Published in final edited form as:
FEBS Lett. 2013 April 17; 587(8): 1053–1061. doi:10.1016/j.febslet.2013.01.064.

Influences of Crowded Cellular Environments on Protein Folding, Binding, and Oligomerization: Biological Consequences and Potentials of Atomistic Modeling

Huan-Xiang Zhou*
Department of Physics and Institute of Molecular Biophysics, Florida State University, Tallahassee, Florida 32306, USA

Abstract

Recent experiments inside cells and in cytomimetic conditions have demonstrated that the crowded environments found therein can significantly reshape the energy landscapes of individual protein molecules and their oligomers. The resulting shifts in populations of conformational and oligomeric states have numerous biological consequences, including on the efficiency of replication and transcription, the development of aggregation-related diseases, and the efficacy of small-molecule drugs. Some of the effects of crowding can be anticipated from hard-particle theoretical models, but the *in vitro* and *in vivo* measurements indicate that these effects are often subtle and complex. These observations, coupled with recent computational studies at the atomistic level, suggest that the latter detailed modeling may be required to yield a quantitative understanding on the influences of the crowded cellular environments.

Keywords

macromolecular crowding; protein folding; protein binding; protein aggregation; postprocessing

Motivation and scope

It is now widely recognized that the crowded conditions found in cellular environments can significantly impact the equilibria and kinetics of biochemical processes such as protein folding, binding, and oligomerization. Pioneering work by Minton [1], Kornberg [2], Ellis [3], and others as well as many recent studies have contributed to this recognition. As more and more researchers pay attention to and start to study the effects of crowded conditions, questions of general interest include: What has been established by studies of crowding? How best can we advance the quantitative understanding on the influences of the crowded cellular environments? This review presents a personal view on these and related questions.

A few studies have presented dramatic effects of macromolecular crowding. In Arthur Kornberg's lab, many failed attempts finally led to success in replicating the *oriC* plasmid in a cell-free condition, upon including a high concentration of polyethylene glycol (PEG) in the incubation mixture [4]. As Kornberg [2] lucidly explained later, "the PEG gel occupies most of the aqueous volume and excludes a small volume into which large molecules are crowded. This concentration is essential when several proteins are needed in the consecutive steps of a pathway." Hence Kornberg made "Thou Shalt Correct for Extract Dilution with Molecular Crowding" one of his ten commandments as he reflected on his studies of DNA replication.

*Address correspondence to hzhou4@fsu.edu.

More recently several labs observed that crowding agents such as PEG, Ficoll, and dextran drastically increased the aggregation rates of α-synuclein [5, 6] and apolipoprotein C-II [7], and enabled the efficient assembly of the HIV-1 capsid protein [8] and the alignment of FtzZ filaments into ribbons [9]. Red blood cells contain high concentrations (~320 g/l) of hemoglobin, and such a level of crowding is an essential condition for sickle hemoglobin (HbS) polymerization and sickle cell anemia [10]. A 20% reduction in the HbS concentration could reduce the rate of homogeneous nucleation of HbS into polymers by as much as 10^{10}fold. Fetal hemoglobin (HbF) is unable to polymerize and replacement of HbS by HbF presents a potential therapy for sickle cell anemia. Ironically, as the HbF replacement essentially preserves the crowding effects, a 20% HbS-to-HbF replacement results in only a 10^3-fold reduction in the rate of homogeneous nucleation, which would render HbF replacement an ineffective therapy.

In contrast to the foregoing studies, the effects of crowding found in many other studies are modest. For example, a number of papers [11–16] reported small increases, of the order of 1–2 $k_B T$ (k_B: Boltzmann constant; T: absolute temperature), in protein folding stability by crowding agents. Similar results were found for the effects of crowding agents on the binding stability of protein-protein heterodimers [17, 18]. Two issues then arise: Can the dramatic effects of crowding observed in some studies be reconciled with the subtle effects found in other studies? Do these subtle effects of crowding nevertheless have biological significance?

Statistical thermodynamics provides a conceptual framework for addressing these issues and for a fundamental understanding of how crowding affects various biochemical processes. In principle, the equilibrium properties of these processes are determined by the energies of interactions within and between the “reactant” molecules and between the reactant molecules and bystander molecules such as crowders; kinetic properties requires in addition information on the dynamics of the reactant molecules in the solvent environment [19]. In practice, until recently, calculations of the effects of crowding on thermodynamic and kinetic properties were only possibly using simple models based on a hard-particle representation of reactant and crowder molecules, with excluded-volume interactions playing a dominant role [20, 21]. Molecular simulations have now opened a powerful way to model the effects of crowding, and promise to enable quantitative predictions on the influences of the crowded cellular environments.

The following sections present a critical assessment of some recent studies of biochemical processes in cells and in crowded solutions, with an eye toward answering the two questions posed in the opening paragraph. I mostly limit to studies that appeared in the last four years. Earlier studies are covered in several reviews [22–26], and some are already mentioned in the preceding paragraphs. In addition, the focus is on equilibrium properties; effects of crowding on kinetic properties are assessed elsewhere [19, 21, 23, 27–30]. Experimental studies, divided into three sections, are followed by computational studies in a single section.

Protein folding

Upon synthesis, most, not all, proteins fold into defined 3-dimensional structures before gaining biological functions (Figure 1). Some others exist as unstructured in one functional state and structured in another (e.g., when bound to a biological target). Extending other studies [11–14, 16], the Pielak lab has assessed how crowded cytomimetic conditions affect the folding stability of proteins [15, 31–34]. These recent studies are notable both for the experimental technique used and for their findings, presenting additional subtleties and complexities of the effects of crowding. The technique is NMR-detected amide proton

exchange, which probes residue-level local stability, i.e., the stability of individual residues to withstand opening up for hydrogen/deuterium exchange [35]. Opening of the residues with the highest local stability involves global unfolding; hence the highest local stability corresponds to the global folding stability.

Regarding the influence of macromolecular crowding on global folding stability, Pielak and co-workers concluded: (1) Two polymer crowders, poly(vinylpyrrolidone) (PVP) and Ficoll, resulted in modest stabilization of chymotrypsin inhibitor 2 (CI2), in line with other studies [11–14, 16], but two protein crowders, lysozyme and bovine serum albumin (BSA), resulted in mild destabilization; (2) The temperature dependence of the stabilizing effect of Ficoll indicated enthalpy, not entropy, as the dominant component, contrary to expectations based on consideration of excluded-volume interactions alone; (3) Temperaturedependent data for the effects of PVP, Ficoll, lysozyme, and BSA on the stability of ubiquitin again suggest a dominant enthalpic component.

These conclusions cast serious doubt on the conceptual framework for understanding the effects of macromolecular crowding on protein folding stability. Recently, the published data of the Pielak lab were re-analyzed, with an emphasis for a unified mechanism of action for all macromolecular crowders [36]. It was recognized that all intermolecular interactions consist of hard-core repulsion and longer-ranged (mostly) attraction (Figure 2a). The hardcore repulsion between a protein molecule and the crowder molecules leads to an entropic component that favors the folded state of the protein, whereas the longer-ranged attraction leads to an enthalpic component that favors the unfolded state (Figure 2b). Because in the folding free energy the entropic component is weighted by the temperature, the net effect of macromolecular crowding is destabilizing at lower temperatures but becomes stabilizing at higher temperatures (Figure 2c). This predicted general trend is exactly what was observed for both polymer crowders and protein crowders on the folding stability of ubiquitin [33].

According to the general trend just noted, for a given protein, every macromolecular crowder is predicted to have a crossover temperature at which the net effect of crowding changes from destabilizing to stabilizing. Hence the experimental temperature plays an important role in determining the sign (and magnitude) of the crowding effect. In line with this expectation, the stabilizing effects of PVP and Ficoll on CI2 were measured at a higher temperature, 37 °C, whereas the destabilizing effects of lysozyme and BSA were measured at a lower temperature, 20 °C. The prediction is that PVP and Ficoll will destabilize CI2 at lower temperatures whereas lysozyme and BSA will stabilize CI2 at higher temperatures. While this prediction on CI2 has yet to be tested, the Gruebele lab has now published results for the increased folding stability of λ_{6-85} by a protein crowder (subL) [37] and of phosphoglycerate kinase inside cells [38], both at high temperatures.

