

ARTICLE

# Symptomatic lipid storage in carriers for the *PNPLA2* gene

This article has been corrected since online publication and a corrigendum also appears in this issue.

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Neutral lipid storage disease comprises a heterogeneous group of inherited disorders characterized by severe accumulation of cytoplasmic triglyceride droplets in several tissues and neutrophils. A novel type of autosomal recessive lipid myopathy due to *PNPLA2* mutations was recently described with associated cardiac disease, myopathy and frequent infections, but without ichthyosis. Here we describe the clinical and biochemical characteristics of a long surviving patient and report on four carrier family members with diverse clinical involvement. Interestingly, heterozygous patients show neutral lipid storage in muscle and in the keratocytes of the skin, Jordans' bodies, mild myopathy and frequent infections. Biochemical analysis of fibroblasts obtained from patients revealed increased triglyceride storage and reduced lipid droplet-associated triglyceride hydrolase activity. Together, our data implicate that the wild-type allele cannot fully compensate for the mutated dysfunctional allele of *PNPLA2* leading to triglyceride accumulation in muscle and mild myopathy in *PNPLA2* mutation carriers. The presence of neutral lipid droplets in the skin in *PNPLA2* mutation carriers strengthens the link between NLS and other neutral lipid storage diseases with ichthyosis. *European Journal of Human Genetics* (2013) 21, 807–815; doi:10.1038/ejhg.2012.256; published online 12 December 2012

**Keywords:** symptomatic carriers; deafness; lipid myopathy; neutral lipid storage; insulin resistance; implantable cardioverter-defibrillator

## INTRODUCTION

Neutral lipid storage disease (NLS) comprises a heterogeneous group of autosomal recessive disorders characterized by severe accumulation of triglyceride (TG) cytoplasmic droplets in several tissues.<sup>1,2</sup> The most common finding in these patients is the presence of May–Grunwald–Giemsa-negative lipid droplets (LDs) (Jordans' bodies) in otherwise normal neutrophils and eosinophils from peripheral blood.<sup>3</sup> Most of the patients with NLS have lipid storage in skeletal muscle and liver, associated with severe ichthyosis. A distinctive type of autosomal recessive NLS was defined recently, with prominent myopathy but without ichthyosis. Fischer *et al*<sup>4</sup> described three affected women, from three families of Dutch, French and Algerian origin, who showed marked TG storage in muscle and delays in walking, variable cardiac abnormalities and hepatomegaly. These patients had no mutations in the *CGI-58* gene, but they harbored four distinct mutations in the *PNPLA2* gene. This form of NLS, caused by *PNPLA2* mutations has been delineated as NLS with myopathy.<sup>4,5</sup> Several patients have been discovered with NLS with myopathy in the past few years, and in most cases the different mutations in the *PNPLA2* gene resulted in cardiomyopathy and early cardiac lethality in young adulthood (Table 1).<sup>4,5</sup>

*PNPLA2* catalyzes the initial step in the breakdown of intracellular TGs and, as expected, *PNPLA2*-defective patients show impaired degradation of cytoplasmic TGs.<sup>4</sup> Recent studies have provided new insights into the genetic basis of a group of disorders of lipid metabolism characterized pathologically by TG accumulation in muscle.<sup>6,16–22</sup> The identification of causative genes offers new tools for diagnosis and genetic counseling. Although novel drugs have shown promising results *in vitro*, therapy for these disorders remains inadequate. Additional cases are needed to further define the clinical spectrum and genetic heterogeneity of this rare condition. In the current paper, we describe the phenotype and disease course of the Dutch index patient and her family and study the clinical consequences of *PNPLA2* heterozygosity.

## SUBJECTS AND METHODS

Patient II-3 (described as a child with a compound heterozygous *PNPLA2* mutation)<sup>4</sup> was born to non-consanguineous, apparently healthy parents (for pedigree, see Figure 1a). She was investigated as a child for myopathy, fatigue and recurrent infections. She had normal intelligence. At the age of 4 years she was diagnosed with a lipid myopathy. Peripheral blood leukocytes showed typical cytoplasmic LDs (Jordans' bodies; Figures 1b and 2a). Biochemical, histological and electron microscopy investigations of skin and muscle revealed

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Received 3 May 2012; revised 12 October 2012; accepted 26 October 2012; published online 12 December 2012

