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## UNC1062, a new and potent Mer inhibitor

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### Abstract

Abnormal activation of Mer kinase has been implicated in the oncogenesis of many human cancers including acute lymphoblastic and myeloid leukemia, non-small cell lung cancer, and glioblastoma. We have discovered a new family of small molecule Mer inhibitors, pyrazolopyrimidine sulfonamides, that potently inhibit the kinase activity of Mer. Importantly, these compounds do not demonstrate significant hERG activity in the PatchXpress assay. Through structure-activity relationship studies, **35 (UNC1062)** was identified as a potent ( $IC_{50} = 1.1$  nM) and selective Mer inhibitor. When applied to live tumor cells, **UNC1062** inhibited Mer phosphorylation and colony formation in soft agar. Given the potential of Mer as a therapeutic target, **UNC1062** is a promising candidate for further drug development.

### Keywords

Mer inhibitors; pyrazolopyrimidines; sulfonamides; leukemia; non-small cell lung cancer; glioblastoma

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## INTRODUCTION

Mer belongs to the TAM (Tyro3, Axl, and Mer) family of receptor tyrosine kinases (RTKs).<sup>1</sup> Under normal physiological conditions, Mer kinase promotes cell proliferation and survival, platelet aggregation, macrophage clearance of apoptotic cells, and cytokine release.<sup>2</sup> Abnormal activation of Mer has been implicated in the oncogenesis of many human cancers and is often associated with poor prognostic indicators.<sup>2-3</sup> For example, Mer is ectopically expressed in both B- and T-cell acute lymphoblastic leukemias (ALL)<sup>4</sup> but not in normal mouse and human T- and B-lymphocytes at any stage of development. Similarly, Mer is expressed in the majority of acute myeloid leukemia (AML) cell lines and patient samples but not in normal bone marrow myeloid cells.<sup>5</sup> Finally, Mer is frequently overexpressed in solid tumors, including non-small cell lung cancer (NSCLC) cell lines<sup>6</sup> and patient samples, even though normal human bronchial epithelial (NHBE) cells and lung tissue adjacent to tumors do not express Mer protein.<sup>6</sup> Similar findings have been reported in other solid tumors, including glioblastoma (GBM)<sup>3b, 7</sup> and metastatic melanoma.<sup>8</sup> This tumor specific expression pattern may confer a large therapeutic window - at least in these tumor types. Expression of a Mer transgene in hematopoietic cells leads to development of leukemia and/or lymphoma in mice.<sup>9</sup> In addition, when Mer is inhibited by sh-RNA knockdown in leukemia,<sup>4c</sup> NSCLC,<sup>6</sup> or GBM cells,<sup>7a</sup> they are more susceptible to apoptotic death and exhibit reduced colony formation in soft agar. Moreover, in orthotopic ALL and AML xenograft mouse models, onset and progression of disease were delayed and survival was significantly increased in mice transplanted with Mer knockdown leukemia cell lines relative to mice transplanted with wild-type cell lines.<sup>4c, 5</sup> Similarly, in a NSCLC xenograft mouse model growth of tumors is markedly slowed by Mer knockdown. In addition, the combination of standard cytotoxic chemotherapies with shRNA-mediated Mer knockdown results in synergistic tumor cell killing. Taken together, these data demonstrate important roles of Mer for oncogenesis and anti-apoptotic activity in multiple tumor types. Therefore, Mer provides a novel therapeutic target for the treatment of ALL, AML, NSCLC, and other Mer-related diseases. Consequently, Mer inhibitors are expected to both mediate direct anti-tumor, proapoptotic effects and augment chemosensitivity in combination with standard therapies.

Few small molecule Mer inhibitors have been reported,<sup>10</sup> among them compound **1** (**UNC569**) (Figure 1) is the most promising one demonstrating inhibition of Mer at sub-nanomolar concentrations.<sup>11</sup> However, **UNC569** has undesirable activity toward the human *ether-a-go-go*-related gene potassium channel (hERG) in the conventional PatchXpress assay ( $EC_{50} = 1.7 \mu\text{M}$ ).<sup>12</sup> Inhibition of hERG could lead to long QT syndrome and sudden death and is the reason for the withdrawal of several FDA-approved drugs.<sup>13</sup> In an effort to decrease the hERG activity of **UNC569** through analog synthesis, we discovered that sulfonamide **2** (Figure 1) had similar activity against Mer kinase with better physical properties. Consequently, compound **2** is a better starting point than **UNC569** to develop potent Mer inhibitors as potential therapeutics. Herein we report the structure-activity relationship (SAR) of a series of compounds related to **2** on inhibiting the TAM family kinases and the discovery of potent Mer inhibitors with minimal hERG activity and excellent cellular activity.

## SYNTHESIS

The synthetic routes to sulfonamide analogues (Scheme 1) are similar to what we have previously developed for the synthesis of analogues of **UNC569**.<sup>11</sup> Briefly, 3-chlorobenzoperoxoic acid (*m*CPBA) oxidation of the known compound **3**<sup>14</sup> followed by the displacement of the methyl sulfinyl/sulfonyl group by amines provided the intermediate **4**,

which was subsequently alkylated at the N1 position of the pyrazole ring to yield **5**. Suzuki coupling between **5** and a boronic acid provided the final product **6**.

## RESULTS AND DISCUSSION

Most hERG inhibitors contain a basic amine group and we thus speculate that the hERG activity of **1** may be due to the presence of a primary amine (NH<sub>2</sub>) at the R<sup>2</sup> position (Table 1). Consequently, we first synthesized a focused library of compounds with different R<sup>2</sup> groups while keeping R<sup>1</sup> fixed as *N*-methyl benzenesulfonamide. Inhibition of Mer kinase activity by analogues was tested at the Km for ATP using a microfluidic capillary electrophoresis (MCE) assay<sup>15</sup> in which phosphorylated and unphosphorylated substrate peptides were separated and analyzed using a LabChip EZ Reader. For comparison, inhibition of other members in the TAM RTK family, Tyro3 and Axl, was also measured. As shown in Table 1, when the NH<sub>2</sub> group at the R<sup>2</sup> position (**2**) was replaced by a hydroxyl (OH) group (**7**), the activity of the analogue **7** was decreased 6-fold. Increasing the length of the linker between the pyrazole ring and the cyclohexyl group also decreased the activity of compounds (**8**, **9** vs **2**). In contrast, when the length of the linker was shortened and R<sup>2</sup> was replaced with a *trans*-4-hydroxycyclohexyl group, the corresponding analogue **10** had enhanced activity compared to the parent **7**. More specifically, the IC<sub>50</sub> value was changed from 18 nM for **7** to 2.4 nM for **10**, suggesting that the conformational rigidity and the size of R<sup>2</sup> was important for Mer inhibition. Consistent with this notion, analogs **12** and **13**, which featured open chain versions of cyclohexanol at the R<sup>2</sup> position, had decreased Mer inhibition. Using a tetrahydropyran to replace the 4-hydroxycyclohexyl group provided analogues that were less active (**14**, **15** & **16**). Furthermore, a cyclohexyl group alone at the R<sup>2</sup> position resulted in a 28-fold weaker inhibitor **17**, which further confirmed our previous observation: a hydrogen bond between the polar substituent at the R<sup>2</sup> position with the carbonyl of Arg727 of Mer protein was important.<sup>11</sup> The configuration of the hydroxyl group was not critical, as the *cis*-analogue **11** had similar activity as *trans*-**10**.

We also tested the activity of the synthetic analogues against Axl and Tyro3 to monitor their selectivity for TAM family members and develop a pan-TAM SAR. In general, these analogues showed modest selectivity within the TAM family and are generally less potent against Axl and Tyro3. In addition, to test our hypothesis regarding on the cause of the hERG activity, compounds **2** & **10** were characterized in the hERG binding assay. Indeed, **10** had no hERG activity in the PatchXpress assay while **2** was active with an EC<sub>50</sub> of 3.2 μM. Removal of the primary amine at R<sup>2</sup> position eliminated the binding of our compounds to the hERG channel.

Next, the SAR at the R<sup>1</sup> site was explored (Table 2). In general, these analogues are very potent against Mer (at sub-nanomolar concentrations) and have good selectivity for Mer over Axl (> 16 fold) and Tyro3 (> 9 fold). A reversed sulfonamide functional group (**18**), sulfonic acid (**19**) and unsubstituted sulfonamide (**20**) at the R<sup>1</sup> site were tolerated. When one of the acidic hydrogens in sulfonamide was substituted with a small alkyl group such as propyl (**21**) or iso-propyl (**22**), the activity of the analogues remained the same. In contrast, a cycloalkyl group such as cyclopropyl (**23**), cyclopentyl (**24**) and cyclohexyl (**25**) at this position yielded a more active analogue. A tetrahydropyran (**26**) or 4-piperidinylmethyl (**28**) group was also well tolerated at this position while a *para*-fluorophenyl ring (**27**) was less suitable. Interestingly, **29**, the precursor of **28** with a Boc protection group at piperidinyl nitrogen, was equally potent, which could be explained by the fact that the R<sup>1</sup> position was exposed to solvents as observed in the X-ray structure of **UNC569** complexed with Mer.<sup>11</sup> To further confirm this explanation, we introduced a large side chain at this position and found that the corresponding derivative **30** was still active. Finally, when the acidic

hydrogen on the sulfonamide functional group was replaced by a variety of alkyl groups, the activity and TAM RTK-selectivity of the resulting analogues **31–36** were maintained.<sup>16</sup>

Compared to **UNC569**, **35 (UNC1062)** had improved selectivity and activity against Mer relative to other TAM family RTKs (78-fold selectivity over Axl and 36-fold over Tyro3). Morrison tight-binding inhibition studies revealed that **UNC1062** had a  $K_i$  of 0.33 nM against Mer.<sup>17</sup> These outstanding inhibitory profiles of **UNC1062** prompted us to further investigate its activity in intact cell systems.

The ability of **UNC1062** to inhibit Mer auto-phosphorylation in different cell lines was first tested. Briefly, human pre-B leukemia 697 cells were grown in suspension and treated with various concentrations of **UNC1062** for 1 h prior to addition of a phosphatase inhibitor to stabilize phospho-Mer. Mer protein was immunoprecipitated from lysates and phospho-Mer and total Mer proteins were detected by western blot. In this cell-based assay, the  $IC_{50}$  value for inhibition of phospho-Mer by **UNC1062** was 6.4 nM (Figure 2). In comparison, the  $IC_{50}$  of **UNC569** in the same assay was 141 nM,<sup>11</sup> suggesting that **UNC1062** was significantly more potent than **UNC569** both with purified Mer kinase and in intact cells.

To determine if **UNC1062** inhibits Mer phosphorylation in adherent cells, similar studies were performed using a human, adherent, pediatric rhabdoid brain tumor cell line (BT-12) and two NSCLC cell lines (A549 and Colo699). Inhibition of Mer phosphorylation was evident in cells when the concentration of **UNC1062** was above 300 nM (Figure 3A). Similarly, phosphorylation of Mer was stably inhibited over 72 h of culture in A549 and Colo699 NSCLC cells treated with 250 nM or 500 nM **UNC1062** (Figure 3B). Thus, while solid tumor cells require significantly higher concentrations of **UNC1062** compared to non-adherent leukemia cells, possibly as a result of decreased exposure across adherent cell membranes, treatment with **UNC1062** at sub-micromolar concentrations was sufficient to inhibit Mer phosphorylation in adherent cell lines.