While the existence of a crossover temperature is predicted to be universal, its exact value of course depends on the size, shape, and chemical nature of the crowders. It is possible that common polymer crowding agents such as Ficoll and dextran may exert weaker soft attraction than protein crowders toward test proteins. Pielak and co-workers measured the effects of polymer and protein crowders on the translational and rotational diffusion and the NMR relaxation of CI2 [39]. The results are consistent with the protein crowders exerting stronger soft attraction toward CI2. Stronger soft attraction will produce a larger enthalpic component favoring the unfolded state of the test protein, leading to a higher crossover temperature (assuming the entropic component is fixed).

NMR-detected amide proton exchange yields information not only on the global folding stability but also on the local stability against opening up. The latter information can

potentially be a rich source for understanding how crowding affects local conformational sampling. Theories and computations have yet to make quantitative predictions that can be directly tested by experiments, so this source is still under-exploited.

The effects of crowding on protein folding stability revealed by all the *in vitro* and *in vivo* studies seem modest. Nevertheless these effects can have significant biological consequences. For example, a $2k_BT$ increase in folding stability will change a marginally stable protein from 75% unfolded into 75% folded. Searches for such cases have been carried out. A recent in-cell NMR study on a destabilized mutant of protein L did not find evidence of folding in *E. coli* [40]. This could mean that the effect of the cellular environment was too modest under the experimental conditions; it may also be possible that conformational exchange makes it difficult for a folded population to be detected by in-cell NMR spectroscopy. An earlier *in vitro* study [11] along the same line is instructive. With increasing concentrations of dextran, the unfolding free energy of a destabilized variant of ribonuclease T1 became less and less negative, extrapolating to a 22% folded population at 400 g/l dextran. At this crowder concentration, the fluorescence spectrum of the protein still resembled largely that of the unfolded state and little that of the folded state; nevertheless the protein gained 16% of the expected enzymatic activity of the folded state. Finally the prediction that temperature plays an important role in determining the net effect of crowding leads one to conclude that crowding effects make greater contributions to the *in vivo* folding stability of proteins in thermophilic organisms [36].

Conformational change of intrinsically disordered proteins

Some proteins, or fragments thereof, are thought to exist in the unfolded state and participate in cellular functions. A notable example is α-synuclein, a presynaptic protein linked to Parkinson's disease and whose normal functions putatively include regulation of neurotransmitter release, in particular acting as a chaperone for SNARE-complex assembly in presynaptic terminals [41]. The classification of α-synuclein and others as "intrinsically disordered proteins" (IDPs) is usually based on conformational probes in dilute solution [42–47]. The conformational flexibility and the propensity to oligomerize and aggregate and to bind with different partner proteins and biological membranes make IDPs particularly susceptible to the effects of crowding under cellular conditions. For α-synuclein, recent studies have uncovered evidence that inside cells it exists as a stable α-helical tetramer [48] or as a dynamic tetramer with high α-helix content [49]. Other studies [50, 51], however, continue to suggest that native α-synuclein is an unfolded monomer. N-terminal acetylation, a posttranslational modification in eukaryotic cells, could contribute to the difference in conformational and oligomeric states [52].

In dilute solution of α-synuclein, raising temperature induces helix formation [45] and expansion in size [53], as indicated by circular dichroism, ^{1}H-^{15}N heteronuclear single quantum coherence (HSQC) spectra, and pulse field gradient measurement of hydrodynamic radius. The expansion and helix formation are suppressed by 300 g/l of BSA and in the *Escherichia coli* periplasm [54], as shown by *in vitro* and in-cell NMR spectroscopy. Compaction under crowding has also been observed for other IDPs, including the N protein of bacteriophage λ (crowded by bovine pancreatic trypsin inhibitor (BPTI) and observed by small-angle neutron scattering) [55] and $p21^{Cip1}$ (crowded by dextran and observed by fluorescence correlation spectroscopy) [56]. Similarly, compression of the polymer PEG by Ficoll as a crowding agent has been observed [57], suggesting excluded-volume interactions as a common cause for the contraction in size. Reinforcing this contention, it was reported that Ficoll resulted in compaction of the unfolded state of a globular protein (as assessed by iodide quenching of tryptophan fluorescence and extent of cysteine modification with PEG-maleimide) [58] and an increase in the population of a more closed conformation in the

folded state of phosphoglycerate kinase (indicated by decreased donor-to-acceptor fluorescence ratio) [59]. The increase in closed population is even more pronounced inside cells [38, 60].

Some IDPs gain structures upon binding a biological target. An example is FlgM, a ~10 kD transcriptional regulator involved in the ordered synthesis of proteins for bacterial flagellar assembly [61]. It binds to σ^{28}, the flagellar σ subunit of the RNA polymerase holoenzyme, thus occluding the catalytic core. Once the transmembrane flagellar base is assembled, FlgM is released through its lumen, allowing σ^{28} to bind the RNA polymerase catalytic core and initiate late gene expression for completion of flagellar assembly. In dilute solution, FlgM is mostly unfolded, with transient helix formation in the C-terminal half [62, 63]. Upon binding the three-domain σ^{28}, FlgM adopts an extended configuration on the σ^{28} surface, with two C-terminal helices interacting with the third σ^{28} domain and a middle helix interacting with the first σ^{28} domain [64]. The Pielak lab [65] has reported HSQC spectra showing that, in the presence of 400 g/l BSA or when overexpressed in *E. coli*, the C-terminal half gains structure, reminiscent of the situation with binding σ^{28} [62]. Given that the helices of σ^{28}-bound FlgM are well separated and have few tertiary contacts, it is doubtful that the structure induced by crowding is the same as that formed on the σ^{28} surface. However, it is entirely possible that, relative to the conformational ensemble in dilute solution, the conformational ensemble under crowding is more similar to the bound structure and has higher affinity for σ^{28}. Retaining conformational flexibility inside the cell may actually be beneficial, since FlgM has to be secreted when its inhibitory activity is no longer needed.

More generally, the crowded conditions in cellular environments could alter the relative stability of IDPs among different conformational and homo- and hetero-oligomeric states (Figure 3). As noted above, crowding agents like Ficoll and dextran were found to drastically increase the aggregation rates of α-synuclein [5, 6], presumably by stabilizing oligomers that nucleate the aggregation process. The tendency of IDPs to form homo- and hetero-oligomers, especially under crowding, can potentially be harmful to cells [66]. Perhaps as a defense mechanism, the expression levels of IDPs are lower relative to those of structured proteins, through increased degradation and decreased translation rate [67]. Oligomerizationrelated processes are further discussed next.

Binding, oligomerization, and aggregation

The modest effects of macromolecular crowding on protein folding stability commented above is in line with similarly modest effects on the binding stability of heterodimers formed by single-domain proteins [17, 18]. A 1–2 $k_B T$ increase in binding stability corresponds to a ~5-fold increase in binding affinity. While it is difficult to directly ascertain whether such an increase has any biological significance, one can infer a positive answer based on biological consequences of mutations that cause similar changes in binding affinities. For example, *in vivo* assays have identified a number of FlgM mutants as defective in inhibiting σ^{28} [62]. These mutants were confirmed to be significantly less effective in suppressing *in vitro* transcription, and yet their binding affinities for σ^{28} were reduced by only 4–10 fold [68].