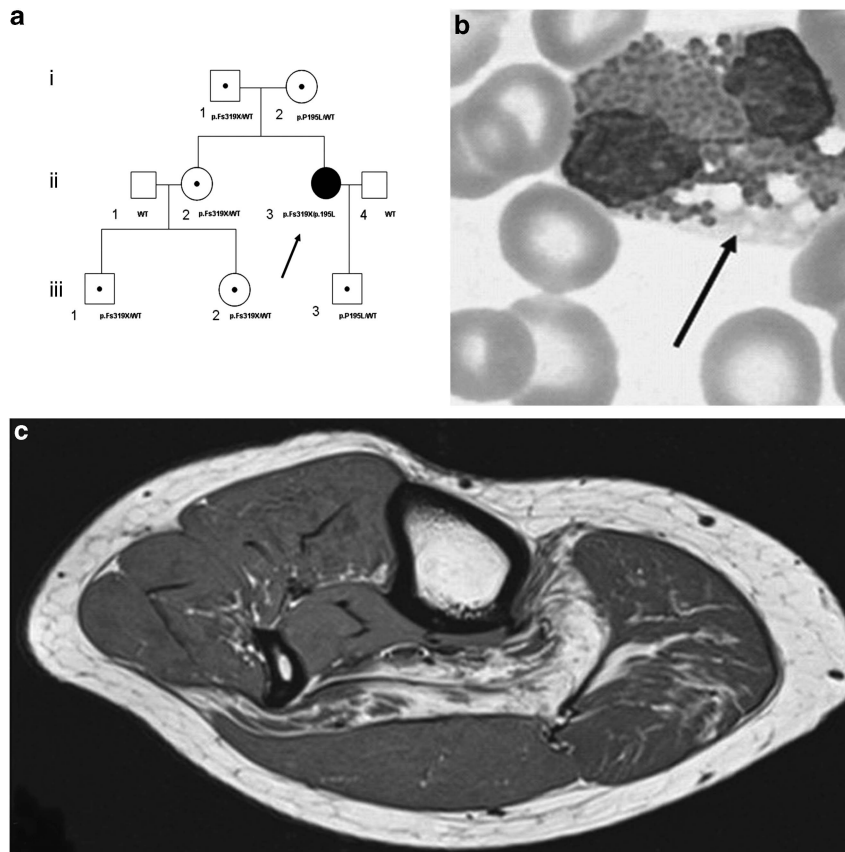
**Table 1 Clinical features in recessive NLSDM and mutation carriers**

	Genetic alteration	Myopathy muscle weakness	Cardiac finding	Liver involvement	Recurrent infections	Hearing impairment	Skin abnormalities	Insulin resistance	Ethnicity
Patient II-3 (Fischer <i>et al</i> <sup>4</sup> , Lefevre <i>et al</i> <sup>6</sup> )	Exon 5 P195L (584C-T)/ Exon 7 319X (808delC)	Progressive	Arrhythmia, cardiac dyskinesia, decreased ejection fraction	Hepatomegaly	Mostly as child	Conductive	Psoriasis	Yes, no diabetes	Dutch
Patient III-3 heterozygous	Exon 5 P195L (584C-T)	No Decreased muscle tone	No	No	Yes (ear infections)	No	No	NA	Dutch
Patient I-2 heterozygous	Exon 5 P195L (584C-T)	Scapula alatae Exercise intolerance ance Episodes of muscle pain	No	No	Yes	Mixed:conductive and sensineural	No	No	Dutch
Patient I-1 heterozygous	Exon 7 319X (808delC)	No	No	No	Mostly as child (ear infections)	Conductive	Psoriasis	NA	Dutch
Patient II-2 heterozygous	Exon 7 319X (808delC)	Progressive	No	No	Mostly as child	No	No	No	Dutch
Patient III-1 heterozygous	Exon 7 319X (808delC)	No Exercise intolerance ance Scapula alatae	No	No	Yes	No	No	No	Dutch
Patient III-2 heterozygous	Exon 7 319X (808delC)	No	No	No	Yes	No	No	No	Dutch
Patient A (Fischer <i>et al</i> ) <sup>4</sup>	Exon 7 319X (808delC)/ Exon 7 319X (808delC)	Progressive	Cardiomyopathy	Hepatomegaly Steatosis	NA	NA	No	Yes, diabetes	French
Patient B1 (Hirano <i>et al</i> ) <sup>7</sup>	Exon 7 Q289X (865C-T)/ Exon 7 Q289X (865C-T)	Yes	NA	No	NA	NA	No	NA	Algerian
Patient B2 (Hirano <i>et al</i> ) <sup>7</sup>	Exon 7 Q289X (865C-T)/ Exon 7 Q289X (865C-T)	Yes	Cardiomyopathy	NA	NA	NA	No	NA	Algerian
Patient C (Hirano <i>et al</i> ) <sup>7</sup>	Exon 4 178X (475dupCTCC)/ Exon 4 178X (475dupCTCC)	Yes	NA	NA	NA	NA	No	NA	Japanese

Table 1 (Continued)

