



Published in final edited form as:

J Clin Immunol. 2012 August ; 32(4): 698–708. doi:10.1007/s10875-012-9664-5.

Additional diverse findings expand the clinical presentation of DOCK8 deficiency

Ozden Sanal^a, Huie Jing^b, Tuba Ozgur^a, Deniz Ayvaz^a, Dara M. Strauss-Albee^b, Sibel Ersoy-Evans^c, Ilhan Tezcan^a, Gulden Turkkan^a, Helen F. Matthews^d, Goknur Haliloglu^e, Aysel Yuce^f, Bilgehan Yalcin^g, Ozay Gokoz^h, Kader K. Oguzⁱ, and Helen C. Su^b

^aDepartment of Pediatric Immunology, Hacettepe University Faculty of Medicine, Ankara 06100, Turkey

^bLaboratory of Host Defenses, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, MD 20892

^cDepartment of Dermatology, Hacettepe University Faculty of Medicine, Ankara 06100, Turkey

^dLaboratory of Immunology, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, MD 20892

^eDepartment of Pediatric Neurology, Hacettepe University Faculty of Medicine, Ankara 06100, Turkey

^fDepartment of Pediatric Gastroenterology, Hacettepe University Faculty of Medicine, Ankara 06100, Turkey

^gDepartment of Pediatric Oncology, Hacettepe University Faculty of Medicine, Ankara 06100, Turkey

^hDepartment of Pathology, Hacettepe University Faculty of Medicine, Ankara 06100, Turkey

ⁱDepartment of Radiology, Hacettepe University Faculty of Medicine, Ankara 06100, Turkey

Abstract

We describe seven Turkish children with DOCK8 deficiency who have not been previously reported. Three patients presented with typical features of recurrent or severe cutaneous viral infections, atopic dermatitis, and recurrent respiratory or gastrointestinal tract infections. However, four patients presented with other features. Patient 1-1 featured sclerosing cholangitis and colitis; patient 2-1, granulomatous soft tissue lesion and central nervous system involvement, with primary central nervous system lymphoma found on follow-up; patient 3-1, a fatal metastatic leiomyosarcoma; and patient 4-2 showed no other symptoms initially besides atopic dermatitis. Similar to other previously reported Turkish patients, but in contrast to patients of non-Turkish ethnicity, the patients' lymphopenia was primarily restricted to CD4⁺ T cells. Patients had homozygous mutations in *DOCK8* that altered splicing, introduced premature terminations, destabilized protein, or involved large deletions within the gene. Genotyping of remaining family members showed that DOCK8 deficiency is a fully penetrant, autosomal recessive disease. In our patients, bone marrow transplantation resulted in rapid improvement followed by disappearance of viral skin lesions, including lesions resembling epidermodysplasia verruciformis, atopic dermatitis, and recurrent infections. Particularly for patients who feature unusual clinical

Address correspondence to Ozden Sanal, MD, Department of Pediatric Immunology, Hacettepe University Faculty of Medicine, Ankara 06100, Turkey: sanaloz@tr.net; or to Helen Su, MD, PhD, Building 10CRC, Room 5-3940, 10CRC Center Drive, MSC 1456, Bethesda, MD 20892, hsu@niaid.nih.gov.

manifestations, immunological testing, in conjunction with genetic testing, can prove invaluable in diagnosing DOCK8 deficiency and providing potentially curative treatment.

Keywords

DOCK8; combined immunodeficiency; epidermodysplasia verruciformis; sclerosing cholangitis; CNS lymphoma; leiomyosarcoma

INTRODUCTION

DOCK8 deficiency is an autosomal recessive combined immunodeficiency that includes the autosomal recessive form of hyperimmunoglobulinemia E syndrome (HIES). [1-4] Loss-of-function mutations in the *DOCK8* gene, which encodes for a poorly understood atypical guanine nucleotide exchange factor on human chromosome 9, underlie this disease. Like the autosomal dominant form of HIES caused by *STAT3* mutations, which is also known as Job's syndrome, DOCK8 deficiency exhibits eczema, recurrent pneumonias, elevated serum IgE, and sometimes boils and mucocutaneous candidiasis. However, unlike Job's syndrome, DOCK8 deficiency features intractable cutaneous viral infections with associated cancers, widespread infections (sinusitis, otitis media, sepsis), and lacks dental, skeletal, or connective tissue abnormalities. Neurologic abnormalities such as CNS vasculitis have been reported [2, 5]. Immunological abnormalities include lymphopenia, defective antibody function, and variable serum immunoglobulin levels.

Despite distinctive clinical features, patients may present with unusual findings or may only be mildly affected, leading to diagnostic uncertainty. The cases reported here illustrate instances where additional immunological and genetic testing were useful.

METHODS

Clinical and Laboratory Evaluations

Seven patients with high clinical suspicion for DOCK8 deficiency but negative HIV testing were studied, along with their immediate family members. Subjects provided written informed consent and were investigated under Institutional Review Board approved research protocols in accordance with the Helsinki Declaration. Lymphocyte immunophenotyping by flow cytometry was performed on whole blood followed by RBC lysis. Lymphocyte proliferation to mitogens (PHA, ConA) was measured by ³H-thymidine incorporation. Serum immunoglobulin isotypes were quantitated by nephelometry, except for IgE which was measured by radioallergosorbent test. Antibody titers were measured by ELISA. All tests were performed in the Clinical Pathology Laboratory and the Pediatric Immunology Laboratory at Hacettepe University Hospital.

Mutational Analyses

DOCK8 sequencing and array comparative genomic hybridization (CGH) were performed on genomic DNA from peripheral blood mononuclear cells (PBMC) of probands and all living immediate family members, as described [1]. Where indicated, total RNA was isolated from PBMC or proliferating T cells and reverse transcribed into cDNA using SuperScript III© First Strand Synthesis SuperMix kit (Invitrogen). PCR amplification of cDNA spanning exons 19 to 24 was performed using primers 5'-TGTACACACCCAGGACAACC-3' and 5'-ACCAGAAGCTCAAAGAAGAACC-3'. Amplified products were separated by gel electrophoresis, or used for cDNA sequencing [1]. Variants were named using GenBank Reference Sequences NC_000009.10 (DNA), NM_001193536.1 (mRNA; previously NM_203447.1), and NP_0011800465.1 (protein;

previously NP_982272.1) based upon NCBI Build 36.3, which correspond to the isoform that is predominantly expressed in primary T cells (Genbank NM991479.1, HM991480.1). The deleterious effects of missense mutations were predicted using the PolyPhen-2 HumDiv, PolyPhen-2 HumVar, and SIFT software packages. Mutations were not found in 50 healthy Turkish normal controls.

