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Th17 cell based vaccines in mucosal immunity

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Abstract

Vaccination is proven to be effective in controlling many infections including small pox, influenza and hepatitis, but strain-specific factors may limit vaccine efficacy. All of these vaccines work through the generation of neutralizing antibodies but for some pathogens there may be roles for serotype-independent immunity. Recently several groups using murine vaccine models have shown that induced T helper cell responses including Th17 responses have shown the potential for CD4+ T-cell dependent vaccine responses. Th17 mediated protective responses involve the recruitment of neutrophils, release of anti-microbial peptides and IL-17-driven Th1 immunity. These effector mechanisms provide immunity against a range of pathogens including the recently described antibiotic-resistant metallo-beta-lactamase 1 *Klebsiella pneumoniae*. Continued elucidation of the mechanism of Th17 responses may lead to successful Th17 based vaccines. Here we summarize the recent advances in understanding the role of Th17 in vaccine induced immunity. We also discuss the current status and future challenges in Th17-based mucosal vaccine development.

Introduction

T cells and B cells, the two major lymphocyte populations, mediate the generation of adaptive immunity. Activation of naïve T cells by their cognate antigens presented by antigen presenting cells (APCs) in the presence of various cytokines leads to the generation of distinct effector T helper (Th) cell-subsets including Th1, Th2 and Th17 [1]. In general, the Th1 subset regulates IFN γ -dependent immunity against most intracellular pathogens. Th1 cell differentiation can be inhibited by IL-4, which subsequently induces another T cell subset: Th2. Th2 cells produce IL-4, IL-5, IL-13 and are required for protection against helminth infection [2, 3]. Th17 cell differentiation requires TGF β and IL-6 in mice, and IL-1 and IL-23 in humans [1]. Th17-derived effector cytokines especially IL-17A, IL-17F and IL-22 are critical for host defense against bacterial and fungal infection [4–7]. In contrast these Th17 cells can play a pathological role in auto-immune diseases [8]. B cell clonal expansion, differentiation and isotype switch responses are result of interaction between cognate antigen specific activated T cells and B cells in the secondary lymphoid organ [9,

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10]. Production of IgG2a antibodies are controlled by Th1 responses, whereas Th2 cell responses promote IgG1 and IgE antibody class switching. Recent studies have documented role of Th17 cells in regulating B cells antibody generation, germinal center and ectopic inducible bronchus-associated lymphoid tissue (iBALT) formation [11–13]. Vaccination refers to administrating specific antigenic substance to induce protective immune responses (antibody dependent or independent) against the microbes. Many mucosal vaccination approaches can induce robust Th17 responses, suggesting Th17 cells may be useful targets for vaccine induced immunity. Here, we discuss recent advances in understanding of Th17 cells in vaccination-induced immunity.

Th17 cells and vaccines: where do we stand now?

Most, if not all U.S. Food and Drug Administration (FDA) approved vaccines are antibody based, such as small pox, influenza and hepatitis, and pneumococcal vaccines. These vaccines have successfully eliminated and controlled many infectious diseases. Only a limited number of vaccines mediate protection via T-cell dependent immunity. Both Th1 responses induced by whole cell pertussis vaccines and Th2-biased responses induced by pertussis acellular vaccines protect against *Bordetella pertussis* infection. Protection by immunization with whole cells pertussis vaccine could also be mediated by enhancement of Th17 dependent bactericidal activity of macrophages [14]. Antibody based vaccines usually have limited protective spectrum since the responses are restricted to the vaccination strains. For example, a recent clinical trial (conducted in Europe, USA, Philippines and South Africa) showed the efficacy of 7-valent, 9-valent or 11-valent pneumococcal conjugate vaccine (PCV) in preventing invasive pneumococcal and World Health Organization (WHO) radiographically defined pneumonia is approximately 80% and 27% respectively [15]. Protection is predominantly mediated by T-cell independent antibody responses, but failure to mount antibody response to some serogroup (PCV7 is minimally effective against 6B and 19F serotype) could be the reason for the poor efficacy or inconsistent response of vaccine [16]. Furthermore, in order to enhance immunogenicity against pneumococcal strains that cause meningitis or pneumonia, polysaccharide conjugate (with Diphtheria proteins) vaccines have been engineered. The role of T cell immunity induced by these conjugate vaccines remains to be determined but clearly these conjugate vaccines can elicit strong T-cell responses [17]. In contrast, T cell based vaccines have the potential to provide serotype-independent protection by recognizing antigens conserved cross species and thus have been investigated by many researchers most recently [18].

