



Published in final edited form as:

Curr Opin Gastroenterol. 2001 September ; 17(5): 434–440.

TGF β -mediated signaling and transcriptional regulation in pancreatic development and cancer

Volker Ellenrieder, MD, Martin E. Fernandez Zapico, MD, and Raul Urrutia, MD
Mayo Clinic, Rochester, Minnesota, USA

Abstract

Transforming growth factor- β (TGF β) plays a critical role in pancreatic development and cell proliferation. Binding of TGF β to its membrane receptor kinases activates the Smad signaling proteins, allowing them to translocate to the nucleus and participate in the transcriptional control of TGF β target genes. In addition, there is an increasing number of cellular mechanisms affecting the final response of a cell to TGF β . This includes crosstalk with other signaling pathways and the induction of TGF β early response genes, such as the TGF β -inducible early response gene (TIEG) family of transcription factors. Like the Smads, TIEGs behave as downstream effector proteins in TGF β -mediated pancreatic growth control. The discovery of the Smads and TIEGs has provided new insights into TGF β -regulated functions. Their significance in pancreatic development and cancer is discussed in this review.

The transforming growth factor- β (TGF β) family consists of multifunctional cytokines that regulate a broad spectrum of biologic functions including wound healing, cellular differentiation, and deposition of extracellular matrix proteins [1]. Among the most dramatic effects of TGF β , however, are those associated with cell differentiation during development and inhibition of cell proliferation, mediated largely by the induction of cell cycle arrest and apoptosis [2,3–5]. TGF β exerts its biologic function mainly by its effects on regulation of gene expression. Elucidating its signal transduction pathway has become a subject of intense investigation in recent years [6]. This has resulted in the identification of a family of membrane receptor protein kinases, TGF β receptor kinase I (T β RI) and TGF β receptor kinase II (T β RII), and the subsequent discovery of their intracellular mediators, the Smads [7,8]. Although Smad signaling appears to be involved in most actions of TGF β , recent evidence suggests the existence of additional TGF β downstream mediators. For example, TGF β has been shown to rapidly activate the mitogen-activated protein kinases (MAPK) extracellular regulated kinase (Erk), p38 and JNK, which regulate gene expression by activating specific downstream transcription factors [9,10]. In the same line of evidence, we have previously reported the identification of the TGF β -inducible early response gene (TIEG) family of TGF β -inducible transcription factors, the expression of which is enriched in the exocrine pancreas. Like the Smads, TIEG proteins are TGF β -regulated effector proteins that significantly affect homeostasis of the pancreas. Expression of both TIEG1 and TIEG2 is rapidly upregulated by TGF β stimulation, leading to the control of pancreatic cell growth through inhibition of cell proliferation and induction of apoptosis [11,12].

Deregulation of TGF β function is a common feature of pancreatic cancer and is frequently associated with genetic disturbances of either the TGF β receptor kinases (T β R) or the

Smads [13]. During carcinogenesis, many tumor cells harbor mutations in TGF β signaling, rendering them unresponsive to TGF β -stimulated cell growth inhibition.

In this review we focus on the significance of TGF β and its downstream mediators in pancreas development, and we particularly emphasize its important functional role in the early and late stages of pancreatic cancer.

Transforming growth factor- β signaling through the Smad proteins

The TGF β exhibits its antiproliferative functions by activating a signaling pathway that mediates cell cycle arrest and induction of apoptosis. Signaling is initiated by binding of TGF β to the T β RII cell surface receptor. This, in turn, recruits the T β RI kinase, which phosphorylates the receptor R-Smads proteins, Smad2 and Smad3 (Fig. 1) [14••]. Activated R-Smads form a complex with the Co-Smad, Smad4, which shuttles directly to the nucleus. Here, the complex can either directly bind to DNA, interact with other DNA-binding proteins, or recruit other transcription factors such as AP1 to regulate the transcription of target genes involved in cell growth control (*eg*, p15, p21, c-myc). These partner proteins are crucial for the selection of Smad-regulated target genes and therefore have an enormous impact on the biologic outcome of TGF β stimulation [14••]. TGF β signaling is further controlled by a third class of Smads, the inhibitory Smad6 and Smad7 proteins, which negatively regulate R-Smad activation. Smad7 inhibits signaling from the serine/threonine kinase receptors by binding the T β RI, thereby preventing the phosphorylation of Smad2 and Smad3 [15–19]. Evidence demonstrates distinct crosstalk and feedback between the Smads and other signaling pathways, resulting in either stimulation or inhibition of Smad signaling [20–25]. The signaling network of the cell at large also modulates the expression and activity of the Smads' nuclear partner proteins, thereby fine-tuning the final selection of target genes [26••].

Together, the current data describe a simple TGF β signal transduction pathway through the Smad proteins, under the tight control of a complex web of regulator proteins, the ultimate effect on gene expression being determined by intranuclear coproteins. In addition to this well-defined pathway, growing evidence suggests the existence of Smad-independent TGF β effector pathways.