Overexpression Studies

Full-length wildtype DOCK8, cloned in the pCMV6-Entry vector that has C-terminal fusion Myc and Flag epitope tags, was purchased from Origene (cat# CW100311). A C1447R mutant version of this construct was generated by site-directed mutagenesis, with confirmation by Sanger sequencing. In brief, PCR amplification of full length DOCK8 plasmid with primers containing the point mutation was performed using AccuPrime™ Pfx SuperMix (Invitrogen), and PCR products were treated with Dpn1 (New England Biolabs) followed by transformation into XL10-Gold Ultracompetent Cells (Stratagene). Plasmid DNA was purified by PureLink HiPure Filter Plasmid Kit (Invitrogen) and 1.5 µg transfected into 0.8 million human embryonic kidney 293T cells using Lipofectamine 2000 (Invitrogen). Cells were lysed in 2% SDS buffer, and 8 µg of protein lysate was separated per lane. Immunoblotting for DOCK8 or β-actin was otherwise performed as described [1]. DOCK8 expression was also confirmed by immunoblotting using the M2 monoclonal antibody specific to the Flag tag (Sigma-Aldrich).

Literature Review and Analyses

We analyzed data from DOCK8-deficient patients in this report and in previous publications^[1, 2, 6–11]. We only included patients with DOCK8 mutations defined on both alleles (ARH001, 2, 3, 4, 5, 8, 9, 10.2, 10.4, 10.8, 10.9, 11.3, 12, 13.3, 13.4, 14, 15, 16.6, and 16.7) [2]. For estimation of disease penetrance, we analyzed data from all living family members of our additional patients, as well as data from a previous publication for the families of ARH008, 10, 11, and 12 [2]. The latter dataset included all living family members except for the parents of ARH010.2/10.3/10.4, ARH010.3, and ARH010.6. The younger sister of ARH011.3, ARH011.4, and ARH011.5 was genotyped as a heterozygous carrier for inclusion in the present analysis (H. Jing and H. C. Su, unpublished). For contingency table analyses of lymphopenia in Turkish versus non-Turkish populations, we analyzed data from DOCK8-deficient patients in this report and in previous publications where ethnicity was reported [1, 2, 7, 9–11]. We classified a patient as Turkish if this ethnicity was indicated, or as non-Turkish if a different ethnicity was indicated, and then used the two-tailed Fisher's exact test as part of the Prism software package.

RESULTS

Patient Presentations

Our seven additional Turkish patients are from four consanguineous families, whose pedigrees are shown in Figure 1. Their demographics and clinical features are described in Table I. In general, patients had early onset of disease, which first manifested as atopic dermatitis between 3 months to 2 years of age. For very young infants, such as patient 4-2, atopic dermatitis was the only initial clinical manifestation. Atopic dermatitis was soon followed by recurrent otitis externa, otitis media, respiratory tract infections, gastrointestinal tract infections, and food allergies. Patients developed recurrent herpes simplex virus (HSV) infections including eczema herpeticum, disseminated molluscum contagiosum, or severe varicella-zoster virus infection. However, in four patients, additional skin lesions resembling epidermodysplasia verruciformis (EV) were diagnosed. Those patients presented with pinkish hypopigmented flat or atrophic papules (patient 1-1) (Figure 2), or tan to pink-colored macules and hyperkeratotic papules (patients 2-1, 3-1, and 3-2), over the trunk and

extremities. Histopathology of representative lesions showed hyperkeratosis and acanthosis consistent with EV (Figure 2). This particular manifestation of human papillomavirus infections has not been previously described for DOCK8 deficiency.

Some cases initially drew clinical attention away from the immune system. Patients 1-1 and 3-1 had prominent gastrointestinal symptoms. Patient 1-1 had poor growth, chronic diarrhea, and active colitis as revealed by endoscopy. Magnetic resonance (MR) cholangiopancreatography showed portal vein dilatation, sclerosing cholangitis, and thickened pancreatic parenchyma, which may be caused by persistent giardiasis. Patient 4-1 also developed sclerosing cholangitis in the setting of persistent cryptosporidium and giardia infections. Patient 3-1, who was previously reported without a molecular diagnosis, was 6 years old when admitted with chronic abdominal pain, bloody stools, weight loss, and hepatomegaly [12]. Imaging revealed multiple nodules within liver lobes, gallbladder wall, left suprarenal retroperitoneum, right lower quadrant intestines, and lungs, and she was diagnosed with metastatic multifocal leiomyosarcomatosis. She died 13 months later after limited surgical resection plus treatment with cisplatin, adriamycin, vincristine, and cyclophosphamide.

Another unusual case was illustrated by Patient 2-1 who had prominent neurological features. Patient 2-1 presented at age 9 with a 3x4 cm granulomatous lesion on her buccal mucosa. Histopathology of buccal mucosa showed chronic granulomatous inflammation and ulceration. Magnetic resonance imaging (MRI) demonstrated a T2-hyperintense lesion in the left buccal region and retromolar trigone. The lesion had an ill-defined margin and showed enhancement on post-Gadolinium (Gd) series (Figure 3A, B). The oral lesion improved after debridement and treatment with corticosteroids. However, during follow-up she developed dizziness and unsteadiness while walking. She displayed nystagmus, spasticity, increased deep tendon and pathological reflexes bilaterally, and a wide-based spastic-ataxic gait. Cranial MRI revealed bilateral pulvinar T2 hyperintensities on T2-weighted imaging and cerebellar and cerebral volume loss (Figure 3C). BK and JC viral studies were negative. Over the subsequent nine months she partially improved without treatment, but repeat cranial MRI disclosed a lesion that showed rim enhancement and restricted diffusion on the left temporo-occipital parenchyma (Figure 3D, 3E). Imaging studies were suggestive of an abscess and she was treated with wide-spectrum antibiotics and antifungal agents. Despite these treatments, her course was progressive. Histopathologic examination of excisional biopsy via craniotomy performed 1.8 years after the onset of neurological symptoms demonstrated an EBNA⁺ diffuse large B cell lymphoma. The patient received systemic chemotherapy with high-dose methotrexate, cyclophosphamide, vincristine, prednisolone, intrathecal chemotherapy, and cranial irradiation. She died four months after the diagnosis of CNS lymphoma due to progressive disease.