To determine if **UNC1062** can mediate functional anti-tumor effects, BT-12, A549, and Colo699 cells were cultured in soft agar overlaid with medium containing **UNC1062** or vehicle only. Treatment with 1.0  $\mu$ M **UNC1062** was sufficient to completely abrogate growth of BT-12 rhabdoid tumor colonies in soft agar (Figure 4A). Similarly, treatment with **UNC1062** inhibited colony formation in both A549 and Colo699 NSCLC cultures. Moreover, inhibition of colony formation in long-term soft agar assays and inhibition of Mer phosphorylation in short-term assays occurred at similar concentrations of **UNC1062** in these cell lines, consistent with a causal relationship.

## CONCLUSIONS

In summary, we have identified and synthesized a new family of small molecule Mer inhibitors, pyrazolopyrimidine sulfonamides, which potently inhibit the kinase activity of Mer. Importantly, this new generation of inhibitors do not show significant hERG activity in the PatchXpress assay. Through structure-activity relationship studies, **35 (UNC1062)** was identified as both a potent ( $K_i = 0.33$  nM) and selective Mer inhibitor. When applied to live cells, **UNC1062** inhibited Mer phosphorylation and colony formation in soft agar. These results provide the first evidence of anti-tumor activity mediated by a member of this novel class of inhibitors, thus further validate Mer as a drug target for cancer.

## EXPERIMENTAL

### General

Microwave reaction was carried out using a Discover-S reactor with a vertically-focused IR external temperature sensor and an Explorer 72 autosampler. The dynamic mode was used to set up the desired temperature and hold time with the following fixed parameters: PreStirring, 1 min; Pressure, 200 psi; Power, 200 W; PowerMax, off; Stirring, high. Flash chromatography was carried out on pre-packed silica gel disposable columns. Analytical thin-layer chromatography (TLC) was performed with silica gel 60 F<sub>254</sub>, 0.25 mm pre-coated TLC plates. TLC plates were visualized using UV<sub>254</sub> or phosphomolybdic acid with charring. All <sup>1</sup>H NMR spectra were obtained with a 400 MHz spectrometer and <sup>13</sup>C NMR spectra were obtained with a 100 MHz spectrometer. Preparative HPLC was performed with the UV detection at 220 or 254 nm. LC-MS was performed with the UV detection at 220 nm, 254 nm, and 280 nm, and a single quadrupole mass spectrometer using electrospray ionization (ESI) source. High-resolution (positive ion) mass spectra (HRMS) were acquired using a LCMS-TOF mass spectrometer.

### Synthesis

**3-Bromo-*N*-butyl-1*H*-pyrazolo[3,4-*d*]pyrimidin-6-amine (4)**—To a mixture of 3-bromo-6-(methylthio)-1*H*-pyrazolo[3,4-*d*]pyrimidine (24.5 g, 100 mmol) in THF (100 mL) was added *meta*-chloroperoxybenzoic acid (33.7 g, 77%, 150 mmol) in portions at room temperature. The white mixture was stirred for 2 h and transferred into a THF (50 mL) solution of *n*-butylamine (49.4 mL, 500 mmol) at 0 °C. The resulting solution was heated at 60 °C for 2.0 h. After removal of the solvent under reduced pressure, MeOH was added and the mixture was filtered. The white solid was washed with MeOH (3x) and dried to afford the title compound **4** (23.6 g, 87%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sup>6</sup>) δ 8.59 (s, 1H), 7.63 (s, 1H), 3.37–3.11 (m, 3H), 1.54–1.41 (m, 2H), 1.29 (dq, *J* = 14.4, 7.3 Hz, 2H), 0.84 (t, *J* = 7.3 Hz, 3H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sup>6</sup>) δ 162.0, 157.4, 153.1, 120.5, 107.1, 41.0, 31.0, 20.1, 14.2; MS *m/z* 270.1 [M+H]<sup>+</sup>.

**4-(1-((*trans*-4-Aminocyclohexyl)methyl)-6-(butylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-methylbenzenesulfonamide (2)**—A mixture of **4** (0.054 g, 0.20 mmol), K<sub>2</sub>CO<sub>3</sub> (0.11 g, 0.8 mmol), DMF (2.0 mL), and *tert*-butyl (*trans*-4-(chloromethyl)cyclohexyl)carbamate (0.062 g, 0.25 mmol) in a 10 mL microwave tube was heated under microwave irradiation at 150 °C for 10 min. After cooling to room temperature, (4-(*N*-methylsulfamoyl)phenyl)boronic acid (0.065 g, 0.30 mmol), Pd(PPh<sub>3</sub>)<sub>4</sub> (0.023 g, 0.020 mmol), and H<sub>2</sub>O (1.0 mL) were added sequentially. The resulting mixture was stirred at room temperature for 1.0 min and then heated under microwave irradiation at 150 °C for 15 min. After cooling to room temperature, the mixture was quenched with H<sub>2</sub>O and extracted with EtOAc (3x). The combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue was purified by an ISCO silica gel column to provide *tert*-butyl (*trans*-4-((3-(4-(*N*-methylsulfamoyl)phenyl)-6-(butylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)cyclohexyl)carbamate as a white solid. This white solid was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (2.0 mL), then was added TFA (0.5 mL) at room temperature. The resulting solution was stirred for 1.0 h and was concentrated. The residue was purified by prep-HPLC to provide the title compound **2** as a white solid (0.053 g, 48% over 3 steps). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.17 (s, 1H), 8.20–8.14 (m, 2H), 8.01–7.95 (m, 2H), 4.26 (d, *J* = 6.8 Hz, 2H), 3.52 (t, *J* = 7.1 Hz, 2H), 3.13–3.00 (m, 1H), 2.81 (s, 3H), 2.15–2.01 (m, 3H), 1.86 (d, *J* = 12.0 Hz, 2H), 1.68 (dt, *J* = 12.7, 7.4 Hz, 2H), 1.48 (dt, *J* = 14.8, 7.3 Hz, 2H), 1.41–1.21 (m, 4H), 1.00 (t, *J* = 7.4 Hz, 3H); LC-MS: >97% purity, *t*<sub>R</sub> = 4.379 min; MS *m/z* 472.3 [M+1]<sup>+</sup>.

**4-(1-((*trans*-4-Hydroxycyclohexyl)methyl)-6-(butylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-methylbenzenesulfonamide (7)**—(General Procedure A) A mixture of **4** (0.054 g, 0.20 mmol), K<sub>2</sub>CO<sub>3</sub> (0.11 g, 0.8 mmol), DMF (2.0 mL), and *tert*-butyl((*trans*-4-(chloromethyl)cyclohexyl)oxy)dimethylsilane (0.066 g, 0.25 mmol) in a 10 mL microwave tube was heated under microwave irradiation at 150 °C for 10 min. After the reaction was cooled to room temperature, (4-(*N*-methylsulfamoyl)phenyl)boronic acid (0.065 g, 0.30 mmol), Pd(PPh<sub>3</sub>)<sub>4</sub> (0.023 g, 0.020 mmol), and H<sub>2</sub>O (1.0 mL) were added sequentially. The mixture was stirred at room temperature for 1.0 min and then heated under microwave irradiation at 150 °C for 15 min. After cooling to room temperature, the mixture was quenched with H<sub>2</sub>O and extracted with EtOAc (3x). The combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue was purified by an ISCO silica gel column to provide 4-(1-((*trans*-4-((*tert*-butyldimethylsilyl)oxy)cyclohexyl)methyl)-6-(butylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-methylbenzenesulfonamide as a white solid. This white solid was dissolved in THF (2.0 mL) and was added TBAF (1.0 M in THF, 1 mL) at room temperature. The resulting solution was refluxed for 2.0 h and was concentrated. The residue was purified by prep-HPLC to provide the title compound **7** (0.040 g, 44% over 3 steps) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.30 (s, 1H), 8.20 (d, *J* = 8.0 Hz, 2H), 8.01 (d, *J* = 8.0 Hz, 2H), 4.26 (d, *J* = 7.0 Hz, 2H), 3.58 (t, *J* = 7.2 Hz, 2H), 3.55–3.45 (m, 1H), 2.58 (s, 3H), 2.11–1.93 (m, 3H), 1.73 (td, *J* = 14.9, 7.9 Hz, 4H), 1.49 (dq, *J* = 14.8, 7.5 Hz, 2H), 1.32–1.14 (m, 4H), 1.02 (t, *J* = 7.7 Hz, 3H); LC-MS: >97% purity, *t*<sub>R</sub> = 6.629 min; MS *m/z* 473.3 [M+1]<sup>+</sup>.

**4-(1-(2-(*trans*-4-Hydroxycyclohexyl)ethyl)-6-(propylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-methylbenzenesulfonamide (8)**—The title compound **8** (0.040 g, 42%) was prepared according to general procedure A from *tert*-butyl((*trans*-4-(2-chloroethyl)cyclohexyl)oxy)dimethylsilane (0.069 g, 0.25 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.79 (s, 1H), 7.92 (d, *J* = 8.6 Hz, 2H), 7.83 (d, *J* = 8.6 Hz, 2H), 4.24 (t, *J* = 7.0 Hz, 2H), 3.45–3.30 (m, 3H), 2.50 (s, 3H), 1.81 (t, *J* = 11.5 Hz, 3H), 1.71 (dd, *J* = 13.7, 6.8 Hz, 2H), 1.53 (dt, *J* = 14.9, 7.3 Hz, 2H), 1.40–1.27 (m, 2H), 1.08 (dd, *J* = 23.1, 12.5 Hz, 4H), 1.00–0.89 (m, 2H), 0.86 (t, *J* = 7.9 Hz, 3H); LC-MS: >97% purity, *t*<sub>R</sub> = 5.759 min; MS *m/z* 487.3 [M+1]<sup>+</sup>.

**4-(1-(3-(*trans*-4-Hydroxycyclohexyl)propyl)-6-(propylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-methylbenzenesulfonamide (9)**—The title compound **9** (0.055 g, 56%) was prepared according to general procedure B from *tert*-butyl((*trans*-4-(3-chloropropyl)cyclohexyl)oxy)dimethylsilane (0.069 g, 0.25 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.03 (s, 1H), 8.17–8.09 (m, 2H), 7.99–7.92 (m, 2H), 4.32 (t, *J* = 6.9 Hz, 2H), 3.48 (t, *J* = 7.0 Hz, 2H), 3.42 (dt, *J* = 10.8, 4.2 Hz, 1H), 2.58 (s, 3H), 2.02–1.84 (m, 4H), 1.76 (d, *J* = 11.6 Hz, 2H), 1.66 (dt, *J* = 14.8, 7.3 Hz, 2H), 1.52–1.39 (m, 2H), 1.29–1.13 (m, 5H), 0.99 (t, *J* = 7.4 Hz, 3H), 0.96–0.86 (m, 2H); LC-MS: >97% purity, *t*<sub>R</sub> = 5.856 min; MS *m/z* 501.3 [M+1]<sup>+</sup>.