Transforming growth factor- β -inducible early response genes: new players in transforming growth factor- β -induced cell growth inhibition in the pancreas

The antiproliferative response of a cell to TGF β requires the transcriptional regulation of a set of target genes, including TGF β signal transduction components, cell cycle regulators (p15, p21, and p27), and transcription factors (*eg*, c-myc). The identification and characterization of TGF β early response genes involved in growth control of exocrine pancreatic cell populations is a focus of our laboratory. We recently reported the cloning of TIEG1 and TIEG2 (TGF β -inducible early response genes) from a rat pancreas cDNA library [12,27]. TIEG1 and TIEG2 comprise a novel family of pancreas-enriched Sp1-like transcription factors and are early-response genes for TGF β that inhibit epithelial cell proliferation [28•]. TIEG1 and TIEG2 share overall structural homology within both DNA-binding and transcriptional regulatory domains and, similarly to other Sp1-like proteins, bind GC-rich promoter sequences found in a large number of genes. Many of these target genes are involved in the control of cell proliferation, including cell cycle regulators (p15, p21, p27), MAPK, mitogenic GTPases (H-ras), and DNA synthesis proteins [11]. In contrast to most Sp1-like proteins, the TIEG proteins behave as transcriptional repressors. The TIEGs transcriptional-regulatory region contains three domains, R1, R2, and R3 [29], separated by linker regions, which may be targets for different signaling pathways. In fact, the most recent data from our laboratory demonstrates that epidermal growth factor (EGF)

signaling through the proliferative Ras-MEK-ERK-MAPkinase pathway antagonizes the transcriptional activity of TIEG2 by phosphorylation of sites in the linker region between R1 and R2 (unpublished data). Such inhibition of TIEG2 activity is likely to interfere with its antiproliferative function. Our data support a model in which TGF β -induced expression of TIEGs stimulates binding to GC-rich promoter sequences, where they regulate the transcription of genes necessary for epithelial cell growth control (Fig. 2). We have shown that overexpression of TIEGs inhibits the proliferation of TGF β -sensitive pancreatic cells and induces apoptosis in a variety of epithelial cell systems following the same mechanistic pattern as TGF β , characterized by the formation of reactive oxygen species and loss of mitochondrial membrane potential [12,27,30]. The significant role TIEGs play in pancreatic growth control is further supported by data from our laboratory using a transgenic mouse model. Targeted overexpression of TIEG2 in the acinar cells of the exocrine organ resulted in increased apoptosis associated with significant weight loss of the pancreas (unpublished data).

Together, these data strongly suggest that in addition to the Smad proteins, the TIEGs function as downstream mediators of TGF β , involved in the regulation of pancreatic growth. It is not yet clear whether the TIEG proteins work with or independently of the Smads in TGF β -mediated pancreatic cell growth control.

Transforming growth factor- β in pancreas development

In recent years, several converging lines of evidence have indicated a crucial role for TGF β in regulating pancreatic morphogenesis. Interestingly, TGF β affects the development of both the endocrine and the exocrine pancreas and also has an impact on the extracellular composition of the organ. Evidence suggests that TGF β plays a significant role in the balance between the exocrine and endocrine composition of the developing pancreas [31••, 32].

The TGF β effectively controls cell proliferation of the exocrine organ by imposing a strong antiproliferative signal on acinar cells. This is strikingly seen on supraphysiologic stimulation of cultured embryonic pancreatic buds with TGF β , resulting in increased apoptosis of acinar cells and reflecting significant regression of the acinar compartment in the developing pancreas [32]. Also, recent *in vivo* work by Sanvito *et al.* [31••] reported dramatic morphologic alterations in the exocrine pancreas of transgenic mice overexpressing TGF β . The architecture and composition of the exocrine portion of the pancreatic gland was heavily changed in TGF β overexpressing mice, characterized by loss of acinar cells and replacement of acini by fibrotic tissue.

Mice overexpressing TGF β also exhibit severe morphologic changes in the endocrine pancreas, indicating a significant role of TGF β in both acinar and islet formation. Multifocal fibrosis was observed throughout the organ, with centered clusters of endocrine cells and invading exocrine tissue. Additionally, the morphology of the islets of Langerhans themselves was affected. Although the islet cells were small and appeared fragmented, the number and distribution of insulin, glucagon, somatostatin, and pancreatic polypeptide-positive cells were normal, and the transgenic mice showed neither hyperglycemia nor changes in viability and overall health [31••]. As shown by Miralles *et al.* [33], TGF β strongly upregulates the expression and activation of matrix metalloproteinases. This crucial step during the development of pancreatic islets enables activated matrix metalloproteinases (eg, matrix metalloproteinase-2) to degrade extracellular matrix components and thereby allows endocrine cells to migrate into the surrounding mesenchyme to form mature islets of Langerhans. Application of specific TGF β -neutralizing antibodies abolished the islet morphogenesis without affecting endocrine cell differentiation.