Pathogen-specific Th17 vaccines

Th17 cells are described as an initiator of pro-inflammatory responses in many autoimmune disease conditions [8, 19]. More recently, it has been appreciated that Th17 responses can also induce protective immunity against many bacterial and fungal pathogens [6, 20–22]. Indeed, vaccination in many mouse models induced significant Th17 responses in the lung and neutralization of IL-17 or blocking its downstream signaling pathways resulted in higher pathogen burden and mortality [5, 21, 23]. Pneumonia is most common cause of death induced by many infectious agents (CDC). *Klebsiella pneumoniae* infection commonly occurs in immune-compromised patients and is a concern for increasing resistance to carbapenem antibiotics. *K. pneumoniae* infection induces IL-17, resulting in production of IL-17-targeted cytokines in the lung [6, 24]. Furthermore, overexpression of IL-17 by adenovirus resulted in enhanced clearance of bacteria [24], suggesting the induction of IL-17 can effectively vaccinate against *K. pneumoniae*. Indeed, immunization with heat killed *K. pneumoniae* induces antibody response against capsular polysaccharides as well as a concomitant Th17 response. However, antibody response offers little or no protection against heterologous strains having different polysaccharide serotypes, whereas Th17 cells

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are sufficient and required for serotype independent heterologous protection [6]. Vaccination with highly conserved outer membrane proteins of *K. pneumoniae* also elicits a strong Th17 response and provides heterologous protection against a range of different strains including the newly described metallo-beta-lactamase 1 strain [6]. Vaccine-induced immunity against *K. pneumonniae* required neutrophils, but could also have involved the generation of Th17-dependent anti-microbial proteins. Both mechanisms require a functional heterodimeric IL-17 receptor, formed by IL-17RA and IL-17RC. Further studies using IL-17 receptor conditional knockout mice are useful to explore the molecular mechanism and cellular targets of Th17 mediated immunity against *K. pneumoniae*.

Both antibody-dependent and -independent mechanisms are involved for immunity against Streptococcus pneumonia. Antibody confers protection against capsular polysaccharide antigens; however, antibody-independent CD4 responses are generated against cell wall polysaccharide antigens [25]. Because polysaccharide antigens are poorly immunogenic in children (< 2 years), newer polysaccharide-based vaccine include a carrier protein (immunogenic non-pneumococcal protein) to induce adaptive immune responses. Moreover, conjugate (covalently attached carrier protein to polysaccharides) pneumococcal vaccine (PCV13) have been developed, and are able to provide protection against prevalent serotypes for use in children 6 weeks to 17 years age; however, other strains of pneumococcus also impose significant public health threats. Recent studies suggest that Tcell responses may also be required for vaccine-induced protection. Indeed, anti-capsular antibody titers did not correlate with experimental pneumococcal carriage [26, 27]. Thus, there is a need to identify surface antigens expressed in all major pathogenic pneumococcal strains, which may be capable of eliciting antibody independent protection against all the pathogenic serotypes. Indeed, antigen specific CD4 T cells limit nasopharyngeal colonization of S. pneumoniae[28]. Furthermore, CD4 T cell-derived IL-17, but not IFNy or IL-4, is required for the clearance of Pneumococcal colonization [23]. The role of CD4derived IL-17 in clearance of pneumococcal colonization was further demonstrated in another study, where IL-17 dependent recruitment of monocytes/macrophages and neutrophils limited the bacteria burden [22]. Furthermore, immunization with pneumococcal whole cell antigen and several derivatives provided IL-17-mediated, but not antibody dependent, protection [25, 29]. Indeed, Th17 effector responses provide serotype independent immunity by recruitment of monocytes, macrophages and neutrophils, thus having the potential to be used for designing novel pneumococcal vaccines [22].

