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Suppression of hypothalamic-pituitary-adrenal axis by acute heroin challenge in rats during acute and chronic withdrawal from chronic heroin administration

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Abstract

It is known that heroin dependence and withdrawal are associated with changes in the hypothalamic-pituitary-adrenal (HPA) axis. The objective of these studies in rats was to systematically investigate the level of HPA activity and response to a heroin challenge at two time points during heroin withdrawal, and to characterize the expression of associated stress-related genes 30 minutes after each heroin challenge. Rats received chronic (10-day) intermittent escalating-dose heroin administration ($3 \times 2.5 \text{ mg/kg/day}$ on day 1; $3 \times 20 \text{ mg/kg/day}$ by day 10). Hormonal and neurochemical assessments were performed in acute (12 hours after last heroin injection) and chronic (10 days after the last injection) withdrawal. Both plasma ACTH and corticosterone levels were elevated during acute withdrawal, and heroin challenge at 20 mg/kg (the last dose of chronic escalation) at this time point attenuated this HPA hyperactivity. During chronic withdrawal, HPA hormonal levels returned to baseline, but heroin challenge at 5 mg/kg decreased ACTH levels. In contrast, this dose of heroin challenge stimulated the HPA axis in heroin naïve rats. In the anterior pituitary, pro-opiomelanocortin (POMC) mRNA levels were increased during acute withdrawal and retuned to control levels after chronic withdrawal. In the medial hypothalamus, however, the POMC mRNA levels were decreased during acute withdrawal, and increased after chronic withdrawal. Our results suggest a long-lasting change in HPA abnormal responsivity during chronic heroin withdrawal.

Keywords

POMC; orexin; mu opioid receptor; gene expression; CRF; AVP

Introduction

Heroin, morphine and other short-acting opiates regulate the activity of endogenous opioid systems. In humans, long-term exposure to opiates leads to relative deficiency in betaendorphin/mu opioid receptor (MOP-r) system [1]. Pro-opiomelanocortin (POMC) is a large peptide precursor that gives rise to several biologically active neuropeptides, including betaendorphin and ACTH. In the hypothalamic-pituitary-adrenal (HPA) axis, stress increases both corticotropin-releasing factor (CRF) and arginine vasopressin (AVP) release into the pituitary portal circulation from terminals of hypothalamic paraventricular nucleus. Both

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CRF type I receptor (CRF-R1) and AVP V1b receptor (V1b) are located on corticotropes in the anterior pituitary and drive the processing and release of the POMC peptides. POMC neurons are also identified in the rodent hypothalamus (arcuate nucleus) and some extra-hypothalamic brain regions [2,3]. During spontaneous or naloxone-precipitated opiate withdrawal, the presumed relative deficiency in endogenous beta-endorphin could lead to a hyperactive HPA axis and corresponding increases in hormone secretion. During acute opiate withdrawal, increases in HPA hormonal levels have been observed both in humans [4-7] and rats [8-10]. Furthermore, long-lasting alterations in HPA responses to various stressors may play an important role in continued vulnerability to relapse during chronic withdrawal [11,12].

Although early studies reported a lack of effect on beta-endorphin-immunoreactivity levels in many regions of the rodent brain [13,14], studies of POMC gene expression indicated that chronic morphine exposure causes significant decreases in mRNA levels in the hypothalamus [15,16]. In the arcuate nucleus, the MOP-r is a pre-synaptic autoreceptor in beta-endorphin neurosecretory neurons that negatively modulates the release of betaendorphin from these cells [17]. Furthermore, in the lateral hypothalamus, about 50% of orexin/dynorphin (Dyn) neurons express MOP-r [18], and these are under tonic inhibition by endogenous opioids [19]. Recently, it has been demonstrated that early withdrawal from opiates is associated with increased orexin mRNA expression in this region [10,18]. Together, these several hypothalamic stress responsive systems may be important neurobiological substrates of intense cravings and heightened relapse vulnerability observed in heroin dependent individuals during acute withdrawal [20].

