

## Review Article

# The pleiotropic role of statins: Could it be the imminent host modulation agent in periodontics?

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## ABSTRACT

Periodontal disease is a chronic inflammatory disease which represents a primarily anaerobic Gram-negative oral infection that results in gingival inflammation, loss of attachment, bone destruction. Bacterial endotoxins in the form of lipopolysaccharides (LPS) that are instrumental in generating a host-mediated tissue destructive immune response by mobilizing their defensive cells and releasing cytokines like Interleukin-1 $\beta$  (IL-1 $\beta$ ), Tumor Necrosis Factor- $\alpha$  (TNF- $\alpha$ ), and Interleukin-6 (IL-6), which lead to tissue destruction by stimulating the production of the collagenolytic enzymes: Matrix metalloproteinases (MMPs). Since the host-mediated tissue destruction is to be controlled, various means have been employed for modulating this response. Statins, 3-hydroxy-3-methylglutarylcoenzyme A (HMG CoA) reductase inhibitors, besides having lipid-lowering abilities also have antioxidant, antithrombotic, anti-inflammatory, immunomodulatory and osteomodulatory properties. All of these pleiotropic effects of statins point out to it perhaps becoming the novel host modulation agent in periodontics.

**Key Words:** Anti-inflammatory, host modulation, immunomodulation, osseo-modulatory, periodontics, statins

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## INTRODUCTION

Periodontal disease is a chronic inflammatory disease which represents a primarily anaerobic Gram-negative oral infection that results in gingival inflammation, loss of attachment, bone destruction.<sup>[1]</sup> Certain organisms within the microbial flora of dental plaque are the major etiologic agents of periodontitis which produce endotoxins in the form of lipopolysaccharides (LPS) that are instrumental in generating a host-mediated tissue destructive immune response by mobilizing their defensive cells and releasing cytokines like Interleukin-1 $\beta$  (IL-1 $\beta$ ), Tumor

Necrosis Factor- $\alpha$  (TNF- $\alpha$ ), and Interleukin-6 (IL-6), which lead to tissue destruction by stimulating the production of the collagenolytic enzymes: Matrix metalloproteinases (MMPs).<sup>[2]</sup>

Since the host-mediated tissue destruction is to be controlled, various means have been employed for modulating this response. These include -Inhibition of MMP's with antiproteinases, blocking the proinflammatory cytokines and prostaglandins by use of anti-inflammatory drugs, and by inhibiting the osteoclasts activity by use of bone sparing agents.<sup>[3]</sup>

Statins, 3-hydroxy-3-methylglutarylcoenzyme A (HMG CoA) reductase inhibitors, can be fermentation derived statins include simvastatin, pravastatin whereas atorvastatin, cerivastatin, fluvastatin, pitavastatin and rosuvastatin are synthetic statins. Synthetic statins have a higher potency as compared to the fermented statins. Statins were primarily approved as lipid lowering agent to prevent cardiovascular events. Statins lower low density lipoprotein-C (LDL-C), but recent studies

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provide compelling evidence that statins also possess anti-inflammatory activity, independent of their lipid-lowering effects.<sup>[4,5]</sup>

Statins differ in terms of their chemical structures, pharmacokinetic profiles, and lipid-modifying efficacy. The chemical structures of statins govern their water solubility, which in turn influences their absorption, distribution, metabolism and excretion. Lovastatin, pravastatin and simvastatin are derived from fungal metabolites and have elimination half-lives of 1-3 h. Atorvastatin, fluvastatin and rosuvastatin are fully synthetic compounds, with elimination half-lives ranging from 1 h for fluvastatin to 19 h for rosuvastatin. Atorvastatin, simvastatin, lovastatin and fluvastatin are relatively lipophilic compounds. Lipophilic statins are more susceptible to metabolism by the cytochrome *P* (450) system. Pravastatin and rosuvastatin are relatively hydrophilic and not significantly metabolized by cytochrome *P* (450) enzymes. All statins are selective for effect in the liver, largely because of efficient first-pass uptake. The bioavailability of the statins differs greatly, from 5% for lovastatin and simvastatin to 60% or greater for cerivastatin.<sup>[6]</sup>

These structures can be broadly divided into three parts: An analogue of the target enzyme substrate, HMG-CoA; a complex hydrophobic ring structure that is covalently linked to the substrate analogue and is involved in binding of the statin to the reductase enzyme; side groups on the rings that define the solubility properties of the drugs and therefore many of their pharmacokinetic properties.<sup>[7]</sup>

