A New Human Colonization Model for Nontypeable *Haemophilus influenzae*

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(See the major article by Winokur et al on pages 728-38.)

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Nontypeable Haemophilus influenzae are small gram-negative bacteria that colonize the upper respiratory tract of humans, beginning at a very early age [1]. Although these organisms are normally commensals, when host defenses are compromised by underlying medical conditions such as malnutrition, immunodeficiency, chronic lung disease, or acute viral infection, H. influenzae-associated disease may ensue [2-4]. The contribution of nontypeable H. influenzae to the disease burden of children with otitis media is substantial. Among children in the developed world, this organism is currently responsible for an estimated 40%-50% of the cases of acute otitis media and an even higher percentage of cases of chronic and recurrent disease [5, 6]. In the adult population, particularly among patients with chronic obstructive pulmonary disease, nontypeable H. influenzae are major contributors to the ongoing disease process, particularly during the

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acute exacerbations that characterize the disease in many patients [4, 7]. Much has been learned about the molecular pathogenesis of disease caused by nontypeable H. influenzae in the past 2 decades, using a variety of in vitro models and in vivo models [8]. The long-term goal of much of this work has been to gain sufficient knowledge about the disease process, such that vaccines or other novel therapies can be developed to prevent nontypeable H. influenzae-associated disease in the future [9]. However, one criticism of this work has been that the findings may not be truly relevant to understanding human disease because many of these earlier studies have been conducted in nonhuman systems. Thus, the study reported by Winokur et al in this issue of the Journal, in which they describe the development of a new human model of nasopharyngeal colonization with nontypeable *H. influenzae*, is particularly timely and has the potential to be very important for the field [10].

In their newly described human colonization model, Winokur et al used a single well-characterized strain of nontypeable *H. influenzae* to intranasally inoculate a small group of human volunteers. The challenge strain was genetically modified to make it streptomycin resistant. This allowed the investigators to efficiently recover the organism from nasopharyngeal specimens collected from subjects over the course of the study. The investigators also used a carefully planned dosing algorithm that consisted of stepwise up-and-down dosing inocula, such that doses of bacteria for later volunteers were dependent on colonization success or failure in earlier study subjects. This approach allowed the investigators to quickly determine the dose that would be expected to lead to successful colonization of most study subjects and minimized the overall number of volunteers required for the project.

Fifteen volunteers took part in the study and, depending on their place in the study sequence, were scheduled to receive 1000-100 000 colony-forming units (CFU) of the nontypeable H. influenzae test strain that was administered by nose drops to each nares. The actual inoculum received by each volunteer was monitored and in most instances reasonably approximated the intended dose. Nine of the fifteen study subjects were successfully colonized with the bacteria, and the chances of success generally correlated with increasing doses of the bacteria. The investigators estimated a human colonizing dose 50 (HCD₅₀) of 1991 CFU and a HCD₉₀ of 150 314 CFU, although each value was associated with relatively wide confidence intervals, reflecting the variability observed in the study group as a whole. Nasal wash specimens were collected from each volunteer on days 3-6

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following inoculation. In individuals who were successfully colonized, positive results were generally observed for nasal wash specimens obtained by days 3 or 4 after inoculation, and results for specimens obtained on successive monitoring days were also positive. Bacterial concentrations in the wash specimens appeared to increase with time, although precise quantitation of bacterial densities in the specimens was not performed. All subjects received antibiotic treatment with levofloxacin to eliminate colonization after the day 6 specimens were obtained and recovered uneventfully.

Although nasopharyngeal acquisition of common respiratory tract bacteria has generally been thought to be an asymptomatic process, it was notable that a majority of study subjects experienced mild-to-moderate upper respiratory tract symptoms following inoculation. None of the study subjects developed a documented fever, but 5 subjects developed headache, 6 subjects developed rhinorrhea or nasal congestion, and 1 subject developed mild-to-moderate sore throat. There appeared to be a close temporal relationship between the development of sore throat and the recovery of nontypeable H. influenzae from the nasal wash specimens. Furthermore, resolution of sore throat symptoms occurred in the majority of colonized subjects coincident with receipt of levofloxacin, again supporting a causative role for bacterial colonization in the development of symptoms.