The significance of TGF β in pancreas development is further emphasized by recent reports describing disorders of the endocrine pancreas related to Smad mutations. For instance, hypoplasia of the β -cell, hypoinsulinemia, impaired glucose tolerance, and significant insulin resistance were observed in mice with heterozygous Smad2 mutations [34••]. Although recent studies provide increasing evidence for a critical role of TGF β in multiple steps of pancreatic development, many effects of TGF β in pancreatic development are still unknown and require further study to better understand the role of TGF β signaling in this process.

Transforming growth factor- β in carcinogenesis

The TGF β plays a dual role in carcinogenesis, acting early as a tumor suppressor but promoting tumor progression in later stages [35]. Its antiproliferative functions allow TGF β to act as a strong tumor suppressor in early phases of tumorigenesis [36]. During tumor development, however, many tumor cells lose their growth inhibitory responses to TGF β [37••]. This results from low expression levels of TGF β receptors, mutations of Smad proteins, or the induction of TGF β resistance by oncogenes. Additionally, genetic alterations within cellcycle regulators and activating mutations within proliferative crosstalk signaling pathways have been reported in association with reduced TGF β cell growth inhibition [37••, 38•].

Alterations of the transforming growth factor- β signaling pathway in pancreatic cancer

A loss of sensitivity to the antiproliferative effects of TGF β is frequently associated with a reduced expression or inactivation of TGF β receptors [39]. T β RII is a frequent locus of inactivating mutations and is particularly common in colon and gastric carcinomas, along with microsatellite instability as a result of defects in the DNA mismatch repair system. By contrast, there are only sporadic reports of mutations or deletions in T β RI in gastrointestinal tumors. A missense mutation in the kinase domain of T β RI has been identified in breast cancers [40], and deletions of the T β RI occur at a very low frequency in pancreatic cancer [41].

Inactivating mutations of the *Smad4* tumor suppressor gene, located on 18q 21.1, have been described in more than 40% of pancreatic cancers. This is usually associated with complete loss of the remaining Smad4 allele and is strongly correlated with loss of growth inhibition [42]. Biallelic loss of Smad4 is also common in metastatic colon cancer (30%) and is occasionally described in other tumors of the gastrointestinal tract. Interestingly, functionally inactivating germline mutations of Smad4 have been reported in patients with familial juvenile polyposis, associated with an increased risk of gastrointestinal cancer. Like *Smad4*, the *Smad2* and *Smad7* genes are located on chromosome 18q21.1, the allelic loss of which is relatively common in gastrointestinal cancers. However, mutational analyses reveals that unlike *Smad4*, the remaining allele of *Smad2* is rarely mutated in colorectal tumors and has not been described in pancreatic cancer [26••,37••]. Although increased expression of the candidate *Smad7* oncogene has recently been reported in pancreatic cancer, no activating mutations or amplifications of the *Smad7* gene have been identified in pancreatic cancers [43]. Nonetheless, increased expression of Smad7 was associated with a more malignant phenotype in pancreatic cancer, suggesting its role in tumorigenesis.

Finally, there is no evidence that either *Smad3* or *Smad6* is the locus of homozygous deletions, functionally inactivating mutations, or amplifications in any given human malignancy.

Altered expression of transforming growth factor- β effector proteins by mutational alterations of crosstalk proteins

Many tumor cells lacking known mutations of the Smad signaling pathway become resistant to TGF β -induced growth inhibition during carcinogenesis. Recent data demonstrate that altered expression of crosstalk signaling proteins or downstream Smad partner proteins may have a significant impact on TGF β -induced growth inhibition [44•]. It has been shown that epithelial cells harboring oncogenic Ras mutations frequently lose TGF β antimitogenic responses. Oncogenic Ras and its downstream mediator, ERK-MAPkinase, compromise TGF β signaling in mammary epithelial cells by inhibiting TGF β -induced nuclear accumulation of Smad2/Smad3, thereby antagonizing TGF β -mediated growth inhibition [20]. This suggests a mechanism whereby hyperactive Ras silences antimitogenic TGF β functions in cancer cells. Similarly, in the above-described work, we showed that EGF signaling antagonizes the transcriptional repression activity of TIEG2 in pancreatic cancer cells. These findings support a model wherein oncogenic Ras mutations antagonize the antiproliferative effects of TGF β through inhibiting the Smad and the TIEG families of TGF β -effector proteins (Fig. 3). This may be of particular relevance in pancreatic cancer, because activating *k-Ras* mutations and loss of TGF β growth inhibition are both found in the majority of pancreatic cancer cells, even without inactivating mutations of the TGF β pathway.