Potential pleiotropic effects of statins involve immunomodulatory, antioxidant, antithrombotic and endothelium stabilization actions,<sup>[8]</sup> as well as angiogenesis promotion and increase of osteoblastic differentiation, inducing bone formation.<sup>[9]</sup> In addition, statins can inhibit tumor cells growth and enhance intracellular calcium mobilization. It was observed that inhibitors of HMG-CoA reductase induce a reduction of the formation of osteoclasts in rodents.<sup>[10]</sup> Human subjects treated with statins have shown a reduction in the number of bone fractures.<sup>[11]</sup>

## MECHANISM OF PLEIOTROPIC ACTION OF STATINS

### Anti-inflammatory

Continuously growing evidences have recognized inflammation as a major component of atherosclerosis.

When atherosclerotic process is at work, each characteristic lesion of atherosclerosis represents a different stage in a chronic inflammatory process in the artery; if unabated and excessive, this process will result in an advanced, complicated lesion.

Certain cell and cytokine mediated markers like C-reactive protein (CRP) are associated both with atherosclerosis and periodontal disease, proving a link between cardiovascular disease and periodontal disease.

CRP, a proinflammatory marker, is produced in liver and it induces release of IL-1, IL-6, and tumor necrosis factor by macrophages. CRP calcium-dependently binds to low density lipoproteins (LDL)<sup>[12]</sup> and induces foam cell formation. Kleemann and colleagues have shown that atorvastatin at doses higher than those required for cholesterol lowering, decrease basal and IL-1 $\beta$ -induced plasma human C-Reactive Protein (huCRP) levels.<sup>[13]</sup> Statins also inhibits Intercellular Adhesion Molecule-1 (ICAM-1) upregulation and chemotaxis in adherent human monocytes.<sup>[14]</sup>

Simvastatin lowers high sensitivity (hsCRP) by 14 days, independent of its effect on LDL cholesterol.<sup>[15]</sup> This rapid impact of a statin on hsCRP has potential implications in the management of acute coronary syndromes.

JUPITER trial showed that statins could be beneficial in the primary prevention of cardiovascular disease in patients with elevated hs CRP, but relatively low cholesterol levels and other cardiovascular risks.<sup>[16]</sup>

### Effect of statin on bone

Statins have been found to be a potent activator of bone morphogenetic protein (BMP-2) which accounts for major osteoinductive potential of bone. Simvastatin, mevastatin and atorvastatin stimulates BMP-2 transcription.

Mechanism of action involves reduction in mevalonate pathway intermediates and inhibition of prenylation by statins is responsible for a large proportion of the pleiotropic effects of these drugs. Mevalonate, farnesyl pyrophosphate and geranyl-geranyl pyrophosphate all inhibited statin-stimulated bone formation. Furthermore, because geranylgeranyl pyrophosphate inhibited statin stimulated bone formation, inhibition of prenylation due to geranylgeranylation must play a major role in the stimulation of bone formation by these drugs.<sup>[17]</sup>

Also statins influence the production of Receptor activator of nuclear factor kappa-B ligand (RANKL)

**Table 1: Different studies done on animal to explore the beneficial effects of different statins in animal models of dental conditions**