Patients 1-1, 3-2, and 4-1 were treated with intravenous immunoglobulin (IVIG), which markedly decreased their frequency of infections and improved their skin lesions, including those associated with HSV. Additionally, patients 4-2 and 1-1, who had matched related donors available, underwent bone marrow transplantation 10 and 2 months ago, respectively. Patient 4-2 is now healthy with resolution of atopic dermatitis, viral and other recurrent infections. In patient 1-1, skin lesions started to improve during pre-conditioning chemotherapy and continued to clear after bone marrow transplantation. For both patient 4-2 and 1-1, details of their clinical progress after transplantation will be reported in a separate manuscript. Patient 2-2 underwent bone marrow transplantation at another medical center several weeks ago. Patients 3-2 and 4-1 lack matched donors and have not yet been transplanted.

Immunological Evaluations

Standard immunological evaluations were performed on all patients with the exception of patient 2-2, for whom we were unable to obtain peripheral blood. Values are shown in Figure 4 and summarized in Table II. Determinations for Table II were made based upon the majority of readings for any given patient, or if only two readings were available, their average. All tested patients had low numbers of CD4⁺ T cells and high serum IgE (peaks ranged from 412 to 20,000 IU/mL). Nearly all had decreased serum IgM, variably impaired specific antibodies, and mild to moderate eosinophilia. Several patients had T cell lymphopenia. However, lymphocyte proliferation to T cell mitogens was within normal limits, except for patient 3-1 who already had multifocal leiomyosarcomatosis when tested. Serum IgG and IgA were elevated in several patients. NK cell numbers were decreased in one patient, but neither CD8⁺ T cells nor B cells were consistently decreased in any patient, and in a few patients B cells were instead increased. Retrospective statistical analysis of all patients in the literature in which ethnicity was reported showed that lymphopenia affecting CD4⁺ T cells, CD8⁺ T cells, and B cells occurred less frequently in Turkish as compared to non-Turkish patients (Table III). Overall, immunological findings were similar to previous observations in other DOCK8-deficient patients, except that in the Turkish cohort lymphopenia occurred less frequently and was primarily restricted to CD4⁺ T cells, and T lymphocyte proliferation to mitogens was normal.

Genetic Evaluations

We found novel homozygous *DOCK8* mutations and large deletions in the genomic DNA of our seven additional patients (Table II). These were predicted to alter splicing (family 1), introduce premature termination with probable nonsense-mediated decay (family 2), alter protein structure (family 3), or lead to loss of protein expression (family 4). For families 1 and 3, in which effects were not obvious, we performed additional experimental validation. We found that in family 1, the mutation in the consensus 5' splice donor site of intron 20 yielded an aberrantly spliced, smaller product (Figure 5A). Sequencing of cDNA showed that this product corresponded to an in-frame deletion of 55 amino acids just after the conserved DHR1 domain (Table II). In family 3, the cysteine to arginine missense mutation in exon 35 between the conserved DHR1 and DHR2 domains was predicted to exert protein destabilizing effects. When equivalent amounts of plasmid encoding either mutant or wildtype DOCK8 were overexpressed in 293T cells (which do not endogenously express DOCK8), the level of mutant DOCK8 protein was markedly decreased (Figure 5B and data not shown). Thus, in all cases, the mutations resulted in loss of normal DOCK8 expression with predicted loss of function, as is characteristic of this disease.

We next studied the immediate families of the seven patients (Figure 1). All living family members were healthy, with no history of serious viral infections, bacterial infections, skin infections, atopic dermatitis, asthma, allergies, or malignancy. The living family members were either homozygous for the normal DOCK8 gene or heterozygous carriers, and these genotyping results are shown in the pedigrees below each corresponding family member (Figure 1). A sibling of patient 1-1 died at 40 days of age with vomiting and abdominal distension. Because of a lack of additional clinical features to support a diagnosis of DOCK8 deficiency, we considered this death to be unrelated. By contrast, a sibling of patients 2-1 and 2-2 who had similar skin lesions died at age 10 years because of an intra-abdominal tumor, whom we judged as likely to have been affected with DOCK8 deficiency. No DNA samples were available from these two deceased individuals for genotyping.

It has been assumed that the autosomal recessive inheritance and lack of discernable symptoms in tested carriers are consistent with a fully penetrant model of disease. However, prior studies have been limited to four fully genotyped families, raising the possibility of

ascertainment bias [2]. When data from those families are combined with data from our four additional families, we observed that all individuals either homozygous for the normal *DOCK8* allele (n = 6) or heterozygous for the mutant *DOCK8* allele (n = 26), who ranged from ages 4 to 49 years old, were healthy. Furthermore, of 15 individuals who were homozygous for the mutant *DOCK8* allele, all 15 developed disease, supporting a disease penetrance of ~100%. Nevertheless, as illustrated by our patients, the range of disease manifestations varied, depending upon patient age.

DISCUSSION

DOCK8 mutations are responsible for a combined immunodeficiency that was formerly characterized as autosomal recessive HIES. A total of 52 patients reported in the literature, including our seven additional patients, had homozygous or compound heterozygous *DOCK8* mutations [1, 2, 6–11]. Their most common findings are summarized in Table IV. Patients typically present with intractable cutaneous viral infections, atopic dermatitis often accompanied by otitis externa or *Staphylococcus aureus* skin infections, recurrent respiratory tract infections, mucocutaneous candidiasis, allergic disease, as well as high serum IgE, low serum IgM, eosinophilia, and lymphopenia [1, 2, 5]. Gastrointestinal infections, malignancies, and neurological abnormalities have been sometimes reported in *DOCK8* deficiency. However, the EV-like skin lesions, liver and pancreatic abnormalities, leiomyosarcoma, and CNS lymphoma in our additional patients have not been previously described in this disease. Interestingly, the increasing constellation of features seemed to reflect increasing patient age, consistent with the progressive nature of this disease (Table I).