**4-(1-(*trans*-4-Hydroxycyclohexyl)-6-(propylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-methylbenzenesulfonamide (10) and 4-(1-(*cis*-4-Hydroxycyclohexyl)-6-(propylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-methylbenzenesulfonamide (11)**—(General Procedure B) A mixture of **4** (0.054 g, 0.20 mmol), K<sub>2</sub>CO<sub>3</sub> (0.11 g, 0.8 mmol), DMF (2.0 mL), and 4-chlorocyclohexanol (0.081 g, 0.6 mmol) in a 10 mL microwave tube was heated under microwave irradiation at 200 °C for 30 min. After cooling to room temperature, (4-(*N*-methylsulfamoyl)phenyl)boronic acid (0.065 g, 0.30 mmol), Pd(PPh<sub>3</sub>)<sub>4</sub> (0.023 g, 0.020 mmol), and H<sub>2</sub>O (1.0 mL) were added sequentially. The resulting mixture was stirred at room temperature for 1.0 min and then heated under microwave irradiation at 150 °C for 15 min. After cooling to room

temperature, the mixture was quenched with H<sub>2</sub>O and extracted with EtOAc (3x). The combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue was purified by prep-HPLC to provide the title compounds **10** (0.011 g, 13% over 3 steps) and **11** (0.020 g, 23% over 3 steps) as white solids. **10**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.78 (s, 1H), 7.92 (d, *J* = 8.4 Hz, 2H), 7.82 (d, *J* = 8.3 Hz, 2H), 4.51 (s, 1H), 3.64 (s, 1H), 3.37 (t, *J* = 7.0 Hz, 2H), 2.50 (s, 3H), 2.17–1.86 (m, 6H), 1.61–1.29 (m, 6H), 0.87 (t, *J* = 7.3 Hz, 3H); LC-MS: >97% purity, t<sub>R</sub> = 5.402 min; HRMS (TOF, ESI+) *m/z*: [M+H]<sup>+</sup> calculated for C<sub>22</sub>H<sub>31</sub>N<sub>6</sub>O<sub>3</sub>S, 459.2178; found 459.2155. **11**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub> + CD<sub>3</sub>OD) δ 8.82 (s, 1H), 7.95 (dd, *J* = 8.2, 6.2 Hz, 2H), 7.84 (dd, *J* = 8.5, 1.8 Hz, 2H), 4.66–4.48 (m, 1H), 3.44–3.33 (m, 3H), 2.53 (s, 3H), 2.46–2.32 (m, 1H), 1.99–1.81 (m, 3H), 1.79–1.61 (m, 3H), 1.55 (dt, *J* = 14.8, 7.4 Hz, 2H), 1.36 (td, *J* = 14.8, 7.4 Hz, 3H), 0.88 (t, *J* = 7.3 Hz, 3H); LC-MS: >97% purity, t<sub>R</sub> = 5.590 min; MS *m/z* 459.2 [M+1]<sup>+</sup>.

**4-(1-(2-Hydroxyethyl)-6-(propylamino)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)-N-methylbenzenesulfonamide (12)**—(General Procedure C) A mixture of **4** (0.054 g, 0.20 mmol), K<sub>2</sub>CO<sub>3</sub> (0.11 g, 0.8 mmol), DMF (2.0 mL), and 2-bromoethanol (0.038 g, 0.30 mmol) in a 10 mL microwave tube was heated under microwave irradiation at 150 °C for 10 min. After cooling to room temperature, (4-(*N*-methylsulfamoyl)phenyl)boronic acid (0.065 g, 0.30 mmol), Pd(PPh<sub>3</sub>)<sub>4</sub> (0.023 g, 0.020 mmol), and H<sub>2</sub>O (1.0 mL) were added sequentially. The resulting mixture was stirred at room temperature for 1.0 min and then heated under microwave irradiation at 150 °C for 15 min. After cooling to room temperature, the mixture was quenched with H<sub>2</sub>O and extracted with EtOAc (3x). The combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue was purified by an ISCO silica gel column to provide the title compound **12** (0.044 g, 57%) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.86 (s, 1H), 8.00–7.92 (m, 2H), 7.91–7.83 (m, 2H), 4.42 (t, *J* = 5.0 Hz, 2H), 4.01 (t, *J* = 5.0 Hz, 2H), 3.38 (t, *J* = 7.1 Hz, 3H), 2.56 (s, 3H), 1.63–1.50 (m, 2H), 1.44–1.30 (m, 2H), 0.90 (t, *J* = 7.3 Hz, 3H); LC-MS: >97% purity, t<sub>R</sub> = 5.244 min; MS *m/z* 405.2 [M+1]<sup>+</sup>.

**4-(1-(3-Hydroxypropyl)-6-(propylamino)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)-N-methylbenzenesulfonamide (13)**—The title compound **13** (0.045 g, 56%) was prepared according to general procedure C from 3-chloropropan-1-ol (0.029 g, 0.30 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.88 (s, 1H), 7.98 (d, *J* = 8.6 Hz, 2H), 7.90 (d, *J* = 8.6 Hz, 2H), 4.43 (t, *J* = 6.2 Hz, 2H), 3.50 (t, *J* = 5.8 Hz, 2H), 3.40 (t, *J* = 7.1 Hz, 2H), 2.59 (s, 3H), 2.10–2.00 (m, 2H), 1.60 (dt, *J* = 14.9, 7.4 Hz, 2H), 1.40 (dq, *J* = 14.5, 7.3 Hz, 2H), 0.92 (t, *J* = 7.3 Hz, 3H); LC-MS: >97% purity, t<sub>R</sub> = 5.336 min; MS *m/z* 419.2 [M+1]<sup>+</sup>.

**N-Methyl-4-(6-(propylamino)-1-(tetrahydro-2H-pyran-4-yl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)benzenesulfonamide (14)**—(General Procedure D) A mixture of **4** (0.054 g, 0.20 mmol), K<sub>2</sub>CO<sub>3</sub> (0.11 g, 0.8 mmol), DMF (2.0 mL), and 4-chlorotetrahydro-2H-pyran (0.072 g, 0.60 mmol) in a 10 mL microwave tube was heated under microwave irradiation at 200 °C for 30 min. After cooling to room temperature, (4-(*N*-methylsulfamoyl)phenyl)boronic acid (0.065 g, 0.30 mmol), Pd(PPh<sub>3</sub>)<sub>4</sub> (0.023 g, 0.020 mmol), and H<sub>2</sub>O (1.0 mL) were added sequentially. The resulting mixture was stirred at room temperature for 1.0 min and then heated under microwave irradiation at 150 °C for 15 min. After cooling to room temperature, the mixture was quenched with H<sub>2</sub>O and extracted with EtOAc (3x). The combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure. The residue was purified by an ISCO silica gel column to provide the title compound **14** as a white solid (0.052 g, 61% over 2 steps). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.96 (s, 1H), 8.07 (d, *J* = 8.5 Hz, 2H), 7.97 (d, *J* = 8.5 Hz, 2H), 5.33 (s, 1H), 4.92–4.79 (m, 1H), 4.32 (dd, *J* = 10.8, 5.4 Hz, 1H), 4.17 (dd, *J* = 11.7, 3.3 Hz, 2H), 3.64 (dd, *J* = 11.9, 10.2 Hz, 2H), 3.52 (dd,

$J = 12.9, 6.9$  Hz, 2H), 2.71 (d,  $J = 5.4$  Hz, 3H), 2.48 (ddd,  $J = 25.0, 12.5, 4.7$  Hz, 2H), 1.97 (d,  $J = 10.8$  Hz, 2H), 1.66 (dt,  $J = 14.8, 7.2$  Hz, 2H), 1.47 (dq,  $J = 14.5, 7.3$  Hz, 2H), 0.99 (t,  $J = 7.3$  Hz, 3H); LC-MS: 96% purity,  $t_R = 5.689$  min; MS  $m/z$  445.2 [M+1]<sup>+</sup>.

***N*-Methyl-4-(6-(propylamino)-1-((tetrahydro-2*H*-pyran-4-yl)methyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)benzenesulfonamide (15)**—The title compound **15** (0.062 g, 70% over 2 steps) was prepared according to general procedure C from 4-(bromomethyl)tetrahydro-2*H*-pyran (0.045 g, 0.25 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.95 (s, 1H), 8.10–8.03 (m, 2H), 8.00–7.93 (m, 2H), 5.37 (bs, 1H), 4.41–4.30 (m, 1H), 4.24 (d,  $J = 7.1$  Hz, 2H), 4.03–3.91 (m, 2H), 3.51 (dd,  $J = 13.0, 6.9$  Hz, 2H), 3.38 (td,  $J = 11.5, 2.6$  Hz, 2H), 2.71 (d,  $J = 5.4$  Hz, 3H), 2.40–2.24 (m, 1H), 1.71–1.61 (m, 2H), 1.60–1.56 (m, 1H), 1.54–1.38 (m, 5H), 0.99 (t,  $J = 7.3$  Hz, 3H); LC-MS: >97% purity,  $t_R = 5.714$  min; MS  $m/z$  459.3 [M+1]<sup>+</sup>.

***N*-Methyl-4-(6-(propylamino)-1-(2-(tetrahydro-2*H*-pyran-4-yl)ethyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)benzenesulfonamide (16)**—The title compound **16** (0.058 g, 63% over 2 steps) was prepared according to general procedure C from 4-(2-bromoethyl)tetrahydro-2*H*-pyran (0.048 g, 0.25 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.95 (s, 1H), 8.10–8.03 (m, 2H), 7.99–7.93 (m, 2H), 5.38 (bs, 1H), 4.47–4.33 (m, 3H), 3.95 (dd,  $J = 11.5, 3.5$  Hz, 2H), 3.49 (dd,  $J = 13.1, 6.8$  Hz, 2H), 3.33 (td,  $J = 11.7, 1.8$  Hz, 2H), 2.71 (d,  $J = 5.4$  Hz, 3H), 1.97–1.86 (m, 2H), 1.76 (d,  $J = 12.7$  Hz, 2H), 1.65 (dt,  $J = 12.7, 7.4$  Hz, 2H), 1.55–1.30 (m, 5H), 0.98 (t,  $J = 7.3$  Hz, 3H); LC-MS: 97% purity,  $t_R = 5.835$  min; MS  $m/z$  473.3 [M+1]<sup>+</sup>.

**4-(1-Cyclohexyl-6-(propylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-methylbenzene sulfonamide (17)**—The title compound **17** (0.035 g, 40% over 2 steps) was prepared according to general procedure D from chlorocyclohexane (0.071 g, 0.60 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.94 (s, 1H), 8.10–8.03 (m, 2H), 7.99–7.92 (m, 2H), 5.30 (bs, 1H), 4.68–4.56 (m, 1H), 4.33 (q,  $J = 5.4$  Hz, 1H), 3.52 (dd,  $J = 12.9, 7.0$  Hz, 2H), 2.70 (d,  $J = 5.4$  Hz, 3H), 2.16–1.89 (m, 6H), 1.77 (d,  $J = 12.7$  Hz, 1H), 1.66 (dt,  $J = 14.8, 7.3$  Hz, 2H), 1.54–1.28 (m, 5H), 0.99 (t,  $J = 7.3$  Hz, 3H); LC-MS: >97% purity,  $t_R = 6.358$  min; MS  $m/z$  443.2 [M+1]<sup>+</sup>.