Transforming growth factor- β in tumor progression

Reduced TGF β growth inhibition in pancreatic cancer cells is often associated with disease progression and accompanied by increased expression of TGF β itself and its type II receptor [45,46]. In fact, increased TGF β secretion has been shown to promote tumor progression both through direct effects on the tumor cells themselves and effects on accessory cells. For example, TGF β can stimulate angiogenesis, repress immune surveillance, and induce desmoplasia—all characteristics of pancreatic cancer [46–48]. Its direct effects on the tumor cells include reduction of cell-cell contacts, upregulation and activation of matrix-degrading proteinases, and induction of epithelial-mesenchymal transdifferentiation, leading to increased tumor cell invasion and metastasis [49–51]. The tumor-promoting effects of TGF β on the tumor cells themselves are observed particularly in cells possessing activating Ras mutations in which the TGF β signaling pathways remain functional despite loss of growth control by TGF β [52•,53].

Future biochemical and functional studies investigating the biologic relevance of crosstalk between TGF β and other signaling cascades in normal and neoplastic epithelial cells are required for a better understanding of the mechanisms of TGF β -induced cell growth inhibition, the loss of antiproliferation, and the switch to tumor progression in late stages of pancreatic cancer.

Conclusions

Current information about TGF β signaling and transcriptional regulation in the developing and transformed pancreatic cell populations is discussed here. In summary, TGF β functions mainly through a relatively simple signal transduction pathway, composed of the Smad proteins. The Smads transduce the TGF β signal from the cell membrane to the nucleus to regulate the transcription of target genes. It is now apparent, however, that this simple signaling system is tightly controlled by an increasing number of cytoplasmic (*eg*, Smad7) and nuclear (*eg*, coproteins) mechanisms that integrate the TGF β signal within a regulatory network of the cell. Furthermore, TGF β activates a distinct pattern of signaling pathways (*eg*, PI3K, JNK) and transcription factors such as the TIEG proteins, which may work

concurrently or independently of the Smads in TGF β -mediated pancreatic growth control. Loss of growth responsiveness to TGF β is a common feature in advanced stages of pancreatic cancer and can result from inactivating mutations within the TGF β pathway or from genetic alterations of crosstalk pathways (eg, Ras), leading to profound changes in TGF β signal transduction.

Thus, we are optimistic that this knowledge will provide the theoretical framework for future studies aimed at developing effective therapeutic strategies to treat this dismal disease.

Acknowledgments

The authors thank Dr. Umraan Ahmad for his invaluable contributions to our writing, Dr. Abigail Conley for her very helpful editorial assistance, and Dr. Vijay Shah for critical reading of the manuscript.

References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- Of special interest
 - Of outstanding interest
1. Moses HL, Serra R. Regulation of differentiation by TGF β . *Curr Opin Genet Dev.* 1996; 6:581–586. [PubMed: 8939725]
 - 2•• Edlund H. Pancreas: how to get there from the gut? *Curr Opin Cell Biol.* 1999; 11:663–668. This excellent review provides a clear outline of pancreatic development, in particular the molecules that might influence pancreatic morphogenesis during the initiation, cell-type specification, and progression of development. [PubMed: 10600706]
 3. Wells JM, Melton D. Vertebrate endoderm development. *Ann Rev Cell Dev Biol.* 1999; 15:393–410. [PubMed: 10611967]
 4. Watanabe M, Whitman M. The role of transcription factors involved in TGF β superfamily signaling during development. *Cell Mol Biol.* 1999; 45:537–543. [PubMed: 10512186]
 5. Zimmerman CM, Padgett RW. Transforming growth factor beta signaling mediators and modulators. *Gene.* 2000; 16:17–30. [PubMed: 10831835]
 6. Massague J. TGF β signal transduction. *Ann Rev Biochem.* 1998; 67:753–791. [PubMed: 9759503]
 7. Piek E, Heldin CH, Ten Dijke P. Specificity, diversity, and regulation in TGF β superfamily signaling. *FASEBJ.* 1999; 13:2105–2124.
 8. Derynck R, Zhang Y, Feng XH. Smads: transcriptional activators of TGF-beta responses. *Cell.* 1998; 95:737–740. [PubMed: 9865691]
 - 9•• Hocevar BA, Brown TL, Howe PH. TGF-beta induces fibronectin synthesis through a c-Jun N-terminal kinase-dependent, Smad4-independent pathway. *EMBO J.* 1999; 18:1345–1356. This paper is the first report describing the regulation of an endogenous gene by TGF β in a Smad4-independent manner. [PubMed: 10064600]
 10. Dai JL, Schutte M, Bansal RK, et al. Transforming growth factor-beta responsiveness in DPC4/SMAD4-null cancer cells. *Mol Carcinog.* 1999; 26:37–43. [PubMed: 10487520]
 11. Cook T, Gebelein B, Urrutia R. Sp1 and its likes: biochemical and functional predictions for a growing family of zinc finger transcription factors. *Ann NY Acad Sci.* 1999; 880:94–102. [PubMed: 10415854]
 12. Cook T, Gebelein B, Mesa K, et al. Molecular cloning and characterization of TIEG2 reveals a new subfamily of transforming growth factor-beta-inducible Sp1-like zinc finger-encoding genes involved in the regulation of cell growth. *J Biol Chem.* 1998; 273:25929–25936. [PubMed: 9748269]

