtained before and during danazol therapy revealed no apparent alteration of the α 1-antitrypsin's electrophoretic microheterogeneity during the course of therapy in any of the seven "responders" (example shown in Fig. 2). Thus, the five of six PiZ patients who responded to danazol did so with increases in the Z protein. Planimetric analysis of crossed immunoelectrophoretic patterns of serum before and after therapy showed that the patient with the PiM_{Duarte}Z phenotype responded with equivalent increases in the M_{Duarte} and Z proteins. Similar analysis demonstrated

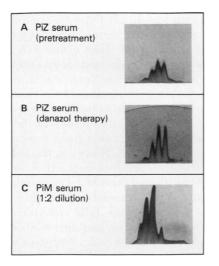


FIGURE 2 Crossed immunoisoelectrophoretic analysis of PiZ serum before (A) and 30 d after danazol therapy (B). The anode is to the left for the first dimension (isoelectric focusing). With danazol therapy the pattern remained the same but the amplitude of the peaks increased, reflecting an increase in the concentration of the PiZ protein. For comparison, a pattern for normal serum (PiM) at a 1:2 dilution is provided (C).

that in the PiSZ patient, the Z protein contributed only 30% of the total increase in serum α 1-antitrypsin, whereas the S protein comprised nearly 70% (data not shown).

Quantitation of the other major serum protease inhibitors before and during danazol administration revealed no significant alteration of the group mean or individual α 2-macroglobulin or antithrombin III levels in any of the homozygous or heterozygous patients (P < 0.2, all comparisons; Fig. 3). In comparison,

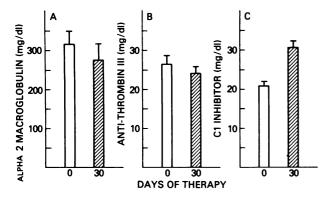


FIGURE 3 Effect of danazol administration on serum levels of antiproteases other than α 1-antitrypsin in the eight study patients; (A) α 2-macroglobulin, (B) antithrombin III, and (C) C1 inhibitor. Serum levels were determined before therapy (day 0) and at the completion of the 30-d danazol trial (day 30) and are expressed as the mean ±SEM. There were no differences between the pre- and the posttreatment sera for α 2macroglobulin and antithrombin III (P > 0.2, both determinations) but C1 inhibitor levels increased significantly with danazol therapy (P < 0.02).

serum C1 inhibitor levels increased in response to danazol; each patient demonstrated an increase in serum C1 inhibitor with group mean levels rising from 21 to 31 mg/dl, representing an average increment of 50% (P < 0.02). This response of serum C1 inhibitor is similar to that observed in "normal" individuals during danazol therapy (14, 15).

Not only did danazol increase antigenic levels of serum α l-antitrypsin in the majority of study patients, but the increased serum protein appeared to be functional as well. Assessment of serum trypsin inhibitory capacity in three of the PiZ patients (R.J., H.P., and L.E.) showed increases in α l-antitrypsin function paralleling increases in their antigenic levels (0.12±0.02 to 0.17±0.02). The one PiZ patient whose antigenic levels did not change (M.G.), also had no change in functional level. Like the PiZ patients who responded to therapy, the two heterozygous patients increased their functional serum inhibitory capacity as well as their antigenic levels.

DISCUSSION

Current concepts of the pathogenesis of the emphysema associated with serum α 1-antitrypsin deficiency suggest that lung destruction in these patients results from a protease-antiprotease imbalance within the lung. In this context, such destruction potentially could be checked by therapeutic maneuvers designed to improve protease-antiprotease homeostasis within the lung parenchyma. The present study suggests that this approach may be partially realized through the administration of an impeded androgen, danazol, which produces a significant increase in serum antigenic and functional α 1-antitrypsin in patients with severe deficiency of this antiprotease. In addition, the effect of danazol on α 1-antitrypsin serum levels of these patients does not appear to be a general, nonspecific increased stimulation of the liver; although serum levels of C1 inhibitor were also increased, α^2 macroglobulin and antithrombin III levels remained unchanged.

Significance of danazol therapy in α l-antitrypsin deficiency. Several considerations suggest that danazol may be a useful therapeutic agent in the treatment of severe α l-antitrypsin deficiency. Because the concept of lung destruction in these individuals is based on the theory of protease-antiprotease imbalance, it can be argued that any increase in circulating antiprotease may be beneficial to these individuals. Danazol increased serum levels of the PiZ patients an average of 37% and increased the serum levels of the Pi M_{Duarte}Z and PiSZ individuals 85 and 87%, respectively. Although danazol did not fully restore α l-antitrypsin levels to normal, these increases may at least partially reestablish protease-antiprotease homeostasis in these patients.