Theories [20, 69] predict that, under the same crowding condition, the effects of crowding on binding stability increase when the sizes of the two reactant species increase, as in the cases of multi-domain or multimeric proteins (Figure 3). In the binding of the large and small subunits to form an intact ribosome (Figure 1) and the subsequent formation of a ribosome dimer, even a low concentration, 60 g/l, of dextran and Ficoll, resulted in ~10-fold increases in the two binding constants [70]. Similarly, 75 g/l of PEG 12K resulted in a 55-fold increase in the binding affinity between the trimeric gene 45 protein and the gene 44/62

protein complex (the latter having a 4:1 stoichiometry) [71], which are part of the DNA replication complex of bacteriophage T4. Jarvis et al. [71] further observed that PEG also significantly enhanced the stability of the replication complex formed on single-stranded DNA, such that a primer is extended much longer than in the absence of PEG. Echoing the words of Kornberg [2], Jarvis et al. noted that, given the μM cellular concentrations of T4 replication proteins, the ~μM dissociation constant between gene 45 and gene 44/62 proteins measured in dilute solution would be much too weak for holding the replication complex together on a template DNA strand for extended replication.

Yi et al. [72] recently studied the effects of dextran, Ficoll, and PEG on the binding affinity of two multimeric proteins, superoxide dismutase and catalase, at a range of temperatures. They found that the stabilizing effects of the crowding agents diminished when the temperature was lowered. This temperature dependence is very similar to that for crowding effects on protein folding stability noted above (Figure 2c) [36], and was indeed accorded a similar explanation, i.e., by invoking the balance of entropic and enthalpic components that favor the bound and unbound states, respectively.

It has also been recognized that the stabilization due to crowding is nonspecific, in that it is applicable to every binding step, regardless of the identities of the reactants [18]. Therefore, the stabilizing effects are cumulative when forming a higher oligomer (Figure 3) [18, 20, 73]: each addition of a monomer is accompanied by an increase in stability due to crowding. This cumulative nature can lead to substantial stabilization of the oligomer. Such stabilization could explain the drastic increases in protein aggregation rates by crowding agents [5–7], if the aggregations are nucleated, i.e., rate-limited by the formation of appropriately-sized oligomers. Recently, dextran 70K at 200 g/l was found to induce the formation of prion protein oligomers with high β-sheet content [74], which putatively are the infectious agents for prion diseases [75]. These results suggest that the crowded environments inside cells play a critical role in the development of protein-aggregation related diseases.

Many structured proteins form oligomers with defined sizes, and effects of crowding on the stability of some of them have been studied. As expected, stabilization much greater than that for dimers formed by single-domain proteins is observed. For example, using magnetic relaxation dispersion, Snoussi and Halle [76] found a 17000-fold increase in the association constant for BPTI decamer by 230 g/l of dextran 10K. Similarly, Aguilar et al. [77], based on tryptophan fluorescence, found that 300 g/l Ficoll resulted in a 300-fold increase in the association constant for heptamer formation by mitochondrial cochaperonin protein 10.

The significant stabilizing effects of crowding on protein oligomers suggest that many proteins that are identified as monomeric based on dilute-solution assays may actually be oligomeric inside cells. The α-synuclein tetramer recently found inside cells [48, 49] could be such an example, though as mentioned above the native oligomeric state is still controversial.

The increase in the effects of crowding with increasing physical or oligomeric sizes of reactant species potentially leads to an important downside: small-molecule drugs that are effective in blocking the formation of large complexes or high oligomers in dilute solution may lose their efficacy under crowded conditions. That is because crowding disproportionately stabilizes the higher oligomer over the competing complex formed by the drug molecule and the monomer (Figure 3). This expectation is borne out by a recent study on the inhibition of the assembly of the HIV-1 capsid protein by a capsid protein fragment or a short peptide [78]. In dilution solution, both inhibitors were found to be effective in slowing down the assembly and reducing the total assembled mass. Both Ficoll and dextran

were found to speed up the assembly in the absence of inhibitors [8]. In the presence of either inhibitor, the two crowding agents were still able to speed up the assembly and nearly maintain the total assembled mass. In other words, the inhibitors lost their efficacy in the presence of the crowding agents.

Toward quantitative predictions: theoretical, coarse-grained, and atomistic modeling

In a number of the experimental studies reviewed above [7, 10, 12, 16, 57, 70, 72, 76], comparison was made to theoretical models based on representing reactant and crowder molecules as hard convex particles. Although these models often give qualitatively correct trends, the adjustable parameters in these models, such as on the shapes and sizes of the molecules, erode their predictive power. Recently molecular simulations have opened the possibility of making quantitative predictions and subjecting these predictions to direct interrogation by experiments.

To calculate the effects of crowding on the thermodynamic properties of biochemical processes, one can emulate what is done experimentally and simulates the reactant proteins along a biochemical process, both in the absence of crowders and in the presence of crowders [79–82] (horizontal paths in Figure 4a). The advantage of this direct simulation approach is that one can visualize how interactions with crowders reshape the energy landscapes of the reactant molecules. However, because one has to sample rare transitions between stable states along the biochemical process in order to calculate free energy differences, and has to do so in the presence of a large number of crowder molecules (Figure 4b), the direct simulation approach inherently carries a formidable computational cost.

The alternative approach is to follow the vertical paths in Figure 4a [73, 83, 84]. That is, one simulates just the stable states in dilute solution and then calculates the free energies of transferring these stable-state conformational ensembles to crowded solution (Figure 4c). In this “postprocessing” approach, one circumvents the need for sampling rare transitions between stable states. However, one does have to take the additional step of calculating the transfer free energies. Since the horizontal paths and vertical paths form a closed thermodynamic cycle, both approaches yield $\delta\Delta G$, the effect of crowding on the free energy difference between the reactant and product states.

To reduce the formidable computational cost of the direct simulation approach, it has only been implemented by representing the reactant protein molecules at the coarse-grained level, e.g., one bead per residue. This representation potentially limits the predictive power of the direct simulation approach, and so far it has mostly been used for calculating qualitative trends. The first such simulation study was by Cheung et al. [79], who calculated the folding stability and kinetics of the 34-residue WW domain in the presence of purely repulsive spherical crowders. Conformational sampling was helped with the replica exchange method. At a crowder volume fraction (Φ) of 0.25, the melting temperature was predicted to increase by as much as 24 °C. In a similar study, Mittal and Best [82] calculated the effects of crowding on the folding free energy surfaces of three small proteins. The reaction coordinate was the fraction of native contacts, and the free energy surfaces were obtained by combining the replica exchange method with umbrella sampling. Also using direct simulation, Minh et al. [80] studied the effects of crowding on the flap opening equilibrium of the HIV-1 protease dimer. This opening motion is essential for substrate and inhibitor binding. They found that repulsive crowders decreased the flap open population.

Recently Feig and Sugita [85] carried out all-atom explicit solvent molecular dynamics simulations of CI2 in the presence of either lysozyme or BSA as crowders. These

simulations were motivated by the experiments of Miklos et al. [32], who, as noted above, found that the protein crowders decreased the local stability of many of the monitored CI2 residues. Interestingly, the residues showing decreased local stability largely overlapped with residues showing increased conformational fluctuations in the simulations in the presence of the protein crowders. It should be noted that the experiments of Miklos et al. present measurements on the equilibria between the folded state and the locally or globally unfolded state, whereas the simulations of Feig and Sugita probe nanosecond conformational fluctuations in the folded state.

The postprocessing approach uses only crowder-free simulations of proteins in their reactant and product states. Therefore the proteins can be represented at the all-atom level and the simulations done in explicit solvent. The atomistic representation makes it possible to make quantitative predictions on the effects of crowding. The additional step to calculate the transfer free energy ($\Delta\mu$) of the reactant proteins requires a separate simulation of the crowder molecules to generate a configurational ensemble for the crowder solution. The latter simulation can be expensive, but needs to be done only once and then can be used over and over for different reactant proteins in various states.

