Editorial

 Published online: February 21, 2013 **Journal of Innate Immunity**

 J Innate Immun 2013;5:97–99 DOI: [10.1159/000347135](http://dx.doi.org/10.1159%2F000347135)

Systems Biology and Innate Immunity

Scott D. Kobayashi Frank R. DeLeo

 Laboratory of Human Bacterial Pathogenesis, Rocky Mountain Laboratories, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Hamilton, Mont. , USA

 The innate immune system is the first line of host defense against invading microorganisms and is essential for maintenance of host health. The innate immune response is largely mediated by soluble host factors – such as complement – and phagocytic leukocytes. Cells of the innate immune system are equipped to recognize a diversity of pathogens through pattern recognition receptors present on the cell surface. In addition, deposition of complement and antibody on the microbial surface enhance the phagocytic process. The ability of phagocytes to ingest and kill pathogenic microorganisms is immediate, nonspecific and not dependent on previous exposure to microbes. The innate immune response plays a pivotal role in initiating inflammation, and regulation of this critical process is highly complex. Over the past decade, our understanding of complex biological processes – including the molecular mechanisms of innate host defense – has increased dramatically through the application of systems biology-level approaches. For example, genome-wide transcript analyses and proteomics studies have been instrumental in dissecting complex signal transduction pathways involved in recognition and killing of bacterial and fungal pathogens by cells of the innate immune system. In addition, similar strategies have been used to elucidate a diversity of host pathways involved in defense against viral pathogens. A thematic focus section in this issue of *Journal of Innate Immunity* highlights the

use of systems biology approaches to better understand innate immunity.

 Inflammatory disorders or syndromes are readily amenable to investigation by systems biology approaches, largely because an extensive repertoire of antibodies, reagents and assays exists for the evaluation of inflammatory cells and molecules. Moreover, inflammatory disorders often have systemic sequelae such as changes in acute inflammatory molecules, which can be measured quantitatively in blood, plasma or serum. Rheumatoid arthritis is a chronic inflammatory disease associated with damage to joint cartilage and bone, as reviewed by Scott et al. [1] . Tumor necrosis factor and cytokines such as interleukin 6 are known contributors to inflammation in rheumatoid arthritis [1]. In the first article of this thematic focus section, Masi et al. [2] utilize serum samples from a relatively large human cohort to identify host inflammatory molecules that might be predictive for rheumatoid arthritis. Analysis of the data from the study cohort using statistical modeling permitted construction of an integrative model of serum inflammatory molecules. The model is a step toward a comprehensive understanding of immune networks and patterns of inflammatory molecule expression that precede or predict the onset of rheumatoid arthritis.

 The complement system is one of the first components of the host innate immune system to respond to invading

KARGER

Fax +41 61 306 12 34 E-Mail karger@karger.ch www.karger.com

 © 2013 S. Karger AG, Basel 1662–811X/13/0052–0097\$38.00/0

 Accessible online at: www.karger.com/jin Dr. Frank R. DeLeo

Laboratory of Human Bacterial Pathogenesis, Rocky Mountain Laboratories National Institute of Allergy and Infectious Diseases, National Institutes of Health South 4th Street, Hamilton, MT 59840 (USA) E-Mail fdeleo @ niaid.nih.gov

microorganisms. There are numerous functions of complement proteins, including the opsonization of microbes for host recognition and activation of the inflammatory response [3]. Complement activation produces protein fragments known as anaphylatoxins – e.g. C5a – which are known regulators of inflammation [3] . C5a receptors are present on many cell types, including neutrophils, and C5a is a known neutrophil chemoattractant and has been reported to either directly activate these leukocytes or prime them for enhanced function [3, 4]. Notably, previous studies in mice have demonstrated that the C5a receptor is important for innate host defense against infection [5]. The ability of complement components such as C5a to diffuse to-and-from bacteria, especially those in biofilm matrices, is likely critical for the function of these molecules in the context of host defense. However, there is limited knowledge of this process. To that end, Conrad et al. [6] used a mathematical modeling approach to predict how bacterial biofilm matrices influence production and diffusion of C5a.

 Neutrophils are critical in the defense against bacterial infections. These host cells are the most abundant leukocytes in humans and are recruited rapidly to sites of infection. Most bacteria are ingested and killed readily by neutrophils and this process ultimately leads to neutrophil apoptosis or phagocytosis-induced cell death, a process important for the resolution of the inflammatory response. However, some bacteria have evolved means to circumvent killing by neutrophils and thereby cause infections. The ability of bacteria to delay neutrophil apoptosis and turnover or cause some other form of cell death (e.g. cytolysis) is an important component of pathogenesis [7]. Inasmuch as neutrophils have a relatively short lifespan (9–10 days, with ∼ 1 day in circulation), they are not especially well suited as hosts for intracellular pathogens. Indeed, macrophages, which are long-lived phagocytes, are targeted as appropriate host cells by many bacterial pathogens. That said, there are a few bacterial pathogens that survive and replicate within neutrophils, including *Anaplasma phagocytophilium* [8] and *Francisella tularensis* [9] . Although progress has been made, our understanding of the molecular mechanisms that permit these pathogens to prolong neutrophil survival remains incompletely characterized. In this special focus section, Schwartz et al. [10] use a microarray-based approach to gain a better understanding of the mechanisms used by *F. tularensis* to delay neutrophil apoptosis. The authors demonstrate that the pathogen triggers specific antiapoptotic and prosurvival mechanisms to survive within neutrophils.