	Genetic alteration	Myopathy muscle weakness	Cardiac finding	Liver involvement	Recurrent infections	Hearing impairment	Skin abnormalities	Insulin resistance	Ethnicity
Patient D (Akiyama <i>et al</i> ) <sup>8</sup>	Exon 5 L255X (695delC) Exon 5 L255X (695delC)	Yes	NA	Hepatomegaly	NA	NA	No	NA	Iranian
Patient E (Campagna) <sup>9</sup>	Exon 5 I212X (542delAC)/ Exon 5 I212X (542delAC)	Yes	Cardiomyopathy	No	NA	NA	No	NA	Italian
Patient F (Campagna) <sup>9</sup>	Exon 5 I212X (542delAC) Exon 5 I212X (542delAC)/ Exon 5 I212X (542delAC)	Yes	Cardiomyopathy	No	NA	NA	No	NA	Italian
Patient G (Kobayashi <i>et al</i> ) <sup>10</sup>	Exon 7 318X (799-802delGCC)/ Exon 7 318X (799-802delGCC)	Yes	Cardiomyopathy	Hepatomegaly	NA	No	No	NA	NA
Patient H (Akman <i>et al</i> ) <sup>11</sup>	Exon 3 1.8 kb ins/ Exon 3 1.8 kb ins	No	No	No	No	No	Dermatomyositis	NA	American
Patient I (Chen <i>et al</i> ) <sup>5</sup>	IVS2 + 1G>A/ IVS2 + 1G>A	Yes	No	NA	NA	Conductive	No	NA	Japanese
Patient J (Chen <i>et al</i> ) <sup>5</sup>	IVS2 + 1G>A/ IVS2 + 1G>A	Yes	Cardiomyopathy	NA	NA	Conductive	No	NA	Japanese
Patient K (Reilich <i>et al</i> ) <sup>12</sup>	Exon 5 P195L(584C-T) Exon 10 G483R(1447G-C)	Yes	Cardiomyopathy	No	No	No	No	Yes, diabetes	English
Patient L (Schweiger <i>et al</i> ) <sup>13</sup> and Akiyama <i>et al</i> ) <sup>14</sup>	Exon 4 178X(475dupCTCC)/ Exon 4 178X(475dupCTCC)	Yes	NA	No	No	NA	No	NA	Japanese
Patient M (Ohkuma <i>et al</i> ) <sup>15</sup>	Exon 4 178X(475dupCTCC)/ Exon 4 178X(475dupCTCC)	Yes	Cardiomyopathy	No	No	NA	No	NA	Japanese

NA, not assessed.



**Figure 1** (a) Pedigree of the family with segregation of the genetic findings. (b) Jordans' bodies in an eosinophil cell in peripheral blood smear (arrow: LDs). (c) MRS of the legs in the compound heterozygous index patient (II-3) at adult age shows extreme fat deposition in the Soleus muscle.

a generalized triglyceridosis with intracellular lipid accumulation. Muscle biopsy demonstrated lipid storage (Figures 2b–d). She had no clinical signs of lipid accumulation in the brain, lungs or heart. She had no ichthyosis, but she did have neutral lipid storage in a skin biopsy. Hormonal investigations were also normal. In young adulthood, she was found to be a compound heterozygote for mutations in the *PNPLA2* gene (1-bp deletion, c.808delC, leading to a premature stop codon at amino-acid position 319 (p.Frame-shift319\*) and a missense mutation c.584C>T, changing proline to leucine (p.P195L). In her 30s, the patient developed hearing loss and progressive proximal muscle weakness. She developed psoriasis and suffered from unexplained, recurrent episodes of short collapses.

Patient I-2 (heterozygous carrier) is the mother of the patient II-3, and she is 61 years old. As a child, she was reported to have frequent infections. She has been diagnosed with a mixed hearing loss and uses a hearing aid since the age of 5 years. She has no symptoms of myopathy or cardiac disease, liver abnormalities or diabetes. She has recurrent episodes of migraine and occasional muscle pain episodes. At neurological investigation, she has no detectable abnormalities.

Patient I-1, (heterozygous carrier) the father of the patient has hearing problems because of recurrent otitis as a child. At this moment, he uses a hearing aid owing to conductive hearing loss. He has no myopathy or fatigue, cardiac disease or diabetes. Since 2008, he has had psoriasis.

Patient III-3 (heterozygous carrier with symptoms) is the son of patient II-3, and he is 7 years old. He has recurrent infections and he is known to have a history of exercise intolerance and concentration problems at school. He cannot fully participate in regular school activities and gymnastics owing to easy fatigue. Neurological examination reveals a decreased muscle tone and winged scapula, no Gowers' sign of decreased strength and normal reflexes. On regular exercise, his muscle strength improved significantly, but his muscle tone remains somewhat decreased.

Patient II-2, (heterozygous carrier with symptoms) the 36-year-old sister of patient 1 has a mild myopathy. She also has a history of recurrent infections, but no cardiac disease or diabetes. She is reported to have exercise intolerance. Neurological examination reveals a positive Gowers sign and decreased strength and normal reflexes. The patient is married to a third-degree relative on the father's side, who is not a *PNPLA2* mutation carrier. They have two children.