The differential diagnosis for *DOCK8* deficiency includes autosomal dominant HIES caused by *STAT3* mutations, as well as other conditions that can be associated with high serum IgE, with or without lymphopenia, such as Comèl-Netherton syndrome, Wiskott-Aldrich syndrome (WAS), IPEX (Immunodysregulation, Polyendocrinopathy, Enteropathy, X-linked) syndrome, acquired immunodeficiency syndrome (AIDS), and severe combined immunodeficiency (SCID). Comèl-Netherton syndrome, due to autosomal recessive *SPINK5* mutations, features a distinctive “bamboo” appearance of the hair shaft (trichorrhexis invaginata), as well as eczema, allergic disease, recurrent bacterial infections of the skin, respiratory and gastrointestinal tracts, particularly with *Staphylococcus aureus*, high IgE, and impaired functional antibodies [13]. WAS, due to mutations in the X-linked gene *WAS*, can present in boys with eczema, bloody diarrhea associated with thrombocytopenia, recurrent sinopulmonary infections, skin viral infections including HSV and molluscum contagiosum, autoimmunity, hematological malignancy, and immune abnormalities including high IgE, low IgM, variably impaired functional antibodies, and progressive T cell lymphopenia with decreased T cell proliferation [14]. IPEX, due to mutations in the X-linked gene *FOXP3*, also presents in boys with eczema, allergic disease, *Staphylococcus aureus* sepsis, high IgE, and eosinophilia [15]. AIDS can present with pruritic dermatitis, *Staphylococcus aureus* skin infections, mucocutaneous candidiasis, eosinophilia, high IgE, and CD4⁺ T cell and sometimes B cell lymphopenia [16]. “Leaky” SCID causing Omenn’s syndrome, usually due to autosomal recessive *RAG1/RAG2* mutations, presents with erythroderma, recurrent infections, autoimmunity, eosinophilia, high IgE, low IgM, and variable lymphopenia [17]. Opportunistic infections such as CMV, *Pneumocystis jirovecii*, or EV-like skin lesions are also remarkable in WAS, AIDS, or SCID [18] and can occur in *DOCK8* deficiency.

Other considerations in the differential diagnosis include EV and WHIM (Warts, Hypogammaglobulinemia, Infections, Myelokathexis) syndrome, which do not present with high serum IgE. WHIM syndrome, due to autosomal dominant *CXCR4* mutations, can also present with lymphopenia involving B cells and sometimes T cells [19]. EV, an HPV

genodermatosis, due to autosomal recessive *EVER1/EVER2* mutations, generally does not feature cell-mediated immune defects [20]. It is possible that other case reports of EV with high IgE, low IgM, and lymphopenia, but lacking confirmatory mutational analyses, were instead cases of DOCK8 deficiency [21–23].

Given that the clinical phenotype of DOCK8 deficiency overlaps with many other disorders and also shows varied features among patients even within the same family, additional immunological testing can be a useful adjunct for diagnosis. However, our results, expanding upon previously published work, also demonstrates heterogeneity of the cellular phenotype among patients. For example, although CD4⁺ T cell lymphopenia appeared nearly universal in all patients, involvement of other lymphocyte subsets occurred less frequently in Turkish patients. Thus, in cases where immunological testing remains equivocal, genetic sequencing is essential to establish a diagnosis. Although most patients have large deletions, or smaller point mutations or indels that cause nonsense-mediated decay of mRNA or premature terminations, mutations that result in abnormal or destabilized protein with loss of normal DOCK8 expression also can occur. Identification of additional cases and research for disease-modifying genes or environmental factors will aid in future management of this clinically heterogeneous disease.

CONCLUSIONS

Our seven additional patients extend the clinical phenotype and help establish that DOCK8 deficiency is highly penetrant with variable multi-organ disease manifestations, and that disease complications, which include CNS involvement and development of malignancy, are associated with increasing patient age. Immunologically, our patients provide additional evidence of gene modifier or environmental factors that modulate the effects of lymphopenia on different cellular subsets. Furthermore, our additional patients establish for the first time that disease can be associated with *DOCK8* mutations that alter protein structure or stability. Together, this knowledge will improve the diagnosis of patients and aid in their management. Once patients are diagnosed, aggressive antimicrobial treatment, treatment with IVIG, and cancer surveillance are indicated. Furthermore, allogeneic hematopoietic cell transplantation, which was reported as curative in four patients, also resulted in rapid healing and clearance of viral and other infections in our patients, and should be considered as early as possible given the progressive course of the disease [6–9].

Acknowledgments

We thank the physicians who referred their patients to us and the patients and their relatives who participated in this study. We also thank Baran Erman for isolating RNA and performing the RT-PCR reaction on PBMC from patient 1-1. This work was supported in part by the Intramural Research Program of the National Institutes of Health, the National Institute of Allergy and Infectious Diseases, through the Lymphocyte Molecular Genetics Unit (LMGU) program. Sequencing was performed by the Genomics Unit of the Rocky Mountain Laboratories Research Technologies Section of the National Institute of Allergy and Infectious Diseases.

References

1. Zhang Q, Davis JC, Lamborn IT, Freeman AF, Jing H, Favreau AJ, Matthews HF, Davis J, Turner ML, Uzel G, Holland SM, Su HC. Combined immunodeficiency associated with DOCK8 mutations. *N Engl J Med*. 2009; 361:2046–2055. [PubMed: 19776401]
2. Engelhardt KR, McGhee S, Winkler S, Sassi A, Woellner C, Lopez-Herrera G, Chen A, Kim HS, Lloret MG, Schulze I, Ehl S, Thiel J, Pfeifer D, Veelken H, Niehues T, Siepermann K, Weinspach S, Reisli I, Keles S, Genel F, Kutuculer N, Camcioglu Y, Somer A, Karakoc-Aydiner E, Barlan I, Gennery A, Metin A, Degerliyurt A, Pietrogrande MC, Yeganeh M, Baz Z, Al-Tamemi S, Klein C, Puck JM, Holland SM, McCabe ER, Grimbacher B, Chatila TA. Large deletions and point