Demonstration of interaction of the nasopharyngeal bacteria with the host immune system is important in validating the model and in justifying use of the model in future investigations. Not surprisingly, given the ubiquity of non-typeable *H. influenzae* in the normal nasopharyngeal flora, a screening enzymelinked immunosorbent assay detected antibody to whole bacterial cells in preinoculation serum samples from all subjects. However, only colonized subjects demonstrated \geq 4-fold increases in titers of the major antibody classes, with 7 of 9

subjects demonstrating such an increase and with IgG being the class most likely to show a 4-fold rise.

The work presented by Winokur et al represents an important first step in the development and application of this unique model. Many questions that could not be directly addressed previously can now be examined in this model system. The investigators appropriately chose to begin their studies by using a previously well-characterized bacterial strain [11, 12]. In future studies, evaluation of mutants derived from the parent strain for their ability to colonize human volunteers should allow the investigators to quickly assess the relative importance of the many surface adhesins and other H. influenzae virulence factors that have been previously defined in other model systems [13-15]. Other investigators studying the related organism Haemophilus ducreyi have had great success defining the relative importance of its potential virulence factors for humans by evaluating H. ducreyi mutants in a different human challenge model [16].

The nontypeable H. influenzae colonization model would also seem to hold great promise for the evaluation and comparison of the many vaccine candidates being studied for prevention of nontypeable Haemophilus-associated disease [9]. Despite many years of research, there is still no well-recognized correlate of protection against nontypeable H. influenzae-associated disease in humans. This has made it very difficult to objectively assess and compare the many vaccine candidates that have been studied over the years. The colonization model developed by Winokur et al may finally give investigators the ability to critically evaluate the protective potential of the leading vaccine candidates prior to progressing to human clinical trials [17, 18]

Despite the promise of the model, there are still many questions and challenges that will need to be addressed and further explored as the work moves forward. Some of these relate to the nontypeable *H. influenzae* organisms themselves and others relate to the human subjects that would form the basis of future investigations. Nontypeable H. influenzae are a very diverse group of organisms with the ability to rapidly change their genetic makeup via natural transformation [19, 20]. One must always be cautious about drawing broad conclusions about experimental results on the basis of studies of a single strain, which may be more or less representative of the bacterial population as a whole. While the data derived from the colonization model is valuable in its own right even with the one prototype strain described in the article by Winokur et al, the validity and generalizability of future studies would be strengthened by the inclusion of additional strains. This concern would apply not only for studies of virulence traits, but also for studies of potential vaccine candidates whose antigenic properties and expression levels may vary considerably from strain to strain [9].

Another potential challenge in the development and interpretation of the colonization model concerns the effect of preexisting immunity on the success or failure of colonization. The investigators allude to this point in their discussion, noting that one volunteer who received an inoculum well above the predicted HCD₉₀ challenge dose was not successfully colonized. Without good understanding at this point as to what specific antibodies or other immune factors are most important in preventing colonization, it may be difficult to reliably prevent this same sort of problem from occurring in future studies. Hopefully, as their work goes forward, these investigators will be able to better define correlates of protection against colonization on the basis of either analysis of preexisting antibody profiles or assessment of protection offered by individual candidate vaccines.

Even with the potential problems that must still be overcome, the development of the nontypeable *H. influenzae* human colonization model must still be seen as a significant advance for the field. The model will, for the first time, provide investigators with a system in which experimental questions related to nontypeable *H. influenzae*-associated disease pathogenesis and vaccine efficacy can be objectively addressed with confidence that the results may have direct applicability to understanding and preventing disease in humans. Winokur et al are to be commended for this important piece of work.

Notes

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References

- Faden H, Waz MJ, Bernstein JM, Brodsky L, Stanievich J, Ogra PL. Nasopharyngeal flora in the first three years of life in normal and otitis-prone children. Ann Otol Rhinol Laryngol 1991; 100:612–5.
- Foxwell AR, Kyd JM, Cripps AW. Nontypeable *Haemophilus influenzae*: pathogenesis and prevention. Microbiol Mol Biol Rev 1998; 62:294–308.
- Sethi S, Murphy TF. Infection in the pathogenesis and course of chronic obstructive pulmonary disease. N Engl J Med 2008; 359:2355–65.
- Sethi S, Evans N, Grant BJ, Murphy TF. New strains of bacteria and exacerbations of chronic obstructive pulmonary disease. N Engl J Med 2002; 347:465–71.