Recently its has also been shown that a live attenuated vaccine against *Y. pestis* induces a robust Th17 response in the lung [21]. In this setting, the majority of IL-17 producing CD4 cells also produce IFN γ and TNF α . Neutralization of IL-17 or IFN γ resulted into reduced survival, suggesting IL-17 contributes to vaccine induced immunity against *Y. pestis*. Interestingly, IL-17 did not reduce bacterial burden in the lung tissue, and was effective only after booster vaccination. Moreover, in this study IL-17-mediated immunity was independent of neutrophil recruitment to the mucosal site; but was dependent on an IL-17 mediated increase in the antimicrobial protein lipocalin 2 [21].

Pseudomonas aeruginosa is a gram negative rod responsible for many cases of hospitalacquired pneumonia. Primary experimental infection with the lab strain of *P. aeruginosa*, PA01 resulted in rapid IL-17 induction [30] and neutralizing IL-17 worsened lung pathology and enhanced bacteria burden, suggesting a protective role of IL-17 in this model [30]. Interestingly, fewer neutrophils were recruited to the mucosal sites after anti-IL-17 treatment, although it was not investigated if neutrophils were required for protection in this study. In contrast, Dubin et al found that although both IL-23 and IL-17RA signaling was critical in regulating early neutrophil emigration into the lung in response to PA01 *P. aeruginosa* infection, both IL-17 and IL-17RA signaling were dispensable for controlling

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bacterial burden [31]. A live-attenuated vaccine against *P. aeruginosa* using PA14 Δ aroA (an aroA deletion in the PA14 strain), has been shown to induce a robust IL-17 response, and neutralization of IL-17 or studies in IL-17R knockout resulted in less neutrophil recruitment and higher mortality in immunized mice against LPS heterologous strain (PAO1 ExoU+) of *P. aeruginosa*[32]. A recent study identified several antigens from *P. aeruginosa* that were capable of inducing Th17 responses, and have demonstrated Th17 dependent immunity after immunization with one of these antigens, PopB [33]. However, a recent study documented a pro-inflammatory role of IL-17 in *P. aeruginosa* induced in an ulcerative keratitis model [34], suggesting protective or inflammatory roles of Th17 cells may be context dependent.

Other microbes for which Th17 cells are protective in vaccine-induced mucosal immunity include Mycobacterium tuberculosis, Bordetella pertussis, Helicobactor pylori and influenza virus. The protective role of IL-17 during the memory recall phase of *M. tuberculosis* infection has been thoroughly investigated and reviewed elsewhere [35]. Vaccination with the mycobacterial peptide (ESAT-6(1-20) followed by a subsequent aerosol *M. tuberculosis* challenge resulted in the early generation of IL-17-producing CD4+ T cells and late generation of IFNy producing CD4 T cells. Interestingly, the early IL-23/IL-17 axis is required for the recruitment of IFNy producing T cells by chemokines (CXCL9, CXCL10 and CXCL11 [36]. The World Health Organization has estimated the prevalence of H. pylori infection in fifty percent of World and 70% of Asian populations, which poses a significant health care and economic burden. Current treatment includes usage of proton-pump inhibitors and antibiotics. Recombinant *H pylori* antigens such as VacA, CagA and NAP are currently in clinical trials. In an experimental animal model, vaccination with Hp-SS1 lysate with cholera toxin adjuvant confers protection against H. pylor/[37]. Furthermore, elevated levels of IL-17 have been reported in the gastric mucosa of *H. pylori* infected patients [38], suggesting that IL-17 is a physiological response to *H. pylori* colonization. Indeed, immunization with H.pylori lysate, confers IL-17 and neutrophil dependent protection against a subsequent infectious challenge [37]. Furthermore, IL-17 dependent protection has also been reported in a mouse model of immunization with H. pylori urease [39], although this study did not specify the underlying mechanism of IL-17-dependent protection.