Patient III-1 (heterozygous carrier with symptoms) is the son of patient II-2, and he is 7 years old. He has recurrent infections and he is known with a history of exercise intolerance, confirmed by the 'six minutes' walk test. Neurological examination reveals a decreased muscle tone and scapula alatae, but normal reflexes.

Patient III-2 (heterozygous carrier) is the daughter of patient II-2, and she is 4 years old. She has recurrent infections but no obvious exercise intolerance. Neurological examination is normal.

#### Genetic studies

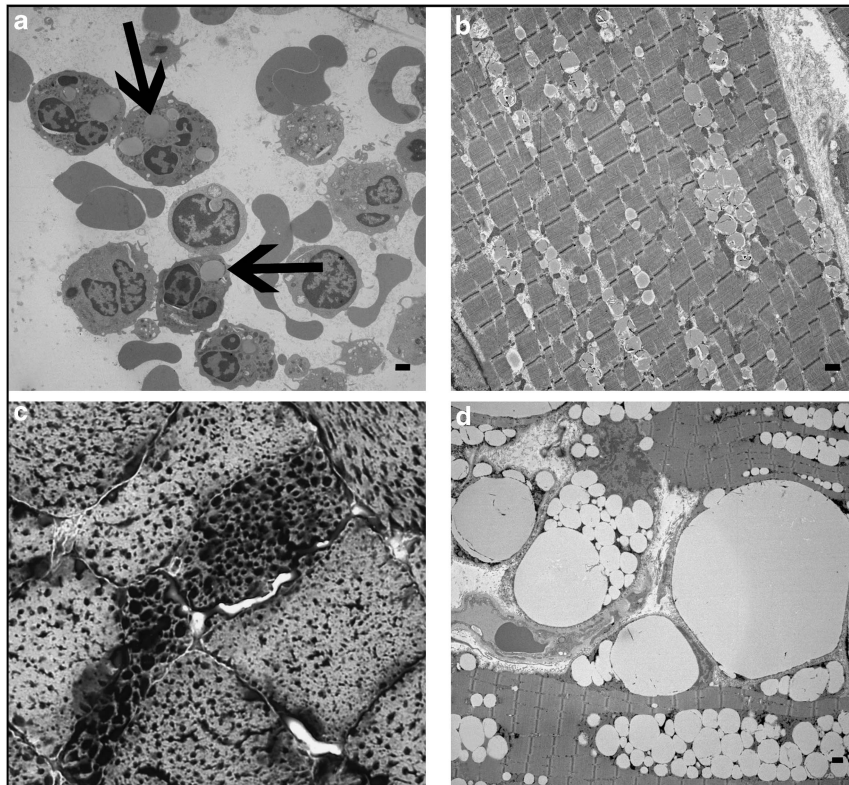
Sequence analysis of the *PNPLA2* gene was performed as described by Fischer *et al.*<sup>4</sup>

#### Laboratory investigations and histology

Histological evaluation and electron microscopy on skin biopsy and muscle were performed according to standard clinical protocols. Light microscopy of frozen transverse sections of skeletal muscle biopsy was stained by Sudan Black B.

Jordans' bodies were assessed by light microscopy of peripheral blood smear and by electron microscopy of leukocytes after centrifuging citrate blood.

The insulin resistance test was done with a glucose infusion rate 3.3 mg/kg/min, normal range 4–8 mg/kg/min.<sup>23</sup>



**Figure 2** (a) Neutral lipid storage in peripheral blood, and (b) in muscle of the compound heterozygous index patient (II-3) at childhood and (c and d) in muscle of patient II-3 at age 21. (a,b,d electron microscopy bar = 1  $\mu$ m, arrows: LDs. (c) Sudan Black B staining).

#### Biochemical studies on TG content of fibroblasts

Fibroblasts were cultivated in Dulbecco's modified Eagle's medium containing 10% FCS, 100 IU/ml penicillin and 100  $\mu$ g/ml streptomycin at 37 °C, 7.5% CO<sub>2</sub>, 95% humidified atmosphere until confluence and incubated for 24 h in the presence or in the absence of 500  $\mu$ M oleic acid complexed to BSA at a FFA/BSA ratio of 3:1.<sup>16</sup> Thereafter, cells were washed with phosphate-buffered saline (PBS) and total cell lipid was extracted using hexane/isopropanol (3:2). The extracts were dried and resuspended in 1% Triton X-100. TG were determined using the Infinity TG kit (Thermo Electron Corporation, Noble Park, VIC, Australia). Cells were lysed using 0.1% SDS/0.3 N NaOH and protein content was measured using the BCA reagent (Pierce, Rockford, IL, USA).