13. Goggins M, Shekher M, Turnacioglu K, et al. Genetic alterations of the transforming growth factor beta receptor genes in pancreatic and biliary adenocarcinomas. *Cancer Res.* 1998; 58:5329–5332. [PubMed: 9850059]
- 14••. Massague J, Chen YG. Controlling TGF β signaling. *Genes Dev.* 2000; 14:627–644. A multitude of regulatory mechanisms control access of TGF β signaling to their receptor, and regulate the activity of their receptor and the nuclear function of the transcriptional complexes. The regulatory mechanisms that are central for understanding TGF β signaling are discussed in this review. [PubMed: 10733523]
15. Brodin G, Ahgren A, ten Dijke P, et al. Efficient TGF β induction of the Smad7 gene requires cooperation between AP-1, Sp1, and Smad proteins on the mouse Smad7 promoter. *Biol Chem.* 2000; 275:29023–29030.
16. Imamura T, Takase M, Nishihara A, et al. Smad6 inhibits signalling by the TGF β superfamily. *Nature.* 1997; 389:622–626. [PubMed: 9335505]
17. Hayashi H, Abdollah S, Qiu Y, et al. The MAD-related protein Smad7 associates with the TGF β receptor and functions as an antagonist of TGF β signaling. *Cell.* 1997; 89:1165–1173. [PubMed: 9215638]
18. Nakao A, Afrakhte M, Moren A, et al. Identification of Smad7, a TGF β -inducible antagonist of TGF β signaling. *Nature.* 1997; 389:631–635. [PubMed: 9335507]
19. Heldin CH, Miyazono K, ten Dijke P. TGF β signaling from cell membrane to nucleus through SMAD proteins. *Nature.* 1997; 390:465–471. [PubMed: 9393997]
20. Kretzschmar M, Doody J, Timokhina I, et al. A mechanism of repression of TGF β /Smad signaling by oncogenic Ras. *Genes Dev.* 1999; 13:804–816. [PubMed: 10197981]
21. de Caestecker MP, Parks WT, Frank CJ, et al. Smad2 transduces common signals from receptor serine-threonine and tyrosine kinases. *Genes Dev.* 1998; 12:1587–1592. [PubMed: 9620846]
22. Brown JD, DiChiara MR, Anderson KR, et al. MEKK-1, a component of the stress (stress-activated protein kinase/c-Jun N-terminal kinase) pathway, can selectively activate Smad2-mediated transcriptional activation in endothelial cells. *J Biol Chem.* 1999; 274:8797–8805. [PubMed: 10085121]
23. Mulder KM. Role of Ras and MAPKs in TGF β signaling. *Cytokine Growth Factor Rev.* 2000; 11:23–35. [PubMed: 10708950]
24. Hartsough MT, Frey RS, Zipfel PA, et al. Altered transforming growth factor beta signaling in epithelial cells when ras activation is blocked. *J Biol Chem.* 1996; 271:22368–22375. [PubMed: 8798398]
25. Engel ME, McDonnell MA, Law BK, et al. Interdependent SMAD and JNK signaling in transforming growth factor-beta-mediated transcription. *J Biol Chem.* 1999; 274:37413–37420. [PubMed: 10601313]
- 26••. Massague J, Blain SW, Lo RS. TGF β signaling in growth control, cancer, and heritable disorders. *Cell.* 2000; 103:295–309. This excellent review summarizes the most current knowledge in TGF β signaling and its control by cellular events. It emphasizes the role of TGF β family members in the control of cell growth and differentiation, and focuses on the significance of TGF β pathway mutations in human disorders. [PubMed: 11057902]
27. Tachibana I, Imoto M, Adjei PN, et al. Overexpression of the TGF β -regulated zinc finger encoding gene, TIEG, induces apoptosis in pancreatic epithelial cells. *J Clin Invest.* 1997; 99:2365–2374. [PubMed: 9153278]
- 28•. Cook T, Urrutia R. TIEG proteins join the Smads as TGF β -regulated transcription factors that control pancreatic cell growth. *Am J Physiol Gastrointest Liver Physiol.* 2000; 278:513–521. This is the first review describing the structure of the TIEG proteins and their role in the regulation of pancreatic cell growth.
29. Cook T, Gebelein B, Belal M, et al. Three conserved transcriptional repressor domains are a defining feature of the TIEG subfamily of Sp1-like zinc finger proteins. *J Biol Chem.* 1999; 274:29500–29504. [PubMed: 10506214]
30. Ribeiro A, Bronk SF, Roberts PJ, et al. The transforming growth factor beta (1)-inducible transcription factor TIEG1 mediates apoptosis through oxidative stress. *Hepatology.* 1999; 30:1490–1497. [PubMed: 10573529]