- mutations involving the dedicator of cytokinesis 8 (DOCK8) in the autosomal-recessive form of hyper-IgE syndrome. *J Allergy Clin Immunol.* 2009; 124:1289–1302.e1284. [PubMed: 20004785]
3. Zhang Q, Su HC. Hyperimmunoglobulin E syndromes in pediatrics. *Curr Opin Pediatr.* 2011; 23:653–658. [PubMed: 21970826]
 4. Su HC, Jing H, Zhang Q. DOCK8 deficiency. *Ann N Y Acad Sci.* 2011; 1246:26–33. [PubMed: 22236427]
 5. Renner ED, Puck JM, Holland SM, Schmitt M, Weiss M, Frosch M, Bergmann M, Davis J, Belohradsky BH, Grimbacher B. Autosomal recessive hyperimmunoglobulin E syndrome: a distinct disease entity. *J Pediatr.* 2004; 144:93–99. [PubMed: 14722525]
 6. Gatz SA, Benninghoff U, Schutz C, Schulz A, Honig M, Pannicke U, Holzmann KH, Schwarz K, Friedrich W. Curative treatment of autosomal-recessive hyper-IgE syndrome by hematopoietic cell transplantation. *Bone Marrow Transplant.* 2010
 7. Bittner TC, Pannicke U, Renner ED, Notheis G, Hoffmann F, Belohradsky BH, Wintergerst U, Hauser M, Klein B, Schwarz K, Schmid I, Albert MH. Successful long-term correction of autosomal recessive hyper-IgE syndrome due to DOCK8 deficiency by hematopoietic stem cell transplantation. *Klin Padiatr.* 2010; 222:351–355. [PubMed: 21058221]
 8. McDonald DR, Massaad MJ, Johnston A, Keles S, Chatila T, Geha RS, Pai SY. Successful engraftment of donor marrow after allogeneic hematopoietic cell transplantation in autosomal-recessive hyper-IgE syndrome caused by dedicator of cytokinesis 8 deficiency. *J Allergy Clin Immunol.* 2010
 9. Barlogis V, Galambrun C, Chambost H, Lamoureux-Toth S, Petit P, Stephan JL, Michel G, Fischer A, Picard C. Successful allogeneic hematopoietic stem cell transplantation for DOCK8 deficiency. *J Allergy Clin Immunol.* 2011
 10. Chu EY, Freeman AF, Jing H, Cowen EW, Davis J, Su HC, Holland SM, Turner ML. Cutaneous Manifestations of DOCK8 Deficiency Syndrome. *Arch Dermatol.* 2011
 11. Dasouki M, Okonkwo KC, Ray A, Folmsbeel CK, Gozales D, Keles S, Puck JM, Chatila T. Deficient T Cell Receptor Excision Circles (TRECs) in autosomal recessive hyper IgE syndrome caused by DOCK8 mutation: Implications for pathogenesis and potential detection by newborn screening. *Clin Immunol.* 2011
 12. Boybeyi O, Akcoren Z, Oguz B, Akyuz C, Sanal O, Ergin S, Ersoy-Evans S, Tanyel FC. Multifocal leiomyosarcomatosis in a 6-year-old child with epidermodysplasia verruciformis and immune defect. *J Pediatr Surg.* 2009; 44:e5–8. [PubMed: 19573649]
 13. Renner ED, Hartl D, Rylaarsdam S, Young ML, Monaco-Shawver L, Kleiner G, Markert ML, Stiehm ER, Belohradsky BH, Upton MP, Torgerson TR, Orange JS, Ochs HD. Comel-Netherton syndrome defined as primary immunodeficiency. *J Allergy Clin Immunol.* 2009; 124:536–543. [PubMed: 19683336]
 14. Thrasher AJ. New insights into the biology of Wiskott-Aldrich syndrome (WAS). *Hematology Am Soc Hematol Educ Program.* 2009:132–138. [PubMed: 20008191]
 15. Halabi-Tawil M, Ruummele FM, Freitag S, Rieux-Laucat F, Neven B, Brousse N, De Prost Y, Fischer A, Goulet O, Bodemer C. Cutaneous manifestations of immune dysregulation, polyendocrinopathy, enteropathy, X-linked (IPEX) syndrome. *Br J Dermatol.* 2009; 160:645–651. [PubMed: 18795917]
 16. Paganelli R, Scala E, Mezzaroma I, Pinter E, D'Offizi G, Fanales-Belasio E, Rosso RM, Ansotegui IJ, Pandolfi F, Aiuti F. Immunologic aspects of hyperimmunoglobulinemia E-like syndrome in patients with AIDS. *J Allergy Clin Immunol.* 1995; 95:995–1003. [PubMed: 7751521]
 17. Villa A, Notarangelo LD, Roifman CM. Omenn syndrome: inflammation in leaky severe combined immunodeficiency. *J Allergy Clin Immunol.* 2008; 122:1082–1086. [PubMed: 18992930]
 18. Rogers HD, Macgregor JL, Nord KM, Tying S, Rady P, Engler DE, Grossman ME. Acquired epidermodysplasia verruciformis. *J Am Acad Dermatol.* 2009; 60:315–320. [PubMed: 19150275]
 19. Kawai T, Malech HL. WHIM syndrome: congenital immune deficiency disease. *Curr Opin Hematol.* 2009; 16:20–26. [PubMed: 19057201]
 20. Ramoz N, Rueda LA, Bouadjar B, Montoya LS, Orth G, Favre M. Mutations in two adjacent novel genes are associated with epidermodysplasia verruciformis. *Nat Genet.* 2002; 32:579–581. [PubMed: 12426567]

21. Guilhou JJ, Malbos S, Barneon S, Habib A, Baldet P, Meynadier J. Epidermodysplasia verruciformis (2 cases). Immunological study (author's transl). *Ann Dermatol Venereol*. 1980; 107:611–619. [PubMed: 6254427]
22. Azzimonti B, Mondini M, De Andrea M, Gioia D, Dianzani U, Mesturini R, Leigheb G, Tiberio R, Landolfo S, Gariglio M. CD8+ T-cell lymphocytopenia and lack of EVER mutations in a patient with clinically and virologically typical epidermodysplasia verruciformis. *Arch Dermatol*. 2005; 141:1323–1325. [PubMed: 16230580]
23. Gul U, Soylu S, Yavuzer R. Epidermodysplasia verruciformis associated with isolated IgM deficiency. *Indian J Dermatol Venereol Leprol*. 2007; 73:420–422. [PubMed: 18032865]

