

REVIEW

Stem cells: novel players in the treatment of erectile dysfunction

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Stem cells are defined by their capacity for both self-renewal and directed differentiation; thus, they represent great promise for regenerative medicine. Historically, stem cells have been categorized as either embryonic stem cells (ESCs) or adult stem cells (ASCs). It was previously believed that only ESCs hold the ability to differentiate into any cell type, whereas ASCs have the capacity to give rise only to cells of a given germ layer. More recently, however, numerous studies demonstrated the ability of ASCs to differentiate into cell types beyond their tissue origin. The aim of this review was to summarize contemporary evidence regarding stem cell availability, differentiation, and more specifically, the potential of these cells in the diagnosis and treatment of erectile dysfunction (ED) in both animal models and human research. We performed a search on PubMed for articles related to definition, localisation and circulation of stem cells as well as the application of stem cells in both diagnosis and treatment of ED. Strong evidence supports the concept that stem cell therapy is potentially the next therapeutic approach for ED. To date, a large spectrum of stem cells, including bone marrow mesenchymal stem cells, adipose tissue-derived stem cells and muscle-derived stem cells, have been investigated for neural, vascular, endothelial or smooth muscle regeneration in animal models for ED. In addition, several subtypes of ASCs are localized in the penis, and circulating endogenous stem cells can be employed to predict the outcome of ED and ED-related cardiovascular diseases.

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INTRODUCTION

Stem cells are by definition capable of self-renewal, meaning that they can make exact copies of themselves indefinitely, and of differentiation into a variety of phenotypes.^{1,2} They have been shown to be able to functionally regenerate damaged tissues, depending on the stimuli or signals that they receive.^{1,2} When a stem cell divides, both daughter cells have the potential either to remain stem cells or to become a more specialized type of cell, such as a muscle cell, a red blood cell or a brain cell. Stem cells are classified into totipotent, pluripotent, multipotent and unipotent in a hierarchical order based on the number of cell lineages to which they may potentially differentiate.³ Totipotent stem cells, like the zygote and its offspring cells of the morula, have the greatest differentiation potential and are capable of forming cells of the ectoderm, mesoderm and endoderm.⁴ Pluripotent stem cells give rise to the three germ layers, but not to extra-embryonic tissues. Embryonic stem cells (ESCs), isolated from the inner cell mass of blastocysts,⁵ are the most widely acknowledged example of pluripotent cells.⁶ Multipotent stem cells, such as haematopoietic stem cells (HSCs) and mesenchymal stem cells (MSCs), isolated from the developing germ layer and their descended adult organs, are capable of self-renewal, and they can differentiate into any cell type within their germ layer.⁴ Unipotent cells are progenitor cells or precursor cells with a limited capacity for self-renewal, and they can differentiate into only one defined cell type, such as epithelial cells.^{4,7}

Harvesting ESCs requires the destruction of human embryos and has raised significant ethical and political concerns. These barriers have prompted the search for alternative stem cell sources, including amniotic fluid-derived stem cells and adult stem cells (ASCs).⁷ ASCs are gaining popularity, as researchers are finding a more extensive differentiation potential than was previously thought to exist, with some studies demonstrating pluripotency of certain ASCs.^{8,9}

Erectile dysfunction (ED) is defined as the inability to attain and maintain an erection with sufficient rigidity to permit satisfactory sexual intercourse.¹⁰ Vascular diseases, which are correlated with smoking, aging, hyperlipidemia, diabetes and hypertension, are major causes of ED. Injury to the cavernous nerves during pelvic surgery, such as radical prostatectomy, comprises an appreciable number of ED cases as well.⁶ Other causes of ED include direct trauma to the genitals, endocrine disorders, and fibrosis of the penile vasculature and corporal smooth muscle.¹¹ The aetiological theme linking both vasculogenic and neurogenic ED is the loss of normal cellular function or death of the cells themselves.

While oral pharmacotherapies, such as phosphodiesterase-5 inhibitors (PDE5is), have clear benefits, their actions are necessarily ephemeral, and treatment is relatively costly.⁶ Furthermore, they do not provide a cure, and a number of patients have tissue damage that is so extensive that the response to either oral or local pharmacotherapy is minimal. Consequently, researchers have been investigating stem cells

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as a substitute therapeutic strategy. Compared with other fields, the application of cell-based therapy for ED is relatively new.¹² The potentially curative nature of stem cells recently prompted a number of studies with varying stem cell populations and strategies.⁶ Currently, it appears to be one of the most studied potential future treatments of ED.

STEM CELL SOURCES, THEIR LOCATION AND CIRCULATION

Stem cells and sources

Currently, there are three ways to obtain a pluripotent stem cell, including ESC isolation, somatic cell nuclear transfer and somatic cell reprogramming. In 1981, pluripotent ESCs were first discovered in the inner cell mass of the mouse embryo.¹³ Their ability to differentiate into many cell types aroused tremendous hope for their potential application in cell therapy and regenerative medicine. However, as stated above, ESCs present an ethical burden. To overcome this limitation, new stem cell technologies, such as the induction of pluripotency by somatic cell nuclear transfer and reprogramming, have been established.⁷ Over the past few years, the successful generation of induced pluripotent stem cells (iPSCs) brought about new hope for regenerative therapies.¹⁴ Many groups have now shown that somatic cells can be reprogrammed by overexpression of variable sets of few transcription factors in cells that share various characteristics with ESCs.^{14–17} To date, several novel reprogramming protocols are available,^{18–22} which have broadened the spectrum of cell types and species involved in iPSCs generation. Similar to ESCs, iPSCs also show the potential for differentiation into numerous cell types. However, the degree of molecular similarity between iPSCs and ESCs has not been completely elucidated. To date, it is not clear whether these small differences are the result of interexperiment variability or whether reprogramming of somatic cells generates a state that is unique to ESCs.²³ Thus, iPSCs seem to hold promise for regeneration medicine once ascribed solely to ESC.

To avoid ethical dilemmas, ASCs have been given increasing attention. ASCs tend to be tissue specific, self-renewing populations of cells that can differentiate into cell types associated with the organ system in which they reside.^{24–26} While they used to be considered multipotent at the most, a continuously expanding body of literature suggests that certain ASCs appear to possess pluripotent differentiation capacity.² ASCs are easily identified in tissues with high cell turnover. Within the last decade, many niches of ASCs were discovered in many tissues, such as the brain, liver, skin, skeletal muscle, the gastrointestinal tract, adipose tissue, pancreas, eye, blood and dental pulp.^{24–28} Among them, the most notable exception to the tissue specificity of ASCs are the MSCs, which are defined as (multipotent) MSCs. They are capable of differentiating *in vitro* into various mesenchymal/mesodermal cells and differentiating developmentally if injected into a blastocyst.²⁶ MSCs have been extensively tested and proven effective in preclinical studies, and they currently are being tested in US FDA-approved clinical trials for the treatment of myocardial infarction, stroke, meniscus injury, limb ischemia, graft-versus-host disease and autoimmune disorders.²⁹ Although the mechanisms are not completely clear, MSCs are known to secrete a broad range of cytokines and growth factors that have both paracrine and autocrine effects on damaged tissues. MSCs have been isolated from bone marrow, adipose tissue, skeletal muscle, dental pulp and cord blood.^{30–34} Among them, the most studied are bone marrow-derived stem cells (BMSCs), which are derived from bone marrow stroma (hence their name). In the early 1960s, BMSCs were first determined to be responsible for marrow reconstitution due to their ability to renew themselves and their ability

to differentiate into various cell types.^{35,36} Recently, MSCs derived from the stromal vascular fraction of adipose tissue, termed as adipose tissue-derived stem cells (ADSCs), represent an abundant and easily accessible source of stem cells.³⁷ While bone marrow is obtainable in the gram range by a painful marrow aspiration procedure, adipose tissue can be obtained in the range of hundreds of grams with a minimally invasive procedure. The possibility of harvesting hundreds of grams of adipose tissue excludes the need for MSC isolation and culture steps and allows for the direct re-injection of the stromal vascular fraction during the same surgical procedure in which they were harvested. ADSCs bear a strong resemblance to BMSCs, as demonstrated by the expression of common cell surface markers, similar gene expression profiles and similar differentiation potentials. Therefore, it is reasonable to expect that ADSCs will become the preferred choice of ASCs for future clinical applications.³⁷

Localisation of ASCs in their native tissue—stem cell niche

The stem cell niche is a microenvironment that maintains stem cells in a quiescent state. After tissue injury, the niche promotes either self-renewal or differentiation to form new tissues.³⁸ Several components of this microenvironment regulate stem cell characteristics within the niche: cell–cell interactions between stem cells and other stem cells or neighbouring differentiated cells; interactions of stem cells with adhesion molecules and the extracellular matrix; the presence and active secretion of a multitude of cytokines and growth factors; and oxygen tension and other physiochemical determinants. The stem cell niche is often located in the perivascular space of various tissues, thus providing direct access to the systemic circulation into which endogenous stem cells are recruited during tissue injury. In spite of the perivascular location of the niche, this microenvironment appears to be in a continuous state of relative hypoxia.^{39,40} For example, the mesenchymal stem cell niche is exposed to oxygen tension as low as 2%.⁴⁰ The latter finding has driven researchers to culture stem cells in hypoxic conditions, thereby replicating their physiological environment. It has been shown that various stem cells may benefit from these culture conditions, replicating the niche *in vitro*: there have been observations of increased growth factor secretion and increased engraftment potential after hypoxic culturing.⁴¹

The niche supports stem cell activity through complex but not completely known mechanisms, including: (i) the secretion of soluble factors, such as growth factors and cytokines;⁴¹ (ii) a strong interaction and self-renewal potential between stem cells and their niches; for example, hair follicle bulge stem cells expanded in culture and grafted into nude mice generated new bulge niches;⁴² and (iii) the activation of a variety of signals and cytokines when stem cell reinforcements are needed for tissue repair or regeneration, transforming the niche from a resting place to a command centre.^{43–45}