- 31•• Sanvito F, Herrera PL, Huarte J, et al. TGF β influences the relative development of the exocrine and endocrine pancreas *in vitro*. *Development*. 1994; 120:3451–3462. This paper describes the role that TGF β plays during pancreatic development *in vitro*. It shows that TGF β regulates the balance between endocrine and exocrine portions of the pancreas. TGF β promotes the development of the endocrine cells but inhibits the development of the exocrine pancreas. [PubMed: 7821214]
32. Lee MS, Gu D, Feng L, et al. Accumulation of extracellular matrix and developmental dysregulation in the pancreas by transgenic production of transforming growth factor-beta 1. *Am J Pathol*. 1995; 147:42–52. [PubMed: 7604884]
33. Miralles F, Battelino T, Czernichow P, et al. TGF β plays a key role in morphogenesis of the pancreatic islets of Langerhans by controlling the activity of the matrix metalloproteinase MMP-2. *J Cell Biol*. 1998; 143:827–836. [PubMed: 9813100]
- 34•• Kim SK, Hebrok M. Intercellular signals regulating pancreas development and function. *Genes Dev*. 2001; 15:111–127. This review describes the interactions between intercellular signals that mediate pancreatic development. The authors emphasize the relationship between cell-extrinsic pancreatic signaling and transcriptional regulators of pancreatic gene expression and their implications in the mechanism underlying human pancreatic exocrine and endocrine diseases. [PubMed: 11157769]
35. Akhurst RJ, Balmain A. Genetic events and the role of TGF β in epithelial tumour progression. *J Pathol*. 1999; 187:82–90. [PubMed: 10341708]
36. Gold LI. The role for transforming growth factor-beta (TGF-beta) in human cancer. *Crit Rev Oncol*. 1999; 10:303–360.
- 37•• de Caestecker MP, Piek E, Roberts AB. Role of transforming growth factor-beta signaling in cancer. *J Natl Cancer Inst*. 2000; 92:1388–1402. The authors discuss the significant role of recently identified proteins that modulate the Smad-signaling pathway and illustrate the relevance of signaling interactions between the Smads and other signaling cascades in carcinogenesis. [PubMed: 10974075]
- 38• Markowitz SD, Roberts AB. Tumor suppressor activity of the TGF β pathway in human cancers. *Cytokine Growth Factor Rev*. 1996; 7:93–102. This excellent review describes the frequency and functional implication of TGF β receptor mutations in human cancer diseases. In addition, it discusses the therapeutic potential of receptor restoration in gene therapy approaches. [PubMed: 8864357]
39. Chen T, Carter D, Garrigue-Antar L, et al. Transforming growth factor beta type I receptor kinase mutant associated with metastatic breast cancer. *Cancer Res*. 1998; 58:4805–4810. [PubMed: 9809982]
40. Goggins M, Shekher M, Turnacioglu K, et al. Genetic alterations of the transforming growth factor beta receptor genes in pancreatic and biliary adenocarcinomas. *Cancer Res*. 1998; 58:5329–5332. [PubMed: 9850059]
41. Hahn SA, Schutte M, Hoque AT, et al. DPC4, a candidate tumor suppressor gene at human chromosome 18q21.1. *Science*. 1996; 271:350–353. [PubMed: 8553070]
42. Miyaki M, Iijima T, Konishi M, et al. Higher frequency of Smad4 gene mutation in human colorectal cancer with distant metastasis. *Oncogene*. 1999; 18:3098–3103. [PubMed: 10340381]
43. Kleeff J, Ishiwata T, Maruyama H, et al. The TGF β signaling inhibitor Smad7 enhances tumorigenicity in pancreatic cancer. *Oncogene*. 1999; 18:5363–5372. [PubMed: 10498890]
- 44• Oft M, Peli J, Rudaz C, et al. TGF β 1 and Ha-Ras collaborate in modulating the phenotypic plasticity and invasiveness of epithelial tumor cells. *Genes Dev*. 1996; 10:2462–2477. This paper describes the significance of Ha-Ras in TGF β mediated epithelial mesenchymal transdifferentiation and tumor progression in mouse mammary carcinoma cells. [PubMed: 8843198]
45. Geng MM, Ellenrieder V, Wallrapp C, et al. Use of representational difference analysis to study the effect of TGFB on the expression profile of a pancreatic cancer cell line. *Genes Chromosomes Cancer*. 1999; 26:70–79. [PubMed: 10441008]
46. Wagner M, Kleeff J, Friess H, et al. Enhanced expression of the type II transforming growth factor-beta receptor is associated with decreased survival in human pancreatic cancer. *Pancreas*. 1999; 19:370–376. [PubMed: 10547197]