 A comprehensive understanding of the interaction of host and pathogen molecules during infection can provide a potentially important view of the disease process. For example, previous work by Shea et al. [11] determined concurrent host and pathogen transcriptomes – known collectively as an interactome – during infection of non-human primates with *Streptococcus pyogenes* . Such interactome studies can identify biological processes important for both host and pathogen during infection. Using an interactome approach combined with previous protein-protein interaction data and mathematical modeling, Kuo et al. [12] investigated the host and pathogen molecules involved in the infection of zebrafish with *Candida albicans* . Zebrafish are genetically tractable and have become widely used as a host model for infectious disease research. *C. albicans* is an opportunistic fungal pathogen of humans that grows as either a budding yeast or filamentous hyphae, and the ability of the yeast to form hyphae is important for virulence [13, 14]. The studies by Kuo et al. [12] identified a protein-protein interaction network that provides new insight into development of hyphae during *C. albicans* infection. The findings can be used as a springboard for more targeted studies of the specific pathways and molecules involved in the fungal invasion process.

 It is well established that circadian rhythms regulate key daily functions and behaviors, such as metabolism and sleep-wake cycles [15–17] . Indeed, much research focus has been placed on understanding the influence of circadian rhythms on metabolism and dietary health. There is also compelling evidence that circadian rhythms influence or regulate immune system function. For example, a circadian clock in macrophages regulates changes in expression of genes important for pathogen recognition and the innate immune response [18], and phagocytosis is circadian-regulated in *Drosophila* [19] . Plant innate defenses against bacterial pathogens are optimal in daytime [20], a phenomenon related at least in part to circadian control of the recognition of pattern-associated molecular patterns [21] . In mammals, REV-ERBα (a nuclear receptor and transcriptional repressor) and cryptochrome and CLOCK (core clock proteins), have been identified recently as important links between circadian rhythm and immune function [22–24] . Given the importance of circadian rhythms in innate immunity, it is not surprising that disruption of the circadian cycle is associated with inflammatory syndromes [25]. The confluence of innate immunity and circadian cycle is highly complex, and it is within this context that Mavroudis et al. [26] review the use of systems biology-level approaches as a

means to gain an enhanced understanding of the role of circadian rhythms in innate immunity. These authors underscore the use of mathematical models to predict function and mechanisms of clock networks.

 Systems biology and system biology approaches are changing rapidly as technology moves forward. As a result, many areas of basic and applied research, including the field of innate immunity, are now poised to undergo – or have undergone – major scientific breakthroughs. The articles presented in this thematic focus section provide examples of the diverse nature of systems biology approaches in innate immunity.