The present experiments were designed to further characterize the activity of the HPA axis during opiate withdrawal from chronic (10-day) escalating-dose heroin administration. This pattern of opiate exposure has been found to stimulate HPA activity during acute spontaneous withdrawal in rats [21]. The first aim was to investigate the effects on HPA activity of a heroin challenge at two phases of heroin withdrawal: (1) acute 12-hour withdrawal, when elevations of plasma ACTH and corticosterone (CORT) levels were found; and (2) chronic 10-day withdrawal, when plasma ACTH and CORT levels had returned to basal levels. We examined expression levels of several HPA-related genes at both hypothalamic and pituitary levels, including CRF, CRF type I receptor (CRF-R1), AVP, AVP V1b receptor (V1b), and POMC, as measured quantitatively by solution hybridization ribonuclease protection assays. The second aim of these experiments was to investigate alterations in another set of stress responsive genes in the hypothalamus and amygdala, including MOP-r, orexin and Dyn.

Methods and materials

1. Animals

Male Fischer rats (190-220 g, Charles River, Kingston, NY) were housed individually in a stress-minimized facility with free access to water and food. Animals were adapted to a standard 12-hr light/dark cycle (lights off from 7:00 h to 19:00 h) for seven days. All experiments followed the Principles of Laboratory Animal Care (NIH Publication No 86-23, 1996), and the specific protocols were approved by the Rockefeller University Animal Care and Use Committee.

2. Procedures

Animals were randomly assigned to two groups (chronic heroin and chronic saline control), and given either heroin or an equal volume of saline (1 ml/kg) three times daily with two 6-hour intervals and one 12-hour interval, beginning 4 hours after the start of their daily dark

cycle (11:00 h, 17:00 h and 23:00 h) for 10 days. The regimen of chronic (10-day) escalating-dose heroin administration included a dose increase every second day (see **Figure 1**): 7.5 (3×2.5) mg/kg/day on days 1-2, 15 (3×5.0) mg/kg/day on days 3-4, 30 (3×10) mg/kg/day on days 5-6, 45 (3×15) mg/kg/day on days 7-8 and 60 (3×20) mg/kg/day on days 9-10. This pattern of opiate exposure using morphine or heroin has been found to induce physiological dependence in rats [10,22].

In Experiment I, animals in each chronic heroin and saline group were randomly assigned to receive a heroin challenge (HC) or vehicle injection on experimental day 11, at 11:00 h (see **Figure 1**). This created four treatment groups (n = 8-9 each): chronic heroin and 20 mg/kg heroin challenge (Chronic heroin + HC); chronic saline and saline challenge (Chronic saline + Sal); chronic heroin and saline challenge (Chronic heroin + HC). The heroin challenge dose for this latter group had to be reduced because 20 mg/kg would be too high in opiate-naïve animals and result in overdose. All animals were sacrificed 30 min following the heroin (or saline) injection by decapitation after brief exposure to CO₂ (within 15 seconds), and specific brain and pituitary regions, as well as plasma, were collected for subsequent mRNA and hormonal analyses.

Experiment II was performed exactly as described above, but the heroin and saline challenge injections were administered after <u>10 days of withdrawal (i.e., experimental day 21)</u> (**Figure 1**). All animals were sacrificed 30 min following the challenge injection and specific brain regions, pituitary and plasma were collected for subsequent mRNA and hormonal analyses.

3. Preparation of RNA extracts

In each experiment, brains were removed from the skull and placed in a chilled rat brain matrix (ASI Instruments, Houston, Texas). A coronal slice containing the hypothalamus and/or amygdala was removed from the matrix and placed on a chilled petri dish. Dissection was carried out using razor blades and forceps under a dissecting microscope. The brain regions of interest were identified according to The rat brain in stereotaxic coordinates (Paxinos G & Watson C, 1989), as described in detail recently [21]. The lateral hypothalamus (LH), medial portion of the hypothalamus (MH, including the perifornical area, dorsomedial hypothalamus, PVN and arcuate nucleus), amygdala (Amy, including the central nucleus, medial nucleus and basolateral portion) and anterior pituitary (AP) were dissected on ice, homogenized in guanidinium thiocyanate buffer and extracted with acidic phenol and chloroform. After the final ethanol precipitation step, each extract was resuspended in diethylpyrocarbonate (DEPC)-treated H₂O and stored at -80 °C.