There are two classic types of stem cell niches based on their anatomic relationship to stem cells:³⁸ the stromal type and the epithelial type:

1. Stromal type. Niche cells reside in close proximity to their respective stem cell pools. For example, many HSCs and BMSCs reside along the endosteal surface of trabecular bone in close proximity to both bone-forming osteoblasts and vascular endothelial cells (ECs)^{46–48} (Figure 1a), the latter of which may facilitate mobilisation of stem cells from bone marrow into the circulation and their return to these niches.⁴⁹ During the last few years, the vascular wall has been considered as an important reservoir for different types of stem and progenitor cells,^{50,51} in addition, many kinds of stem cells, including ADSCs, HSCs, neural stem cells

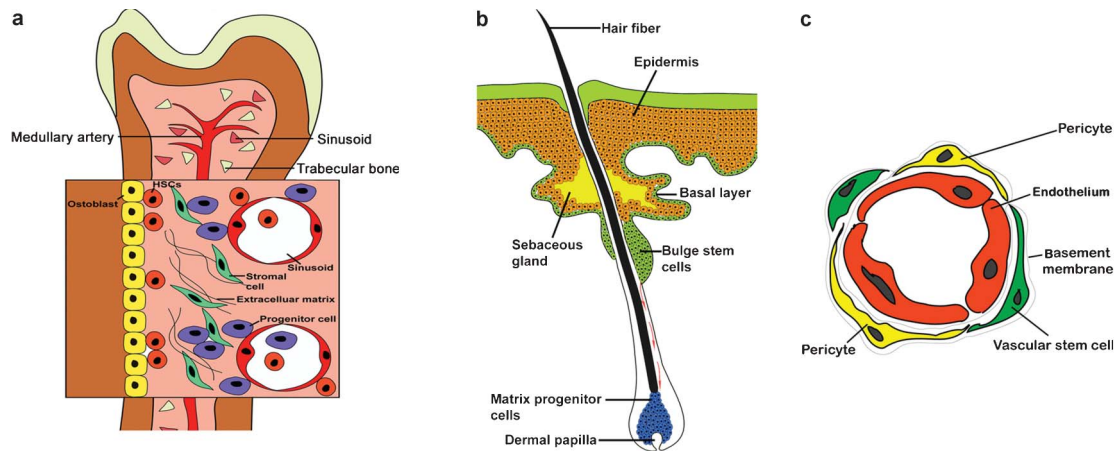


Figure 1 Stem cells niches. **(a)** Stromal type. HSCs are found adjacent to sinusoidal blood vessels as well as at or near the endosteum. Osteoblasts and mesenchymal progenitors have been proposed to produce factors that regulate HSC maintenance. **(b)** Epithelial type. Stem cells located within the bulge region of the hair follicle give rise to new matrix cells located at the base of the hair and adjacent to the dermal papilla that supports hair growth. Bulge cells can also help maintain the sebaceous gland and the epidermal stem cells. **(c)** The vascular wall as a reservoir for stem and progenitor cells. Capillaries are constructed by endothelial cells lining the lumen and pericytes, which cover the endothelial tube. Vascular wall-resident endothelial progenitor cells, HSCs and MSCs, are localized in the subendothelial space and are probably located between or around pericytes. HSCs, haematopoietic stem cells; MSCs, mesenchymal stem cells.

(NSCs), intestinal stem cells and spermatogonial stem cells,^{37,52–55} have been discovered by culturing stromal vascular fractions from their respective native tissue. Generally, perivascular cells, including pericytes in capillaries and adventitial cells around larger vessels, are considered to be able to originate multilineage mesodermal progenitor cells (**Figure 1c**).⁵⁰

2. Epithelial type. The most typical example of this type resides within a specialized region of the outer root sheath of the hair follicle and is known as the follicular bulge stem cell niche. The stem cells in this niche could give rise to new matrix cells located at the base of the hair and adjacent to the dermal papilla that support hair growth⁵⁶ (**Figure 1b**).

Trafficking of exogenously administered stem cells

In the clinical setting, the administration route of stem cells depends on the anatomy and the extent of damage of the involved tissue or organ, offering a choice between two approaches: direct local implantation versus systemic intravascular administration.⁵⁷ Local implantation is an invasive procedure that could also disrupt the highly complex and delicate microenvironment, causing inflammation and multifocality of many disorders.⁵⁷ Therefore, systemic diseases rely on vascular delivery of stem cells. However, the therapeutic efficacy depends on ensuring that sufficient stem cells home to the bone marrow or mobilize from the bone marrow to the damaged area. Homing is thought to be a multistep process that involves: (i) signalling and chemoattraction by chemokines; (ii) activation and attachment to adhesion molecules on the endothelial cell surface; and (iii) transendothelial migration of the stem cells into the diseased tissue.

Chemokines are a family of small cytokines. Their name is derived from their ability to induce directed chemotaxis in nearby responsive cells. Some chemokines are considered pro-inflammatory and can be induced during an immune response to recruit cells of the immune system to a site of infection. While most chemokines are important mediators of inflammation, it was recently discovered that chemokine secretion in sites of tissue damage or inflammation can be used to direct regenerative therapy.⁵⁸ More specifically, certain types of stem

cells have been shown to express mRNA and a large number of chemokine-receptors. Various groups have shown expression of the chemokine stromal cell-derived factor 1 α (CXCL12) in myocardial infarction;⁵⁹ CX3CL1, CCL2, CCL3 and CCL5 in peripheral neural injury;⁶⁰ and CCL7 in rat models of birth injury and stress urinary incontinence.⁶¹ Recent data indicate that CXCL12 may play a role in trafficking ADSCs towards the major pelvic ganglion in rats with an injured cavernous nerve.⁶² In our laboratory, we have performed a comparative analysis between ADSCs and BMSCs and were able to show that these two types of MSCs express a similar panel of chemokine receptors. We identified expression of mRNA for 12 of the currently known 21 chemokine receptors, and found that these receptors were located both on the cell surface as well as intracellularly, suggesting that chemokine receptors are recycled in these stem cells, as has been previously shown in various immune cells (unpublished data). What the exact role of these chemokine receptors is in homing towards sites of tissue injury and what effects this stem cell recruitment has on the host tissue remains largely unknown and is the focus of intense investigation in stem cell biology.

After the recruitment of stem cells towards the injured tissue by chemoattraction, the activation of lymphocyte function-associated antigen 1, very late antigen 4/5 and CD44, cytoskeleton rearrangement, membrane type 1-matrix metalloproteinase activation and the secretion of matrix metalloproteinase-2/9 play important roles in the interaction between stem cells and endothelial cells.^{58–63} Rolling and firm adhesion is followed by transendothelial migration across the physical endothelium/extracellular matrix barrier.

Similar processes play a role in homing of stem cells towards their niche from the circulation. Chemoattraction, mainly by CXCL12, adhesion to the endothelium and transendothelial migration are comparable to those processes in injured tissues. In the last stage, stem cells finalize their 'homing' by anchorage to their specialized niches in the extravascular space of the endosteum region and at perivascular sites.⁶⁰

Reawakening and mobilisation of endogenous stem cells

The development of techniques to reawaken and mobilize endogenous pools of stem cells would represent progress in the field of regenerative

medicine. In general, the recruitment of HSCs from bone marrow into blood is termed 'mobilisation'.⁶⁴ This is a complex mechanism that has not yet been clarified. The haematopoietic niche of the bone marrow expresses a broad array of cell surface receptors, such as the CXC chemokine receptors CXCR4 and CXCR2, lymphocyte function-associated antigen 1, very late antigen 4 and glycoprotein CD44, among others.⁶⁵ Data from a number of preclinical models showed that the inhibition of these receptor–ligand interactions resulted in enhanced progenitor cell mobilisation.^{66–68} Among them, CXCL12/CXCR4 interactions may play key role in the regulation of the routine and active egress of progenitor and maturing cells from the marrow into the blood.⁶⁴ Plerixafor, a novel agent that disrupts the CXCR4–CXCL12 bond, which is the primary haematopoietic stem cell anchor in the bone marrow, has recently been US FDA-approved for mobilizing HSCs from the bone marrow. Another common mobilisation strategy in current clinical applications includes the use of cytokines alone or in conjunction with chemotherapy.⁶⁵ The currently available cytokines include granulocyte colony-stimulating factor, granulocyte macrophage colony-stimulating factor, and erythropoietin and recombinant methionyl human stem cell factor.⁶⁵

PENILE STEM CELLS (PSCS)

Many types of ASCs have been reported in different tissues. In this review, we will focus on the male erectile organ, the penis. As an organ composed of multiple types of tissues, the penis itself contains a variety of stem cells. To date, few types of penile stem cells have been isolated or identified.

Foreskin stem cells

Two types of foreskin stem cells have been isolated to date, including skin-derived progenitors (SKPs) and MSCs. A new and unique multipotent progenitor cell population from adult mammalian dermis, termed SKPs, has been isolated and expanded from rodent and human skin and differentiated into both neural and mesodermal progeny.^{69,70} SKPs did not apparently produce keratinocytes, distinguishing them from epidermal stem cells. These progenitor cells have been reported to reside in the dermal papillae of hair follicles.⁷¹ When the skin cells of neonatal and adult rodents were dissociated to single cells and grown in suspension culture in the presence of the mitogens fibroblast growth factor (FGF) and epidermal growth factor, floating spheres of proliferating cells were generated.⁶⁹ These spheres were positive for nestin, a filament mainly expressed in neural and skeletal muscle progenitors. Moreover, differentiation of SKPs *in vitro* resulted in the generation of separate subpopulations of cells expressing neuronal, glial, smooth muscle and adipocyte markers. In 2005, SKPs from neonatal human foreskin tissue were identified,⁷² and they showed similar biological characteristics to the animal cells mentioned above. Because human neonatal foreskin does not contain hair follicles, the presence of SKPs in foreskin provides evidence for an extrafollicular SKP niche.⁷² However, the difficulty of reliably isolating and expanding comparable populations of SKPs from both whole hairless and hairy adult human skin suggests that this extrafollicular niche may be significantly limited after the neonatal period in humans.^{70,71}

Meanwhile, MSCs were also obtained from low-temperature preserved human foreskin biopsies by adherent culture.⁷³ These cells could differentiate into mesodermal lineages, including adipocytes, osteocytes and myocytes.⁷⁴ MSCs were antigenically distinct from SKPs, and when grown under the same conditions, the MSCs grew adherently (plastic adherence is one of the three hallmarks of MSCs), while SKPs grew as floating spheres.