- Pichichero ME, Casey JR, Hoberman A, Schwartz R. Pathogens causing recurrent and difficult-to-treat acute otitis media, 2003–2006. Clin Pediatr (Phila) 2008; 47: 901–6.
- Casey JR, Pichichero ME. Changes in frequency and pathogens causing acute otitis media in 1995–2003. Pediatr Infect Dis J 2004; 23:824–8.
- Sethi S, Murphy TF. Bacterial infection in chronic obstructive pulmonary disease in 2000: a state-of-the-art review. Clin Microbiol Rev 2001; 14:336–63.
- Erwin AL, Smith AL. Nontypeable Haemophilus influenzae: Understanding virulence and commensal behavior. Trends Microbiol 2007; 15:355–62.
- Bakaletz LO, Klein DL, Pelton SI, et al. Report of the vaccine panel: ninth international otitis media research conference. In: Lim DJ, ed. Recent advances in otitis media. St. Pete Beach, FL: House Research Institute, 2010:94–119. http://www.hei.org/ otitis2007/reportcontent.htm. Accessed 11 June 2013.
- Winokur P, Chaloner K, Doern G, Ferreira J, Apicella M. Safety and immunological outcomes following human inoculation with nontypeable *Haemophilus influenzae*. J Infect Dis 2013; 208:728–38.
- Pang B, Winn D, Johnson R, et al. Lipooligosaccharides containing phosphorylcholine delay pulmonary clearance of nontypeable *Haemophilus influenzae*. Infect Immun 2008; 76:2037–43.
- Swords WE, Ketterer MR, Shao J, Campbell CA, Weiser JN, Apicella MA. Binding of the nontypeable *Haemophilus influenzae* lipooligosaccharide to the PAF receptor initiates host cell signalling. Cell Microbiol **2001**; 3:525–36.
- 13. Schachern PA, Tsuprun V, Wang B, et al. Effect of lipooligosaccharide mutations of

Haemophilus influenzae on the middle and inner ears. Int J Pediatr Otorhinolaryngol **2009**; 73:1757–60.

- Novotny LA, Adams LD, Kang DR, et al. Epitope mapping immunodominant regions of the PilA protein of nontypeable *Haemophilus influenzae* (NTHI) to facilitate the design of two novel chimeric vaccine candidates. Vaccine 2009; 28:279–89.
- St Geme JW 3rd, Kumar VV, Cutter D, Barenkamp SJ. Prevalence and distribution of the *hmw* and *hia* genes and the HMW and Hia adhesins among genetically diverse strains of nontypeable *Haemophilus influenzae*. Infect Immun **1998**; 66:364–8.
- Janowicz DM, Ofner S, Katz BP, Spinola SM. Experimental infection of human volunteers with Haemophilus ducreyi: fifteen years of clinical data and experience. J Infect Dis 2009; 199:1671–9.
- Prymula R, Hanovcova I, Splino M, et al. Impact of the 10-valent pneumococcal nontypeable *Haemophilus influenzae* protein D conjugate vaccine (PHiD-CV) on bacterial nasopharyngeal carriage. Vaccine **2011**; 29: 1959–67.
- Prymula R, Peeters P, Chrobok V, et al. Pneumococcal capsular polysaccharides conjugated to protein D for prevention of acute otitis media caused by both *Streptococcus pneumoniae* and nontypable *Haemophilus influenzae*: A randomised double-blind efficacy study. Lancet 2006; 367:740–8.
- Erwin AL, Sandstedt SA, Bonthuis PJ, et al. Analysis of genetic relatedness of *Haemophilus influenzae* isolates by multilocus sequence typing. J Bacteriol **2008**; 190: 1473–83.
- Shen K, Antalis P, Gladitz J, et al. Identification, distribution, and expression of novel genes in 10 clinical isolates of nontypeable *Haemophilus influenzae*. Infect Immun 2005; 73:3479–91.