To calculate $\Delta\mu$, one fictitiously inserts each protein conformation into the crowder solution (Figure 4c). If the effective interaction energy between a reactant protein and the crowders is $U(\mathbf{X}, \mathbf{R}, \mathbf{\Omega})$, where **X, R**, and **Ω** represent the conformation, position, and orientation, respectively, of the protein, then

$$\exp(-\Delta\mu/k_{\mathrm{B}}T)=<\exp[-U(\mathbf{X},\mathbf{R},\mathbf{\Omega})/k_{\mathrm{B}}T]>_0 \quad (1)$$

where $<\cdots>_0$ means averaging over the conformation, position, and orientation of the protein and over the configuration of the crowders. Both a numerical algorithm [73] and a theoretical formula [69] have been developed for calculating $\Delta\mu$. The theoretical formula, validated by the numerical algorithm, speeds up the $\Delta\mu$ calculations considerably and is essential for cases where the numerical algorithm becomes impractical, such as when the reactant molecules are very large. Notably, the theoretical formula enabled the $\Delta\mu$ calculations for the binding of the large and small subunits of the ribosome [69]. In these calculations, the reactant molecules are represented at the all-atom level.

The postprocessing approach yields the transfer free energy by reweighting the protein conformations, sampled in the absence of crowders, according to the would-be protein-crowder interaction energy. The important conformational region will likely be shifted by the crowders. The postprocessing calculation would be exact if the conformational space of the protein is exhaustively sampled so as to cover the important conformational region in the presence of crowders. For both the flap open population of the HIV-1 dimer [83] and the free energy surfaces of three small proteins [86], the results of the postprocessing approach predicted from the crowder-free simulations were found to agree well with those calculated from the direct simulation approach in the presence of crowders [80, 82].

In several studies, postprocessing predictions have been interrogated by experimental results. In the first such study [73], a modest increase in the folding stability of a four-helix bundle protein by 85 g/l PEG 20K [13] was reproduced when the folded and unfolded states were represented atomistically while the crowder was modeled simplistically as a 20 Å sphere. A subsequent study [87], with the folding transition state represented atomistically, also yielded the experimentally observed small effects of crowding on the folding and unfolding rates. Similarly, modest increases in the binding stability of the ε and θ subunits of *E. coli* DNA polymerase III by dextran of various molecular weights and Ficoll70 were quantitatively rationalized with atomistic representation of the proteins [18]. Using the

postprocessing approach, Johansen et al. [55] was able to explain the compaction of the N protein of bacteriophage λ when crowded by BPTI. While in these applications of the postprocessing approach the crowders were modeled as repulsive spheres, a major step forward was taken by McGuffee and Elcock [84], who represented protein crowders at the coarse-grained level with compositions and concentrations mimicking those of the *E. coli* cytoplasm, and included repulsive as well as attractive interactions with the reactant proteins.

Conclusion and outlook

Various *in vitro* and *in vivo* studies have now demonstrated effects of crowding that range from modest to drastic. On the modest side are protein folding stability and binding stability of single-domain proteins. On the drastic side are the efficiency of replication and transcription, the rates of protein aggregations linked to Parkinson's and other diseases, and the efficacy of small-molecule drugs. Regardless of magnitudes of the effects, the biological consequences of crowding in all these cases are important.

Two emerging trends will significantly contribute to the progress of our understanding on the influences of the crowded cellular environments. The first is the convergence of *in vitro* and *in vivo* observations on the effects of crowding. For example, the Pielak lab has observed similar compaction or structure formation inside cells and when crowded by BSA [54, 65]. Likewise the Gruebele lab has observed similar increases in the closed population of a folded protein inside cells [38, 60] and when crowded by Ficoll [59]. These convergent observations strengthen the confidence in our understanding on the nature of the cellular environments and in our ability to make inferences on biochemical processes inside cells based on experiments under cytomimetic conditions.

The second emerging trend is the convergence of experimental and computational studies on the same reactant proteins under the same crowding conditions. In this regard, the postprocessing approach, with the ability to represent reactant proteins and crowders at the atomistic level, will have a significant role to play. Unlike the previous generation of theoretical models based on a hard-particle representation, the new computational approach potentially holds truly predictive power, because the parameters of intermolecular interactions, once set, should be applicable to all reactant proteins and all crowders. There is hope that computations will be able to capture the subtle and complex effects observed in recent experiments, and the integration of the two will bring us toward a quantitative understanding on the influences of the crowded cellular environments.

Acknowledgments

This work was supported in part by Grant GM88187 from the National Institutes of Health.

ABBREVIATIONS

BPTI	bovine pancreatic trypsin inhibitor
BSA	bovine serum albumin
CI2	chymotrypsin inhibitor 2
HbF	fetal hemoglobin
HbS	sickle hemoglobin
HSQC	^{1}H-^{15}N heteronuclear single quantum coherence

IDP	intrinsically disordered protein
PEG	polyethylene glycol
PVP	poly(vinylpyrrolidone)

REFERENCES

1. Zimmerman SB, Minton AP. Macromolecular crowding: biochemical, biophysical, and physiological consequences. Annu Rev Biophys Biomol Struct. 1993; 22:27–65. [PubMed: 7688609]
2. Kornberg A. Ten commandments: lessons from the enzymology of DNA replication. J Bacteriol. 2000; 182:3613–3618. [PubMed: 10850972]
3. Ellis RJ. Macromolecular crowding: obvious but underappreciated. Trends Biochem Sci. 2001; 26:597–604. [PubMed: 11590012]
4. Fuller RS, Kaguni JM, Kornberg A. Enzymatic replication of the origin of the Escherichia coli chromosome. Proc Natl Acad Sci U S A. 1981; 78:7370–7374. [PubMed: 6278471]
5. Shtilerman MD, Ding TT, Lansbury PT Jr. Molecular crowding accelerates fibrillization of alpha-synuclein: could an increase in the cytoplasmic protein concentration induce Parkinson's disease? Biochemistry. 2002; 41:3855–3860. [PubMed: 11900526]
6. Uversky VN, E MC, Bower KS, Li J, Fink AL. Accelerated alpha-synuclein fibrillation in crowded milieu. FEBS Lett. 2002; 515:99–103. [PubMed: 11943202]
7. Hatters DM, Minton AP, Howlett GJ. Macromolecular crowding accelerates amyloid formation by human apolipoprotein C-II. J Biol Chem. 2002; 277:7824–7830. [PubMed: 11751863]
8. del Alamo M, Rivas G, Mateu MG. Effect of macromolecular crowding agents on human immunodeficiency virus type 1 capsid protein assembly in vitro. J Virol. 2005; 79:14271–14281. [PubMed: 16254362]
9. Gonzalez JM, Jimenez M, Velez M, Mingorance J, Andreu JM, Vicente M, Rivas G. Essential cell division protein FtsZ assembles into one monomer-thick ribbons under conditions resembling the crowded intracellular environment. J Biol Chem. 2003; 278:37664–37671. [PubMed: 12807907]
10. Rotter M, Aprelev A, Adachi K, Ferrone FA. Molecular crowding limits the role of fetal hemoglobin in therapy for sickle cell disease. J Mol Biol. 2005; 347:1015–1023. [PubMed: 15784260]
11. Qu Y, Bolen DW. Efficacy of macromolecular crowding in forcing proteins to fold. Biophys Chem. 2002; 101–102:155–165.
12. Spencer DS, Xu K, Logan TM, Zhou H-X. Effects of pH, salt, and macromolecular crowding on the stability of FK506-binding protein: an integrated experimental and theoretical study. J Mol Biol. 2005; 351:219–232. [PubMed: 15992823]
13. Ai X, Zhou Z, Bai Y, Choy WY. ^{15}N NMR spin relaxation dispersion study of the molecular crowding effects on protein folding under native conditions. J Am Chem Soc. 2006; 128:3916–3917. [PubMed: 16551092]
14. Roberts A, Jackson SE. Destabilised mutants of ubiquitin gain equal stability in crowded solutions. Biophys Chem. 2007; 128:140–149. [PubMed: 17434659]
15. Charlton LM, Barnes CO, Li C, Orans J, Young GB, Pielak GJ. Residue-level interrogation of macromolecular crowding effects on protein stability. J Am Chem Soc. 2008; 130:6826–6830. [PubMed: 18459780]
16. Batra J, Xu K, Zhou H-X. Nonaddtive effects of mixed crowding on protein stability. Proteins. 2009; 77:133–138. [PubMed: 19408299]
17. Phillip Y, Sherman E, Haran G, Schreiber G. Common crowding agents have only a small effect on protein-protein interactions. Biophys J. 2009; 97:875–885. [PubMed: 19651046]
18. Batra J, Xu K, Qin S, Zhou H-X. Effect of macromolecular crowding on protein binding stability: modest stabilization and significant biological consequences. Biophys J. 2009; 97:906–911. [PubMed: 19651049]