4. Solution hybridization ribonuclease (RNase) protection-trichloroacetic acid (TCA) precipitation assay

The solution hybridization RNase protection-TCA precipitation protocol has been described in detail in earlier reports [21]. A 538 base pair (bp) fragment from the rat POMC cDNA, a 2100 bp fragment from the rat MOP-r cDNA, a 1700 bp fragment from the rat Dyn cDNA or a 760 bp fragment from the rat CRF cDNA was cloned into the polylinker region of either pSP64 or pSP65 plasmids (Promega, Madison, WI) in both the sense and antisense orientations. A 531 bp fragment from the rat orexin (or hypocretin) cDNA was cloned into the polylinker region of pBC SK+ (Stratagene, La Jolla, CA). A 2.5 k base fragment from the rat CRF-R1 cDNA was cloned into the polylinker region of pcDNA (Promega). A 502 bp fragment from the rat AVP cDNA and a 1201 bp fragment from the rat V1b cDNA were cloned into the polylinker region of pCR II (Invitrogen, Carlsbad CA). The plasmid pS/E (a pSP65 derivative) was used to synthesize riboprobe for the 18S rRNA to determine total RNA. ³³P-labeled cRNA antisense probes and unlabeled cRNA sense standards were synthesized using a SP6, T3 or T7 transcription system. A denaturing agarose gel containing

1.0 M formaldehyde showed that a single full-length transcript had been synthesized from each plasmid.

RNA extracts were dried in 1.5 ml Eppendorf tubes and resuspended in 30 μ l of 2 × TESS (10 mM N-Tris[hydroxy-methyl]methyl-2-aminoethane sulfonic acid, pH 7.4; 10 mM ethylenediaminetetraacetic acid [EDTA]; 0.3 M NaCl; 0.5% sodium dodecyl sulfate [SDS]) that contained 150 to 300 K cpm of a probe. Samples were covered with mineral oil and hybridized overnight at 75 °C. For RNase treatment, 250 μ l of a buffer containing 0.3 M NaCl; 5 mM EDTA; 10 mM Tris-HCl (pH 7.5), 40 μ g/ml RNase A (Worthington, Biochemicals, Freehold, NJ) and 2 μ g/ml RNase T1 (Calbiochem, San Diego, CA) was added and each sample was incubated at 30 °C for 1 hour. TCA precipitation was effected by the addition of 1 ml of a solution that contained 5% TCA and 0.75% sodium pyrophosphate. Precipitates were collected onto a filter in sets of 24 using a cell harvester (Brandel, Gaithersburg, MD) and were measured in a scintillation counter with liquid scintillant (Beckman, Palo Alto, CA).

The procedure to measure mRNA levels involved a comparison of values obtained from experimental samples (brain extracts) to those obtained for a set of calibration standards. The calibration standards had known amounts of an in vitro sense transcript whose concentration was determined by optical absorbance at 260 nm. The set of calibration standards included those with no added sense transcript and those that contained between 1.25 and 80 pg of the sense transcript. To determine the total attomoles of each mRNA in each extract, the amounts calculated from the standard curves were multiplied by 2.73 for POMC, 0.25 for MOP-r, 4.3 for orexin, 1.25 for Dyn, 5.00 for AVP, 2.41 for AVP V1b, 2.15 for CRF or 1.11 for CRF-R1 to correct for the difference in length between the sense transcript and the full-length mRNA. A new standard curve was generated each time experimental samples were analyzed and all extracts of a particular tissue were assayed for each mRNA as a group in a single assay.

Total cellular RNA concentrations were measured by hybridization of diluted extracts to a 33 P-labeled probe complementary to 18S rRNA at 75 °C. The calibration standards for this curve contained 10 µg of E. coli tRNA plus either 0.0, or from 2.5 to 40 ng of total RNA from rat brain whose concentration was determined by optical absorbance at 260 nm.

Selected samples were subjected to solution hybridization and RNase treatment followed by gel electrophoresis. The protected RNA:RNA hybrids were phenol-chloroform extracted, precipitated with 100% ethanol, and electrophoresed through nondenaturing 4% polyacrylamide gels, with ³²P-labeled ΦX 174 RF DNA digested with *Hae*lll as molecular weight markers. The gels were then dried and exposed to X-film. The protected species for each mRNA was approximately at the size, corresponding to an RNA:RNA hybrid formed by hybridization of the full-length cRNA probe with total cytoplasmic RNA samples extracted from the tissues (data not shown).

5. Radioimmunoassays

At the time of decapitation, blood from each rat was collected in tubes, placed on ice, and was spun in a refrigerated centrifuge. Plasma was separated and stored at -40 °C for hormonal measurements by radioimmunoassay. Corticosterone (CORT) levels were assayed using a rat corticosterone ¹²⁵I kit from MP Biomedicals (Costa Mesa, CA). ACTH immunoreactivity levels were assayed from unextracted plasma by using a kit from DiaSorin Inc. (Stillwater, MN). All values were determined in duplicate in a single assay.