S. No	Statin used	Indication of use	Animals used in experiment	Results effective/ not effective	Route of administration	Authors
1.	Atorvastatin	Effect on radiographic density of alveolar bone	Wistar rats	E	Oral	Goes <i>et al.</i> [26]
2.	Simvastatin	Bone repair	Rats	NE	Topical	Lima <i>et al.</i> [27]
3.	Lovastatin or simvastatin	Bone stimulatory effect	Mouse periodontal ligament cells	E	( <i>In vitro</i> study)	Kim <i>et al.</i> [28]
4.	Simvastatin	Relapse and periodontal tissue remodeling after experimental tooth movement	Wistar rats	E	Oral	Han <i>et al.</i> [29]
5.	Simvastatin	Augmentation of facial jaw bone	Beagle dogs	E	Topical + Systemic	Rutledge <i>et al.</i> [30]
6.	Simvastatin	Calvarial bone healing	Rats	NE	Local	Calixto <i>et al.</i> [31]
7.	Simvastatin	Distraction osteogenesis	New Zealand White Rabbits	E	Local + Systemic	Kilic <i>et al.</i> [32]
8.	Simvastatin	i. Augmentation of bone thickness	Beagle dogs	E	Topical	Morris <i>et al.</i> [33]
9.		ii. Periodontal defects		NE		
10.	Simvastatin	Alveolar bone loss	Rat	E	Topical	Seto <i>et al.</i> [34]
11.	Fluvastatin	Bone healing around implants	Rat	E	Topical	Moriyama <i>et al.</i> [35]
12.	Simvastatin	Bone augmentation and preservation of alveolar ridge after tooth extraction	Rat	E	Topical	Nishimura <i>et al.</i> [36]
13.	Simvastatin	Extent of relapse after orthodontic treatment	Rat	E	Systemic	Chen <i>et al.</i> [37]
14.	Simvastatin	Mandibular bone formation	Rat	E	Systemic	Lee <i>et al.</i> [38]
15.	Simvastatin	Remodelling of the alveolar bone following tooth extraction	Wistar Rat	E	Topical	Wu, Liu, Zang, Sun <i>et al.</i> [39]
16.	Simvastatin	Residual ridge resorption after tooth extraction	Wistar rats	E	Topical	Wu <i>et al.</i> [40]
17.	Simvastatin	Bone regeneration of tibial defects and blood cholesterol level	Rats	NE	Systemic	Anbinder <i>et al.</i> [41]
18.	Simvastatin	Mandibular bone growth and inflammation	Rats	E	Topical	Stein <i>et al.</i> [42]
19.	Simvastatin	Promotion of osteogenesis around titanium implants	Rats	E	Systemic	Ayukawa <i>et al.</i> [43]
20.	Simvastatin	Guided bone regeneration in mandible	Rats	E	Oral	Junqueira <i>et al.</i> [44]

NE: Not effective, E: Effectiv

and Osteoprotegrin by human gingival fibroblasts to favor bone catabolism under non-inflammatory conditions [Table 1].<sup>[18]</sup>

### Immunomodulatory

Various experiments have shown statins to have immunomodulatory effects. Primary effects of statins include inhibition of major histocompatibility complex II expression. Antigen presenting cells such as endothelial cells and monocytes requires co-stimulation by Interferon gamma. This is done by inhibition of promoter IV of class II transcription activator which is an important regulator in this pathway. Another effect on immune system is mediated by binding of statins to leucocyte function associated antigen and preventing its binding to ICAM1. This leads to inhibition of its function in leucocyte adhesion and extravasation.<sup>[19]</sup>

Another mode of exerting immunomodulatory effects by statins include inhibition of release of proinflammatory cytokines like IFN- $\gamma$ , TNF- $\alpha$ , IL-1 $\beta$ , IL-6 in various cell types like microglia,

astrocytes, and mononuclear cells. These cytokines play an important role in T cell differentiation. Thus disrupting cytokine production can adversely affect the immune reactions of the body.<sup>[19]</sup>

### Antioxidant effects of statins

Antioxidant effect of statins is proposed as mechanism of its various pleiotropic effects. Many studies done to assess the antioxidant potential of statins have concluded favorably. Various mechanisms of its antioxidant activity are being proposed. Production of Nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, a major source of oxidant production is inhibited by statins. Simvastatin has been reported most effective in this regard.<sup>[20]</sup>

Few animal studies have demonstrated the upregulation and also increased activity of antioxidant enzyme catalase.<sup>[21]</sup> Studies with other antioxidant enzymes have shown inconsistent activity. Other studies have also demonstrated the free radical scavenging activity of statins.<sup>[22,23]</sup> Direct scavenging reactive oxygen species (ROS) prevents its interaction

with lipids, proteins and deoxyribonucleic acid. In this regard simvastatin and atorvastatin were more active against hydroxyl radical, and that fluvastatin was more active against peroxy radical.<sup>[24]</sup>

Statins have been shown in both human studies and animal models to upregulate Nitric oxide synthase-3 (NOS-3) expression<sup>[25]</sup> thereby increasing the levels of Nitric oxide (NO). Inhibition of formation of NO further inhibits conversion of superoxide ion to highly ROS peroxy nitrite.