19. Zhou HX. Rate theories for biologists. Q Rev Biophys. 2010; 43:219–293. [PubMed: 20691138]
20. Minton AP. Excluded volume as a determinant of macromolecular structure and reactivity. Biopolymers. 1981; 20:2093–2120.
21. Minton AP. Molecular crowding: analysis of effects of high concentrations of inert cosolutes on biochemical equilibria and rates in terms of volume exclusion. Methods Enzymol. 1998; 295:127–149. [PubMed: 9750217]
22. Ellis RJ, Minton AP. Protein aggregation in crowded environments. Biol Chem. 2006; 387:485–497. [PubMed: 16740119]
23. Zhou HX, Rivas G, Minton AP. Macromolecular crowding and confinement: biochemical, biophysical, and potential physiological consequences. Annu Rev Biophys. 2008; 37:375–397. [PubMed: 18573087]
24. Elcock AH. Models of macromolecular crowding effects and the need for quantitative comparisons with experiment. Curr Opin Struct Biol. 2010; 20:196–206. [PubMed: 20167475]
25. Gershenson A, Gierasch LM. Protein folding in the cell: challenges and progress. Curr Opin Struc Biol. 2011; 21:32–41.
26. Ebbinghaus S, Gruebele M. Protein Folding Landscapes in the Living Cell. J Phys Chem Lett. 2011; 2:314–319.
27. Minton, A. Holobiochemistry: an integrated approach to the understanding of biochemical mechanism that emerges from the study of proteins and protein associations in volume-occupied solutions. In: Srere, P.; Jones, M.; Mathews, C., editors. Structural and Organizational Aspects of Metabolic Regulation. New York: Liss; 1989. p. 291-306.
28. Zhou H-X. Protein folding and binding in confined spaces and in crowded solutions. J Mol Recognit. 2004; 17:368–375. [PubMed: 15362094]
29. Ferrone FA, Rotter MA. Crowding and the polymerization of sickle hemoglobin. J Mol Recognit. 2004; 17:497–504. [PubMed: 15362110]
30. Zhou HX, Qin S. Simulation and modeling of crowding effects on the thermodynamic and kinetic properties of proteins with atomic details. Biophys Rev. 2013 in press.
31. Miklos AC, Li CG, Sharaf NG, Pielak GJ. Volume exclusion and soft interaction effects on protein stability under crowded conditions. Biochemistry. 2010; 49:6984–6991. [PubMed: 20672856]
32. Miklos AC, Sarkar M, Wang Y, Pielak GJ. Protein crowding tunes protein stability. J Am Chem Soc. 2011; 133:7116–7120. [PubMed: 21506571]
33. Wang Y, Sarkar M, Smith AE, Krois AS, Pielak GJ. Macromolecular crowding and protein stability. J Am Chem Soc. 2012; 134:16614–16618. [PubMed: 22954326]
34. Benton LA, Smith AE, Young GB, Pielak GJ. Unexpected effects of macromolecular crowding on protein stability. Biochemistry. 2012; 51:9773–9775. [PubMed: 23167542]
35. Englander SW, Kallenbach NR. Hydrogen exchange and structural dynamics of proteins and nucleic acids. Q Rev Biophys. 1983; 16:521–655. [PubMed: 6204354]
36. Zhou HX. Polymer crowders and protein crowders act similarly on protein folding stability. FEBS Lett. 2013 in press.
37. Denos S, Dhar A, Gruebele M. Crowding effects on the small, fast-folding protein lambda6-85. Faraday Discuss. 2012; 157:451–462. discussion 475–500. [PubMed: 23230782]
38. Guo MH, Xu YF, Gruebele M. Temperature dependence of protein folding kinetics in living cells. Proc Natl Acad Sci U S A. 2012; 109:17863–17867. [PubMed: 22665776]
39. Wang Y, Li C, Pielak GJ. Effects of proteins on protein diffusion. J Am Chem Soc. 2010; 132:9392–9397. [PubMed: 20560582]
40. Schlesinger AP, Wang Y, Tadeo X, Millet O, Pielak GJ. Macromolecular crowding fails to fold a globular protein in cells. J Am Chem Soc. 2011; 133:8082–8085. [PubMed: 21534566]
41. Burre J, Sharma M, Tsetsenis T, Buchman V, Etherton MR, Sudhof TC. Alpha-synuclein promotes SNARE-complex assembly in vivo and in vitro. Science. 2010; 329:1663–1667. [PubMed: 20798282]
42. Weinreb PH, Zhen W, Poon AW, Conway KA, Lansbury PT Jr. NACP, a protein implicated in Alzheimer's disease and learning, is natively unfolded. Biochemistry. 1996; 35:13709–13715. [PubMed: 8901511]

43. Eliezer D, Kutluay E, Bussell R Jr, Browne G. Conformational properties of alpha-synuclein in its free and lipid-associated states. J Mol Biol. 2001; 307:1061–1073. [PubMed: 11286556]
44. Morar AS, Olteanu A, Young GB, Pielak GJ. Solvent-induced collapse of alpha-synuclein and acid-denatured cytochrome c. Protein Sci. 2001; 10:2195–2199. [PubMed: 11604526]
45. Uversky VN, Li J, Fink AL. Evidence for a partially folded intermediate in alpha-synuclein fibril formation. J Biol Chem. 2001; 276:10737–10744. [PubMed: 11152691]
46. Ulmer TS, Bax A, Cole NB, Nussbaum RL. Structure and dynamics of micelle-bound human alpha-synuclein. J Biol Chem. 2005; 280:9595–9603. [PubMed: 15615727]
47. Sandal M, Valle F, Tessari I, Mammi S, Bergantino E, Musiani F, Brucale M, Bubacco L, Samori B. Conformational equilibria in monomeric alpha-synuclein at the single-molecule level. PLoS Biol. 2008; 6:e6. [PubMed: 18198943]
48. Bartels T, Choi JG, Selkoe DJ. alpha-Synuclein occurs physiologically as a helically folded tetramer that resists aggregation. Nature. 2011; 477:107–110. [PubMed: 21841800]
49. Wang W, Perovic I, Chittuluru J, Kaganovich A, Nguyen LT, Liao J, Auclair JR, Johnson D, Landeru A, Simorellis AK, Ju S, Cookson MR, Asturias FJ, Agar JN, Webb BN, Kang C, Ringe D, Petsko GA, Pochapsky TC, Hoang QQ. A soluble alpha-synuclein construct forms a dynamic tetramer. Proc Natl Acad Sci U S A. 2011; 108:17797–17802. [PubMed: 22006323]
50. Fauvet B, Mbefo MK, Fares MB, Desobry C, Michael S, Ardah MT, Tsika E, Coune P, Prudent M, Lion N, Eliezer D, Moore DJ, Schneider B, Aebischer P, El-Agnaf OM, Masliah E, Lashuel HA. alpha-Synuclein in central nervous system and from erythrocytes, mammalian cells, and Escherichia coli exists predominantly as disordered monomer. J Biol Chem. 2012; 287:15345–15364. [PubMed: 22315227]
51. Lashuel HA, Overk CR, Oueslati A, Masliah E. The many faces of alphasynuclein: from structure and toxicity to therapeutic target. Nat Rev Neurosci. 2013; 14:38–48. [PubMed: 23254192]
52. Trexler AJ, Rhoades E. N-Terminal acetylation is critical for forming alpha-helical oligomer of alpha-synuclein. Protein Sci. 2012; 21:601–605. [PubMed: 22407793]
53. McNulty BC, Tripathy A, Young GB, Charlton LM, Orans J, Pielak GJ. Temperature-induced reversible conformational change in the first 100 residues of alpha-synuclein. Protein Sci. 2006; 15:602–608. [PubMed: 16452621]
54. McNulty BC, Young GB, Pielak GJ. Macromolecular crowding in the Escherichia coli periplasm maintains alpha-synuclein disorder. J Mol Biol. 2006; 355:893–897. [PubMed: 16343531]
55. Johansen D, Jeffries CMJ, Hammouda B, Trewhella J, Goldenberg DP. Effects of macromolecular crowding on an intrinsically disordered protein characterized by small-angle neutron scattering with contrast matching. Biophys J. 2011; 100:1120–1128. [PubMed: 21320458]
56. Szasz C, Alexa A, Toth K, Rakacs M, Langowski J, Tompa P. Protein Disorder Prevails under Crowded Conditions. Biochemistry. 2011; 50:5834–5844. [PubMed: 21634433]
57. Le Coeur C, Teixeira J, Busch P, Longeville S. Compression of random coils due to macromolecular crowding: scaling effects. Phys Rev E. 2010; 81:061914.
58. Hong J, Gierasch LM. Macromolecular crowding remodels the energy landscape of a protein by favoring a more compact unfolded state. J Am Chem Soc. 2010; 132:10445–10452. [PubMed: 20662522]
59. Dhar A, Samiotakis A, Ebbinghaus S, Nienhaus L, Homouz D, Gruebele M, Cheung MS. Structure, function, and folding of phosphoglycerate kinase are strongly perturbed by macromolecular crowding. Proc Natl Acad Sci U S A. 2010; 107:17586–17591. [PubMed: 20921368]
60. Dhar A, Girdhar K, Singh D, Gelman H, Ebbinghaus S, Gruebele M. Protein stability and folding kinetics in the nucleus and endoplasmic reticulum of eucaryotic cells. Biophys J. 2011; 101:421–430. [PubMed: 21767495]
61. Hughes KT, Gillen KL, Semon MJ, Karlinsey JE. Sensing structural intermediates in bacterial flagellar assembly by export of a negative regulator. Science. 1993; 262:1277–1280. [PubMed: 8235660]
62. Daughdrill GW, Chadsey MS, Karlinsey JE, Hughes KT, Dahlquist FW. The C-terminal half of the anti-sigma factor, FlgM, becomes structured when bound to its target, sigma 28. Nat Struct Biol. 1997; 4:285–291. [PubMed: 9095196]