Tunica albuginea stem cells

Vernet and colleagues⁷⁵ investigated whether cultures of cells from normal tunica albuginea and Peyronie's disease plaques undergo osteogenesis, express markers of stem cells, and originate other cell lineages *via* processes modulated by transforming growth factor- β 1. They found that both cultures express the stem cell marker CD34 and the SMCs markers smoothelin and transgelin. Cells expressing CD34 and Sca-1 were also found *in vivo* in the normal tunica albuginea, as well as in the corpora cavernosa.⁷⁶ They were identified as potential endogenous stem cells because they are likely the ones that, in the *in vitro* models, undergo multiple lineage differentiation, and in the Peyronie's disease plaque, they may convert into myofibroblasts and osteoblasts. In addition, shaft penile tissue sections from the rat and wild type mouse were immunostained for Oct-4, an ESC marker.⁷⁷ Results showed that Oct-4⁺ cells were detected in tunical and corporal tissues and that they could differentiate into SMCs, myofibroblasts and cardiomyocytes. This is the first report of the isolation and characterisation of embryonic-like endogenous stem cells in penile tissues.

Perivascular stem cells

Although perivascular stem cells have been extracted from multiple organs, such as bone marrow, dental pulp, placenta, fat and umbilical cord,⁵¹ the penis, as a part of a systematic circulation tree, has not yet received attention in this regard. We hypothesize that a ubiquitous reserve of multilineage progenitor cells in the capillaries, veins and arteries of the penis exists.

Endothelial progenitor cells (EPCs)

ED, cardiovascular disease and male hypogonadism share the common denominator of endothelial dysfunction, which has a well-established role in the pathogenesis of both atherosclerosis and plaque instability.^{78–80} One of the most exciting discoveries in vascular biology came in 1997 with the apparent detection of bone marrow-derived EPCs,⁸¹ which have the capacity to migrate into the peripheral circulation and to differentiate into mature ECs, thus providing a circulating pool of the cells that may contribute to ongoing endothelial repair.⁸² This process, called vasculogenesis,⁸³ is impaired in ED, as documented by low levels of circulating EPCs.⁸² Meanwhile, at least one research group believes that EPCs are native inhabitants of certain blood vessel walls, and it would be instructive to determine whether EPCs are embedded within the penile vasculature and can be stimulated to repair dysfunctional penile ECs.⁸⁴

Recently, several groups demonstrated that in ED patients, the reduced level of EPCs could be restored by administration of PDE5i,^{85,86} such as vardenafil and tadalafil, resulting in an effective vasculoprotection and prevention of the initiation and progression of endothelial dysfunction. The mechanisms remain unknown; however, a possible hypothesis involves the presence of PDE5 in bone marrow that, when inhibited, may magnify the local effect of nitric oxide, thus leading to the mobilisation of stem and progenitor cells.⁸⁵

APPLICATION OF STEM CELLS FOR PREDICTING ED

EPCs show a wide heterogenic antigenic profile, including expression of CD34, CD133 and KDR. CD34 is an adhesion molecule expressed on HSCs and is typically considered a marker of immaturity; CD133 is a surface antigen of unknown function that identifies more immature progenitor cells than CD34 alone; and KDR represents the type 2 vascular endothelial growth factor (VEGF) receptor and indicates early endothelial differentiation.⁸⁷

There are several types of circulating EPCs related to ED. In particular, circulating levels of CD34⁺CD133⁺ cells were recently shown to be decreased in patients with ED with or without cardiovascular risk factors,⁸² while CD34⁺KDR⁺ cells showed no relationship with ED.⁸⁸

Meanwhile, increased levels of osteocalcin⁺ EPCs could be a predictive marker of subsequent coronary artery disease as well as ED.⁸⁹ An increasing number of authors have described a population of EPCs expressing osteocalcin, a typical osteo-related protein.⁹⁰ Carlo's group⁸⁹ investigated the correlation between osteocalcin⁺ EPCs and cavernous atherosclerotic lesions in ED patients. They found that osteocalcin⁺ EPC levels increased significantly with cavernous atherogenesis progression.

We should note that decreased levels of EPCs are not specific indicators for ED. Cardiovascular risk factors are known to decrease EPCs with a subsequent increase of cardiovascular events and cardiovascular deaths.⁹¹ Thus, circulating levels of EPCs are thought to be a link between cardiovascular risk factors and endothelial dysfunction, with a potential influence on erectile function.

APPLICATION OF STEM CELLS AS A THERAPY OF ED

Current pharmacological agents for vasculogenic ED, such as PDE5i, lack efficacy in treating ED patients with advanced diabetes or patients suffering from ED following radical prostatectomy.^{92,93} In the latter condition, Wallerian degeneration of the nerve and target apoptosis and fibrosis in the corpus cavernosum result in an irreversible loss of function.^{6,94} Stem cell-based therapy has been proposed in the management of ED by complete replacement of lost or damaged cells or by protecting threatened host cells *via* immunomodulatory effects, the provision of trophic factors or gene delivery.⁹⁵ In recent years, a number of reports related to the progress of stem cell-based ED therapy have been published. Different therapeutic forms of stem cells have been developed, including multiple sources of stem cells or progenitor cells, gene-transfected stem cells, stem cell lysates and stem cells seeded on tissue matrices (Table 1).

While some of the more extravagant claims are excessively optimistic, there is clear validity to the notion that stem cells may lead to novel and potentially curative therapies. Nonetheless, clinical application is a long way off. Prior to clinical use of stem cells, it will be necessary to thoroughly investigate the malignant potential of stem cells, especially for ESCs. Although ASCs seem to be more stable than ESCs and are not as prone to forming tumours, many studies have observed that ASCs can also form malignant tumours when transplanted *in vivo*.⁹⁶ Another consideration is the rejection of allogeneic stem cells by the host immune system.⁹⁷ The use of ASCs and tissues derived from the patient's own ASCs would mean that the cells are less likely to be rejected by the immune system. This represents a significant advantage, as immune rejection can be circumvented only by continuous administration of immunosuppressive drugs, and the drugs themselves may cause deleterious side effects. To date, nine different types of stem cells have been reported in the therapy of ED in the laboratory.

BMSCs

The preclinical applications of BMSCs in treating ED were initially based on their capacity to home to damaged tissues, differentiate into the necessary mature phenotypes, and their amenability to genetic manipulation.^{6,98}

In 2003, Deng *et al.*⁹⁹ demonstrated successful adenoviral gene transfer of endothelial nitric oxide synthase (eNOS) to *ex vivo* expanded rat BMSCs. The transfected BMSCs expressed high levels

of eNOS that persisted in culture for more than 21 days. The cells retained their multipotential differentiation capability after transduction. Moreover, intracavernous injection of eNOS-BMSCs increased the expression of eNOS in the corpus cavernosum and improved erectile function in aged rats.

In 2007, it was shown that rat BMSCs were able to reverse age-associated ED both with and without eNOS transfection through mechanisms involving increased penile eNOS, improved eNOS/cGMP signalling, and apparent differentiation into penile cells expressing endothelial and smooth muscle markers.¹⁰⁰ It also has been suggested that human BMSCs are capable of differentiating toward ECs and SMCs in the corpus cavernosum of rats.¹⁰¹ However, the efficacy of these cells for the treatment of penile erectile function should be confirmed in immunocompromised animals because human stem cells are not compatible in non-immunocompromised animals. In addition, although a broad differentiation potential of BMSCs has been observed, it may also be explained as the result of spontaneous fusion of host and donor cells;^{102,103} therefore, it is difficult to prove the differentiation.

In 2010, Kendirci and colleagues¹⁰⁴ showed that transplantation of BMSCs isolated with p75 nerve growth factor (NGF) receptor into the rat penis could rescue erectile function following cavernous nerve injury. The authors suggested that these effects were mediated by FGF, NGF, brain-derived neurotrophic factor (BDNF), VEGF and insulin-like growth factor 1 (IGF-1) secreted by the stem cells. In a rat model of diabetes mellitus type 1, Qiu *et al.*¹⁰⁵ indicated that transplantation of BMSCs restored erectile function by increasing the content of endothelium and smooth muscle in the corporal cavernosum, though they did not study the paracrine action of transplanted BMSCs or fusion between BMSCs and host smooth muscle cells or endothelial cells. For age-associated ED, BMSC transplantation improved erectile function during a long follow-up by improving eNOS/cGMP signalling and, potentially, by also differentiation into ECs and SMCs.¹⁰⁶ In 2011, a study suggested that VEGF-gene-modified BMSCs resulted in enhanced regeneration of smooth muscle and endothelium in the corpora cavernosa of type I diabetic rats.¹⁰⁷ The efficacy and safety of BMSC treatment over a long duration needs to be investigated. Unexpected side effects of VEGF caused by angiogenesis, such as angioma formation, may occur in a clinical trial.¹⁰⁸ Thus, the possible side effects of VEGF need to be assessed before a clinical trial is conducted.