***trans*-4-(3-Bromo-6-(propylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)cyclohexanol**—A solution of **4** (2.70 g, 10 mmol) and *cis*-4-chlorocyclohexanol (4.04 g, 30 mmol) in DMF (50 mL) was added K<sub>2</sub>CO<sub>3</sub> (5.52 g, 40 mmol). The reaction mixture was heated at 150 °C for overnight, quenched with water at room temperature and extracted with EtOAc (3x). The combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue was purified by an Isco silica gel column to afford the title compound (1.83 g, 52%) as a white solid. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$  8.49 (s, 1H), 4.59–4.41 (m, 1H), 3.69–3.56 (m, 1H), 3.42 (t,  $J = 7.1$  Hz, 2H), 2.13–2.00 (m, 4H), 1.98–1.89 (m, 2H), 1.66–1.56 (m, 2H), 1.52–1.36 (m, 4H), 0.96 (t,  $J = 7.4$  Hz, 3H); <sup>13</sup>C NMR (100 MHz, dms-*d*<sup>6</sup>)  $\delta$  161.6, 155.0, 153.4, 119.1, 110.3, 107.7, 68.3, 55.5, 34.6, 30.9, 29.8, 20.0, 14.1; MS  $m/z$  369.1 [M+H]<sup>+</sup>.

***trans*-*N*-(4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)phenyl)methanesulfonamide (18)**—The title compound **18** (0.008 g, 18%) was prepared according to general procedure B from 4-(methylsulfonamido)phenylboronic acid (0.033 g, 0.15 mmol) as a white solid. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>)  $\delta$  8.76 (s, 1H), 7.76–7.72 (m, 2H), 7.28–7.24 (m, 2H), 4.56–4.48 (m, 1H), 3.71–3.61 (m, 1H), 3.42–3.39 (m, 1H), 3.29–3.27 (m, 2H), 2.92 (s, 3H), 2.14–2.02 (m, 4H), 1.97–1.90 (m, 2H), 1.61–1.53 (m, 2H), 1.47–1.32 (m, 6H), 0.90 (t,  $J = 7.36$  Hz, 3H); LC-MS: >97% purity,  $t_R = 5.538$  min; MS  $m/z$  459.3 [M+1]<sup>+</sup>.



**4-(1-(*trans*-4-Hydroxycyclohexyl)-6-(propylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)benzenesulfonic acid (19)**—(General Procedure E) A mixture of *trans*-4-(3-bromo-6-(propylamino)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)cyclohexanol (0.055 g, 0.15 mmol), K<sub>3</sub>PO<sub>4</sub> (0.063 g, 0.30 mmol), 4-boronobenzenesulfonic acid (0.045 g, 0.225 mmol), Pd(PPh<sub>3</sub>)<sub>4</sub> (0.017 g, 0.015 mmol), dioxane (2.0 mL) and H<sub>2</sub>O (0.5 mL) in a 10 mL microwave tube was heated under microwave irradiation at 150 °C for 10 min. After cooling to room temperature, the mixture was quenched with H<sub>2</sub>O and extracted with EtOAc (3x). The combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue was filtered through a plug of Celite and then purified by prep-HPLC to afford the title compound **19** (0.027 g, 30%) as a white solid. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ 9.19 (s, 1H), 8.05–7.93 (m, 4H), 4.77–4.61 (m, 1H), 3.73 (tt, *J* = 10.7, 4.1 Hz, 1H), 3.57 (t, *J* = 7.1 Hz, 2H), 2.31–2.02 (m, 6H), 1.77–1.67 (m, 2H), 1.61–1.44 (m, 4H), 1.03 (t, *J* = 7.4 Hz, 3H); LC-MS: >97% purity, t<sub>R</sub> = 4.890 min; MS *m/z* 444.2 [M-1]<sup>-</sup>.

***trans*-4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)benzenesulfonamide (20)**—The title compound **20** (0.064 g, 71%) was prepared according to general procedure E from 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzenesulfonamide (0.086 g, 0.306 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD+CDCl<sub>3</sub>) δ 8.74 (s, 1H), 7.87–7.78 (m, 4H), 4.46–4.43 (m, 1H), 3.59–3.55 (m, 1H), 3.33–3.28 (m, 2H), 2.03–1.93 (m, 4H), 1.90–1.85 (m, 2H), 1.50–1.45 (m, 2H), 1.39–1.33 (m, 2H), 1.30–1.25 (m, 2H), 1.08–1.06 (m, 3H), 0.80 (t, *J* = 7.4 Hz, 3H); LC-MS: 97% purity, t<sub>R</sub> = 5.147 min; MS *m/z* 445.2 [M+1]<sup>+</sup>.

***trans*-4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-propylbenzenesulfonamide (21)**—The title compound **21** (0.019 g, 13%) was prepared according to general procedure B from (4-(*N*-propylsulfamoyl)phenyl)boronic acid (0.109 g, 0.45 mmol) as a white solid. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>) δ 8.83 (s, 1H), 7.98 (s, 4H), 4.68–4.62 (m, 1H), 4.42 (t, *J* = 6.16 Hz 1H), 3.87–3.81 (m, 1H), 3.57–3.51 (m, 2H), 2.98 (dd, *J* = 6.95, 13.46 Hz, 2H), 2.25–2.16 (m, 4H), 2.10–2.07 (m, 2H), 1.71 (dt, *J* = 7.43, 14.83 Hz, 2H), 1.60–1.42 (m, 6H), 1.00 (t, *J* = 7.36 Hz, 3H), 0.89 (t, *J* = 7.40 Hz, 3H); LC-MS: >97% purity, t<sub>R</sub> = 6.217 min; MS *m/z* 487.3 [M+1]<sup>+</sup>.

***trans*-4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-isopropylbenzenesulfonamide (22)**—The title compound **22** (0.016 g, 13%) was prepared according to general procedure B from (4-(*N*-isopropylsulfamoyl)phenyl)boronic acid (0.095 g, 0.39 mmol) as a white solid. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>) δ 8.91 (s, 1H), 8.00 (dd, *J* = 8.62, 20.77 Hz, 4H), 4.69–4.61 (m, 1H), 4.26 (d, *J* = 7.61 Hz, 1H), 3.88–3.80 (m, 1H), 3.57–3.48 (m, 2H), 2.27–2.18 (m, 4H), 2.07 (d, *J* = 10.85 Hz, 2H), 1.71–1.53 (m, 6H), 1.47 (dd, *J* = 7.36, 14.90 Hz, 2H), 1.11 (d, *J* = 6.52 Hz, 6H), 0.99 (t, *J* = 7.34 Hz, 3H); LC-MS: >97% purity, t<sub>R</sub> = 5.826 min; MS *m/z* 487.3 [M+1]<sup>+</sup>.

**4-(6-(Butylamino)-1-(*trans*-4-hydroxycyclohexyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-3-yl)-*N*-cyclopropylbenzenesulfonamide (23)**—The title compound **23** (0.003 g, 4%) was prepared according to general procedure E from (4-(*N*-cyclopropylsulfamoyl)phenyl)boronic acid (0.090 g, 0.375 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD). δ 9.11 (s, 1H), 8.17 (d, *J* = 8.6 Hz, 2H), 8.00 (d, *J* = 8.6 Hz, 2H), 4.73–4.63 (m, 1H), 3.79–3.67 (m, 1H), 3.51 (t, *J* = 7.1 Hz, 2H), 2.30–2.01 (m, 7H), 1.69 (td, *J* = 14.7, 7.4 Hz, 2H), 1.61–1.42 (m, 4H), 1.01 (t, *J* = 7.4 Hz, 3H), 0.62–0.47 (m, 4H); LC-MS: 95% purity, t<sub>R</sub> = 5.957 min; MS *m/z* 485.0 [M+1]<sup>+</sup>.

**trans-4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)-N-cyclopentylbenzenesulfonamide (24)**—The title compound **24** (0.015 g, 10%) was prepared according to general procedure B from 4-(*N*-cyclopentylsulfamoyl)phenyl)boronic acid (0.121 g, 0.45 mmol) as a white solid. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>) δ 8.82 (s, 1H), 7.99 (q, *J* = 8.67 Hz, 4H), 4.68–4.64 (m, 1H), 4.44 (d, *J* = 7.34 Hz, 1H), 3.86–3.82 (m, 1H), 3.67 (dd, *J* = 6.82, 13.53 Hz, 1H), 3.55–3.50 (m, 2H), 2.22–2.16 (m, 4H), 2.10–2.07 (m, 2H), 1.84 (dd, *J* = 5.70, 12.36 Hz, 2H), 1.71 (dd, *J* = 7.63, 14.79 Hz, 2H), 1.63–1.57 (m, 4H), 1.54–1.48 (m, 4H), 1.40–1.35 (m, 2H), 1.00 (t, *J* = 7.35 Hz, 3H); LC-MS: 95 % purity, *t*<sub>R</sub> = 6.186 min; MS *m/z* 513.3 [M+1]<sup>+</sup>.

**trans-4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)-N-cyclohexylbenzenesulfonamide (25)**—The title compound **25** (0.015 g, 11%) was prepared according to general procedure B from 4-(*N*-cyclohexylsulfamoyl)phenyl)boronic acid (0.110 g, 0.39 mmol) as a white solid. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>) δ 8.95 (s, 1H), 8.04 (d, *J* = 8.61 Hz, 2H), 7.97 (d, *J* = 8.59 Hz, 2H), 5.36 (brs, 1H), 4.67–4.64 (m, 1H), 4.34 (d, *J* = 7.55 Hz, 1H), 3.87–3.80 (m, 1H), 3.52 (dd, *J* = 7.03, 12.98 Hz, 2H), 3.24–3.15 (m, 1H), 2.28–2.18 (m, 4H), 2.09–2.06 (m, 2H), 1.82–1.79 (m, 2H), 1.67–1.56 (m, 6H), 1.52–1.46 (m, 4H), 1.27–1.14 (m, 6H), 1.00 (t, *J* = 7.35 Hz, 3H); LC-MS: >97% purity, *t*<sub>R</sub> = 6.171 min; MS *m/z* 527.0 [M+1]<sup>+</sup>.

**trans-4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)-N-(tetrahydro-2H-pyran-4-yl)benzenesulfonamide (26)**—The title compound **26** (0.016 g, 10%) was prepared according to general procedure B from 4-(*N*-(tetrahydro-2H-pyran-4-yl)sulfamoyl)phenyl)boronic acid (0.128 g, 0.45 mmol) as a white solid. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>) δ 9.44 (brs, 1H), 8.92 (s, 1H), 8.00 (q, *J* = 8.67 Hz, 4H), 4.69–4.61 (m, 2H), 3.90–3.84 (m, 4H), 3.55 (t, *J* = 7.04 Hz, 2H), 3.43–3.33 (m, 2H), 2.21–2.07 (m, 6H), 1.79–1.68 (m, 4H), 1.60–1.44 (m, 6H), 1.00 (t, *J* = 7.36 Hz, 3H); LC-MS: 95% purity, *t*<sub>R</sub> = 5.747 min; MS *m/z* 529.3 [M+1]<sup>+</sup>.

**trans-4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)-N-(4-fluorophenyl)benzenesulfonamide (27)**—The title compound **27** (0.009 g, 6%) was prepared according to general procedure B from 4-(*N*-(4-fluorophenyl)sulfamoyl)phenyl)boronic acid (0.133 g, 0.45 mmol) as a white solid. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>) δ 9.38 (s, 1H), 8.82 (s, 1H), 7.94–7.89 (m, 2H), 7.85–7.81 (m, 2H), 7.09–7.05 (m, 2H), 6.98–6.94 (m, 2H), 6.57 (s, 1H), 4.67–4.61 (m, 1H), 3.98–3.80 (m, 1H), 3.57–3.51 (m, 2H), 2.22–2.08 (m, 6H), 1.71 (dt, *J* = 7.29, 14.74 Hz, 2H), 1.62–1.57 (m, 2H), 1.45 (dt, *J* = 7.37, 14.45 Hz, 2H), 0.99 (t, *J* = 7.38 Hz, 3H); LC-MS: >97% purity, *t*<sub>R</sub> = 6.198 min; MS *m/z* 539.3 [M+1]<sup>+</sup>.