#### Labeling and isolation of LDs

For radioactive labeling of TG stores, confluent cell layer was incubated for 24 h in the presence of 0.5 mM oleic acid (4 mCi 3H-9,10-oleate/mmol) (GE Healthcare, Frederick, MD, USA) complexed to BSA at a FFA/BSA ratio of 3:1.<sup>16</sup> For the isolation of LD, cells were washed extensively with PBS and collected using a cell scraper. After centrifugation (200 g, 5 min) the cells were resuspended in buffer A (0.25 M sucrose, 1 mM EDTA, 1 mM dithiothreitol, 20  $\mu$ g/ml leupeptine, 2  $\mu$ g/ml antipain, 1  $\mu$ g/ml pepstatin, pH 7.0) and disrupted by sonication (Virsonic 475, Virtis). The lysate was centrifuged to remove cell debris and nuclei at 1000 g and 4 °C for 15 min. The supernatant was transferred to SW41 tubes, overlaid with buffer B (50 mM potassium phosphate (pH 7.4), 100 mM KCl, 1 mM EDTA, 20  $\mu$ g/ml leupeptine, 2  $\mu$ g/ml antipain, 1  $\mu$ g/ml pepstatin) and centrifuged in a SW41 rotor (Beckman, Fullerton, CA, USA) for 2 h at 100 000 g and 4 °C. LD were collected as a white band from the top of the tubes and concentrated by centrifugation for 15 min at 20 000 g and 4 °C. The underlying solution was removed and the LD were resuspended by brief sonication in buffer B. TG and protein content of the isolated LD was determined using the Infinity TG kit of the Bradford reagent (Biorad Laboratories GmbH, Munchen, Germany).

#### Assay for TG hydrolase activity in purified LDs

To determine LD-associated TG hydrolase activity of fibroblasts 150  $\mu$ l <sup>3</sup>H-9,10-oleate radiolabeled LDs in buffer B were mixed with 50  $\mu$ l fatty acid-free BSA (20%, w/v).<sup>16</sup> After incubating the mixture at 37 °C for 1 h, the reaction was terminated by adding 3.25 ml of methanol/chloroform/heptane (10:9:7) and 1 ml of 0.1 M potassium carbonate, 0.1 M boric acid (pH 10.5). After centrifugation (800 g, 15 min), the radioactivity in 1 ml of the upper phase was determined by liquid scintillation counting.

## RESULTS

### Clinical studies

Echocardiography in patient II-3 (compound heterozygous index patient) demonstrated hypokinesia of the mid- and basal inferior wall of the heart. Magnetic resonance imaging (MRI) of the heart showed diffuse hypokinesia of the left and right ventricles. Lipid accumulation could not be demonstrated using the present software. The ejection fraction was decreased to 43%. Because of suspected cardiac rhythm abnormalities as a cause of the recurrent collapses, a cardiac reveal was implanted for a period of 2 years to register her heart rhythm. Once an episode of ventricular tachycardia was documented, the patient received an implantable cardioverter-defibrillator.

MRI of the legs showed extreme fat deposition in the Soleus muscle (Figure 1c).

MRI and magnetic resonance spectroscopy (MRS) of the liver showed no hepatomegaly nor lipid accumulation, and an asymptomatic liver adenoma was detected.

Insulin sensitivity was measured by a hyperinsulinemic euglycemic clamp (insulin infusion rate 60 mE/m<sup>2</sup>). Glucose infusion rate at steady state conditions (plasma insulin concentration 56 mU/l) was



18.1  $\mu\text{mol/kg/min}$ , which is substantially decreased compared with an age- and body mass-matched control group ( $53.9 \pm 4.3 \mu\text{mol/kg/min}$ , see Tack *et al*<sup>23</sup>). The findings are consistent with frank insulin resistance.

Patient III-3 (heterozygous carrier with symptoms): MRI and MRS of the brain showed no lipid accumulation.

#### Audiometric investigations

In patient II-3, audiometry showed a conductive hearing loss with a Fletcher index of 57 dB.

In patient I-2, audiometry showed a mixed hearing loss at 55 dB. In patient I-1, audiometry showed a bilateral conductive hearing loss at 60 dB.

#### Genetic studies

Patient I-2, the mother of the index patient, was heterozygous for the c.584C>T/p.P195L mutation, whereas the father, patient I-1 and the sister, patient II-2, were heterozygous for c.808delC/p.Frameshift319\* mutation.

Heterozygosity for c.584C>T/p.P195L was detected in patient III-3 and the c.808delC/p.Frameshift319\* was confirmed in patient III-1 and III-2.

#### Laboratory investigations

Patient II-3 had an increased creatine kinase (CK 482 U/l normal < 170 U/l). Serum lactic acid, pyruvic acid and acyl-carnitine profile levels are within the normal range in all patients. Patient II-2, III-1, III-2 and III-3 had a CK level of 138, 57, 62 and 158 U/l, respectively (normal). None of the family members has hyperglycemia or hyperlipidemia (TGs, total cholesterol levels and subfractions were normal).