47. Friess H, Yamanaka Y, Buchler M, et al. Enhanced expression of transforming growth factor beta isoforms in pancreatic cancer correlates with decreased survival. *Gastroenterology*. 1993; 105:1846–1856. [PubMed: 8253361]
48. Ellenrieder V, Adler G, Gress TM. Invasion and metastasis in pancreatic cancer. *Ann Oncol*. 1999; 10:46–50. [PubMed: 10436784]
49. Welch DR, Fabra A, Nakajima M. Transforming growth factor beta stimulates mammary adenocarcinoma cell invasion and metastatic potential. *Proc Natl Acad Sci USA*. 1990; 87:7678–7682. [PubMed: 2217201]
50. Wikstrom P, Stattin P, Franck LI, et al. Transforming growth factor beta1 is associated with angiogenesis, metastasis, and poor clinical outcome in prostate cancer. *Prostate*. 1998; 37:19–29. [PubMed: 9721065]
51. Ellenrieder V, Hendler, Boeck W, et al. TGFβ 1 treatment leads to an epithelial-mesenchymal-transdifferentiation of pancreatic cancer cells requiring ERK2 activation. *Cancer Res*. 2001; 61:4222–4228. [PubMed: 11358848]
- 52••. Oft M, Heider KH, Beug H. TGFβ signaling is necessary for carcinoma cell invasiveness and metastasis. *Curr Biol*. 1998; 8:1243–1252. The authors provide evidence that cell-autonomous TGFβ signaling is required for both the induction and the maintenance of *in vitro* invasiveness during late-stage tumorigenesis. [PubMed: 9822576]
53. Ellenrieder V, Hendler SF, Ruhland C, et al. TGFβ induced invasiveness of pancreatic cancer cells is mediated by MMP-2 and the urokinase plasminogen activator system. *Int J Cancer*. 2001; 93:204–211. [PubMed: 11410867]

Abbreviations

TGFβ	transforming growth factor-β
TIEG	TGFβ-inducible early response gene
MAPK	mitogen-activated protein kinase
EGF	epidermal growth factor
Erk	extracellular regulated kinase
TβRI/II	TGFβ receptor kinases I/II

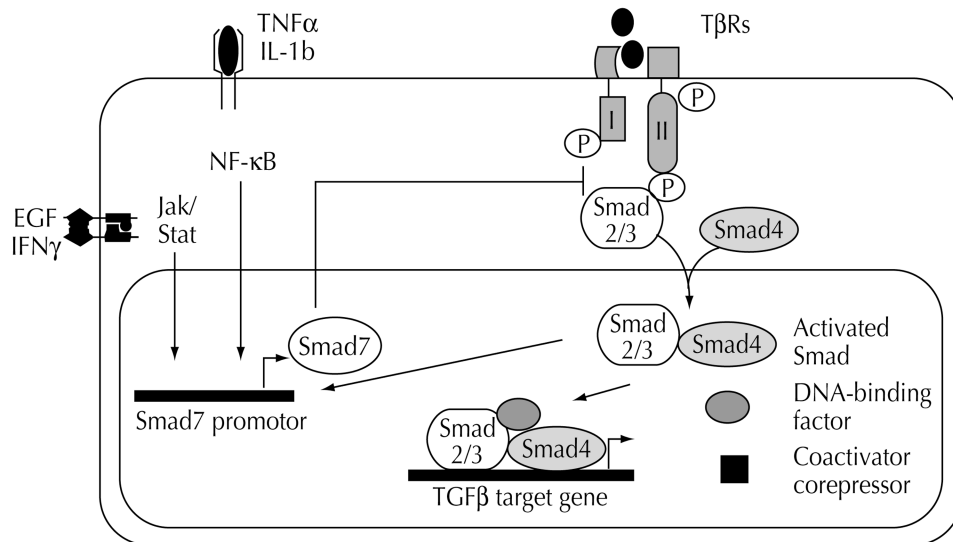


Figure 1. Schematic diagram of transforming growth factor- β (TGF β) signaling and regulation
 The signaling pathway includes two types of membrane Ser/Thr receptor kinases (T β R-I and T β R-II), the receptor Smads2/3, the co-Smad4, the inhibitory Smad7 and nuclear partner proteins (*eg*, DNA-binding factors). The Smad signaling is negatively regulated by Smad7, which itself can be induced by several crosstalks, including epidermal growth factor signaling and NF κ B.