Th17 responses also provide protective immunity against fungal pathogens. A dominant stable antigen specific Th17 recall response is required for clearance of fungus Candida albicans [7]. Interestingly, both Th1 and Th17 response are induced during primary fungal infection but mice lacking IL-17 receptor are more susceptible to Candida infection, suggesting Th17 cells plays a dominant role in anti-fungal immunity [20]. Interestingly, IL-23 driven IL-17 response have also been reported to promote inflammation during infections with *Candida albicans* or *Aspergillus fumigatus*[40]. A recent study shows that Th17 mediated immunity is required for the protection against *Blastomyces dermatitidis, C. posodasii* and *H. Capsulatum*[5]. They further show that IL-17 mediated immunity is conferred by the recruitment and activation of PMN and macrophages [5]. To date, only a limited number of studies implicate the direct role of antigen specific memory Th17 cells in clearance of these fungal pathogens. Future studies require the understanding of plastic nature of Th17 cells *in vivo* during pathogenesis of disease. It is possible that although Th17 cells confer robust protection in most of the diseases, the inflammatory milieu may result in conversion to other T helper subsets.

Mechanisms of protection

Th17-mediated host defense against microorganisms can be classified into many mechanisms and are depicted in Figure 1, including initiating the release of antimicrobial peptides by epithelial cells, recruitment of neutrophils and/or macrophages, initiation of humoral immunity, and augmenting other T helper subsets. Vaccine induced protection can

be achieved in a serotype dependent manner through T-cell independent B-cell/antibody responses (Figure 1A). In contrast, recent studies have shed light on vaccine induced serotype independent protection mediated by Th17 cells (Figure 1B). Receptors for Th17-derived cytokines are mainly expressed by epithelial cells and fibroblasts. IL-17 targeted genes include G-CSF, CXCL-1, CCL2 and various other chemokines, resulting in the recruitment of neutrophils and macrophages to mucosal sites [41]. Importance of IL-17 dependent neutrophil recruitment was further demonstrated by adoptive transfer and depletion strategy, demonstrating that IL-17-dependent neutrophil recruitment is required for immunity against pneumococcus [23] and *H. pylori* infection [39].

Studies have indicated the role of IL-17 in activating B cell antibody response and germinal center formation [11, 12]. Blockade of IL-17 signaling resulted in inhibition of antibody class switch [13]. Furthermore, IL-17 is responsible for lymphotoxin independent inducible bronchus-associated lymphoid tissue (iBALT) formation in the lung [42], promoting B cell and T cell interactions [43]. In addition, Th17-cells enhance influx of B cells in the lung and promote polymeric immunoglobulin receptor (pIgR) expression on bronchial epithelial cells [44]. pIgR regulates the transport of IgA and IgM into the airway lumen [44]. Despite these findings, there is no direct evidence of IL-17 dependent antibody responses after vaccination. Interestingly, conventional influenza vaccines appear less effective for the prevention of clinical illness in the elderly population [45] and circulating IL-17 levels were found decreasing with age [46], suggesting that IL-17 might be involved in influenza vaccine induced antibody responses.

Th17 cells are maintained as effector memory cells in mucosal tissue for a very long period [47]. Indeed, superantigens from *S. aureus* induce a robust IL-17 response from memory Th17 cells in adult human but not from naïve T cells [48]. Interestingly, CD4+ T cells isolated from cord blood or from infants were poor producers of IL-17 *in vitro*, however authors did not provide molecular mechanism for this poor T-cells response [48]. Th17 cells are highly plastic in nature, and the cytokine milieu at mucosal cites is capable of transforming Th17 cells into either the Th1 or Th2 lineage. For example, IFN γ or IL-12 can convert Th17 cells into IFN γ producing Th1 cells [49]. Although Th17 cells can be unstable under Th1 inflammatory conditions, stable long lived memory Th17 cells are induced following vaccination in the absence of inflammation [50].