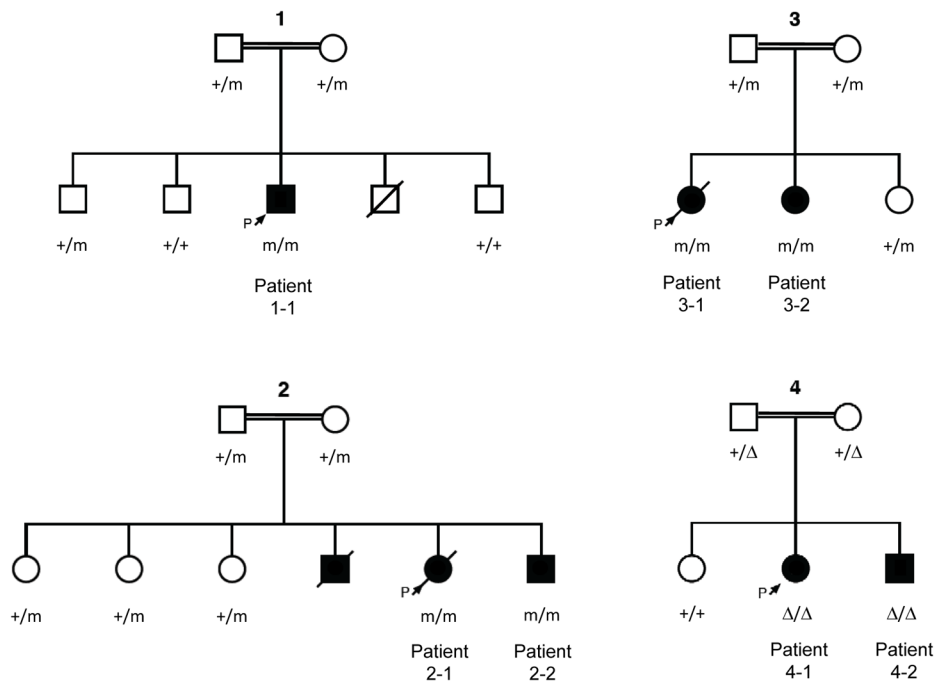


Figure 1. Pedigrees of the families studied. Genotypes are indicated below each individual, where “+” designates normal *DOCK8* allele, and “m” or “Δ” designates mutated or deleted *DOCK8* allele, respectively. Numbers about the pedigrees correspond to the family identifiers, whereas patient identifiers are indicated below the symbols. A closed symbol designates an affected individual, whereas an open symbol designates an unaffected individual. Squares designate males; circles, females; arrows, probands; and slashes, deceased individuals.

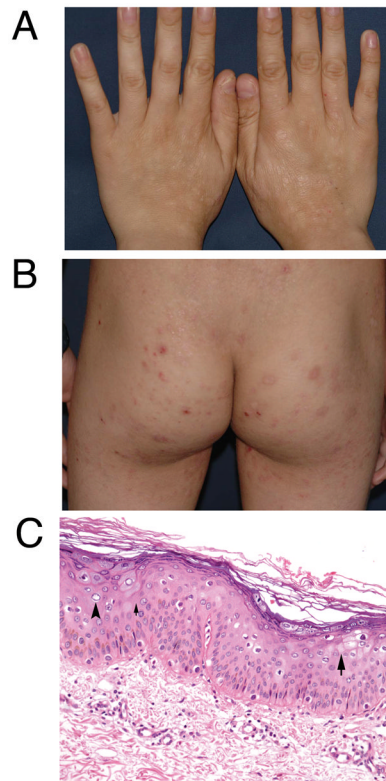


Figure 2. Skin lesions in patient 1-1. Human papillomavirus-associated pinkish, hypopigmented flat papules on dorsal hands (A) and buttocks (B). C) Histopathology shows hyperkeratosis and acanthosis, with vacuolated cells in the stratum malpighii (arrow), keratinocytes having abundant slightly basophilic cytoplasm (small arrow), and occasional large, round, empty-appearing nuclei (arrowhead). Patients 2-1, 2-2, 3-1, and 3-2 had similar lesions.

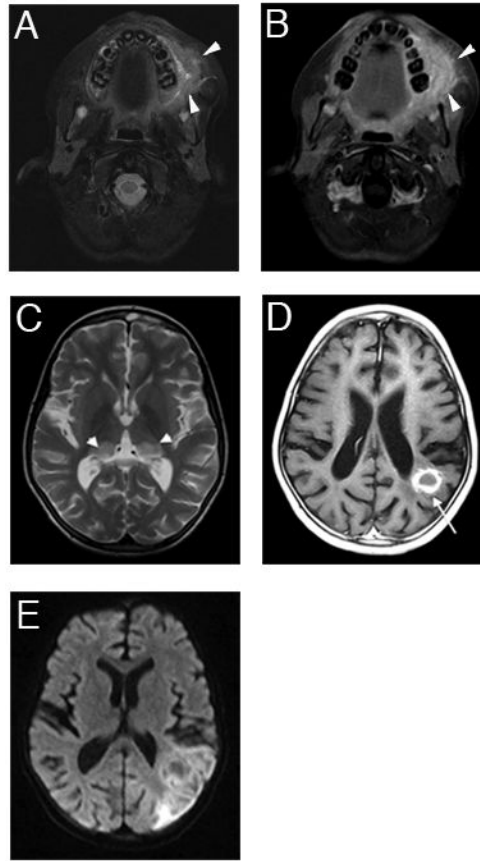


Figure 3.

Cranial MRI in patient 2-1. A) Transverse T2 weighted (W) fat-suppressed (left) and B) contrast enhanced (ce) fat-suppressed T1W (right) spin-echo (SE) images, at initial presentation. A T2-hyperintense and Gadolinium-enhancing lesion with irregular border in the left buccal space and retromolar trigone is seen (arrowheads in A, B). C). Transverse T2W turbo SE images, at five months, show bilateral pulvinar T2 hyperintensities (arrowheads) and cerebral volume loss. D) Axial Gadolinium-enhanced T1W SE shows a lesion with rim enhancement (long arrow). E) Axial diffusion-weighted imaging (DWI), at eighteen months, shows hyperintensity in the parenchyma and in the lesion, especially at the periphery.