References

- 1 Scott DL, Wolfe F, Huizinga TW: Rheumatoid arthritis. Lancet 2010;376:1094–1108.
- 2 Masi AT, Rehman AA, Elmore KB, Aldag JC: Serun acute phase protein and inflammatory cytokine network correlations: comparison of a pre-rheumatoid arthritis and non-rheumatoid arthritis community cohort. J Innate Im- \blacktriangleright 11 mun 2013;5:100–113.
- 3 Guo RF, Ward PA: Role of C5a in inflammatory responses. Annu Rev Immunol 2005;23: 821–852.
- 4 Weisbart RH, Kwan L, Golde DW, Gasson JC: Human GM-CSF primes neutrophils for enhanced oxidative metabolism in response to the major physiological chemoattractants. Blood 1987;69:18–21.
- 5 Hopken UE, Lu B, Gerard NP, Gerard C: The C5a chemoattractant receptor mediates mucosal defence to infection. Nature 1996;383: 86–89.
- 6 Conrad EC, Hsu Y-Y, Bortz DM, Younger JG: Spatiotemporal dynamics of complement C5a production within bacterial extracellular polymeric substance. J Innate Immun 2013;5: 114–123
- 7 Kobayashi SD, Rigby KM, DeLeo FR: Bacteria-induced host cell death; in Locht C, Simonet, M (ed): Bacterial Pathogenesis: Molecular and Cellular Mechanisms. Norwich, Caister Academic Press, 2012, pp 317–362.
- 8 Yoshiie K, Kim HY, Mott J, Rikihisa Y: Intracellular infection by the human granulocytic apoptosis. Infect Immun 2000;68:1125–1133.
- 9 Schwartz JT, Barker JH, Kaufman J, Fayram DC, McCracken JM, Allen LA: *Francisella tularensis* inhibits the intrinsic and extrinsic pathways to delay constitutive apoptosis and prolong human neutrophil lifespan. J Immunol 2012;188:3351–3363.
- 10 Schwartz JT, Bandyopadhyay S, Kobayashi SD, McCracken J, Whitney AR, Deleo FR, Allen LA: *Francisella tularensis* alters human neutrophil gene expression: insights into the molecular basis of delayed neutrophil apoptosis. J Innate Immun 2013;5:124–136.
- 11 Shea PR, Virtaneva K, Kupko JJ 3rd, Porcella SF, Barry WT, Wright FA, Kobayashi SD, Carmody A, Ireland RM, Sturdevant DE, Ricklefs SM, Babar I, Johnson CA, Graham MR, Gardner DJ, Bailey JR, Parnell MJ, Deleo FR, Musser JM: Interactome analysis of longitudinal pharyngeal infection of cynomolgus macaques by group A *Streptococcus* . Proc Natl Acad Sci USA 2010;107:4693–4698.
- 12 Kuo Y-Z, Chuang J-Y, Chao C-C, Liu F-C, Chen B-S: Identification of infection- and defense-related genes via a dynamic host pathogen interaction network using a *Candida albicans* -zebrafish infection model. J Innate Immun 2013;5:137–152.
- 13 Gow NA, van de Veerdonk FL, Brown AJ, Netea MG: *Candida albicans* morphogenesis and host defence: discriminating invasion from colonization. Nat Rev Microbiol 2012; 10:112–122.
- 14 Lo HJ, Kohler JR, DiDomenico B, Loebenberg D, Cacciapuoti A, Fink GR: Nonfilamentous *C. albicans* mutants are avirulent. Cell 1997; 90:939–949.
- 15 Bass J: Circadian topology of metabolism. Nature 2012;491:348–356.
- ehrlichiosis agent inhibits human neutrophil 16 Huang W, Ramsey KM, Marcheva B, Bass J: Circadian rhythms, sleep, and metabolism. J Clin Invest 2011;121:2133–2141.
	- Feng D, Lazar MA: Clocks, metabolism, and 26 the epigenome. Mol Cell 2012;47:158–167.
	- 18 Keller M, Mazuch J, Abraham U, Eom GD, Herzog ED, Volk HD, Kramer A, Maier B: A circadian clock in macrophages controls inflammatory immune responses. Proc Natl Acad Sci USA 2009;106:21407–21412.
	- 19 Stone EF, Fulton BO, Ayres JS, Pham LN, Ziauddin J, Shirasu-Hiza MM: The circadian clock protein Timeless regulates phagocytosis of bacteria in *Drosophila* . PLoS Pathog 2012; 8:e1002445.
- 20 Roden LC, Ingle RA: Lights, rhythms, infection: the role of light and the circadian clock in determining the outcome of plant-pathogen interactions. Plant Cell 2009;21:2546– 2552.
- 21 Bhardwaj V, Meier S, Petersen LN, Ingle RA, Roden LC: Defence responses of *Arabidopsis thaliana* to infection by *Pseudomonas syringae* are regulated by the circadian clock. PLoS One 2011;6:e26968.
- 22 Gibbs JE, Blaikley J, Beesley S, Matthews L, Simpson KD, Boyce SH, Farrow SN, Else KJ, Singh D, Ray DW, Loudon AS: The nuclear receptor REV-ERBα mediates circadian regulation of innate immunity through selective regulation of inflammatory cytokines. Proc Natl Acad Sci USA 2012;109:582–587.
	- 23 Narasimamurthy R, Hatori M, Nayak SK, Liu F, Panda S, Verma IM: Circadian clock protein cryptochrome regulates the expression of proinflammatory cytokines. Proc Natl Acad Sci USA 2012;109:12662–12667.
- 24 Spengler ML, Kuropatwinski KK, Comas M, Gasparian AV, Fedtsova N, Gleiberman AS, Gitlin II, Artemicheva NM, Deluca KA, Gudkov AV, Antoch MP: Core circadian protein CLOCK is a positive regulator of NF-κBmediated transcription. Proc Natl Acad Sci U S A 2012:109:E2457-2465.
- 25 Castanon-Cervantes O, Wu M, Ehlen JC, Paul K, Gamble KL, Johnson RL, Besing RC, Menaker M, Gewirtz AT, Davidson AJ: Dysregulation of inflammatory responses by chronic circadian disruption. J Immunol 2010;185:5796–5805.
	- 26 Mavroudis PD, Scheff JD, Calvano SE, Androulakis IP: Systems biology of circadian-immune interactions. J Innate Immun 2013;5:153–162.