It is worth emphasizing that BMSCs may not simply act by replacing lost or damaged cells. The provision of trophic factors may protect threatened cells from disease or stimulate proliferation of host progenitors. Meanwhile, they could act independently to alter the immune response to limit damage and promote repair and regeneration; thus, their immunomodulatory effects have been employed clinically in the treatment of graft-versus-host disease.¹⁰⁹ It remains controversial which mechanism plays the main role; however, an increasing number of observations support the hypothesis that BMSCs exert their effects on the host tissue by paracrine mechanisms.¹⁰⁴ The beneficial effects of BMSCs in other animal models have been observed within 3 days following transplantation, a time frame that is too small to allow for engraftment and differentiation.¹⁰⁴ Additionally, and perhaps most striking, the beneficial effects of BMSCs have been replicated using cell-free lysates and conditioned BMSC culture medium.^{110,111}

Muscle-derived stem cells (MDSCs)

MDSCs are ASCs found in muscle tissues. They are easily isolated from autologous muscle biopsies, and they pose low immunogenic and

Table 1 Application of stem cells for the therapy of erectile dysfunction

SCs type	SCs form	Model system	Study duration	Functional results	Histology and molecular results
Rat BMSCs ¹⁰²	Cell (gene modified with eNOS)	Aged rats	3 weeks	Enhanced ICP during CN electrostimulation. Better enhancement when combined with eNOS gene therapy	Enhanced eNOS expression, and cGMP levels. Transplanted cells exhibited ECs and SMCs markers
Human BMSC ¹⁰³	Cell	Young adult rats	2 weeks	No functional testing.	Transplanted cells exhibited ECs and SMCs markers
Rat BMSCs ¹⁰⁶	Cell (isolated by p75 NGF receptor)	Young adult rats with CN crush	4 weeks	Enhanced ICP during CN electrostimulation. Better enhancement for p75-derived BMSCs	p75-derived BMSCs secreted significantly more basic NGF than control group
Rat BMSCs ^{107,109}	Cell (gene modified with VEGF)	Young adult rats with DM I	4 weeks	Enhanced ICP during CN electrostimulation. Better enhancement when combined with VEGF gene therapy	Increased content of smooth muscle and endothelium in corporal cavernosum. Transplanted cells exhibited ECs and SMCs markers
Rat MDSCs ¹¹⁵	Cell	Young adult rats with CN transaction	2 weeks, 4 weeks	Enhanced ICP during CN electrostimulation	Increased percent area of PGP 9.5 staining
Mouse MDSCs ⁷⁸	Cell	Young adult rats and aged rats	2 weeks, 4 weeks	Enhanced ICP during CN electrostimulation in aged rats (2 weeks) and young adult rats (4 weeks)	Increased content of smooth muscle in corporal cavernosum. Transplanted cells exhibited SMCs markers
Rabbit MDSC ¹¹⁴	ACCMs seeded with MDSCs	Young adult rabbits	2 months, 4 months, 6 months	No functional testing	Cells expressing ECs and SMCs markers and better arranged growth were prevalent
Rat EPCs ¹³²	Cell (gene modified with VEGF)	Young adult rats with DM I	3 weeks	Enhanced ICP during CN electrostimulation	Enhanced neovascularisation in the corpora cavernosum. Transplanted cells exhibited ECs markers
Human UCBCS ¹³⁴	Cell	Senior human with DM II	11 months	Regained morning erections in some patients. Increased rigidity, but was insufficient for penetration	No relevant testing
Rat ADSCs ¹²³	Cell	Young adult rats	4 weeks	No functional testing	Injected ADSCs were localized to the sinusoid endothelium. Some Transplanted cells exhibited ECs markers
Rat ADSCs ¹²⁶	Cell	Young adult rats with hyperlipid-emia	4 weeks	Enhanced ICP during CN electrostimulation	Increased content of smooth muscle and endothelium in corporal cavernosum, and nNOS-positive nerve fibres in penile dorsal nerves
Rat ADSCs ¹²⁷	Cell	Adult ZDF rats with DM II	4 weeks	Enhanced ICP during CN electrostimulation	Increased nNOS staining area in penile dorsal nerves. Inhibition of apoptosis in corpus cavernosum
Rat ADSCs ¹²⁹	Cell and cell lysate	Young adult rats with CN crush	4 weeks	Enhanced ICP during CN electrostimulation	Increased content of smooth muscle in corporal cavernosum, and nNOS-positive nerve fibres in penile dorsal nerves. Inhibition of fibrosis and apoptosis
Human fetal NCSCs ¹³⁷	Cell	Young adult rats	2 weeks	No functional testing	Transplanted cells exhibited ECs and SMCs markers
Rat fetal BDSCs ¹³⁶	Cell	Young adult rats	6 weeks	No functional testing	Transplanted cells exhibited SMCs markers or VEGF
Rat GRPCs ¹⁴⁰	Cell	Young adult rats with spinal cord injury	12 weeks	Preserved full and partial erectile events per 24 h	GRPCs survived, differentiated and formed extensive transplants that were well integrated with host tissue
Rat neural ESCs ¹⁴¹	Cell	Young adult rats with CN crush	12 weeks	Enhanced ICP during CN electrostimulation	Increased neurofilament staining in the MPG and penile dorsal nerves. Improved morphological characteristics of nerve fibres in the corpora cavernosum

Abbreviations: ACCMs, acellular corporal collagen matrices; ADSCs, adipose tissue derived stem cells; BDSCs, brain-derived stem cells; BMSCs, bone marrow mesenchymal stem cells; CN, cavernous nerve; DM I, diabetes mellitus type I; DM II, diabetes mellitus type 2; ECs, endothelial cells; eNOS, endothelial nitric oxide synthase; EPCs, endothelial precursor stem cells; ESCs, embryonic stem cells; GRPCs, glial restricted progenitor cells; ICP, intracavernous pressure; MDSCs, muscle-derived stem cells; MPG, major pelvic ganglia; NCSCs, neural crest stem cells; nNOS, neuronal nitric oxide synthase; SCs, stem cells; SMCs, smooth muscle cells; UCBCSs, umbilical cord blood stem cells. PGP9.5, protein gene product 9.5; NGF, nerve growth factor; VEGF, vascular endothelial growth factor; ZDF, Zucker diabetic fatty.

carcinogenic risks. Their enzymatic and non-enzymatic antioxidant capacity provides them a critical property for MDSC survival post-transplantation.¹¹² This superior capacity supports their survival advantages.

In 2006, MDSCs were first injected into the corpora cavernosum to treat ED in rats with bilateral cavernous nerve injury.¹¹³ Although the increase in PGP 9.5 neuronal staining suggested that the MDSCs protected the penile nerve from atrophy after cavernous nerve transection, the mechanisms need to be interpreted in the following studies.

In 2008, the ability of MDSCs to convert into SMCs after implantation into the corpora cavernosum of aged rats was reported.⁷⁶ Exogenous MDSCs were able to correct ED, and endogenous cells also expressed stem cell markers in the control group, suggesting that exogenous stem cell implantation as well as endogenous stem cell modulation may be viable therapeutic approaches for ageing-related ED. Similar to BMSCs, when using the rat as a host, a truly allogenic source of MDSCs has to be tested to eliminate the need for pharmacological immunosuppression. Moreover, the effects of MDSCs were comparatively short-lived, and the considerable stimulation of erectile function achieved at 2 weeks was reduced at 4 weeks. Therefore, additional research needs to be conducted to improve the survival of MDSCs and ensure that the salutary functional effects could be prolonged for at least 3–6 months to exclude transient amelioration.

In 2010, acellular corporal collagen matrices seeded with MDSCs were implanted within the albuginea of rabbits.¹¹⁴ Histological analyses of the explants at all time points in the experimental group showed more cells and improved arranged growth compared to the control group. Alpha-smooth muscle actin and eNOS-positive cells were more prevalent in the experimental group. As the authors mentioned in the text, oxygen and nutrition were not available to MDSCs in the deeper regions of the acellular corporal collagen matrices because of their complex structure, resulting in some death of the transplanted MDSCs. Thus, more suitable scaffold materials should be explored.

ADSCs

ADSCs isolated from the stromal vascular fraction of adipose tissue have been recently identified and investigated for their multiple differentiation properties.¹¹⁵ ADSCs share similar properties of other stem cells, such as the ability to divide and renew themselves over long periods of time and to differentiate into specialized cells. They are much easier and safer to obtain in large quantities than BMSC,¹¹⁶ and, therefore, they appear to be a better choice for clinical application for ED regeneration medicine.

In 2006, Ning *et al.*¹¹⁷ reported that ADSCs could be induced by isobutylmethylxanthine to differentiate into neuron-like cells *via* the IGF-1 signalling pathway.¹¹⁸ The significance of these studies is that ADSCs possess the potential to treat degenerative neurological diseases, including neurogenic ED.²

In 2008, a pilot experiment was conducted that demonstrated that ADSCs improved the erectile function in the rat following bilateral cavernous nerve crush injury.¹¹⁹ Several possible mechanisms underlying the treatment are proposed, including cell incorporation and differentiation into native tissue cells versus paracrine signalling and stimulation of the host tissue to regenerate, for instance, by secretion of growth factors such as IGF-1 and GDF-5.^{116,120}

In 2009, Ning *et al.*¹²¹ first reported that ADSCs could differentiate into ECs in the penis and that this differentiation was mediated by FGF2 signalling. Previously published experimental procedures for the differentiation of ADSCs into ECs generally employed culture media that contained VEGF, IGF or FGF2, and VEGF was assumed to be the

responsible factor.¹²² Interestingly, vitamin C was also found to be essential for endothelial differentiation of ADSCs. Its role is most likely to promote the maintenance of a healthy growth environment for endothelial cells by protecting them from oxidative stress.¹²³