Peripheral blood leukocytes show Jordans' bodies in all investigated family members in the range of 6–12%.

#### Pathological examinations

Patient II-3 (compound heterozygous index patient): electron microscopy examination and routine blood smear of blood revealed the presence of large neutral LDs in the granulocytes (Figure 2a). Light microscopy of muscle biopsy at age 6 years showed numerous accumulations of cytoplasmic droplets containing neutral lipids (TGs). Transmission electron microscopy of longitudinal sections of skeletal muscle biopsy confirmed the presence of these LDs of up to more than 200  $\mu\text{m}$  width between the sarcomeric filament. In a muscle biopsy taken at the age of 21 years, the LDs were even greater, and were found between the sarcomeric filaments and under the

muscle membrane, where they reached a width of more than 1 mm. These LDs were present in all muscle fiber types, but the larger ones were found in the type 1 fibers (Figures 2b–d). The LDs could also be found in a Schwann cell of an intramuscular nerve fiber (not shown). Examination of a skin biopsy at age 3 years demonstrated a small layer of empty vacuoles just under the epithelial basal membrane (Figure 3a), that could already be seen on light microscopy level. On electron microscopy the keratocytes just above the basal membrane contained small neutral LDs (Figure 3b). Some dermal fibroblasts also contained small neutral LDs (not shown).

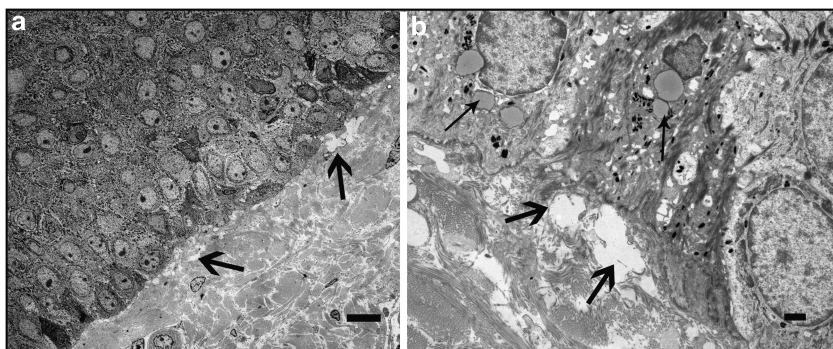
Patient II-2 (heterozygous carrier sister with symptoms): light microscopy showed mild abnormalities of neutral lipid storage in muscle at age 35 years, especially in type 1 fibers. A few type 1 fibers were somewhat atrophic (Figure 4a). Electron microscopy, however, confirmed an increased subsarcolemmal and cytoplasmic storage of LDs that were smaller than in the index patient (Figure 4b). Light and electron microscopy of skin at age 5 years revealed no vacuoles or LDs (Figures 5a and b). Patient III-3 (heterozygous carrier): electron microscopy of peripheral blood at age 4 years revealed a few small LDs in the granulocytes (Figure 4c)

#### Biochemical studies

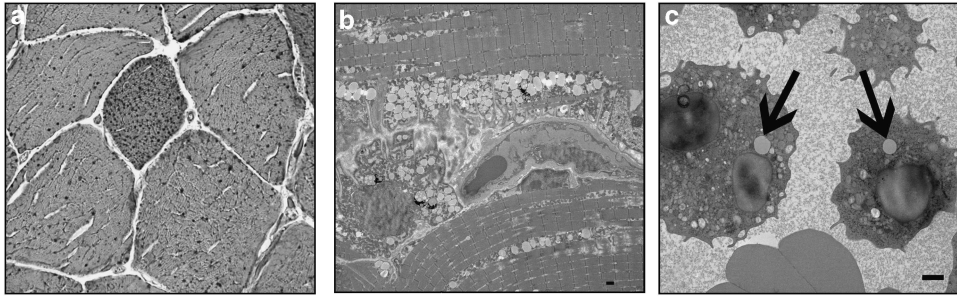
To investigate how heterozygous mutations of *PNPLA2* affect TG metabolism *in vitro*, we determined TG content and TG hydrolase activity in cultured fibroblasts obtained from patients heterozygous for the p.P195L (patient III-3) and the p.Frameshift319\* (patient I-2, patient II-2) mutation. In comparison to wild-type fibroblasts, TG content in mutant fibroblasts was significantly increased under standard culture conditions (2.2-, 1.9- and 2-fold for patient III-3, I-2 and II-2, respectively, (Figure 6a) and in the presence of exogenously added fatty acids (2-, 1.6- and 1.7-fold, for patients III-3, I-2 and II-2, respectively, (Figure 6b). Activity assays revealed that the LD-associated TG hydrolase activity of fibroblasts from patients III-3, I-2 and II-2 amounted to 37%, 41% and 56% of normal, respectively (Figure 6c).