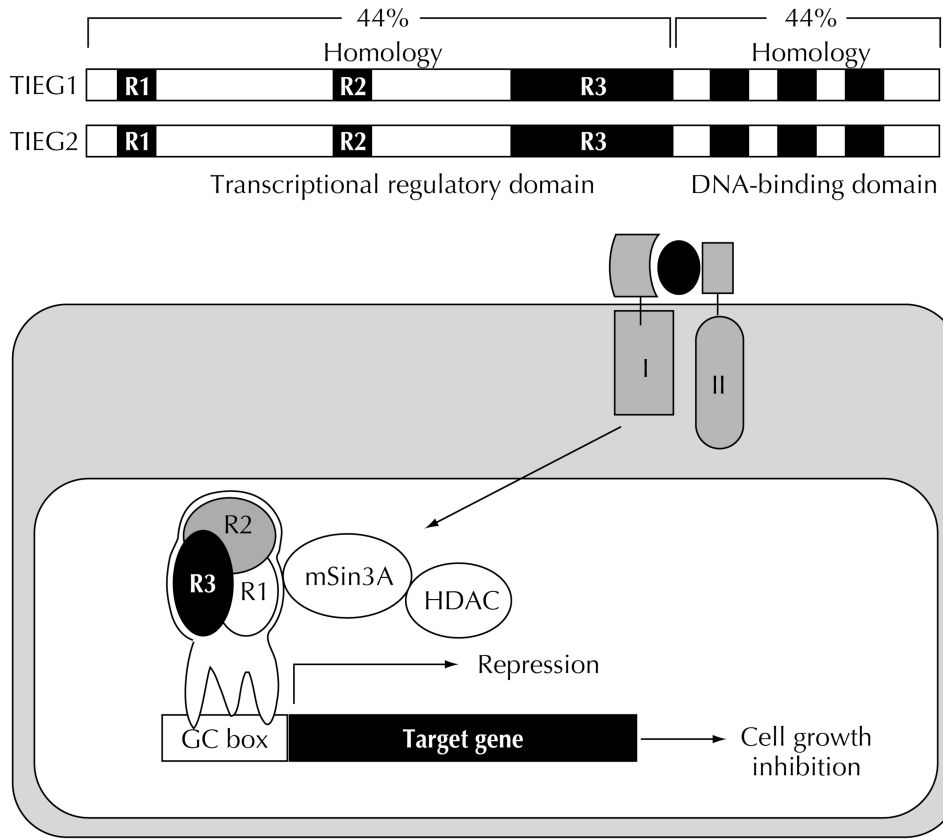


Figure 2. Schematic representation of transforming growth factor- β (TGF- β)-inducible early response gene (TIEG) structure and function
 TIEG1 and TIEG2 share a structural homology of 44% within the proline-rich transcription domain (N terminus) and a 91% similarity within the three Sp1-like zinc finger motifs of the C-terminal domain, which is responsible for DNA binding. Biochemical analysis of the transcription domain revealed the presence of three highly conserved domains that behave as potent transcriptional repression domains (R1, R2, and R3). TGF β signaling rapidly induces TIEG1 and TIEG2 expression in exocrine pancreatic cells. TIEGs then regulate the transcription of their target genes through recruiting the Sin3A corepressor complex, which contains histone deacetylase activity.

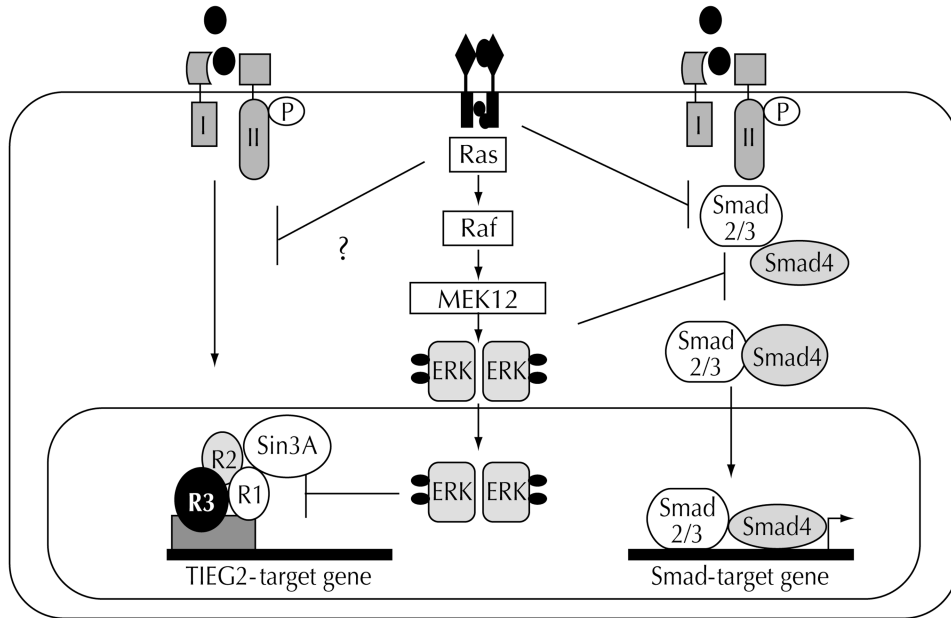


Figure 3. Crosstalk between the epidermal growth factor (EGF)-Ras- extracellular regulated kinase (Erk)-mitogen-activated protein (MAP) kinase pathway and both the transforming growth factor- β -inducible early response gene TIEG and Smad signaling pathway in cancer Hypersensitive Ras and its downstream mediator kinase Erk block transforming growth factor- β (TGF β) signaling through phosphorylation of the receptor Smads S2/S3 and thereby inhibit the complex formation with Smad4. In addition, EGF-activated Erk MAPkinase phosphorylates TIEG2 and thereby antagonizes its transcriptional repression activity through interfering with the binding of the Sin3A corepressor complex.