In 2010, a surge of reports concerning the application of ADSCs as a therapy for ED was published. First, ADSCs were injected into the penis to examine their effects on a rat model of hyperlipidemia-associated ED; elevated erectile function was demonstrated after ADSCs transplantation.¹²⁴ An increase in neuronal nitric oxide synthase (nNOS)-positive nerve fibres and ECs was observed in the experimental group compared to the control animals. The authors indicated that the underlying mechanisms involved cytokine and growth factor secretion rather than stem cell differentiation. However, they failed to indicate the specific growth factors. Another critical question is the long-term safety of ADSCs with respect to the possibility of tumour formation. Investigation of the long-term fate of ADSCs is required before human trials could be considered, although the lack of ADSCs in tissue sections observed for a 28-day follow-up period implies that the potential of these cells to persist and undergo malignant transformation is limited.¹²⁴ Second, Garcia and co-workers¹²⁵ observed improved erectile function in type-2 diabetic rats after autologous ADSC injection. This observation was based on decreased numbers of apoptotic cells in the corpus cavernosum, increased numbers of sinusoid ECs and increased expression of nNOS in the penile nerves. Because only a few of the prelabelled ADSCs were observed within the corporal tissue of the treatment group, the authors suggested that the treatment effect of the ADSCs may not be through direct transformation into local cell types, but rather, *via* a more 'indirect' mechanism, whereby ADSCs improve the extracellular environment and local tissue function within the treatment area. The biggest shortcoming of this study is that the authors did not assess the local retention or net survival of the injected ADSCs. How to improve local retention of the transplanted ADSCs is a challenge for cellular therapy. Improved local retention could lead to greater improvement in erectile function after treatment. The use of bioabsorbable PLGA microspheres is a well-established means by which to improve local retention, survival and possibly the therapeutic effect of transplanted cells.¹²⁶ Third, ADSCs and cell-free lysates derived from ADSCs were injected into the penis in a rat model of cavernous nerve injury, resulting in a significant recovery of erectile function in both treatments compared with the control group.¹²⁷ In the treated rats, nNOS and smooth muscle content were preserved, and less fibrosis occurred in the corpus cavernosum. The underlying mechanisms involved neuron preservation and cytoprotection by inhibition of apoptosis by releasing intracellular preformed substances or by active secretion of certain biomolecules. However, the authors failed to provide direct evidence to support the effects of a paracrine pathway. Moreover, further investigations should be aimed to identify the proteomic characteristics of the cell-free lysate. At last, CXCL5, which is secreted in much larger amounts by ADSCs than by control cells (penile smooth muscle cells),¹²⁸ was discovered to be capable of enhancing chemoattraction and angiogenesis,¹²⁹ promoting rat major pelvic ganglia neurite outgrowth, and activating the JAK/STAT pathway in cultured Schwann cells.¹²⁸ CXCL5 may thus contribute to ADSCs' therapeutic efficacy in cavernous nerve injury-induced ED.

EPCs

While EPCs have been evaluated for predicting vasculogenic ED as discussed above, there are few reports regarding the use of EPCs

to treat ED. In 2010, Gou's team¹³⁰ investigated intracavernosal injection of EPCs overexpressing VEGF. The results showed that modified EPCs could restore the erectile function of rats with diabetic ED by enhancing the expression of VEGF, facilitating the process of neovascularisation and incorporating into the endothelium to improve the function of ECs.

Umbilical cord blood stem cells (UCBSCs)

Umbilical cord blood is an important source of stem cells. UCBSCs are deemed to be the newest and youngest among the stem cells. They also avoid the debates surrounding ESCs because they can be obtained without destroying an embryo. Moreover, UCBSCs do not possess mutations in DNA, which are commonly found in ASCs.¹³¹ In 2010, intracavernous transplantation of human UCBSCs was reported to improve erectile function and blood glucose levels in patients with diabetic ED without immune suppression.¹³² This study is the first clinical report describing the use of unmodified stem cell therapy for the treatment of diabetic ED. However, the exact mechanisms were not elucidated and may involve as-yet-unidentified humoral factors generated by the stem cells themselves. Moreover, the total cell number that was injected in the report was quite small relative to the unit body weights compared with other studies, and the rates of implantation and differentiation of stem cells in the corpus cavernosa during the follow-up remained unknown.

Brain-derived stem cells (BDSCs)

Previous work showed the remarkable plasticity of central nervous system-derived stem cells; they have the capacity to give rise to large, flat smooth muscle-like cells that express phenotypic characteristics of SMCs.¹³³ In 2009, Song *et al.*¹³⁴ injected foetal BDSCs labelled with green fluorescent protein into penile tissue. Six weeks later, there was evidence of transdifferentiation of BDSCs into penile SMCs as the differentiated cells (30%–40%) expressed smooth muscle markers. Although the demonstration of pluripotency and confirmation of the expression of smooth muscle markers confirm that BDSCs are worthy of further examination as a potential resource for penile smooth muscle regeneration, clinical application will be limited regarding the source of these cells.

Neural crest stem cells (NCSCs)

NCSCs are the progenitor cells of cell types constituting the peripheral nervous system, including neurons, Schwann cells and adrenal chromaffin cells as well as SMCs.¹³⁵ NCSCs are closely related to neural tube cells, as both are descendants of the same higher order stem cells.¹³⁶ Importantly, similar to ASCs, intraspinal administration of NCSCs did not result in the formation of tumours or teratomas.¹³⁷ Many studies have shown that transplantation of NCSCs could induce the regeneration of connective tissues, SMCs, skeletal muscle and endothelium. In 2008, the feasibility of using human NCSCs for corpus cavernosum repair and potential differentiation of NCSCs into penile cavernous sinus ECs and SMCs was investigated.¹³⁵ Histological analysis indicated that transplanted NCSCs could possibly differentiate into ECs and SMCs, as shown by their expression of cell type-specific markers. However, we do not know whether NCSCs-based cell therapy could compromise penile erectile function *in vivo* in animals. Furthermore, the differentiation of human NCSCs is unconvincing before excluding the effects of fusion between transplanted cells and host cells. The critical limitation of NCSCs in clinical application, similar to BDSCs, is the lack of a sufficient source of these donor cells.

Glial restricted progenitor cells (GRPCs)

In 2011, Nout *et al.*¹³⁸ delivered GRPCs into a rat model of spinal cord injury, and they found that those animals regained some autonomic functions, such as a similar number of erectile events per 24 h compared to the control group, implying that GRPCs have potential therapeutic efficacy in patients with ED associated with spinal cord injury.

ESCs

Although ESCs demonstrate powerful differentiation potency, research has lagged due to legal and ethical considerations. In 2004, ESCs transfected with BDNF that had differentiated along a neuronal cell line were injected into the corpus cavernosum and found to improve cavernous nerve regeneration and functional erectile status after bilateral crush injury in the rat.¹³⁹ Stem cells may present a cellular substrate at the lesion site to support axonal extension. In addition, stem cells may not require a prolonged presence to function; their mechanism of action may be through growth factor expression (BDNF, NGF and neurotrophin-3),¹⁴⁰ inhibition of demyelination and as an initial lattice of cellular substrate.

PERSPECTIVES

Stem cells, especially the ASCs, are regarded as candidates for the diagnosis and treatment of ED due to many factors. Stem cells possess the potential to undergo long-term proliferation, self-renewal and multipotent differentiation, and they can serve as a vehicle for the release of neurotrophins, such as nerve growth factor, to repair damage caused by diabetes or pelvic surgery.

In addition to using stem cells for treatment of ED, stem cells could be used as a predictor for the diagnosis and progress of ED.

It is worth noting that except for the application of exogenous stem cells, reawakening and mobilizing endogenous stem cells could allow the prevention and treatment of ED in the near future.

Of note, while most of the literature cited in this review concerns animal studies, increasing interest is being given to translation of the observed results into clinical applications of stem cells. Positive and encouraging results have been obtained with the use of BMSC, MDSC and ADSC in various diseases.

With growing evidence from basic research on the use of stem cells for treatment of ED, in 2010, the first clinical trial was conducted in Korea.¹³² In this study, morning erection was restored within one month in three patients and within 3 months in six patients. However, despite having increased penile rigidity, none of the patients was able to achieve vaginal penetration unless aided by taking sildenafil before coitus. In 2011, another clinical trial was carried out in France with intracavernous BMSCs injected into patients with ED post-prostatectomy. The transplanted stem cells were autologous intracavernous bone marrow-mononucleated cells containing mainly MSCs, EPCs and HSCs (<http://www.clinicaltrials.gov/ct2/show/NCT01089387?term=stem±cell±and±ED&rank=1>). As stated above, a significant proportion of preclinical studies investigating the effects of stem cells on ED concern cavernous nerve injury-induced ED, a condition that we observe almost solely after pelvic surgery for malignancies. The application of stem cells in this condition poses the risk of interaction between the stem cells and prostate tumour cell lines,¹⁴¹ and it is difficult to predict the effects of this interaction *in vivo*. Thus, while animal studies are promising, translation to clinical application requires further study on how various types of stem cells and their secretions interact with tumour cells.

Furthermore, it remains largely unknown what the fate of the majority of stem cells is after injection into experimental animals. As investigators focus on penile changes after cellular therapy, the effects that this therapy has on other, non-diseased organ tissues is still a mystery in the field of sexual medicine. This issue warrants further investigations before translating these results and applying these cell types in human patients.

CONCLUSIONS

A broad range of stem cell sources has been investigated preclinically and has shown a high potential in the treatment and prevention of ED of various origins. While preclinical animal data are highly promising, attention is needed to elucidate the mechanisms of action of stem cell therapy for ED and the potential adverse events associated with stem cell application before the translation to clinical application is attempted.

COMPETING FINANCIAL INTERESTS

The authors declare no competing financial interests.