#### DISCUSSION

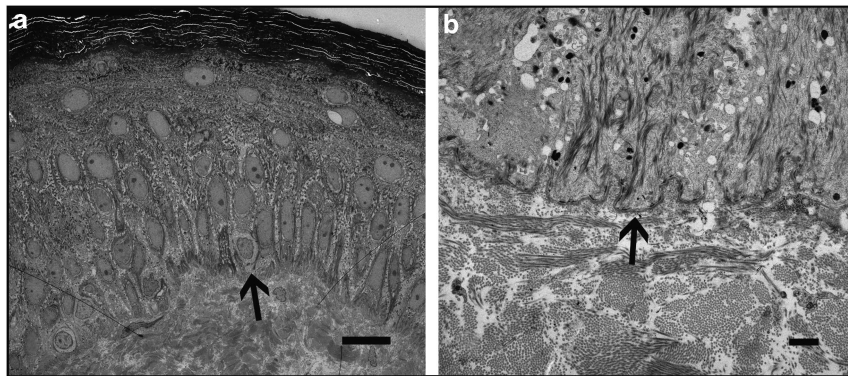
Several patients have been recently reported with recessive *PNPLA2* mutations and myopathy, but no other organ involvement.<sup>5,11,12,14</sup> Here we report on additional unique findings in four carrier family members with diverse clinical involvement, including significant neutral lipid storage in muscle. The clinical features in these individuals included muscle weakness, episodes of muscle pain, sensorineural or conductive deafness and frequent infections in heterozygous individuals, carrying either the p.P195L or the



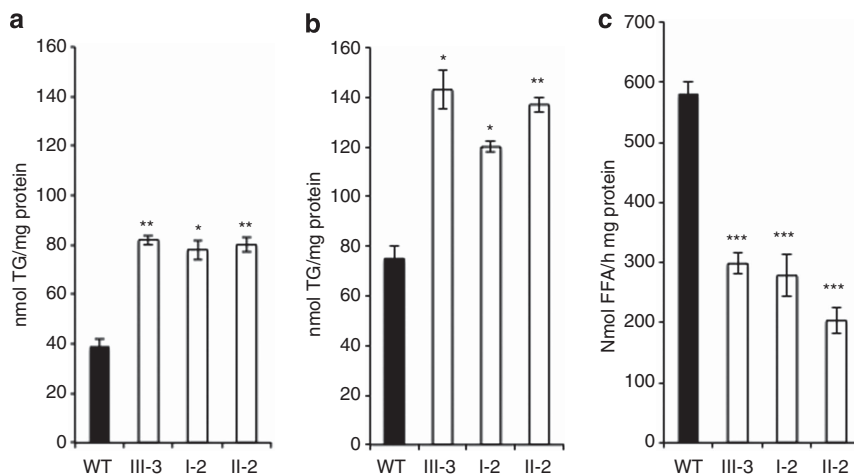
**Figure 3** (a) Vacuolisation (thick arrows) immediately under the basal membrane. (b) Neutral LDs (thin arrows) in the basal keratocytes in a skin biopsy at age 3 years in the compound heterozygous index patient (II-3) ((a) low-power EM, bar = 10  $\mu\text{m}$ ; (b) EM, bar = 1  $\mu\text{m}$ ).



**Figure 4** (a) Muscle biopsy of the heterozygous carrier sister, (II-2) at age 35 years with centrally located small type 1 fiber filled with black-stained LDs. (b) Interfilamentous and subsarcolemmal lipid deposition on EM. (c) Neutral LDs (arrows) in granulocytes of the heterozygous sister (III-3) at age 4 years. ((a) Sudan Black B, (b and c) EM, bar = 1  $\mu$ m).



**Figure 5** Skin biopsy at age 5 years in the heterozygous sister (II-2) does not show any vacuolisation under the basal membrane (thick arrows) or LDs in the keratocytes ((a) low-power EM, bar = 10  $\mu$ m; (b) EM, bar = 1  $\mu$ m).