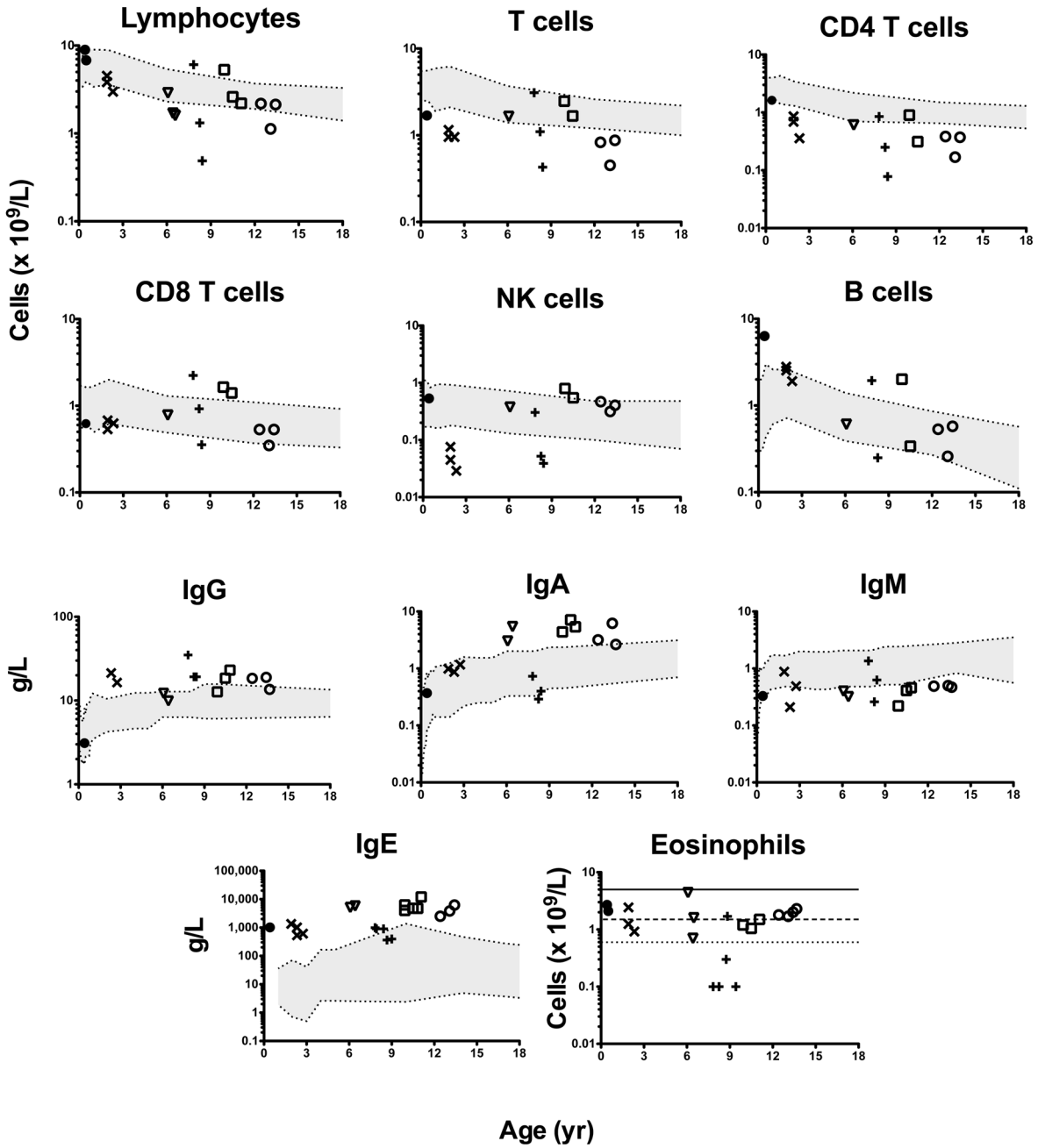


Figure 4. Peripheral blood enumerations of lymphocyte subsets, quantitative serum immunoglobulin isotypes, and eosinophils. Patient 1-1, open circle; patient 2-1, open square; patient 3-1, cross; patient 3-2, X; patient 4-1, open inverted triangle; patient 4-2, filled-in circle. Shaded in grey are the ranges for normal age-matched controls. For lymphocytes and their subsets, published 10th to 90th percentiles are shown in grey. For immunoglobulin isotypes, the 95 % confidence interval is shown. For eosinophils, arbitrarily thresholds for mild, moderate, and severe eosinophilia are as indicated.

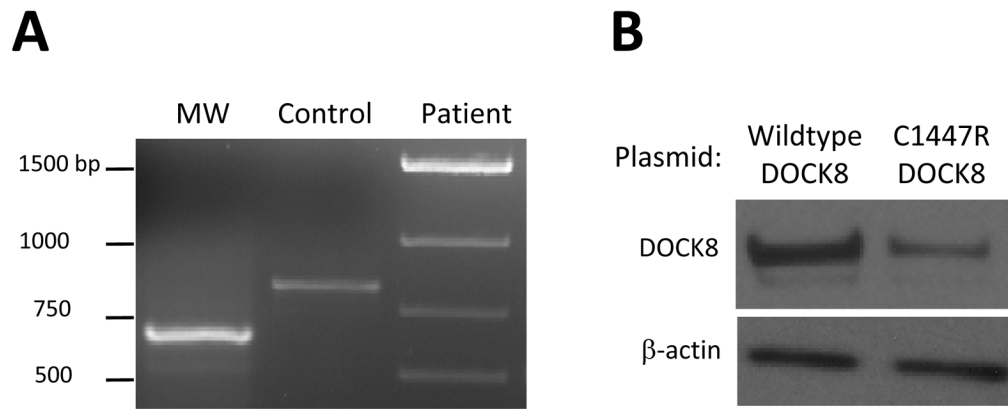


Figure 5.

DOCK8 expression data for mutations in patients from families 1 and 3. A) PCR amplification of exons 19 to 24 of DOCK8 from cDNA shows a smaller product from patient 1-1, which corresponds to an aberrantly spliced mRNA. B) Immunoblotting shows destabilized DOCK8 protein in 293T cells that have been transiently transfected with a plasmid encoding the full-length mutant (C1447R), as compared to wildtype, DOCK8. Patients 3-1 and 3-2 only express this mutant form of DOCK8.