**trans-tert-Butyl 4-((4-(6-(butylamino)-1-(4-hydroxycyclohexyl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)phenylsulfonamido)methyl)piperidine-1-carboxylate (29)**—The title compound **29** (0.011 g, 9%) was prepared according to general procedure B from 4-(*N*-((1-(tert-butoxycarbonyl)piperidin-4-yl)methyl)sulfamoyl)phenyl)boronic acid (0.119 g, 0.30 mmol, 1.5 eq) as a white solid. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>+CD<sub>3</sub>OD) δ 8.78 (s, 1H), 7.90 (d, *J* = 8.44 Hz, 2H), 7.80 (d, *J* = 7.34 Hz, 2H), 4.46–4.43 (m, 1H), 3.91 (d, *J* = 12.95 Hz, 2H), 3.63–3.61 (m, 1H), 2.36 (t, *J* = 7.08 Hz, 2H), 3.22 (dt, *J* = 1.61, 3.24 Hz, 3H), 2.65 (d, *J* = 6.62 Hz, 2H), 2.54–2.51 (m, 2H), 2.08–2.00 (m, 4H), 1.93–1.90 (m, 2H), 1.58–1.46 (m, 4H), 1.31 (s, 12H), 0.97–0.91 (m, 2H), 0.89 (t, *J* = 7.35 Hz, 3H); LC-MS: 96% purity, *t*<sub>R</sub> = 6.305 min; MS *m/z* 642.4 [M+1]<sup>+</sup>.

**trans-4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)-N-(piperidin-4-ylmethyl)benzenesulfonamide (28)**—A mixture

**29** (20 mg, 0.03 mmol) and TFA (2.3 mL) in dichloromethane (4.7 mL) were stirred at room temperature for 2.0 h, then diluted with dichloromethane (20 mL) and washed with a saturated aqueous NaHCO<sub>3</sub> solution, water and brine. The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to afford the title compound **28** (12.7 mg, 75%) as a white solid. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>+CD<sub>3</sub>OD) δ 8.25 (s, 1H), 7.23 (d, *J* = 8.45 Hz, 2H), 7.14 (d, *J* = 8.20 Hz, 2H), 3.85–3.80 (m, 1H), 2.96–2.91 (m, 1H), 2.72 (t, *J* = 7.15 Hz, 2H), 2.54–2.52 (m, 3H), 2.07–1.99 (m, 4H), 1.41–1.32 (m, 4H), 1.26–1.23 (m, 2H), 1.12 (d, *J* = 13.63 Hz, 2H), 0.98–0.94 (m, 1H), 0.91–0.84 (m, 2H), 0.79–0.75 (m, 2H), 0.67–0.56 (m, 4H), 0.17 (t, *J* = 7.35 Hz, 3H); LC-MS: >97% purity, t<sub>R</sub> = 4.975 min; MS *m/z* 542.3 [M+1]<sup>+</sup>.

**trans-1-(4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)phenylsulfonamido)-N-(4-fluorophenyl)cyclopropanecarboxamide (30)**—The title compound **30** (0.009 g, 7%) was prepared according to general procedure B from (4-(*N*-(1-(4-fluorophenyl)carbamoyl)cyclopropyl)sulfamoyl)phenyl)boronic acid (0.160 g, 0.42 mmol, 2.0 eq) as a white solid. <sup>1</sup>H NMR (400MHz, CD<sub>3</sub>OD) δ 9.17 (s, 1H), 8.95 (s, 1H), 8.06 (d, *J* = 8.54 Hz, 2H), 7.96 (d, *J* = 8.33 Hz, 2H), 7.34–7.31 (m, 2H), 6.90–6.86 (m, 2H), 4.68–4.62 (m, 1H), 3.75–3.70 (m, 1H), 3.50 (t, *J* = 7.90 Hz, 2H), 2.24–2.13 (m, 4H), 2.06–2.03 (m, 2H), 1.71–1.64 (m, 2H), 1.58–1.38 (m, 6H), 1.10 (dd, *J* = 4.67, 7.89 Hz, 2H), 1.00 (t, *J* = 7.36 Hz, 3H); LC-MS: 96% purity, t<sub>R</sub> = 6.051 min; MS *m/z* 622.3 [M+1]<sup>+</sup>.

**4-(6-(Butylamino)-1-(trans-4-hydroxycyclohexyl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)-N,N-dimethylbenzenesulfonamide (31)**—The title compound **31** (0.052 g, 74%) was prepared according to general procedure E from (4-(*N,N*-dimethylsulfamoyl)phenyl)boronic acid (0.052 g, 0.225 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.94 (s, 1H), 8.07 (d, *J* = 8.4 Hz, 2H), 7.88 (d, *J* = 8.3 Hz, 2H), 5.34 (bs, 1H), 4.73–4.59 (m, 1H), 3.90–3.76 (m, 1H), 3.52 (dd, *J* = 13.0, 6.8 Hz, 2H), 2.75 (s, 6H), 2.29–2.15 (m, 4H), 2.13–2.02 (m, 2H), 1.66 (dt, *J* = 14.9, 7.3 Hz, 2H), 1.59–1.52 (t, *J* = 10.2 Hz, 3H), 1.47 (dd, *J* = 15.0, 7.4 Hz, 2H), 1.00 (t, *J* = 7.3 Hz, 3H); LC-MS: 97% purity, t<sub>R</sub> = 6.011 min; MS *m/z* 473.3 [M+1]<sup>+</sup>.

**N-(tert-Butyl)-4-(6-(butylamino)-1-(trans-4-hydroxycyclohexyl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)-N-methylbenzenesulfonamide (32)**—The title compound **32** (0.058 g, 76%) was prepared according to general procedure E from ((4-(*N*-(*tert*-butyl)-*N*-methylsulfamoyl)phenyl)boronic acid (0.061 g, 0.225 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.93 (s, 1H), 8.07–7.97 (m, 2H), 7.96–7.89 (m, 2H), 5.40 (bs, 1H), 4.73–4.58 (m, 1H), 3.91–3.76 (m, 1H), 3.52 (dd, *J* = 12.9, 6.9 Hz, 2H), 3.01 (s, 3H), 2.30–2.14 (m, 4H), 2.07 (d, *J* = 10.8 Hz, 2H), 1.68–1.44 (m, 7H), 1.38 (d, *J* = 4.8 Hz, 9H), 0.99 (t, *J* = 7.3 Hz, 3H); LC-MS: 96% purity, t<sub>R</sub> = 6.328 min; MS *m/z* 515.3 [M+1]<sup>+</sup>.

**trans-4-(3-(4-(Azetidin-1-ylsulfonyl)phenyl)-6-(butylamino)-1H-pyrazolo[3,4-d]pyrimidin-1-yl)cyclohexanol (33)**—The title compound **33** (0.054 g, 74%) was prepared according to general procedure E from (4-(azetidin-1-ylsulfonyl)phenyl)boronic acid (0.054 g, 0.225 mmol) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.96 (s, 1H), 8.11 (d, *J* = 8.3 Hz, 2H), 7.95 (d, *J* = 8.2 Hz, 2H), 5.33 (s, 1H), 4.75–4.57 (m, 1H), 3.92–3.74 (m, 5H), 3.53 (dd, *J* = 13.1, 6.7 Hz, 2H), 2.32–2.16 (m, 4H), 2.15–2.03 (m, 4H), 1.66 (dt, *J* = 14.8, 7.2 Hz, 2H), 1.55–1.50 (m, 3H), 1.45 (dd, *J* = 14.6, 7.3 Hz, 2H), 1.00 (t, *J* = 7.3 Hz, 3H); LC-MS: >97% purity, t<sub>R</sub> = 5.992 min; MS *m/z* 485.3 [M+1]<sup>+</sup>.

**trans-4-(6-(Butylamino)-3-(4-(pyrrolidin-1-ylsulfonyl)phenyl)-1H-pyrazolo[3,4-d]pyrimidin-1-yl)cyclohexanol (34)**—The title compound **34** (0.020 g, 27%) was

prepared according to general procedure E from (4-(pyrrolidin-1-ylsulfonyl)phenyl)boronic acid (0.096 g, 0.375 mmol) as a white solid.  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ ).  $\delta$  9.20 (s, 1H), 8.20 (d,  $J = 8.5$  Hz, 2H), 7.98 (d,  $J = 8.5$  Hz, 2H), 4.75–4.64 (m, 1H), 3.78–3.67 (m, 1H), 3.55 (t,  $J = 7.1$  Hz, 2H), 3.32–3.23 (m, 4H), 2.30–2.02 (m, 6H), 1.85–1.64 (m, 6H), 1.62–1.42 (m, 4H), 1.02 (t,  $J = 7.4$  Hz, 3H); LC-MS: >97% purity,  $t_{\text{R}} = 6.100$  min; MS  $m/z$  499.0  $[\text{M}+1]^+$ .

***trans*-4-(6-(Butylamino)-3-(4-(morpholin-sulfonyl)phenyl)-1H-pyrazolo[3,4-d]pyrimidin-1-yl)cyclohexanol (35, UNC1062)**—The title compound **35** (0.063 g, 82%) was prepared according to general procedure E from 4-((4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl)sulfonyl)morpholine (0.079 g, 0.225 mmol) as a white solid.  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ ).  $\delta$  9.20 (s, 1H), 8.26–8.20 (m, 2H), 7.95–7.89 (m, 2H), 4.75–4.64 (m, 1H), 3.79–3.68 (m, 1H), 3.73 (dd,  $J = 10.2, 5.7$  Hz, 4H), 3.55 (t,  $J = 7.1$  Hz, 2H), 3.08–2.97 (m, 4H), 2.36–2.01 (m, 6H), 1.76–1.65 (m, 2H), 1.62–1.42 (m, 4H), 1.02 (t,  $J = 7.4$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{DMSO}-d_6$ )  $\delta$  160.6, 155.2, 154.0, 140.6, 136.9, 133.7, 128.3, 127.0, 105.1, 68.0, 65.3, 55.0, 45.9, 40.3, 34.2, 30.6, 29.4, 19.6, 13.7; LC-MS: >97% purity,  $t_{\text{R}} = 5.657$  min; HRMS (TOF, ESI+)  $m/z$ :  $[\text{M}+H]^+$  calculated for  $\text{C}_{25}\text{H}_{35}\text{N}_6\text{O}_4\text{S}$ , 515.2440; found 515.2421.