Although there is no precise information regarding the threshold of serum α 1-antitrypsin required for maintenance of protease-antiprotease homeostasis within the lung, estimates of this threshold level can be inferred from studies of individuals with different α 1-antitrypsin phenotypes. While the PiM phenotype (serum levels 200-300 mg/dl) is not associated with increased risk of lung disease, it is estimated that 80% of PiZ individuals (levels 10-50 mg/dl) and a significant proportion of PiSZ individuals (levels 60-80 mg/dl) develop panacinar emphysema (6). In comparison, although there are anecdotal reports of PiS (levels 80-100 mg/dl) and PiMZ (levels 100-140 mg/dl) individuals with accelerated emphysema, the association of serum levels from 80-140 mg/dl with increased risk to the development of destructive lung disease remains controversial (23-31). Thus, it is likely that the threshold level of serum α 1-antitrypsin necessary to maintain pulmonary protease-antiprotease homeostasis is in the range of 80 to 140 mg/dl, and that serum α 1antitrypsin levels of greater than 80 mg/dl are partially, if not completely, protective. However, additional considerations suggest that in some α 1-antitrypsin deficient individuals, the serum threshold for proteaseantiprotease balance may even be lower. Recent studies suggest that the protease-antiprotease equation may be influenced by the concentration of elastase in lysosomes of circulating polymorphonuclear leukocytes (32, 33), implying that individuals with relatively low levels of lysosomal elastase activity may require proportionally lower α 1-antitrypsin serum levels in order to effect pulmonary protease-antiprotease homeostasis. It is conceivable, therefore, that some PiZ individuals may have an α 1-antitrypsin threshold as low as 50 mg/dl, a level which is close to that reached during danazol therapy. All of these considerations must be viewed in terms of the recent epidemiologic study of smoking and nonsmoking PiZ individuals (34). Cigarette smoking is clearly a major risk in PiZ individuals and thus smoking PiZ patients are more likely to require higher α 1-antitrypsin levels than nonsmokers to protect their lungs from proteolytic damage.

There has been considerable experience with the administration of danazol to normals (35), women with endometriosis (36), and individuals with hereditary angioedema (14, 15). The most common adverse effects include weight gain, water retention, acne, decreased breast size, monilial vaginitis, mild hirsutism, and menopause-like symptoms, all of which are reversible with cessation of therapy. Although it is important to integrate these adverse effects with a consideration of the risks and benefit of chronic danazol administration to α l-antitrypsin deficient patients, danazol is the first example of an agent that offers practical therapeutic potential to these individuals. Previous studies have shown that the administration of combination

estrogen-progestin oral contraceptives increases serum α 1-antitrypsin levels in normal individuals and those with intermediate serum deficiency of α 1-antitrypsin, but produces no effect in the PiZ individual (37). Similar results are reported for diethylstilbesterol administration (13). The intravenous administration of Salmonella typhi vaccine produced increases in serum α 1-antitrypsin comparable to those achieved by danazol in two PiZ individuals (38), as did the development of acute lobar pneumonia in a patient receiving oral contraceptive therapy (39). However, the enlistment of such inflammatory stimuli is impractical in a context wherein chronic therapy would be required. Although the oral contraceptives theoretically might raise serum α 1-antitrypsin levels in the PiSZ individual, danazol is a nonvirilizing androgen, and thus may be administered to both males and females and does not appear to share a liability for thromboembolic events with the estrogen-progestin agents.

One potential hazard of chronic danazol administration relates to the probability that increases in serum α 1-antitrypsin levels in these patients are the result of stimulation of the hepatocyte. Because α 1-antitrypsin deficiency is associated with an inability of the hepatocyte to transport the variant protein to the extracellular space (3), if danazol stimulates hepatocytes to increase synthesis of this protein, chronic danazol administration may increase intracellular α 1-antitrypsin accumulation and thus potentiate liver disease. For this reason, the present study was specifically designed to last only 30 d, a time by which early liver disease could be detected and reversed. However, no changes in liver enzymes were seen in any study patients, suggesting that this potential hazard may not be important, at least in the short term.

Future approaches. A short-term study such as this cannot answer the question as to whether danazol administration effects any alteration in lung function. However, before long-term studies to resolve this issue can be designed, three major questions must be answered. First, although danazol causes an increase in α 1-antitrypsin levels within 2 wk and maintains this level for an additional 2 wk, it is not known whether these levels will be maintained during chronic therapy. Second, studies must be conducted to evaluate the effect of chronic danazol therapy on the liver function of α 1-antitrypsin deficient individuals. Third, it is possible that higher levels of danazol administration may be associated with even higher levels of serum α 1antitrypsin than demonstrated in this study. In the experience with the use of danazol in the therapy of hereditary angioedema, there appears to be a dose response between the amount of drug given and the resulting serum C1-inhibitor level (15). If the same is true for α 1-antitrypsin in PiZ individuals, it may be possible to raise their serum levels above 50 mg/dl with higher doses.