Adjuvants

Adjuvants are inorganic or organic additives to vaccines for generating optimal immune responses to antigens. Aluminum-based adjuvants are the most commonly used additive in FDA approved vaccines. Hepatitis A, hepatitis B, Haemophilus influenza type B, pneumococcal and diphtheria-tetanus-pertussis vaccine immune responses are boosted by use of alum salts (CDC). Aluminum-based adjuvants have been used in vaccine formulation for more than 90 years; however, the mechanism of actions of adjuvants is still poorly understood. Possible mechanism of adjuvants are enhanced antigen uptake and presentation by activated antigen presenting cells (APCs), which leads to initiation of antigen specific T cells responses [51]. Activation of APCs such as macrophages and dendritic cells involves activation of Toll-like receptors (TLRs) and inflammasome pathways [52-54]. In addition, Th2 cell and their specific B cells antibody response (IgG1 and IgE) are preferentially induced by alum [55]. To date, how inflammasome activation contributes to alum-based Th2 response is ill-defined. Interestingly, recent findings report that alum adjuvant can enhance the release of uric acid, which subsequently activates the inflammasome via both direct and indirect mechanisms [56, 57]. A recent study documented the role of uric acid in inflammasome-dependent Th17 differentiation [58], suggesting a possible mechanism that Th17 cells are promoted in alum formulated vaccines. Furthermore, activation of the

inflammasome has been shown to induce IL-1 β and IL-18 production from APCs and both of these cytokines are known to enhance Th17 responses [59]. Interestingly, non-alum based adjuvants such as a nanoemulsion, incomplete Freund's adjuvants, monophospho lipid A (MPL)-trehalose dimycolate (TDM), heat labile enterotoxin and cholera toxin have been reported to induce Th17 responses in experimental models [36, 60-65]. MPL-TDM is stable oil in water emulsion derived from cell wall component of bacterial and mycobacterium, thus capable of inducing strong adaptive Th1 and Th17 immune responses via activation of TLR4 on innate immune cells [36, 63, 66]. E. coli heat labile enterotoxin have also been reported to induce protective Th17 immune responses against *B. pertussis*[67] and *M. tuberculosis*[65]. Heat labile enterotoxin induced Th17 responses appeared to be mediated through activation of inflammasome and IL-1 and IL-23 release from dendritic cells [67]. Similarly, cholera toxin have been reported to induce Th17 responses by dendritic cells through cAMP dependent secretion of IL-1 β [61]. However, the major limitation of nonalum based adjuvants is concern for toxicity. A strong Th17 cell response could be induced or enhanced by co-administration of antigen and recombinant protein such as IL-1β, IL-6 or IL-23 as adjuvants. Indeed, this strategy has been successfully utilized in pre-clinical models to increase the efficacy of helper function of young and aged CD4 T cells [68].

Vaccine Safety

Despite the requirement of IL-17 in vaccine induced immunity in experimental animal models, its role in vaccination induced protection in humans remains ill-defined. The proinflammatory role of IL-17 is well documented in auto-immune diseases [8, 19]. Thus, induction of Th17 responses in autoimmune patients may aggravate underlying disease conditions. IL-17 has also been shown to be responsible for the immunopathology seen in influenza infected mice [69]. Thus, elevated IL-17 production could be harmful during an active flu infection. Another Th17-derived cytokine, IL-22, has been reported to induce psoriasis like skin inflammation and can be a safety concern for Th17 based vaccines [70]. Indeed, mucosal IL-22 producing Th17 cells are elevated in heat-killed K. pneumoniae immunized mice [6], whether these cells are pathogenic in autoimmune disease settings has not been investigated. IL-22 has also been reported to play a critical role in vaccine induced immunity against Mycobacterium [71], its protective or inflammatory role in other vaccination model has to be determined. Aside from the concern of the pro-inflammatory nature of Th17 effector cytokines, Th17 promoting adjuvants also raise safety concerns. Many experimental adjuvants such as cholera toxin, pertussis toxin and complete Freund's adjuvants induce potent Th-17 responses [61, 72, 73], but underlying toxicity may limit the use of these adjuvants. Recently, TLRs have been shown to respond to endogenous host molecules and trigger inflammatory responses in addition to recognizing pathogens [74]. Thus, adjuvants that may induce endogenous TLR ligands from the host should be carefully examined. Candidate antigens for Th17 based vaccines should also undergo stringent selection processes. Antigens or their homologs found in commensal species may need to be excluded due to the concern of autoimmunity. One recent study showed that not all IL-17 producing Th17 cells can induce pathogenic responses. Endogenous TGFβ3 produced by developing Th17 cells can drive them to a pathogenic IL-17 producing Th17 subset, and the presence of IL-23 was necessary for induction of autoimmunity by this subset [75]. Our increasing knowledge of what controls pathogenic versus protective Th17 responses will greatly aid vaccine design.