***trans*-(1-((4-(6-(Butylamino)-1-(4-hydroxycyclohexyl)-1H-pyrazolo[3,4-d]pyrimidin-3-yl)phenyl)sulfonyl)piperidin-4-yl)(morpholino)methanone (36)**—The title compound **36** (0.065 g, 66%) was prepared according to general procedure E from (4-((4-(morpholine-4-carbonyl)piperidin-1-yl)sulfonyl)phenyl)boronic acid (0.091 g, 0.24 mmol) as a white solid.  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}+\text{CDCl}_3$ )  $\delta$  9.21 (s, 1H), 8.37–8.28 (m, 2H), 8.24–8.16 (m, 2H), 5.04–4.94 (m, 1H), 4.60–4.38 (m, 2H), 4.23–4.07 (m, 3H), 3.99–3.94 (m, 3H), 3.93–3.82 (m, 4H), 3.78–3.68 (m, 2H), 2.91–2.82 (m, 2H), 2.76 (ddd,  $J = 10.4, 7.2, 3.9$  Hz, 1H), 2.61–2.46 (m, 4H), 2.46–2.36 (m, 2H), 2.30–2.17 (m, 2H), 2.15–2.06 (m, 2H), 2.06–1.98 (m, 2H), 1.98–1.86 (m, 2H), 1.85–1.74 (m, 2H), 1.33 (t,  $J = 7.4$  Hz, 3H); LC-MS: 96% purity,  $t_{\text{R}} = 5.468$  min; MS  $m/z$  626.3  $[\text{M}+1]^+$ .

### Microfluidic Capillary Electrophoresis (MCE) Assay

Activity assays were performed in a 384 well, polypropylene microplate with a final volume of 50  $\mu\text{L}$  in 50 mM Hepes, Ph 7.4 containing 10 mM  $\text{MgCl}_2$ , 1.0 mM DTT, 0.01% Triton X-100, 0.1% Bovine Serum Albumin (BSA), using 1.0  $\mu\text{M}$  fluorescent substrate (table 3) and ATP at  $K_m$  for each enzyme (Table 3). All reactions were terminated by addition of 20  $\mu\text{L}$  of 70 mM EDTA. Phosphorylated and unphosphorylated substrate peptides (Table 3) were separated following a 180 min incubation on a LabChip EZ Reader equipped with a 12-sipper chip in separation buffer supplemented with 1 x CR-8 and analyzed using EZ Reader software.

### Morrison Ki Measurement

Inhibition of Mer kinase by **UNC1062** was measured in the MCE assay using the Morrison Tight-Binding Method<sup>17b</sup>. Reactions were performed in a 384 well, polypropylene microplate in a final volume of 80  $\mu\text{L}$  in 50 mM Hepes, Ph 7.4 containing 10 mM  $\text{MgCl}_2$ , 1.0 mM DTT, 0.01% Triton X-100, 0.1% Bovine Serum Albumin (BSA), and 5.0  $\mu\text{M}$  ATP. Compounds were tested in 20 dilution (1.5 fold) dose curves spanning a concentration range of 60 nM to 0.027 nM, 2.0 nM Mer, 1.0  $\mu\text{M}$  Peptide substrate, and 5.0  $\mu\text{M}$  ATP, final. To 40  $\mu\text{L}$  of enzyme (4 nM Mer), 8.0  $\mu\text{L}$  of compound in 10% DMSO was added and allowed to incubate for 10 min followed by addition of 32  $\mu\text{L}$  of substrate mix (containing 2.5  $\mu\text{M}$  peptide substrate and 12.5  $\mu\text{M}$  ATP).

Kinetic reads were taken from 7.0 to 209 min, 12 reads total, on an EZReader, using upstream voltage = -2250 V, downstream voltage = -500 V and pressure = -1.0 psi (sip time 0.2 sec, buffer 30 sec). The steady-state velocity was analyzed by linear regression of the Peptide as percent-conversion/min, and a plot of nM compound (**UNC1062**) vs. velocity was fit to the Morrison equation using GraphPad Prism 5.0.,  $Y = V_o * (1 - (((Et + X + (K_i * (1 + (S / K_m))))^2 - 4 * Et * X)^{0.5}) / (2 * Et))$  (Figure 5). The total enzyme concentration (Et) was fixed to 0.855 nM as determined from previous titration studies (data not shown).<sup>17b</sup> (Y = velocity, X = inhibitor concentration).

For **UNC1062**, the Morrison  $K_i$  was  $0.33 \pm 0.083$  nM (n=3, error reported as S.D.).

### HERG Binding Assay

Assay Buffer: Hank's Balanced Salt Solution, 20 mM HEPES, 2.5 mM probenecid, pH 7.4, membrane potential sensitive dye (Molecular Devices).

To screen for drugs that block or open HERG K<sup>+</sup> channels, we employ a fluorescence-based membrane potential assay (Molecular Devices). HEK293 cells stably expressing recombinant human HERG (provided by Dr. J. Overholt, Case Western Reserve University, Cleveland, OH; cells originally from Drs. A. Brown and E. Ficker, MetroHealth Medical Center, Cleveland, OH) are seeded in poly-L-lysine-coated 96-well plates (45,000 cells/100  $\mu$ L DMEM supplemented with 10% fetal bovine serum/well) one day prior to assay. The next day, the medium is removed and replaced with 30  $\mu$ L/well of assay buffer containing membrane potential dye (Molecular Devices) (the lyophilized dye is reconstituted with 15 mL of assay buffer). After a 15 min incubation at 37 degrees centigrade, 30  $\mu$ L/well of 2X dilutions of terfenadine (a known HERG blocker used as a reference compound) or test compound (final assay concentration ranging from 0.1 nM to 10  $\mu$ M) are added to the cells (each concentration assayed in triplicate). Baseline fluorescence (excitation 530 nm, emission 565 nm) is measured over 15 min, then 140  $\mu$ L of depolarization solution (143 mM KCl in distilled water) containing test or reference compound (1x) are added to the cells and fluorescence is recorded for 3 min. Raw fluorescence data are analyzed by GraphPad Prism 4.0.

### Cell Based Assays for Mer Kinase Inhibition

697, BT-12, A549, or Colo699 cells were cultured in the presence of **UNC1062** or vehicle only for 1.0 h (697, BT-12) or 72 h (A549, Colo699). Pervanadate solution was prepared fresh by combining 20 mM sodium orthovanadate in 0.9x PBS in a 1:1 ratio with 0.3% (w/w) hydrogen peroxide in PBS for 15–20 min at room temperature. Cultures were treated with 120  $\mu$ M pervanadate prior to collection for preparation of whole cell lysates, immunoprecipitation of Mer, and analysis by western blot.

697, A549, and Colo699 cells were treated with pervanadate for 3.0, 5.0, and 1.0 min respectively. Cell lysates were prepared in 50 mM HEPES pH 7.5, 150 mM NaCl, 10 mM EDTA, 10% glycerol, and 1% Triton X-100, supplemented with protease inhibitors (Roche Molecular Biochemicals, #11836153001). Mer protein was immunoprecipitated with a monoclonal anti-Mer antibody (R&D Systems, #MAB8912) and Protein G agarose beads (Invitrogen). Phospho-Mer was detected by western blot using a polyclonal anti-phospho-Mer antibody raised against a peptide derived from the tri-phosphorylated activation loop of Mer. Nitrocellulose membranes were stripped and total Mer protein was detected using a second anti-Mer antibody (Epitomics Inc., #1633-1). For 697 cells, relative phosphorylated and total Mer protein levels were determined by densitometry and IC<sub>50</sub> values were determined by non-linear regression.

Confluent BT-12 cultures were treated with pervanadate for 15 min. Cell lysates were prepared in 20 mM HEPES pH 7.5, 500 mM NaCl, 5.0 mM EDTA, 10% glycerol, and 1% Triton X-100, supplemented with protease inhibitors (10 µg/mL leupeptin, 10 µg/mL phenylmethylsulfonyl fluoride, and 20 µg/ml aprotinin) and phosphatase inhibitors (50 mM sodium fluoride and 1.0 mM sodium orthovanadate) and Mer protein was immunoprecipitated using a custom polyclonal rabbit anti-Mer antisera raised against a peptide derived from the N-terminus of Mer and Protein A agarose beads (Santa Cruz Biotechnology). Phosphotyrosine-containing proteins were detected by western blot with a monoclonal HRP-conjugated anti-phosphotyrosine antibody (Santa Cruz Biotechnology, #sc-508). Antibodies were stripped from membranes and total Mer levels were determined using a custom polyclonal rabbit anti-Mer raised against a peptide derived from the catalytic domain of Mer.

### Soft Agar Colony Formation Assays

BT-12 cells (10,000 cells) were cultured in 2.0 mL of 0.35% soft agar containing 0.5x RPMI medium, 7.5% FBS, and the indicated concentrations of **UNC1062** or DMSO vehicle only and overlaid with 0.5 mL of 1x RPMI medium containing 15% FBS and **UNC1062** or DMSO vehicle only. Medium and **UNC1062** or vehicle were refreshed 2 times per week. Colonies were stained with thiazolyl blue tetrazolium bromide (Sigma Aldrich, #M5655) and counted after 3 weeks.

A549 or Colo699 cells (15,000 cells) were cultured in 1.5 mL of 0.35% soft agar containing 1x RPMI medium and 10% FBS and overlaid with 2.0 mL of 1x RPMI medium containing 10% FBS and the indicated concentrations of **UNC1062** or DMSO vehicle only. Medium and **UNC1062** or vehicle were refreshed 3 times per week. Colonies were stained with nitrotetrazolium blue chloride (Sigma Aldrich, #N6876) and counted after 2 weeks.

### Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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### References

1. Chen J, Carey K, Godowski PJ. Identification of Gas6 as a ligand for Mer, a neural cell adhesion molecule related receptor tyrosine kinase implicated in cellular transformation. *Oncogene*. 1997; 14(17):2033–9. [PubMed: 9160883]
2. Linger, RMA.; Keating, AK.; Earp, HS.; Graham, DK. *Advances in Cancer Research*. Vol. 100. Academic Press; New York: 2008. TAM Receptor Tyrosine Kinases: Biologic Functions, Signaling, and Potential Therapeutic Targeting in Human Cancer; p. 35-83.
3. (a) Linger RMA, Keating AK, Earp HS, Graham DK. Taking aim at Mer and Axl receptor tyrosine kinases as novel therapeutic targets in solid tumors. *Expert Opin Ther Targets*. 2010; 14(10):1073–1090. [PubMed: 20809868] (b) Brandao L, Migdall-Wilson J, Eisenman K, Graham DK. TAM receptors in leukemia: expression, signaling, and therapeutic implications. *Critical reviews in oncogenesis*. 2011; 16(1–2):47–63. [PubMed: 22150307]