REFERENCES

- Laurell, C. B., and S. Eriksson. 1963. The electrophoretic αl-globulin pattern of serum in αl-antitrypsin deficiency. Scand. J. Clin. Lab. Invest. 15: 132-140.
- Kueppers, F., and L. F. Black. 1974. αl-Antitrypsin and its deficiency. Am. Rev. Respir. Dis. 110: 176-194.
- Kueppers, F. 1978. Inherited differences in alpha 1antitrypsin. In Genetic Determinants of Pulmonary Disease. S. D. Litwin, editor. Marcel Dekker, Inc., New York. 23-74.
- Morse, J. O. 1978. Alpha 1-antitrypsin deficiency. N. Engl. J. Med. 299: 1045-1048, 1099-1105.
- 5. Eriksson, S. 1964. Pulmonary emphysema and alpha lantitrypsin deficiency. Acta Med. Scand. 177: 175.
- Larrson, C., H. Dirksen, G. Sundstrom, and S. Eriksson. 1976. Lung function studies in asymptomatic individuals with moderately (PiSZ) and severely (PiZ) reduced levels of α1-antitrypsin. Scand. J. Respir. Dis. 57: 267-280.
- Lieberman, J., L. Gaidulis, and S. D. Klotz. 1976. A new deficient variant of alpha 1-antitrypsin (M_{Duarte}). Am. Rev. Respir. Dis. 113: 31-36.
- Jones, E. A., J. Vergalla, C. J. Steer, P. R. Bradley-Moore, and J. M. Vierling. 1978. Metabolism of intact and desalylated α1-antitrypsin. *Clin. Sci. Mol. Med.* 55: 139–148.
- 9. Yoshida, A., J. Lieberman, and L. Gaidulis. 1976. Molecular abnormality of human alpha 1-antitrypsin variant (PiZZ) associated with plasma activity deficiency. *Proc. Natl. Acad. Sci. U. S. A.* **73**: 1324–1328.
- Jeppson, J. O. 1976. Amino acid substitution Glu-Lys in αl-antitrypsin PiZ. FEBS (Fed. Eur. Biochem. Soc.) Lett. 65: 196-197.
- Owen, M. C., and R. W. Carrell. 1976. Alpha 1-antitrypsin. Molecular abnormality of S variant. Br. Med. J. 1: 130-131.
- Ohlsson, K. 1971. Neutral leucocyte proteases and elastase inhibited by plasma alpha 1-antitrypsin. Scand. J. Clin. Lab. Invest. 28: 251-253.
- 13. Lieberman, J., C. Mittman, and J. R. Kent. 1971. Screening for heterozygous α 1-antitrypsin deficiency III. A provocative test with diethylstilbestrol and effect of oral contraceptives. JAMA (J. Am. Med. Assoc.) 217: 1198-1206.
- Gelfand, J. A., R. J. Sherins, D. W. Alling, and M. M. Frank. 1976. Treatment of hereditary angioedema with danazol: reversal of clinical and biochemical abnormalities. N. Engl. J. Med. 295: 1444-1448.
- Frank, M. M., J. A. Gelfand, R. J. Sherins, and J. Gadek. 1978. The treatment of hereditary angioedema with danazol. *In* Clinical Aspects of the Complement System.
 W. Opferkuch, K. Rother, and D. R. Schultz, editors. Georg Thieme Verlag KG, Stuttgart. 134-137.
- Gadek, J. E., S. W. Hosea, J. A. Gelfand, and M. M. Frank. 1979. Response of variant hereditary angioedema phenotypes to danazol therapy. Genetic implications. J. Clin. Invest. 64: 280-286.
- 17. Fulmer, J. D., W. C. Roberts, E. R. von Gal, and R. G. Crystal. 1977. Small airways in idiopathic pulmonary fibrosis. Comparison of morphologic and physiologic observations. J. Clin. Invest. 60: 595-610.
- Mancini, G., A. O. Carbonara, and J. F. Heremans. 1965. Immunochemical quantitation of antigens by single radial immunodiffusion. *Immunochemistry.* 2: 235–254.
- Fagerhol, M. K. 1972. Acid starch gel electrophoresis for detection of alpha 1-antitrypsin variants (Pi types). Outline of techniques employed currently. *In* Pulmonary Emphysema and Proteolysis. C. Mittman, editor. Academic Press, Inc., New York. 145-149.
- Eriksson, S. 1965. Studies in α1-antitrypsin deficiency. Acta Med. Scand. 177(Suppl.): 175.