Concluding remarks

Generating antibody dependent immune responses by vaccination had saved and is saving millions of lives every year. However, this strategy might fail to provide serotype independent heterologous protection. Recent advances in Th17 biology, especially antibody

independent heterologous protection provided by Th17 cells, open up a new avenue for vaccine development. In experimental models, Th17 cells are effective in providing vaccination induced immunity against a range of pathogens. Identification of Th17 specific antigens for common prevalent pathogens will help to formulate a serotype independent effective vaccination strategy. Modulation of toxic properties and identification of new Th17 specific adjuvants will also have tremendous impact on designing safe and effective vaccines.

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Highlights

- Th17 responses show protection against both bacterial and fungal pathogens
- Mucosal and systemic vaccines can elicit protective Th17 responses
- Th17 responses can mediate serotype immunity against clades of organisms

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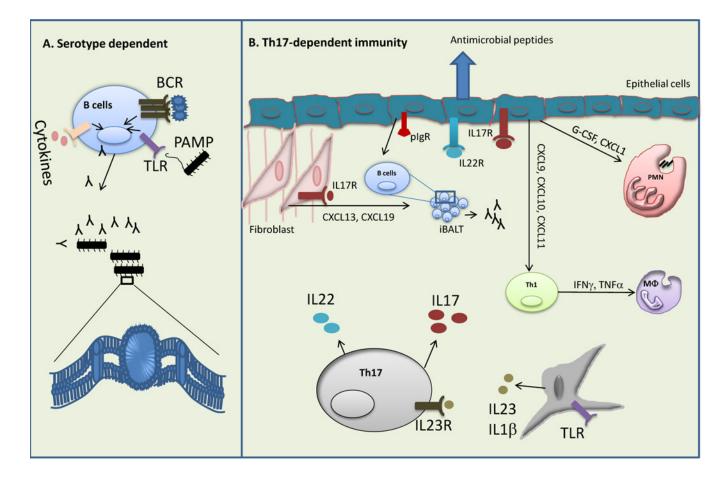


Figure 1. Proposed model of serotype dependent and independent immunity

Vaccination can induce both T cells dependent and independent immune responses. A) T cells independent antibody response generated by strong B cells receptor (BCR) signal by antigens and activation of both Toll-like receptor (TLR) and proinflammatory cytokine response. B) Dendritic cells derived IL-23 and IL-16 induce robust Th17 responses. IL-17 and IL-22 produced by Th17 cells activate epithelial cells mediated immune responses. Th17 targeted cytokines and chemokines (G-CSF and CXCL1) released from epithelial cells regulate neutrophils recruitment to the mucosal sites. Th17-mediated neutrophils response is reported to be essential for the protection against a range of pathogens. Epithelial cells also directly participate in limiting pathogens multiplication by the release of Th17 related cytokines regulated antimicrobial peptides. Apart from lymphotoxin signaling, Th17 cells involve in iBALT formation in the lung tissue by activating chemokines (CXCL13, CXCL19) released from the fibroblast. iBALT mediated local immune responses (both B and T cells response) are critical for clearance of influenza virus. Th17 cells also regulate late Th1 responses by modulating chemokines especially CXCL9, CXCL10 and CXCL11 response from epithelial cells. Th1-regulated cytokines (IFN γ or TNF α) and activation of macrophages are required for the immunity against *M. tuberculosis*. In addition, Th17 cells regulate polymeric immunoglobulin receptor (pIgR) expression on epithelial cells, which is essential for influx of B cells to the mucosal sites, however, the relevance of this pathway in Th17 mediated immunity is yet to determine.