4. (a) Graham DK, Salzberg DB, Kurtzberg J, Sather S, Matsushima GK, Keating AK, Liang X, Lovell MA, Williams SA, Dawson TL, Schell MJ, Anwar AA, Snodgrass HR, Earp HS. Ectopic expression of the proto-oncogene Mer in pediatric T-cell acute lymphoblastic leukemia. *Clin Cancer Res.* 2006; 12(9):2662–9. [PubMed: 16675557] (b) Yeoh EJ, Ross ME, Shurtleff SA, Williams WK, Patel D, Mahfouz R, Behm FG, Raimondi SC, Relling MV, Patel A, Cheng C, Campana D, Wilkins D, Zhou X, Li J, Liu H, Pui CH, Evans WE, Naeve C, Wong L, Downing JR. Classification, subtype discovery, and prediction of outcome in pediatric acute lymphoblastic leukemia by gene expression profiling. *Cancer cell.* 2002; 1(2):133–43. [PubMed: 12086872] (c) Brandao LN, Wings A, Christoph S, Sather S, McGranahan A, Liang X, Gao D, DeRyckere D, Graham DK. Inhibition of MerTK increases chemosensitivity and decreases oncogenic potential in T-cell acute lymphoblastic leukemia. *Blood Cancer J.* 2013; 3:e101.10.1038/bcj.2012.46 [PubMed: 23353780]
5. Lee-Sherick AB, Eisenman KM, Sather S, McGranahan A, Armistead PM, McGary CS, Hunsucker SA, Schlegel J, Martinson H, Cannon C, Keating AK, Earp HS, Liang X, DeRyckere D, Graham DK. Aberrant Mer receptor tyrosine kinase expression contributes to leukemogenesis in acute myeloid leukemia. *Oncogene.* 2013 In press.
6. Linger RM, Cohen RA, Cummings CT, Sather S, Migdall-Wilson J, Middleton DH, Lu X, Baron AE, Franklin WA, Merrick DT, Jedlicka P, Deryckere D, Heasley LE, Graham DK. Mer or Axl receptor tyrosine kinase inhibition promotes apoptosis, blocks growth and enhances chemosensitivity of human non-small cell lung cancer. *Oncogene.* 2012
7. (a) Keating AK, Kim GK, Jones AE, Donson AM, Ware K, Mulcahy JM, Salzberg DB, Foreman NK, Liang X, Thorburn A, Graham DK. Inhibition of Mer and Axl Receptor Tyrosine Kinases in Astrocytoma Cells Leads to Increased Apoptosis and Improved Chemosensitivity. *Mol Cancer Ther.* 2010; 9(5):1298–1307. [PubMed: 20423999] (b) Rogers AE, Le JP, Sather S, Pernu BM, Graham DK, Pierce AM, Keating AK. Mer receptor tyrosine kinase inhibition impedes glioblastoma multiforme migration and alters cellular morphology. *Oncogene.* 2012; 31(38):4171–81. [PubMed: 22179835] (c) Wang Y, Moncayo G, Morin P Jr, Xue G, Grzmil M, Lino MM, Clement-Schatlo V, Frank S, Merlo A, Hemmings BA. Mer receptor tyrosine kinase promotes invasion and survival in glioblastoma multiforme. *Oncogene.* 2012:1–11.
8. (a) Schlegel JS, MJ, Sather S, Moschos SJ, Tan A-C, Wings A, DeRyckere D, Carson CC, Trembath DG, Tentler JJ, Eckhardt SG, Kuan P-F, Hamilton RL, Duncan LM, Alldredge V, Groben PA, Ollila DW, Thomas NE, Liu J, Zhang W, Yang C, Wang X, Frye SV, Earp HS, Shields JM, Graham DK. Mer receptor tyrosine kinase is a novel therapeutic target in melanoma. *J Clin Invest.* In press. (b) Tworokoski K, Singhal G, Szpakowski S, Zito CI, Bacchiocchi A, Muthusamy V, Bosenberg M, Krauthammer M, Halaban R, Stern DF. Phosphoproteomic screen identifies potential therapeutic targets in melanoma. *Molecular cancer research : MCR.* 2011; 9(6):801–12. [PubMed: 21521745]
9. Keating AK, Salzberg DB, Sather S, Liang X, Nickoloff S, Anwar A, Deryckere D, Hill K, Joung D, Sawczyn KK, Park J, Curran-Everett D, McGavran L, Meltesen L, Gore L, Johnson GL, Graham DK. Lymphoblastic leukemia/lymphoma in mice overexpressing the Mer (MerTK) receptor tyrosine kinase. *Oncogene.* 2006; 25(45):6092–100. [PubMed: 16652142]
10. (a) Huang X, Finerty P Jr, Walker JR, Butler-Cole C, Vedadi M, Schapira M, Parker SA, Turk BE, Thompson DA, Dhe-Paganon S. Structural insights into the inhibited states of the Mer receptor tyrosine kinase. *J Struct Biol.* 2009; 165(2):88–96. [PubMed: 19028587] (b) Greshock J, Bachman KE, Degenhardt YY, Jing J, Wen YH, Eastman S, McNeil E, Moy C, Wegrzyn R, Auger K, Hardwicke MA, Wooster R. Molecular Target Class Is Predictive of In vitro Response Profile. *Cancer Res.* 2010; 70(9):3677–3686. [PubMed: 20406975] (c) Howard S, Berdini V, Boulstridge JA, Carr MG, Cross DM, Curry J, Devine LA, Early TR, Fazal L, Gill AL, Heathcote M, Maman S, Matthews JE, McMenamin RL, Navarro EF, O'Brien MA, O'Reilly M, Rees DC, Reule M, Tisi D, Williams G, Vinkovic M, Wyatt PG. Fragment-based discovery of the pyrazol-4-yl urea (AT9283), a multitargeted kinase inhibitor with potent aurora kinase activity. *J Med Chem.* 2009; 52(2):379–88. [PubMed: 19143567] (d) Schroeder GM, An Y, Cai ZW, Chen XT, Clark C, Cornelius LA, Dai J, Gullo-Brown J, Gupta A, Henley B, Hunt JT, Jeyaseelan R, Kamath A, Kim K, Lippy J, Lombardo LJ, Manne V, Oppenheimer S, Sack JS, Schmidt RJ, Shen G, Stefanski K, Tokarski JS, Trainor GL, Wautlet BS, Wei D, Williams DK, Zhang Y, Fargnoli J, Borzilleri RM. Discovery of N-(4-(2-amino-3-chloropyridin-4-yloxy)-3-fluorophenyl)-4-ethoxy-1-(4-

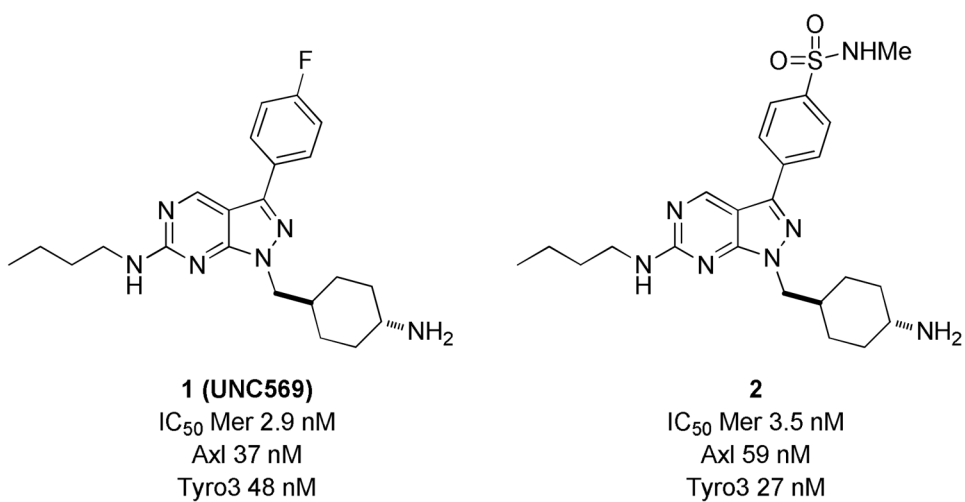
fluorophenyl)-2-oxo-1,2-dihydropyridine-3-carboxamide (BMS-777607), a selective and orally efficacious inhibitor of the Met kinase superfamily. *J Med Chem.* 2009; 52(5):1251–4. [PubMed: 19260711]

11. Liu J, Yang C, Simpson C, DeRyckere D, Van DA, Miley MJ, Kireev D, Norris-Drouin J, Sather S, Hunter D, Korboukh VK, Patel HS, Janzen WP, Machius M, Johnson GL, Earp HS, Graham DK, Frye SV, Wang X. Discovery of Small Molecule Mer Kinase Inhibitors for the Treatment of Pediatric Acute Lymphoblastic Leukemia. *ACS Med Chem Lett.* 2012; 3:129–134. [PubMed: 22662287]
12. (a) Dubin AE, Nasser N, Rohrbacher J, Hermans AN, Marrannes R, Grantham C, Van Rossem K, Cik M, Chaplan SR, Gallacher D, Xu J, Guia A, Byrne NG, Mathes C. Identifying modulators of hERG channel activity using the PatchXpress planar patch clamp. *Journal of biomolecular screening.* 2005; 10(2):168–81. [PubMed: 15799960] (b) Witchel HJ, Milnes JT, Mitcheson JS, Hancox JC. Troubleshooting problems with in vitro screening of drugs for QT interval prolongation using HERG K+ channels expressed in mammalian cell lines and *Xenopus* oocytes. *Journal of pharmacological and toxicological methods.* 2002; 48(2):65–80. [PubMed: 14565563]
13. (a) Priest B, Bell IM, Garcia M. Role of hERG potassium channel assays in drug development. *Channels.* 2008; 2(2):87–93. [PubMed: 18849661] (b) Redfern WS, Carlsson L, Davis AS, Lynch WG, MacKenzie I, Palethorpe S, Siegl PK, Strang I, Sullivan AT, Wallis R, Camm AJ, Hammond TG. Relationships between preclinical cardiac electrophysiology, clinical QT interval prolongation and torsade de pointes for a broad range of drugs: evidence for a provisional safety margin in drug development. *Cardiovascular research.* 2003; 58(1):32–45. [PubMed: 12667944] (c) Picard S, Lacroix P. QT interval prolongation and cardiac risk assessment for novel drugs. *Curr Opin Investig Drugs.* 2003; 4(3):303–8. (d) De Ponti F, Poluzzi E, Cavalli A, Recanatini M, Montanaro N. Safety of non-antiarrhythmic drugs that prolong the QT interval or induce torsade de pointes: an overview. *Drug safety : an international journal of medical toxicology and drug experience.* 2002; 25(4):263–86. [PubMed: 11994029]
14. Liu J, Wang X. Microwave-Assisted, Divergent Solution-Phase Synthesis of 1,3,6-Trisubstituted Pyrazolo[3,4-d]pyrimidines. *ACS Combinatorial Science.* 2011; 13(4):414–420. [PubMed: 21528879]
15. (a) Pommereau A, Pap E, Kannt A. Two simple and generic antibody-independent kinase assays: comparison of a bioluminescent and a microfluidic assay format. *Journal of biomolecular screening.* 2004; 9(5):409–16. [PubMed: 15296640] (b) Dunne J, Reardon H, Trinh V, Li E, Farinas J. Comparison of on-chip and off-chip microfluidic kinase assay formats. *Assay Drug Dev Technol.* 2004; 2(2):121–9. [PubMed: 15165508] (c) Bernasconi P, Chen M, Galasinski S, Popa-Burke I, Bobasheva A, Coudurier L, Birkos S, Hallam R, Janzen WP. A chemogenomic analysis of the human proteome: application to enzyme families. *Journal of biomolecular screening.* 2007; 12(7):972–82. [PubMed: 17942790]
16. Wang, X.; Liu, J.; Yang, C.; Zhang, W.; Frye, S.; Kireev, D. Preparation of pyrazolopyrimidine compounds for the treatment of cancer. WO2011146313A1. 2011.
17. (a) Morrison JF. Kinetics of the reversible inhibition of enzyme-catalysed reactions by tight-binding inhibitors. *Biochimica et biophysica acta.* 1969; 185(2):269–86. [PubMed: 4980133] (b) Copeland RA. Evaluation of enzyme inhibitors in drug discovery. A guide for medicinal chemists and pharmacologists. *Methods of biochemical analysis.* 2005; 46:1–265. [PubMed: 16350889]

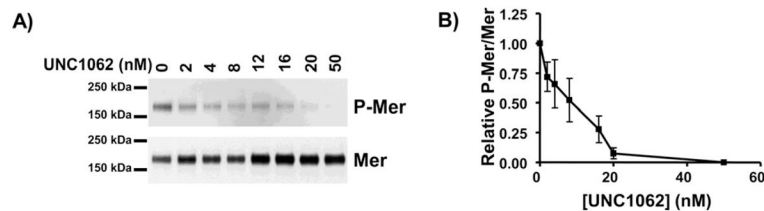


### Highlights

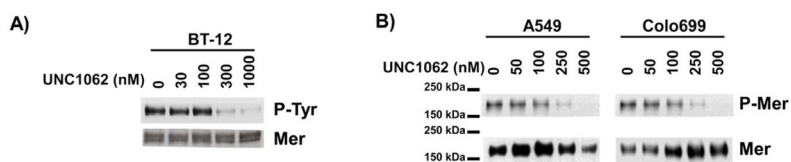
1. Discovery of pyazolopyrimidine sulfonamides as potent and selective Mer inhibitors.
2. **UNC1062** selectively inhibits Mer both in purified enzymes and intact cell systems.
3. **UNC1062** inhibits the colony formation of multiple tumor cell lines in soft agar.
4. The anti-tumor activity of **UNC1062** further validates Mer as a therapeutic target.