**Figure 6** TG content and LD-associated TG hydrolase activity of wild-type and mutant fibroblasts heterozygous for the p.P195L (III-3) and the p.Frameshift319\* (I-2), (II-2) mutation. TG content in fibroblasts was determined under standard culture conditions (a) and in the presence of exogenously added oleic acid (500  $\mu$ M, b). LD-associated TG hydrolase activity was determined by self-digestion of isolated LDs (c). Data are shown as mean  $\pm$  SD ( $*P < 0.05$ ,  $**P < 0.01$ ,  $***P < 0.001$ ). In comparison to wild-type fibroblasts, TG content in mutant fibroblasts was significantly increased under standard culture conditions (2.2-, 1.9- and 2-fold, for patient 2, 3 and 5, respectively, a) and in the presence of exogenously added fatty acids (2-, 1.6- and 1.7-fold for patients 2, 3 and 5, respectively, b). Activity assays revealed that the LD-associated TG hydrolase activity of fibroblasts from patients III-3, I-2 and II-2, amounted to 37%, 41% and 56% of normal, respectively (c).

mutation p.Frameshift319\* in *PNPLA2* (Table 1). The observation of recurrent cardiovascular collapse and other cardiac and endocrine findings in our patient emphasize the need for targeted screening, regular follow-up and preventive treatment in patients with recessive

*PNPLA2* mutations. It is interesting to note that the compound heterozygous index patient (patient II-3) and also her father (patient I-1) developed psoriasis, but no ichthyosis, based on clinical and histological evaluation of the lesions. This is the first observation of

psoriasis in NLS. In the compound heterozygous patient analogous, but less-pronounced lipid deposits, could be found in the basal epidermal layers as in patients with Chanarin–Dorfman disease.<sup>24</sup> In the presence of obvious lipid storage in the different layers of skin in patient II-3, and increased lipid storage in patient I-1, we cannot rule out that psoriasis is related to defective lipolysis in these patients.

Our index patient with compound heterozygosity for mutations in the *PNPLA2* gene is one of the few individuals surviving to adulthood with cardiac intervention with this novel type of NLS showing symptomatic multisystem involvement and insulin resistance. Besides the classic presentation of progressive myopathy, cardiac involvement and insulin resistance, she demonstrates hearing loss and recurrent infections. There seems to be no liver involvement including fat storage, however, the significance of the liver adenoma is not yet clear. An interesting observation is the high frequency of ear infections (in the age range of 2 to 7 years/ 3–5 monthly) in the index patient with compound heterozygous mutation. This might be due to lipid storage in granulocytes and lymphocytes, however, no cellular or humoral immune deficiency could be demonstrated in our patients. Even more intriguing is the decrease in frequency of infections as she got older, which needs further explanation.

In our carrier family members, we could detect a significant TG storage not only in white blood cells but also in fibroblasts of the p.P195L and the p.Frameshift319\* mutation carriers compared with healthy controls. These mutations have been previously shown to be inactive (p.P195L) or to exhibit strongly reduced LD association (p.Frameshift319\*<sup>13</sup>). Mutant variants lacking the C-terminal domain retain *in vitro* lipase activity, which is located in the intact N-terminal region containing a patatin domain and the catalytic site. The C-terminal domain contains a hydrophobic region, which is essential for LD binding.<sup>13,25</sup>

*PNPLA2* catalyzes the initial step in the breakdown of intracellular TGs<sup>24</sup> and, as expected; *PNPLA2*-defective patients showed impaired degradation of cytoplasmic TGs. Experimental knockout of *PNPLA2* in mice leads to cytoplasmic accumulation of TGs in multiple tissues.<sup>26</sup> Interestingly, ATGL-deficient mice exhibit a strong decrease in PPAR $\alpha$  target gene expression tissues with high-energy consumption, such as cardiac muscle and liver. As PPARs have a prominent role in the transcriptional regulation of genes involved in FA uptake and oxidation, ATGL deficiency is associated with impaired mitochondrial respiration. Reconstituting normal PPAR target gene expression by treatment with PPAR $\alpha$  agonist reverses the mitochondrial defect, reduced fat accumulation in cardiac muscle and liver, and prologs life span of mice.<sup>27</sup> It remains to be investigated whether PPAR agonists, such as fibrates, can improve myopathy and counteract TG accumulation in NLS patients.

As the initial reports on *PNPLA2*-related NLS, novel patients have been reported, with milder forms of the disease.<sup>5,11,12,14,15</sup> One might question whether the disorder is not more common than previously thought, and the diagnosis is missed in some of these milder patients owing to missing the diagnostic sign of Jordans' bodies (due to variable percentage of droplets in blood smear or the use of automated blood cell analysis) without specifically looking for the vacuoles by electron microscopy.

In summary, our data on increased TG content in muscle and in cultured fibroblasts of carriers implicate that the wild-type allele cannot fully compensate the mutated dysfunctional allele of *PNPLA2* leading to defective TG catabolism and mild myopathy. Based on clinical and metabolic features of comparable severity expressed in both heterozygous p.P195L and p.Frameshift319\* mutation carriers, we suggest screening patients with a mild form of lipid myopathy and

also patients with unexplained hearing loss for the presence of Jordans' bodies and *PNPLA2*-related NLS.

## CONFLICT OF INTEREST

The authors declare no conflict of interest

## ACKNOWLEDGEMENTS

We thank the family. We acknowledge Professor Dr C Tack for performing the insulin clamp test and Mrs L Eshuis for preparing the electron microscopy figures.

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