- 21. Allen, R. C., R. A. Harley, and R. C. Talamo. 1974. A new method for determination of alpha 1-antitrypsin phenotypes using isoelectric focusing on polyacrylamide gel slabs. *Am. J. Clin. Pathol.* 62: 732-739.
- Laurell, C. B. 1965. Antigen-antibody crossed electrophoresis. Anal. Biochem. 10: 358.
- 23. Kueppers, F., and A. Donhardt. 1974. Obstructive lung disease in heterozygotes for alpha 1-antitrypsin deficiency. Ann. Intern. Med. 80: 209-212.
- 24. Klayton, R., R. Fallat, and A. B. Cohen. 1975. Determinants of chronic obstructive pulmonary disease in patients with intermediate levels of α l-antitrypsin. Am. Rev. Respir. Dis. 112: 71-75.
- Hall, W. J., R. W. Hyde, R. H. Schwartz, G. S. Mudholkar, D. R. Webb, Y. P. Chaubey, and P. L. Townes. 1976. Pulmonary abnormalities in intermediate alpha 1-antitrypsin deficiency. J. Clin. Invest. 58: 1069-1077.
- Cox, D. W., V. H. Hoeppner, and H. Levison. 1976. Protease inhibitors in patients with chronic obstructive pulmonary disease: the alpha 1-antitrypsin heterozygote controversy. Am. Rev. Respir. Dis. 113: 601-606.
- Gelb, A. F., E. Klein, and J. Lieberman. 1977. Pulmonary function in nonsmoking subjects with alpha 1-antitrypsin deficiency (MZ phenotype). Am. J. Med. 62: 93-98.
- Morse, J. O., M. D. Liebowitz, R. J. Knudson, and B. Burrows. 1977. Relation of protease inhibitor phenotypes to obstructive lung diseases in a community. N. Engl. J. Med. 296: 1190-1194.
- Liebowitz, M. D., R. J. Knudson, J. O. Morse, and D. Armet. 1978. Closing volume and flow volume abnormalities in alpha 1-antitrypsin phenotype groups in a community population. Am. Rev. Respir. Dis. 117: 179-181.
- Mittman, C. 1978. The PiMZ phenotype: is it a risk factor for the development of chronic obstructive lung disease? *Am. Rev. Respir. Dis.* 118: 649-652.
- McDonogh, D. J., S. P. Nathan, R. J. Knudson, and M. D. Lebowitz. 1979. Assessment of alpha 1-antitrypsin deficiency heterozygosity as a risk factor in the etiology of emphysema. J. Clin. Invest. 63: 299-310.
- 32. Galdston, M., A. Janoff, and A. L. Davis. 1973. Familial variation of leukocyte lysosomal protease and serum α 1-antitrypsin as determinants in chronic obstructive pulmonary disease. *Am. Rev. Respir. Dis.* 107: 718-727.
- Kidokoro, Y., T. C. Kravis, K. M. Moser, J. C. Taylor, and I. P. Crawford. 1977. Relationship of leukocyte elastase concentration to severity of emphysema in homozygous α1-antitrypsin deficient persons. Am. Rev. Respir. Dis. 115: 793-803.
- 34. Larsson, C. 1978. Natural history and life expectancy in severe α 1-antitrypsin deficiency, PiZ. Acta Med. Scand. **204**: 345-351.
- Wentz, A. C., G. C. Jones, and K. C. Sapp. 1976. Investigation of danazol as a contraceptive agent. *Contraception*. 13: 619–621.
- Greenblatt, R. B., W. P. Dmowski, V. B. Mahesh, and H. F. L. Scholer. 1971. Clinical studies with an antigonadotropin-danazol. *Fertil. Steril.* 22: 102-112.
- 37. Laurell, C. B., S. Kullander, and J. Thorell. 1967. Effect of administration of a combined estrogen-progestin contraceptive on the level of individual plasma proteins. *Scand. J. Clin. Lab. Invest.* 21: 337-343.
- 38. Kueppers, F. 1968. Genetically determined differences in the response of alpha 1-antitrypsin levels in human serum to typhoid vaccine. *Humangenetik.* 6: 207-214.
- Laurell, C. B., 1971. Variation of the alpha 1-antitrypsin level of plasma. *In* Pulmonary Emphysema and Proteolysis. C. Mittman, editor. Academic Press, Inc., New York. 161-167.