**Figure 1.**  
The structures and enzymatic activities of **1** and **2**.



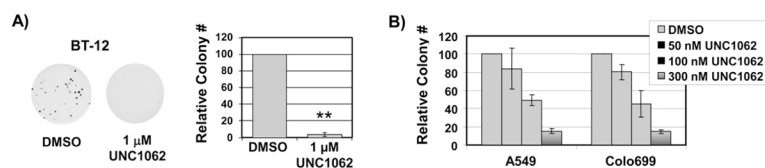
**Figure 2. UNC1062 inhibits accumulation of activated Mer protein in acute leukemia cells** 697 cell cultures were treated with the indicated concentrations of **UNC1062** for 1 h. Pervanadate was added to cultures for 3 min to stabilize the phosphorylated form of Mer. Mer was immunoprecipitated from cell lysates and total Mer protein and Mer phosphoprotein were detected by immunoblot. **A)** Representative western blots. **B)** Relative levels of phospho-Mer and Mer proteins were determined. Mean values  $\pm$  standard error derived from 4 independent experiments are shown.  $IC_{50} = 6.4$  nM with a 95% confidence interval of 3.8 nM to 10.7 nM.



**Figure 3. UNC1062 inhibits Mer activation in adherent tumor cell lines**

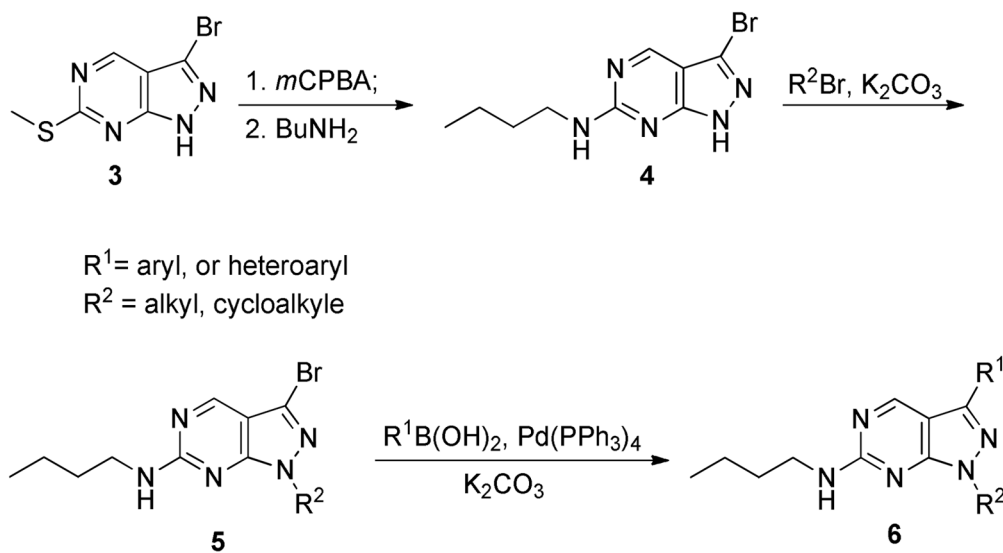
**A)** BT-12 pediatric rhabdoid tumor cells were treated with **UNC1062** or vehicle only for 1 h prior to the addition of pervanadate for 15 min. Cell lysates were prepared, Mer protein was immunoprecipitated, and phospho-tyrosine and Mer protein were detected by western blot.

**B)** The indicated NSCLC cell lines were treated with **UNC1062** or an equivalent volume of vehicle only for 72 h prior to treatment with pervanadate for 1.0 (Colo699) or 5.0 (A549) min. Whole cell lysates were prepared, Mer protein was immunoprecipitated, and phosphorylated and total Mer proteins were detected by western blot. The results shown are representative of 3 independent experiments.



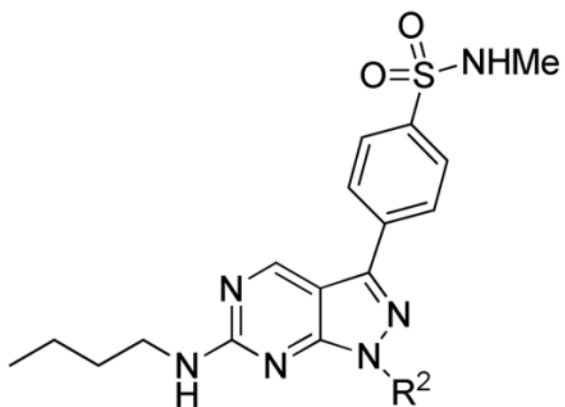
**Figure 4. UNC1062 inhibits colony formation in solid tumor cell lines**

**A)** BT-12 rhabdoid tumor cells were cultured in soft agar containing 1.0 μM **UNC1062** or vehicle only and overlaid with medium containing **UNC1062** or vehicle only. Medium and **UNC1062** were refreshed twice weekly. Colonies were stained and counted. Representative plates are shown. **B)** A549 and Colo699 NSCLC cells were cultured in 0.35% soft agar overlaid with medium containing **UNC1062** or vehicle only. Medium and **UNC1062** were refreshed 3 times per week. Colonies were stained and counted. Mean values ± SEM derived from 3 independent experiments are shown. \*Statistically significant results were determined using the student's paired t test (\*  $p < 0.02$ , \*\*  $p < 0.001$  relative to vehicle only).

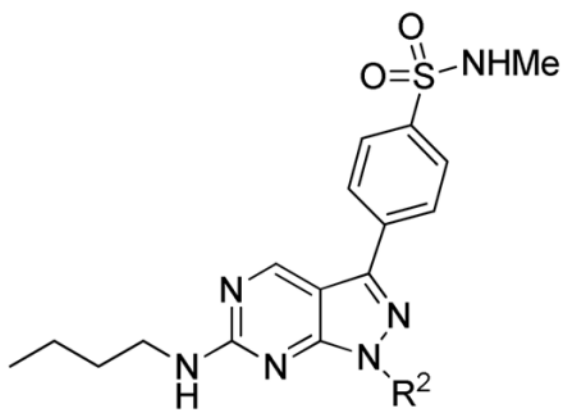


**Scheme 1.**  
The synthetic routes for sulfonamide analogues

Table 1

Preliminary SAR at the R<sup>2</sup> position

Compound	R <sup>2</sup>	IC <sub>50</sub> (nM) <sup>a</sup>		
		Mer	Axl	Tyro3
2		3.5	59	27
7		18	140	130
8		35	120	200
9		50	190	82
10		2.4	74	47
11		4.4	70	44
12		250	2400	860
13		130	5000	730
14		23	450	280

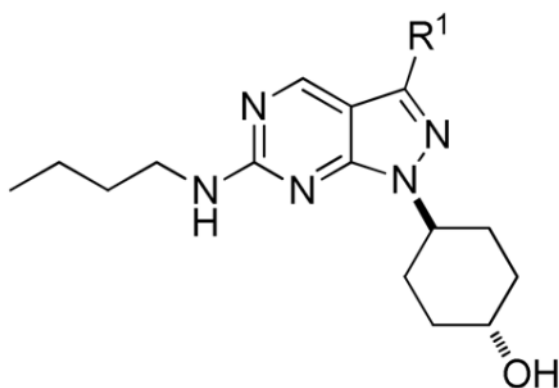


Compound	R <sup>2</sup>	IC <sub>50</sub> (nM) <sup>a</sup>		
		Mer	Axl	Tyro3
15		47	620	810
16		54	360	430
17		100	26000	720

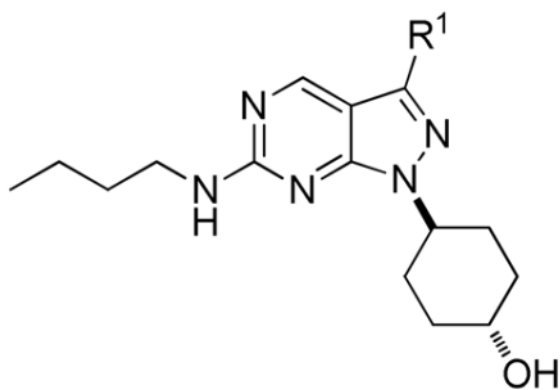
<sup>a</sup>Values are the mean of two or more independent assays performed at the ATP Km.



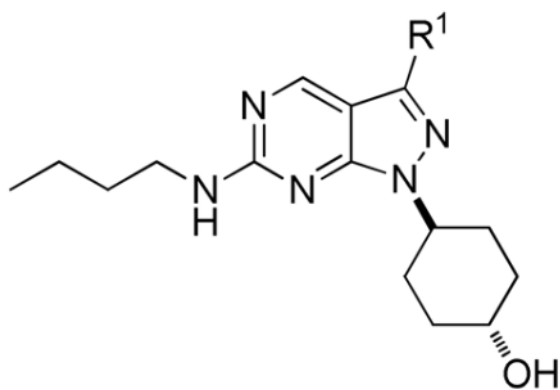
Table 2

SAR Study of R<sup>1</sup>

Compound	R <sup>1</sup>	IC <sub>50</sub> (nM) <sup>a</sup>		
		Mer	Axl	Tyro3
18		4.0	110	35
19		1.8	130	55
20		1.4	29	54
21		1.8	62	35
22		3.2	120	68
23		1.1	75	38
24		0.80	67	31



Compound	R <sup>1</sup>	IC <sub>50</sub> (nM) <sup>a</sup>		
		Mer	Axl	Tyro3
25		1.1	350	45
26		1.2	69	28
27		9.7	6500	290
28		1.1	55	47
29		1.4	73	150
30		18	780	320
31		1.8	71	47
32		1.2	110	160



Compound	R <sup>1</sup>	IC <sub>50</sub> (nM) <sup>a</sup>		
		Mer	Axl	Tyro3
33		1.0	42	27
34		0.65	46	22
35		1.1	85	60
36		1.4	23	76

<sup>a</sup>Values are the mean of two or more independent assays performed at the ATP Km.

**Table 3**

Assay conditions for MCE assays currently used

<b>Kinase</b>	<b>Peptide Substrate</b>	<b>Kinase (nM)</b>	<b>ATP (uM)</b>
Mer	5-FAM-EFPIYDFLPAKKK-CONH <sub>2</sub>	2.0	5.0
Axl	5-FAM-KKKKEEIYFFF-CONH <sub>2</sub>	120	65
Tyro	5-FAM-EFPIYDFLPAKKK-CONH <sub>2</sub>	10	21