- Morrison SJ, Shah NM, Anderson DJ. Regulatory mechanisms in stem cell biology. *Cell* 1997; **88**: 287–98.
- Lin CS, Xin ZC, Deng CH, Ning H, Lin G *et al*. Recent advances in andrology-related stem cell research. *Asian J Androl* 2008; **10**: 171–5.
- Keller G. Embryonic stem cell differentiation: emergence of a new era in biology and medicine. *Genes Dev* 2005; **19**: 1129–55.
- Becker C, Jakse G. Stem cells for regeneration of urological structures. *Eur Urol* 2007; **51**: 1217–28.
- Evans MJ, Kaufman MH. Establishment in culture of pluripotential cells from mouse embryos. *Nature* 1981; **292**: 154–6.
- Strong TD, Gebaska MA, Champion HC, Burnett AL, Bivalacqua TJ. Stem and endothelial progenitor cells in erection biology. *Int J Impot Res* 2008; **20**: 243–54.
- Yamzon JL, Kokorowski P, Koh CJ. Stem cells and tissue engineering applications of the genitourinary tract. *Pediatr Res* 2008; **63**: 472–7.
- Petersen BE, Bowen WC, Patrene KD, Mars WM, Sullivan AK *et al*. Bone marrow as a potential source of hepatic oval cells. *Science* 1999; **284**: 1168–70.
- Vrana KE, Hipp JD, Goss AM, McCool BA, Riddle DR *et al*. Nonhuman primate parthenogenetic stem cells. *Proc Natl Acad Sci USA* 2003; **100**(Suppl 1): 11911–6.
- NIH Consensus Development Panel on Impotence. NIH Consensus Conference: impotence. *JAMA* 1993; **270**: 83–90.
- Burnett AL. Erectile dysfunction. *J Urol* 2006; **175**: S25–31.
- Wessells H, Williams SK. Endothelial cell transplantation into the corpus cavernosum: moving towards cell-based gene therapy. *J Urol* 1999; **162**: 2162–4.
- Martin GR. Isolation of a pluripotent cell line from early mouse embryos cultured in medium conditioned by teratocarcinoma stem cells. *Proc Natl Acad Sci USA* 1981; **78**: 7634–8.
- Yao L, Yu X, Hui N, Liu S. Application of iPS in assisted reproductive technology: sperm from somatic cells? *Stem Cell Rev* 2011; **7**: 714–21.
- Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 2006; **126**: 663–76.
- Takahashi K, Tanabe K, Ohnuki M, Narita M, Ichisaka T *et al*. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell* 2007; **131**: 861–72.
- Yu J, Vodyanik MA, Smuga-Otto K, Antosiewicz-Bourget J, Frane JL *et al*. Induced pluripotent stem cell lines derived from human somatic cells. *Science* 2007; **318**: 1917–20.
- Okita K, Nakagawa M, Hyenjong H, Ichisaka T, Yamanaka S. Generation of mouse induced pluripotent stem cells without viral vectors. *Science* 2008; **322**: 949–53.
- Stadtfield M, Nagaya M, Utikal J, Weir G, Hochedlinger K. Induced pluripotent stem cells generated without viral integration. *Science* 2008; **322**: 945–9.
- Woltjen K, Michael IP, Mohseni P, Desai R, Mileikovsky M *et al*. piggyBac transposition reprograms fibroblasts to induced pluripotent stem cells. *Nature* 2009; **458**: 766–70.
- Yu J, Hu K, Smuga-Otto K, Tian S, Stewart R *et al*. Human induced pluripotent stem cells free of vector and transgene sequences. *Science* 2009; **324**: 797–801.
- Zhou H, Wu S, Joo JY, Zhu S, Han DW *et al*. Generation of induced pluripotent stem cells using recombinant proteins. *Cell Stem Cell* 2009; **4**: 381–4.
- Chin MH, Mason MJ, Xie W, Volinia S, Singer M *et al*. Induced pluripotent stem cells and embryonic stem cells are distinguished by gene expression signatures. *Cell Stem Cell* 2009; **5**: 111–23.
- Spradling A, Drummond-Barbosa D, Kai T. Stem cells find their niche. *Nature* 2001; **414**: 98–104.
- Presnell SC, Petersen B, Heidarman M. Stem cells in adult tissues. *Semin Cell Dev Biol* 2002; **13**: 369–76.
- Hipp J, Atala A. Sources of stem cells for regenerative medicine. *Stem Cell Rev* 2008; **4**: 3–11.
- Jiang Y, Vaessen B, Lenvik T, Blackstad M, Reyes M *et al*. Multipotent progenitor cells can be isolated from postnatal murine bone marrow, muscle, and brain. *Exp Hematol* 2002; **30**: 896–904.
- Schaffler A, Buchler C. Concise review: adipose tissue-derived stromal cells—basic and clinical implications for novel cell-based therapies. *Stem Cells* 2007; **25**: 818–27.
- Joyce N, Annett G, Wirthlin L, Olson S, Bauer G *et al*. Mesenchymal stem cells for the treatment of neurodegenerative disease. *Regen Med* 2010; **5**: 933–46.
- Erices A, Conget P, Minguell JJ. Mesenchymal progenitor cells in human umbilical cord blood. *Br J Haematol* 2000; **109**: 235–42.
- Phinney DG, Prockop DJ. Concise review: mesenchymal stem/multipotent stromal cells: the state of transdifferentiation and modes of tissue repair—current views. *Stem Cells* 2007; **25**: 2896–902.
- Gronthos S, Mankani M, Brahimi J, Robey PG, Shi S. Postnatal human dental pulp stem cells (DPSCs) *in vitro* and *in vivo*. *Proc Natl Acad Sci USA* 2000; **97**: 13625–30.
- Williams JT, Southerland SS, Souza J, Calcutt AF, Cartledge RG. Cells isolated from adult human skeletal muscle capable of differentiating into multiple mesodermal phenotypes. *Am Surg* 1999; **65**: 22–6.
- Rodriguez AM, Elabd C, Amri EZ, Ailhaud G, Dani C. The human adipose tissue is a source of multipotent stem cells. *Biochimie* 2005; **87**: 125–8.
- McCulloch EA, Till JE. Proliferation of hemopoietic colony-forming cells transplanted into irradiated mice. *Radiat Res* 1964; **22**: 383–97.
- Till JE, McCulloch EA, Siminovitch L. A stochastic model of stem cell proliferation, based on the growth of spleen colony-forming cells. *Proc Natl Acad Sci USA* 1964; **51**: 29–36.
- Lin G, Garcia M, Ning H, Banie L, Guo YL *et al*. Defining stem and progenitor cells within adipose tissue. *Stem Cells Dev* 2008; **17**: 1053–63.
- Kiefer JC. Primer and interviews: the dynamic stem cell niche. *Dev Dyn* 2011; **240**: 737–43.
- Becerra J, Santos-Ruiz L, Andrades JA, Mari-Beffa M. The stem cell niche should be a key issue for cell therapy in regenerative medicine. *Stem Cell Rev* 2011; **7**: 248–55.
- Jones DL, Wagers AJ. No place like home: anatomy and function of the stem cell niche. *Nat Rev Mol Cell Biol* 2008; **9**: 11–21.
- Song X, Wong MD, Kawase E, Xi R, Ding BC *et al*. Bmp signals from niche cells directly repress transcription of a differentiation-promoting gene, bag of marbles, in germline stem cells in the *Drosophila* ovary. *Development* 2004; **131**: 1353–64.
- Blanpain C, Lowry WE, Geoghegan A, Polak L, Fuchs E. Self-renewal, multipotency, and the existence of two cell populations within an epithelial stem cell niche. *Cell* 2004; **118**: 635–48.
- Jiang H, Patel PH, Kohlmaier A, Grenley MO, McEwen DG *et al*. Cytokine/Jak/Stat signaling mediates regeneration and homeostasis in the *Drosophila* midgut. *Cell* 2009; **137**: 1343–55.
- Ding BS, Nolan DJ, Butler JM, James D, Babazadeh AO *et al*. Inductive angiocrine signals from sinusoidal endothelium are required for liver regeneration. *Nature* 2010; **468**: 310–5.
- Voog J, Jones DL. Stem cells and the niche: a dynamic duo. *Cell Stem Cell* 2010; **6**: 103–15.
- Zhang J, Niu C, Ye L, Huang H, He X *et al*. Identification of the haematopoietic stem cell niche and control of the niche size. *Nature* 2003; **425**: 836–41.
- Kiel MJ, Yilmaz OH, Iwashita T, Yilmaz OH, Therost C *et al*. SLAM family receptors distinguish hematopoietic stem and progenitor cells and reveal endothelial niches for stem cells. *Cell* 2005; **121**: 1109–21.
- Calvi LM, Adams GB, Weibrecht KW, Weber JM, Olson DP *et al*. Osteoblastic cells regulate the haematopoietic stem cell niche. *Nature* 2003; **425**: 841–6.
- Wright DE, Wagers AJ, Gulati AP, Johnson FL, Weissman IL. Physiological migration of hematopoietic stem and progenitor cells. *Science* 2001; **294**: 1933–6.
- Corselli M, Chen CW, Crisan M, Lazzari L, Peault B. Perivascular ancestors of adult multipotent stem cells. *Arterioscler Thromb Vasc Biol* 2010; **30**: 1104–9.
- Ergun S, Tilki D, Klein D. Vascular wall as a reservoir for different types of stem and progenitor cells. *Antioxid Redox Signal* 2011; **15**: 981–95.
- Garrett RW, Emerson SG. Bone and blood vessels: the hard and the soft of hematopoietic stem cell niches. *Cell Stem Cell* 2009; **4**: 503–6.
- Barker N, van Es JH, Kuipers J, Kujala P, van den Born M *et al*. Identification of stem cells in small intestine and colon by marker gene *Lgr5*. *Nature* 2007; **449**: 1003–7.
- Palmer TD, Willhoite AR, Gage FH. Vascular niche for adult hippocampal neurogenesis. *J Comp Neurol* 2000; **425**: 479–94.
- Yoshida S, Sukeno M, Nabeshima Y. A vasculature-associated niche for undifferentiated spermatogonia in the mouse testis. *Science* 2007; **317**: 1722–6.
- Morrison SJ, Spradling AC. Stem cells and niches: mechanisms that promote stem cell maintenance throughout life. *Cell* 2008; **132**: 598–611.
- Khalidoyanidi S. Directing stem cell homing. *Cell Stem Cell* 2008; **2**: 198–200.
- Sackstein R, Merzaban JS, Cain DW, Dagia NM, Spencer JA *et al*. *Ex vivo* glycan engineering of CD44 programs human multipotent mesenchymal stromal cell trafficking to bone. *Nat Med* 2008; **14**: 181–7.
- Wright N, Hidalgo A, Rodriguez-Frade JM, Soriano SF, Mellado M *et al*. The chemokine stromal cell-derived factor-1 alpha modulates alpha 4 beta 7 integrin-mediated lymphocyte adhesion to mucosal addressin cell adhesion molecule-1 and fibronectin. *J Immunol* 2002; **168**: 5268–77.
- Lapidot T, Dar A, Kollet O. How do stem cells find their way home? *Blood* 2005; **106**: 1901–10.