63. Daughdrill GW, Hanely LJ, Dahlquist FW. The C-terminal half of the anti-sigma factor FlgM contains a dynamic equilibrium solution structure favoring helical conformations. Biochemistry. 1998; 37:1076–1082. [PubMed: 9454599]
64. Sorenson MK, Ray SS, Darst SA. Crystal structure of the flagellar sigma/anti-sigma complex sigma(28)/FlgM reveals an intact sigma factor in an inactive conformation. Mol Cell. 2004; 14:127–138. [PubMed: 15068809]
65. Dedmon MM, Patel CN, Young GB, Pielak GJ. FlgM gains structure in living cells. Proc Natl Acad Sci U S A. 2002; 99:12681–12684. [PubMed: 12271132]
66. Vavouri T, Semple JI, Garcia-Verdugo R, Lehner B. Intrinsic protein disorder and interaction promiscuity are widely associated with dosage sensitivity. Cell. 2009; 138:198–208. [PubMed: 19596244]
67. Gsponer J, Futschik ME, Teichmann SA, Babu MM. Tight regulation of unstructured proteins: from transcript synthesis to protein degradation. Science. 2008; 322:1365–1368. [PubMed: 19039133]
68. Chadsey MS, Karlinsey JE, Hughes KT. The flagellar anti-sigma factor FlgM actively dissociates Salmonella typhimurium sigma28 RNA polymerase holoenzyme. Genes Dev. 1998; 12:3123–3136. [PubMed: 9765212]
69. Qin S, Zhou HX. Generalized fundamental measure theory for atomistic modeling of macromolecular crowding. Phys Rev E. 2010; 81:031919.
70. Zimmerman SB, Trach SO. Effects of macromolecular crowding on the association of E. coli ribosomal particles. Nucleic Acids Res. 1988; 16:6309–6326. [PubMed: 3041372]
71. Jarvis TC, Ring DM, Daube SS, von Hippel PH. “Macromolecular crowding”: thermodynamic consequences for protein-protein interactions within the T4 DNA replication complex. J Biol Chem. 1990; 265:15160–15167. [PubMed: 2168402]
72. Jiao M, Li HT, Chen J, Minton AP, Liang Y. Attractive protein-polymer interactions markedly alter the effect of macromolecular crowding on protein association equilibria. Biophys J. 2010; 99:914–923. [PubMed: 20682270]
73. Qin S, Zhou H-X. Atomistic modeling of macromolecular crowding predicts modest increases in protein folding and binding stability. Biophys J. 2009; 97:12–19. [PubMed: 19580740]
74. Huang LQ, Jin R, Li JR, Luo K, Huang T, Wu D, Wang WX, Chen R, Xiao GF. Macromolecular crowding converts the human recombinant PrPC to the soluble neurotoxic beta-oligomers. FASEB J. 2010; 24:3536–3543. [PubMed: 20400537]
75. Silveira JR, Raymond GJ, Hughson AG, Race RE, Sim VL, Hayes SF, Caughey B. The most infectious prion protein particles. Nature. 2005; 437:257–261. [PubMed: 16148934]
76. Snoussi K, Halle B. Protein self-association induced by macromolecular crowding: A quantitative analysis by magnetic relaxation dispersion. Biophys J. 2005; 88:2855–2866. [PubMed: 15665132]
77. Aguilar X, Weise CF, Sparrman T, Wolf-Watz M, Wittung-Stafshede P. Macromolecular Crowding Extended to a Heptameric System: The Co-chaperonin Protein 10. Biochemistry. 2011; 50:3034–3044. [PubMed: 21375247]
78. Rincon V, Bocanegra R, Rodriguez-Huete A, Rivas G, Mateu MG. Effects of Macromolecular Crowding on the Inhibition of Virus Assembly and Virus-Cell Receptor Recognition. Biophys J. 2011; 100:738–746. [PubMed: 21281589]
79. Cheung MS, Klimov D, Thirumalai D. Molecular crowding enhances native state stability and refolding rates of globular proteins. Proc Natl Acad Sci U S A. 2005; 102:4753–4758. [PubMed: 15781864]
80. Minh DD, Chang CE, Trylska J, Tozzini V, McCammon JA. The influence of macromolecular crowding on HIV-1 protease internal dynamics. J Am Chem Soc. 2006; 128:6006–6007. [PubMed: 16669648]
81. Kim YC, Best RB, Mittal J. Macromolecular crowding effects on protein-protein binding affinity and specificity. J Chem Phys. 2010; 133:205101. [PubMed: 21133453]
82. Mittal J, Best RB. Dependence of protein folding stability and dynamics on the density and composition of macromolecular crowders. Biophys J. 2010; 98:315–320. [PubMed: 20338853]

83. Qin S, Minh DD, McCammon JA, Zhou HX. Method to predict crowding effects by postprocessing molecular dynamics trajectories: application to the flap dynamics of HIV-1 protease. J Phys Chem Lett. 2010; 1:107–110. [PubMed: 20228897]

84. McGuffee SR, Elcock AH. Diffusion, crowding & protein stability in a dynamic molecular model of the bacterial cytoplasm. PLoS Comput Biol. 2010; 6:e1000694. [PubMed: 20221255]

85. Feig M, Sugita Y. Variable interactions between protein crowders and biomolecular solutes are important in understanding cellular crowding. J Phys Chem B. 2012; 116:599–605. [PubMed: 22117862]

86. Qin S, Mittal J, Zhou HX. Folding free energy surfaces of three small proteins under crowding: validation of the postprocessing method by direct simulation. Phys Biol. 2013 submitted.