### Implications of pleiotropic effects of statins in dental conditions

Various animal [Table 1] and human studies have been done to assess the effect of statins. A randomized controlled clinical trial on patients with chronic periodontitis showed that there was a greater decrease in gingival index (GI) and probing depth (PD) and more clinical attachment level gain with significant intrabony defect fill at sites treated with scaling and root planning (SRP) plus locally delivered simvastatin than with SRP alone.<sup>[45]</sup>

Another study showed beneficial effect of atorvastatin on bone alveolar loss and tooth mobility in subjects with periodontal disease.<sup>[46]</sup>

Few other cross sectional studies failed to conclusively state the beneficial effects of statin on periodontal conditions as such.<sup>[47,48]</sup>

### Statins in implants

According to Takeshita, *et al.* (1993) a mesh-like woven bone structure was predominantly formed around the implant 5 days after implantation, and then diminished gradually.<sup>[46]</sup> Ayukawa, *et al.*, showed that simvastatin promotes osteogenesis around titanium implants. Under the administration of statin, a similar mesh-like structure of bone was seen around the implant even at 30 days after the implantation which resulted in an improved implant fixation and that the bone network can successfully disperse the stress applied to the implant.<sup>[43]</sup> Also, topical application of statin affects bone healing around implants<sup>[35]</sup> [Table 1, S.No: 18, 10].

### Adverse effects

Adverse effects of statin therapy includes myalgias and muscle cramps. *SLCO1B1* gene, which participates in the absorption of statins, has been shown to significantly increase the risk of myopathy.<sup>[49]</sup> It is suggested that the risk of myopathy is lowest with pravastatin and fluvastatin probably because they are

more hydrophilic and as a result have less muscle penetration. Coenzyme Q10 (ubiquinone) levels are decreased in statin use;<sup>[50]</sup> Q10 supplements are sometimes used to treat statin-associated myopathy.<sup>[51]</sup>

Liver enzymes derangement is seen in statin therapy which includes increased hepatic transaminases enzyme. Also an increase in creatine phosphokinase, up to 10 times higher than normal, has been reported.<sup>[52]</sup> Cerivastatin was withdrawn from the market in 2001 due to cases of rhabdomyolysis. Combining any statin with a fibrate or niacin, another category of lipid-lowering drugs, increases the risks for rhabdomyolysis to almost 6.0 per 10,000 person-years.<sup>[53]</sup> Consumption of grapefruit or bitter oranges inhibits the metabolism of statins.

Cruz, *et al.*, attempted to describe the oral adverse effects of statins which are often underestimated. They identified dry mouth, itching or paresthesia in tongue, lips or throat, bitterness and cough as major complaints and all of them subsided with interruption of statin therapy. Insomnia was often associated with above mentioned symptoms.<sup>[54]</sup>

A study by Saxlin, *et al.*, has also stated that in patients with no plaque or gingival bleeding statin medication has shown an increased likelihood of deepened periodontal pockets which could be attributed to disruption of immune homeostasis in periodontal tissues as a result of statin therapy predisposing the periodontal tissues to a breakdown.<sup>[47]</sup>

The frequency and severity of adverse effect due to statin treatment is related to their own potency. Atorvastatin being the most potent (per mg of active principle), is maximally associated with the adverse effects whereas Fluvastatin exhibits the lowest efficacy, which correlates with less adverse effect. Simvastatin, pravastatin and lovastatin pose intermediate risk of adverse effect.

## CONCLUSION

Thus multiple effects of statins have been demonstrated in various experiments. The ultimate effect of statins on host is a combination of all these effects. Immunomodulation, anti-inflammatory effects and antioxidants effects are thought to contribute most towards its beneficial effects. Its effect on promotion of osteogenesis makes it a potential blockbuster in periodontics. Other systemic diseases where role of statins are being studied are Alzheimers,

multiple sclerosis, vitiligo, osteoporosis, pulmonary hypertension and many more.

There is a great potential to explore the pleiotropic effects of statins. Various animal and few human studies have demonstrated the effects of statins and the underlying mechanisms thereof. Further controlled clinical trials are needed to establish these facts. Further studies with varying doses of statins in varying disease conditions can help us maximize the beneficial effects. Also studies regarding its long term safety will be needed when statins are prescribed more commonly and for longer duration of actions. Continuing research to first establish and secondly to understand the mechanism of these effects would help not only to effectively use statins for various disorders but also, it will provide an insight to the role of statins as a potential host modulating agent for treatment of periodontal diseases.

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