- 61 Peled A, Kollet O, Ponomaryov T, Petit I, Franitza S *et al*. The chemokine SDF-1 activates the integrins LFA-1, VLA-4, and VLA-5 on immature human CD34⁺ cells: role in transendothelial/stromal migration and engraftment of NOD/SCID mice. *Blood* 2000; **95**: 3289–96.
- 62 Bonig H, Priestley GV, Papayannopoulou T. Hierarchy of molecular-pathway usage in bone marrow homing and its shift by cytokines. *Blood* 2006; **107**: 79–86.
- 63 Heissig B, Hattori K, Dias S, Friedrich M, Ferris B *et al*. Recruitment of stem and progenitor cells from the bone marrow niche requires MMP-9 mediated release of kit-ligand. *Cell* 2002; **109**: 625–37.
- 64 Cottle-Fox MH, Lapidot T, Petit I, Kollet O, DiPersio JF *et al*. Stem cell mobilization. *Hematology Am Soc Hematol Educ Program* 2003; **2003**: 419–37.
- 65 Bensinger W, DiPersio JF, McCarty JM. Improving stem cell mobilization strategies: future directions. *Bone Marrow Transplant* 2009; **43**: 181–95.
- 66 Craddock CF, Nakamoto B, Andrews RG, Priestley GV, Papayannopoulou T. Antibodies to VLA4 integrin mobilize long-term repopulating cells and augment cytokine-induced mobilization in primates and mice. *Blood* 1997; **90**: 4779–88.
- 67 Papayannopoulou T, Priestley GV, Nakamoto B, Zafirooulos V, Scott LM *et al*. Synergistic mobilization of hemopoietic progenitor cells using concurrent beta1 and beta2 integrin blockade or beta2-deficient mice. *Blood* 2001; **97**: 1282–8.
- 68 Nakamura Y, Tajima F, Ishiga K, Yamazaki H, Oshimura M *et al*. Soluble c-kit receptor mobilizes hematopoietic stem cells to peripheral blood in mice. *Exp Hematol* 2004; **32**: 390–6.
- 69 Toma JG, Akhavan M, Fernandes KJ, Barnabe-Heider F, Sadikot A *et al*. Isolation of multipotent adult stem cells from the dermis of mammalian skin. *Nat Cell Biol* 2001; **3**: 778–84.
- 70 Joannides A, Gaughwin P, Schwiening C, Majed H, Sterling J *et al*. Efficient generation of neural precursors from adult human skin: astrocytes promote neurogenesis from skin-derived stem cells. *Lancet* 2004; **364**: 172–8.
- 71 Hunt DP, Morris PN, Sterling J, Anderson JA, Joannides A *et al*. A highly enriched niche of precursor cells with neuronal and glial potential within the hair follicle dermal papilla of adult skin. *Stem Cells* 2008; **26**: 163–72.
- 72 Toma JG, McKenzie IA, Bagli D, Miller FD. Isolation and characterization of multipotent skin-derived precursors from human skin. *Stem Cells* 2005; **23**: 727–37.
- 73 Bartsch G, Yoo JJ, de Coppi P, Siddiqui MM, Schuch G *et al*. Propagation, expansion, and multilineage differentiation of human somatic stem cells from dermal progenitors. *Stem Cells Dev* 2005; **14**: 337–48.
- 74 Chen FG, Zhang WJ, Bi D, Liu W, Wei X *et al*. Clonal analysis of nestin⁺ vimentin⁺ multipotent fibroblasts isolated from human dermis. *J Cell Sci* 2007; **120**: 2875–83.
- 75 Vernet D, Nolazco G, Cantini L, Magee TR, Qian A *et al*. Evidence that osteogenic progenitor cells in the human tunica albuginea may originate from stem cells: implications for Peyronie disease. *Biol Reprod* 2005; **73**: 1199–210.
- 76 Nolazco G, Kovanecz I, Vernet D, Gelfand RA, Tsao J *et al*. Effect of muscle-derived stem cells on the restoration of corpora cavernosa smooth muscle and erectile function in the aged rat. *BJU Int* 2008; **101**: 1156–64.
- 77 Vernet D, Heydarkhan S, Kovanecz I, Lue YH, Rajfer J *et al*. Characterization of endogenous stem cells from the mouse penis that express an embryonic stem cell gene and undergo differentiation into several cell lineages. *J Urol* 2009; **181**(Suppl): 43.
- 78 Widlansky ME, Gokce N, Keane JF Jr, Vita JA. The clinical implications of endothelial dysfunction. *J Am Coll Cardiol* 2003; **42**: 1149–60.
- 79 Sugiyama S, Kugiyama K, Aikawa M, Nakamura S, Ogawa H *et al*. Hypochlorous acid, a macrophage product, induces endothelial apoptosis and tissue factor expression: involvement of myeloperoxidase-mediated oxidant in plaque erosion and thrombogenesis. *Arterioscler Thromb Vasc Biol* 2004; **24**: 1309–14.
- 80 La Vignera S, Condorelli R, Vicari E, D'Agata R, Calogero A. New immunophenotype of blood endothelial progenitor cells and endothelial microparticles in patients with arterial erectile dysfunction and late onset hypogonadism. *J Androl* 2011; **32**: 509–17.
- 81 Asahara T, Murohara T, Sullivan A, Silver M, van der Zee R *et al*. Isolation of putative progenitor endothelial cells for angiogenesis. *Science* 1997; **275**: 964–7.
- 82 Foresta C, Caretta N, Lana A, Cabrelle A, Palu G *et al*. Circulating endothelial progenitor cells in subjects with erectile dysfunction. *Int J Impot Res* 2005; **17**: 288–90.
- 83 Eguchi M, Masuda H, Asahara T. Endothelial progenitor cells for postnatal vasculogenesis. *Clin Exp Nephrol* 2007; **11**: 18–25.
- 84 Ingram DA, Mead LE, Moore DB, Woodard W, Fenoglio A *et al*. Vessel wall-derived endothelial cells rapidly proliferate because they contain a complete hierarchy of endothelial progenitor cells. *Blood* 2005; **105**: 2783–6.
- 85 Foresta C, Ferlin A, de Toni L, Lana A, Vinanzi C *et al*. Circulating endothelial progenitor cells and endothelial function after chronic Tadalafil treatment in subjects with erectile dysfunction. *Int J Impot Res* 2006; **18**: 484–8.
- 86 Foresta C, Caretta N, Lana A, de Toni L, Biagioli A *et al*. Relationship between vascular damage degrees and endothelial progenitor cells in patients with erectile dysfunction: effect of vardenafil administration and PDE5 expression in the bone marrow. *Eur Urol* 2007; **51**: 1411–7; discussion 1417–9.
- 87 Fadini GP, de Kreutzenberg SV, Coracina A, Baesso I, Agostini C *et al*. Circulating CD34⁺ cells, metabolic syndrome, and cardiovascular risk. *Eur Heart J* 2006; **27**: 2247–55.
- 88 Baumhake M, Werner N, Bohm M, Nickenig G. Circulating endothelial progenitor cells correlate with erectile function in patients with coronary heart disease. *Eur Heart J* 2006; **27**: 2184–8.
- 89 Foresta C, de Toni L, Biagioli A, Ganz F, Magagna S *et al*. Increased levels of osteocalcin-positive endothelial progenitor cells in patients affected by erectile dysfunction and cavernous atherosclerosis. *J Sex Med* 2010; **7**: 751–7.
- 90 Matsumoto T, Kawamoto A, Kuroda R, Ishikawa M, Mifune Y *et al*. Therapeutic potential of vasculogenesis and osteogenesis promoted by peripheral blood CD34-positive cells for functional bone healing. *Am J Pathol* 2006; **169**: 1440–57.
- 91 Werner N, Kosiol S, Schiegl T, Ahlers P, Walenta K *et al*. Circulating endothelial progenitor cells and cardiovascular outcomes. *N Engl J Med* 2005; **353**: 999–1007.
- 92 Kendirci M, Bejima J, Hellstrom WJ. Update on erectile dysfunction in prostate cancer patients. *Curr Opin Urol* 2006; **16**: 186–95.
- 93 Aversa A, Bruzziches R, Vitale C, Marazzi G, Francomano D *et al*. Chronic sildenafil in men with diabetes and erectile dysfunction. *Expert Opin Drug Metab Toxicol* 2007; **3**: 451–64.
- 94 Gonzalez-Cadavid NF, Rajfer J. Molecular pathophysiology and gene therapy of aging-related erectile dysfunction. *Exp Gerontol* 2004; **39**: 1705–12.
- 95 Strong TD, Gebaska MA, Burnett AL, Champion HC, Bivalacqua TJ. Endothelium-specific gene and stem cell-based therapy for erectile dysfunction. *Asian J Androl* 2008; **10**: 14–22.
- 96 Jeong JO, Han JW, Kim JM, Cho HJ, Park C *et al*. Malignant tumor formation after transplantation of short-term cultured bone marrow mesenchymal stem cells in experimental myocardial infarction and diabetic neuropathy. *Circ Res* 2011; **108**: 1340–7.
- 97 Li SC, Zhong JF. Twisting immune responses for allogeneic stem cell therapy. *World J Stem Cells* 2009; **1**: 30–5.
- 98 Jiang W, Ma A, Wang T, Han K, Liu Y *et al*. Homing and differentiation of mesenchymal stem cells delivered intravenously to ischemic myocardium *in vivo*: a time-series study. *Pflugers Arch* 2006; **453**: 43–52.
- 99 Deng W, Bivalacqua TJ, Chattergoon NN, Hyman AL, Jeter JR Jr *et al*. Adenoviral gene transfer of eNOS: high-level expression in *ex vivo* expanded marrow stromal cells. *Am J Physiol Cell Physiol* 2003; **285**: C1322–9.
- 100 Bivalacqua TJ, Deng W, Kendirci M, Usta MF, Robinson C *et al*. Mesenchymal stem cells alone or *ex vivo* gene modified with endothelial nitric oxide synthase reverse age-associated erectile dysfunction. *Am J Physiol Heart Circ Physiol* 2007; **292**: H1278–90.
- 101 Song YS, Lee HJ, Park IH, Kim WK, Ku JH *et al*. Potential differentiation of human mesenchymal stem cell transplanted in rat corpus cavernosum toward endothelial or smooth muscle cells. *Int J Impot Res* 2007; **19**: 378–85.
- 102 Terada N, Hamazaki T, Oka M, Hoki M, Mastalerz DM *et al*. Bone marrow cells adopt the phenotype of other cells by spontaneous cell fusion. *Nature* 2002; **416**: 542–5.
- 103 Ying QL, Nichols J, Evans EP, Smith AG. Changing potency by spontaneous fusion. *Nature* 2002; **416**: 545–8.
- 104 Kendirci M, Trost L, Bakondi B, Whitney MJ, Hellstrom WJ *et al*. Transplantation of nonhematopoietic adult bone marrow stem/progenitor cells isolated by p75 nerve growth factor receptor into the penis rescues erectile function in a rat model of cavernous nerve injury. *J Urol* 2010; **184**: 1560–6.
- 105 Qiu X, Lin H, Wang Y, Yu W, Chen Y *et al*. Intracavernous transplantation of bone marrow-derived mesenchymal stem cells restores erectile function of streptozocin-induced diabetic rats. *J Sex Med* 2010; **8**: 427–36.
- 106 Abdel Aziz MT, El-Haggag S, Mostafa T, Atta H, Fouad H *et al*. Effect of mesenchymal stem cell penile transplantation on erectile signaling of aged rats. *Andrologia* 2010; **42**: 187–92.
- 107 Qiu X, Sun C, Yu W, Lin H, Sun Z *et al*. Combined strategy of mesenchymal stem cells injection with VEGF gene therapy for the treatment of diabetes associated erectile dysfunction. *J Androl* 2011; e-pub ahead of print 10 February 2011; doi:10.2164/jandrol.110.012666.
- 108 Carmeliet P. VEGF gene therapy: stimulating angiogenesis or angioma-genesis? *Nat Med* 2000; **6**: 1102–3.
- 109 Ringden O, Uzunel M, Rasmusson I, Remberger M, Sundberg B *et al*. Mesenchymal stem cells for treatment of therapy-resistant graft-versus-host disease. *Transplantation* 2006; **81**: 1390–7.
- 110 Yeghiazarians Y, Zhang Y, Prasad M, Shih H, Saini SA *et al*. Injection of bone marrow cell extract into infarcted hearts results in functional improvement comparable to intact cell therapy. *Mol Ther* 2009; **17**: 1250–6.
- 111 Crisostomo PR, Markel TA, Wang Y, Meldrum DR. Surgically relevant aspects of stem cell paracrine effects. *Surgery* 2008; **143**: 577–81.
- 112 Drowley L, Okada M, Beckman S, Vella J, Keller B *et al*. Cellular antioxidant levels influence muscle stem cell therapy. *Mol Ther* 2010; **18**: 1865–73.
- 113 Kim Y, de Miguel F, Usiene I, Kwon D, Yoshimura N *et al*. Injection of skeletal muscle-derived cells into the penis improves erectile function. *Int J Impot Res* 2006; **18**: 329–34.
- 114 Ji C, Min F, Liang W, Chen Y, Pan S *et al*. Construction of tissue-engineered corpus cavernosum with muscle-derived stem cells and transplantation *in vivo*. *BJU Int* 2010; **107**: 1638–46.
- 115 Zuk PA, Zhu M, Ashjian P, de Ugarte DA, Huang JI *et al*. Human adipose tissue is a source of multipotent stem cells. *Mol Biol Cell* 2002; **13**: 4279–95.
- 116 Lin G, Banie L, Ning H, Bella AJ, Lin CS *et al*. Potential of adipose-derived stem cells for treatment of erectile dysfunction. *J Sex Med* 2009; **6**(Suppl 3): 320–7.
- 117 Ning H, Lin G, Lue TF, Lin CS. Neuron-like differentiation of adipose tissue-derived stromal cells and vascular smooth muscle cells. *Differentiation* 2006; **74**: 510–8.
- 118 Ning H, Lin G, Fandel T, Banie L, Lue TF *et al*. Insulin growth factor signaling mediates neuron-like differentiation of adipose-tissue-derived stem cells. *Differentiation* 2008; **76**: 488–94.
- 119 Anthony J, Bella T, Lin G, Phonsombat S, Lin CS, *et al*. Non-cell line induced autologous adult adipose tissue derived stem cells enhance recovery of erectile function in the rat following bilateral cavernous nerve crush injury. *Sex Med Soc North Am Meet* 2008; **68**: Abstract.