87. Tjong H, Zhou HX. The folding transition-state ensemble of a four-helix bundle protein: helix propensity as a determinant and macromolecular crowding as a probe. Biophys J. 2010; 98:2273–2280. [PubMed: 20483336]

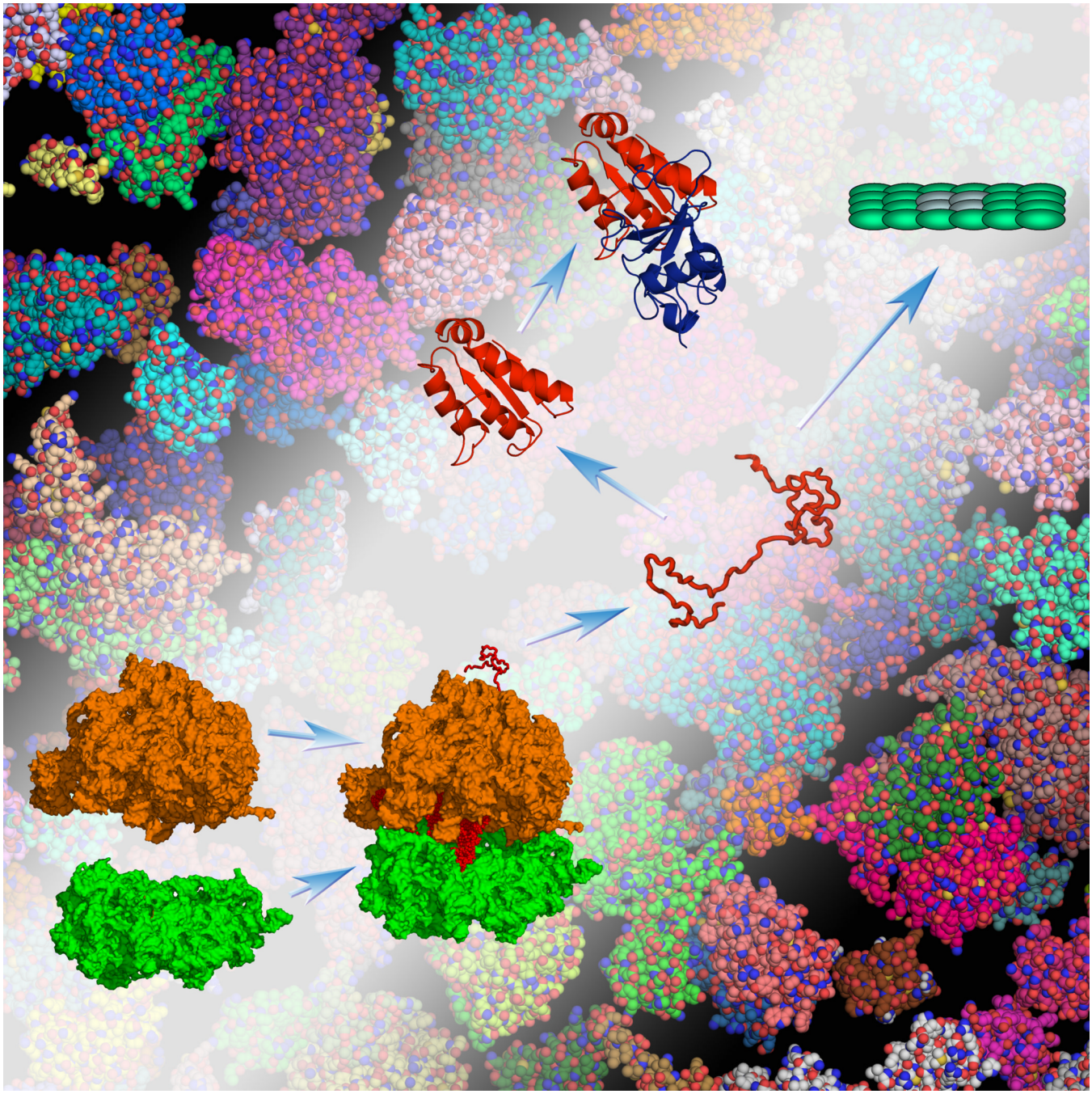

Fig. 1.
Inside cells biochemical processes occur in crowded environments. The depicted processes include the binding of the large and small subunits of the ribosome, the folding of a nascent protein, its binding to another protein, and aggregation. The crowded conditions can significantly change the relative stability among the various species.

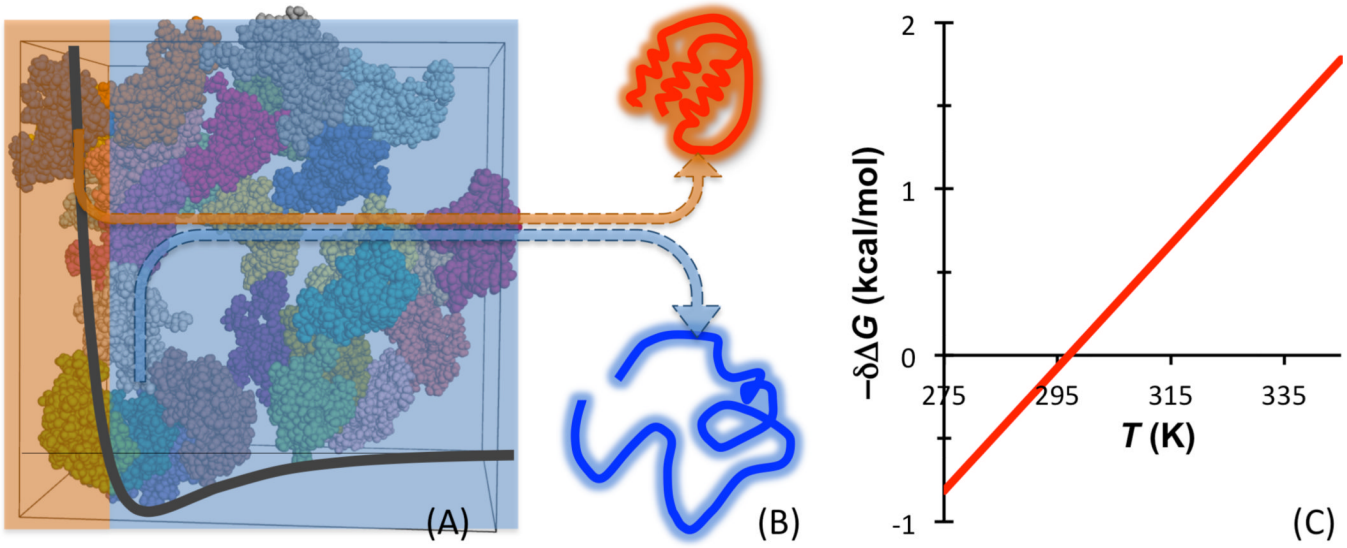

Fig. 2.

Illustration of the effects of macromolecular crowding on protein folding stability. (A) The interactions between a protein and surrounding crowder molecules consist of hard-core repulsion and longer-ranged attraction. (B) The hard-core repulsion leads to an entropic component that favors the folded state of the protein, whereas the longer-ranged attraction leads to an enthalpic component that favors the unfolded state. (C) The net effect of macromolecular crowding has a crossover temperature, where $\delta\Delta G = 0$.