Table I

Summary of the seven additional DOCK8-deficient patients studied

Demographics							
Patient	1-1	2-1	2-2	3-1	3-2	4-1	4-2
Age (yr) [†]	15	12 [†]	8	8 [†]	5	9	2.5
Sex	Male	Female	Male	Female	Female	Female	Male
Age of onset of symptoms (yr)	2	0.5	unknown	0.5	1.75	0.25	0.58
Clinical Features							
Skin							
Atopic dermatitis	(+)	(+)	(+)	(+)	(+)	(+)	(+)
Lesions resembling epidermolytic hyperkeratosis	(+)	(+)	(+)	(+)	(-)	(-)	(-)
Recurrent Herpes Simplex Virus infections	(-)	(+)	(+)	(-)	(+)	(+)	(+)
Severe primary varicella	(-)	(-)	(-)	(-)	(+)	(-)	(-)
Molluscum contagiosum	(-)	(-)	(-)	(-)	(-)	(+)	(+)
Other					Seborrheic dermatitis	Eczema herpeticum	
Respiratory tract							
Recurrent respiratory tract infections	(+)	(+)	(+)	(+)	(+)	(+)	(+)
Otitis media	(-)	(+)	(+)	(-)	(+)	(-)	(-)
Mastoiditis	(-)	(+)	(+)	(-)	(-)	(-)	(-)
Sinusitis	(-)	(+)	(+)	(-)	(-)	(+)	(-)
Pneumonia/ bronchitis	(+)	(+)	(-)	(+)	(+)	(+)	(-)
Bronchiectasis	(-)	(+)	(-)	(-)	(-)	(-)	(-)
Otitis externa	(+)	(-)	unknown	(-)	(+)	(-)	(-)
Gastrointestinal tract							
Poor growth	(+)	(-)	(-)	(-)	(-)	(+)	(-)
Chronic diarrhea	(+)	(-)	(-)	(+)	(-)	(+)	(-)
Giardiasis	(+)	(-)	unknown	(-)	(-)	(+)	(-)

Demographics							
Cryptosporidium infections	(-)	(-)	(-)	(-)	(-)	(-)	(-)
Other	Colitis, sclerosing cholangitis, portal vein dilation	Buccal granulomatous inflammation and ulceration				Oral candidiasis, Sclerosing cholangitis	
Neurological	(-)	Presented with nystagmus, spasticity, ataxia due to CNS lymphoma	(-)	(-)	(-)	(-)	(-)
Allergies or specific IgE	(-)	(-)	Egg	ND	Egg, lentil, milk	Egg, hazelnut, milk, peanut	Egg, hazelnut, milk
Malignancy		Diffuse large B cell CNS lymphoma		Metastatic multifocal leiomyosarcoma			
Other	Recently underwent matched related donor HCT		Recently underwent matched related donor HCT at another center				Underwent matched related donor HCT

[†] Age at death.

ND, not determined

Table II

Laboratory evaluation of additional DOCK8-deficient patients

Patient	1-1	2-1	2-2	3-1	3-2	4-1	4-2
Lymphocyte numbers	Normal	Normal	ND	↓	Normal	↓	Normal
T cells	↓	Normal	ND	↓	↓	Normal	↓
CD4 T cells	↓	↓	ND	↓	↓	↓	↓
CD8 T cells	Normal	↑	ND	Normal	Normal	Normal	Normal
NK cells	Normal	↑	ND	↓	↓	Normal	Normal
B cells	Normal	Normal	ND	Normal	↑	Normal	↑
Lymphocyte proliferation to T cell mitogens (PHA, ConA)	Normal	Normal	ND	↓	Normal	Normal	Normal
Eosinophil numbers	↑	↑	ND	Normal	↑	↑	↑
IgG	↑	↑	ND	↑	↑	Normal	Normal
IgA	↑	↑	ND	↓	Normal	↑	Normal
IgM	↓	↓	ND	Normal	↓	↓	↓
IgE	↑	↑	ND	↑	↑	↑	↑
Hepatitis B virus surface Ab	(-)	ND	ND	(+)	ND	(-)	(+)
Rubella IgG	(+)	ND	ND	ND	(-)	ND	(-)
Cytomegalovirus IgG	(+)	ND	ND	ND	(+)	ND	ND
Epstein-Barr Virus viral capsid antigen IgG	(-)	ND	ND	(+)	ND	ND	(+)
Hepatitis A Virus IgG	(+)	ND	ND	ND	ND	ND	(+)
Herpes Simplex Virus-1 IgG	(+)	(+)*	ND	ND	ND	(+)	(+)*
Varicella-Zoster Virus IgG	(-)	(+)	ND	ND	ND	ND	(-)
Genetic mutation in DOCK8	Homozygous c. 2401G>C → p.A746_M800del	Homozygous c. 4698C>G → p.Thr1566X plus c. 2389A>G → p.Val797Met	Homozygous c. 4698C>G → p.Thr1566X plus c. 2389A>G → p.Val797Met	Homozygous c. 4339T>C → p.Cys1447Arg	Homozygous c. 4339T>C → p.Cys1447Arg	Homozygous c. 4339T>C → p.Cys1447Arg	Chr9:g.(153,190_194,193)_(315,772_322,456) homozygous deletion spanning from exons 1 to 9

* Age at death.

ND, not determined.

* IgG to HSV-2 also detected

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

Table III

Lymphopenia in Turkish as compared to non-Turkish DOCK8-deficient patients

Lymphocyte subset	Turkish		Non-Turkish		P-value*		
	# Affected	# Unaffected	% Affected	# Affected		# Unaffected	% Affected
CD4 T cells	15	8	65	24	0	100	0.0016
CD8 T cells	5	17	23	16	8	67	0.0037
NK cells	8	14	36	12	10	55	NS
B cells	0	23	0	10	14	42	0.0006

Numbers of affected or unaffected DOCK8-deficient patients, who have lymphopenia of the indicated lymphocyte subsets.

* P-value calculated by Fisher's exact test.

NS, not statistically significant.

Table IV

Frequency of findings in DOCK8-deficient patients reported in the literature

Feature	Number of patients affected*	% of patients affected
Virus infections, 1 or more	46	88
HSV	28	54
MCV	19	37
HPV	19	37
VZV	13	25
Respiratory tract infections	49	94
Atopic dermatitis	46	88
Bacterial skin infections	34	65
Mucocutaneous candidiasis	28	54
Gastrointestinal tract infections	6	12
Cancers	9	17
Neurological abnormalities	11	21
Low serum IgM	38 [§]	75
High serum IgE	50 [§]	98
Eosinophilia	46 [¶]	92
CD4 T cell lymphopenia	42 [¶]	84
CD8 T cell lymphopenia	22 [‡]	45
NK cell lymphopenia	21 [‡]	45
B cell lymphopenia	9 [†]	18

* Total number of patients evaluated was 52 except where indicated.

[§] of 51 patients evaluated.

[¶] of 50 patients evaluated.

[‡] of 49 patients evaluated.

[†] of 47 patients evaluated.