- 120 Fandel TM, Bella AJ, Lin G, Tantiwongse K, Lin CS *et al*. Intracavernous growth differentiation factor-5 therapy enhances the recovery of erectile function in a rat model of cavernous nerve injury. *J Sex Med* 2008; **5**: 1866–75.
- 121 Ning H, Liu G, Lin G, Yang R, Lue TF *et al*. Fibroblast growth factor 2 promotes endothelial differentiation of adipose tissue-derived stem cells. *J Sex Med* 2009; **6**: 967–79.
- 122 DiMuzio P, Tulenko T. Tissue engineering applications to vascular bypass graft development: the use of adipose-derived stem cells. *J Vasc Surg* 2007; **45**(Suppl A): A99–103.
- 123 Smith AR, Visioli F, Hagen TM. Vitamin C matters: increased oxidative stress in cultured human aortic endothelial cells without supplemental ascorbic acid. *FASEB J* 2002; **16**: 1102–4.
- 124 Huang YC, Ning H, Shindel AW, Fandel TM, Lin G *et al*. The effect of intracavernous injection of adipose tissue-derived stem cells on hyperlipidemia-associated erectile dysfunction in a rat model. *J Sex Med* 2010; **7**: 1391–400.
- 125 Garcia MM, Fandel TM, Lin G, Shindel AW, Banie L *et al*. Treatment of erectile dysfunction in the obese type 2 diabetic ZDF rat with adipose tissue-derived stem cells. *J Sex Med* 2010; **7**: 89–98.
- 126 Kang SW, Seo SW, Choi CY, Kim BS. Porous poly(lactic-co-glycolic acid) microsphere as cell culture substrate and cell transplantation vehicle for adipose tissue engineering. *Tissue Eng Part C Methods* 2008; **14**: 25–34.
- 127 Albersen M, Fandel TM, Lin G, Wang G, Banie L *et al*. Injections of adipose tissue-derived stem cells and stem cell lysate improve recovery of erectile function in a rat model of cavernous nerve injury. *J Sex Med* 2010; **7**: 3331–40.
- 128 Zhang H, Yang R, Wang Z, Lin G, Lue TF *et al*. Adipose tissue-derived stem cells secrete CXCL5 cytokine with neurotrophic effects on cavernous nerve regeneration. *J Sex Med* 2011; **8**: 437–46.
- 129 Zhang H, Ning H, Banie L, Wang G, Lin G *et al*. Adipose tissue-derived stem cells secrete CXCL5 cytokine with chemoattractant and angiogenic properties. *Biochem Biophys Res Commun* 2010; **402**: 560–4.
- 130 Gou X, He WY, Xiao MZ, Qiu M, Wang M *et al*. Transplantation of endothelial progenitor cells transfected with VEGF165 to restore erectile function in diabetic rats. *Asian J Androl* 2010; **13**: 332–8.
- 131 Rube CE, Fricke A, Widmann TA, Furst T, Madry H *et al*. Accumulation of DNA damage in hematopoietic stem and progenitor cells during human aging. *PLoS One* 2011; **6**: e17487.
- 132 Bahk JY, Jung JH, Han H, Min SK, Lee YS. Treatment of diabetic impotence with umbilical cord blood stem cell intracavernosal transplant: preliminary report of 7 cases. *Exp Clin Transplant* 2010; **8**: 150–60.
- 133 Tsai RY, McKay RD. Cell contact regulates fate choice by cortical stem cells. *J Neurosci* 2000; **20**: 3725–35.
- 134 Song Y, Mehta N, Sheh B, Saljoogue F, U HS *et al*. Transdifferentiation of rat fetal brain stem cells into penile smooth muscle cells. *BJU Int* 2009; **104**: 257–62.
- 135 Song YS, Lee HJ, Park IH, Lim IS, Ku JH *et al*. Human neural crest stem cells transplanted in rat penile corpus cavernosum to repair erectile dysfunction. *BJU Int* 2008; **102**: 220–4; discussion 224.
- 136 Mujtaba T, Mayer-Proschel M, Rao MS. A common neural progenitor for the CNS and PNS. *Dev Biol* 1998; **200**: 1–15.
- 137 Sieber-Blum M, Schnell L, Grim M, Hu YF, Schneider R *et al*. Characterization of epidermal neural crest stem cell (EPI-NCSC) grafts in the lesioned spinal cord. *Mol Cell Neurosci* 2006; **32**: 67–81.
- 138 Nout YS, Culp E, Schmidt MH, Tovar CA, Proschel C *et al*. Glial restricted precursor cell transplant with cyclic adenosine monophosphate improved some autonomic functions but resulted in a reduced graft size after spinal cord contusion injury in rats. *Exp Neurol* 2011; **227**: 159–71.
- 139 Bochinski D, Lin GT, Nunes L, Carrion R, Rahman N *et al*. The effect of neural embryonic stem cell therapy in a rat model of cavernosal nerve injury. *BJU Int* 2004; **94**: 904–9.
- 140 Lu P, Jones LL, Snyder EY, Tuszynski MH. Neural stem cells constitutively secrete neurotrophic factors and promote extensive host axonal growth after spinal cord injury. *Exp Neurol* 2003; **181**: 115–29.
- 141 Lin G, Yang R, Banie L, Wang G, Ning H *et al*. Effects of transplantation of adipose tissue-derived stem cells on prostate tumor. *Prostate* 2010; **70**: 1066–73.