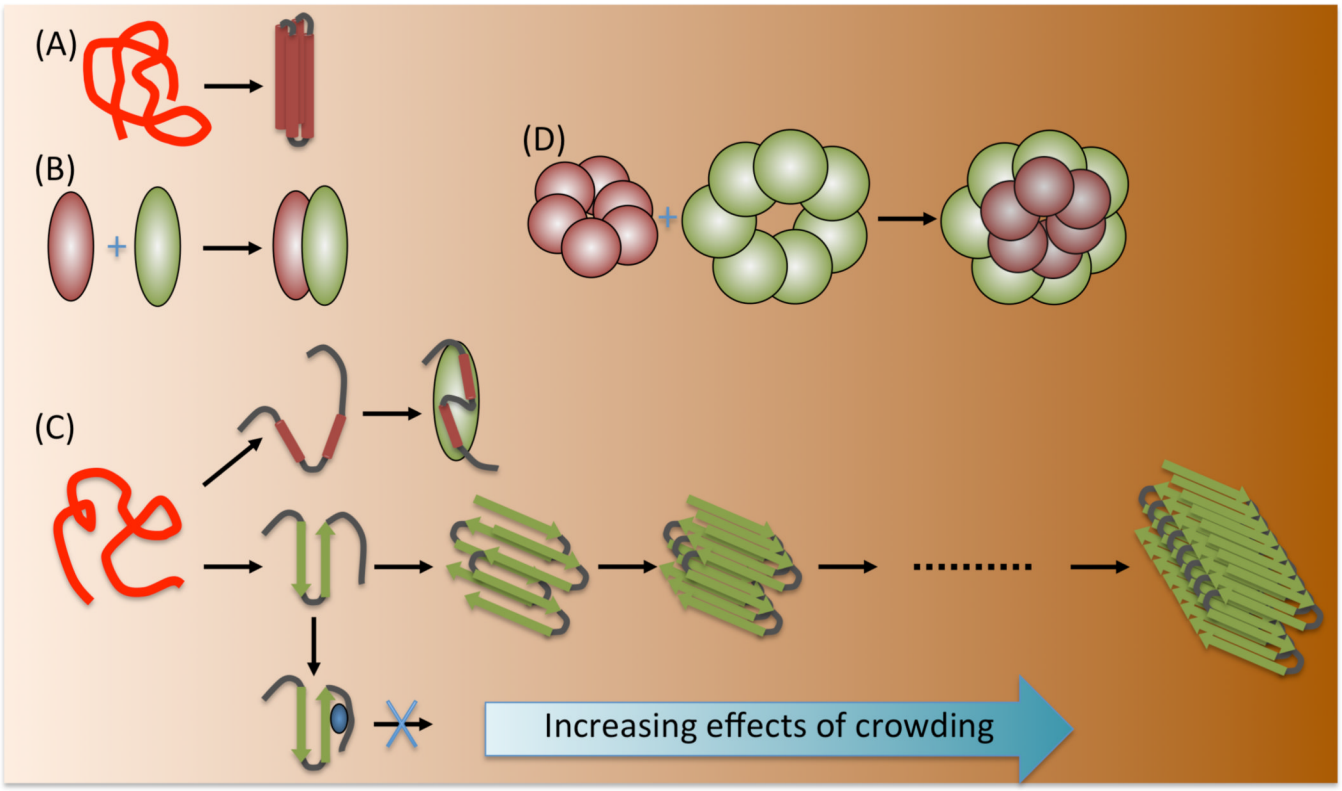

Fig. 3.

Varying magnitudes of the effects of macromolecular crowding on different biochemical processes. The effects are expected to be modest for (A) folding stability and (B) binding stability of single-domain proteins, (C) conformational changes of an IDP and its binding to a structured partner, and the binding of a small-molecule drug to a protein monomer, but dramatic for (D) the binding of multimeric proteins, (C) oligomerization, and aggregation.

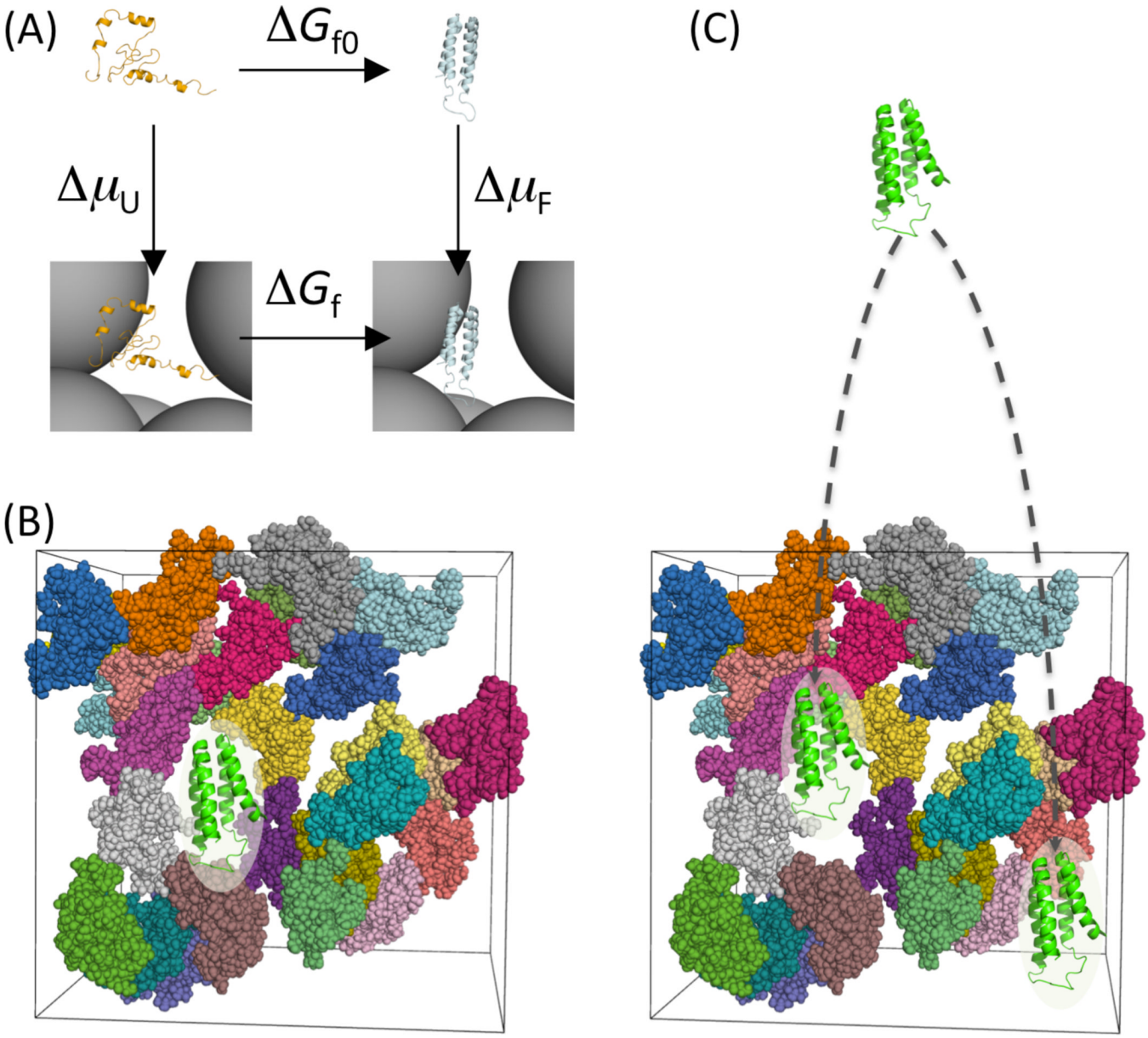

Fig. 4.

Two different approaches for modeling biochemical processes under crowding. (A) The direct simulation approach follows the horizontal paths whereas the postprocessing approach follows the vertical paths, illustrated on the folding process of a protein. The same free energy difference $\delta\Delta G$, here representing the effect of crowding on the folding free energy, is obtained by following either the horizontal paths ($\delta\Delta G = \Delta G_f - \Delta G_{f0}$) or the vertical paths ($\delta\Delta G = \Delta\mu_F - \Delta\mu_U$). (b) In the direct simulation approach, one follows the folding transition in the presence of crowders. Here the protein is shown in a folded conformation. (c) In the postprocessing approach, one simulates only the folded state and the unfolded state, not their inter-conversion, and does so in the absence of crowders [73]. To calculate $\Delta\mu_F$, folded conformations are fictitiously inserted into the crowders. An analogous calculation yields $\Delta\mu_U$.