6. Data analysis

In Experiments I and II, group differences in mRNA levels of each gene in each brain region and in plasma hormonal levels were analyzed using a two-way analysis of variance (ANOVA) with Treatment (chronic heroin or saline in Experiment I and heroin or saline withdrawal in Experiment II) and Challenge (heroin or saline in both Experiments I and II), followed by Newman-Keuls *post-hoc* tests or planned comparisons or Student's t-tests when appropriate. The accepted level of significance for all tests was p < 0.05. All statistical analyses were performed using *Statistica* (version 5.5, StatSoft Inc.).

Results

1. ACTH

In Experiment I, the ANOVA revealed a significant Treatment × Challenge interaction (F $_{(1,21)} = 10.0$, p < 0.01) (**Figure 2A**). Newman-Keuls *post-hoc* tests revealed that plasma ACTH levels were significantly higher in the Chronic heroin + Sal group in comparison to the Chronic saline + Sal group (p < 0.05). Also, plasma ACTH levels after 5 mg/kg heroin challenge in heroin naïve rats (Chronic saline + HC) were significantly higher than in the Chronic saline + Sal group (F $_{(1,21)} = 5.52$, p < 0.05), and plasma ACTH levels in the Chronic heroin + HC group were significantly lower than levels in the Chronic heroin + Sal group (F $_{(1,21)} = 4.53$, p < 0.05).

In Experiment II, the ANOVA revealed a significant main effect of Treatment (F $_{(1,32)}$ = 19.4, p < 0.001), a significant effect of Challenge (F $_{(1,32)}$ = 4.73, p < 0.05), and a significant Treatment × Challenge interaction (F $_{(1,32)}$ = 12.5, p < 0.005; **Figure 2B**). Newman-Keuls *post-hoc* tests revealed that plasma ACTH levels after 5 mg/kg heroin challenge in heroin naïve rats (Saline withdrawal + HC) were significantly higher than those in controls (Saline withdrawal + Sal; p < 0.001). Since other studies have reported suppression of HPA activity in the rat after 5 or more days of withdrawal from chronic administration of morphine [8], a planned comparison was employed to reveal that plasma ACTH levels in the Heroin withdrawal + HC group were significantly lower than in the Heroin withdrawal + Sal group ($t_{(1,17)}$ = 4.55, p < 0.05).

2. CORT

As shown in **Figures 2C** and **2D**, plasma CORT levels showed a pattern of variation similar to plasma ACTH. In Experiment I, the ANOVA revealed a significant Treatment × Challenge interaction (F $_{(1,25)} = 10.2$, p < 0.01; **Figure 2C**). Newman-Keuls *post-hoc* tests indicated that plasma CORT levels in the Chronic heroin + Sal group were significantly higher than those in the Chronic saline + Sal group (p < 0.05), and that plasma CORT levels in the Chronic saline + HC group was significantly higher than those in the Chronic saline + HC group was significantly higher than those in the Chronic saline + HC group was significantly higher than those in the Chronic saline + Sal group (p < 0.05). Further, levels of CORT were reduced in the Chronic heroin + HC, but this was not significantly different from the Chronic Heroin + Sal group (F $_{(1,21)} = 3.57$, p = 0.07).

In Experiment II (**Figure 2D**), the ANOVA revealed a significant main effect of Treatment (F $_{(1,32)} = 4.28$, p < 0.05) and a significant Treatment × Challenge interaction (F $_{(1,32)} = 8.10$, p < 0.01). Newman-Keuls *post-hoc* tests indicated that plasma CORT levels in the Saline withdrawal + HC group were significantly higher than those in the Saline withdrawal + Sal group (p < 0.05). Plasma CORT levels in the Heroin withdrawal + HC group were not significantly different from that of the Heroin withdrawal + Sal group, although plasma CORT levels showed a reduction similar to plasma ACTH (**Figure 2B**).

3. POMC in MH

In Experiment 1, the ANOVA revealed a significant main effect of Treatment (F $_{(1,24)}$ = 4.35, p < 0.05), and Newman-Keuls *post-hoc* tests indicated that POMC mRNA levels were significantly lower in the Chronic heroin + HC and Chronic heroin + Sal groups in comparison to the Chronic saline + Sal and Chronic saline + HC groups (**Figure 3A**).

In Experiment II (**Figure 3B**), the ANOVA revealed only a significant main effect of Treatment (F $_{(1,32)} = 13.5$, p < 0.001). Newman-Keuls *post-hoc* tests indicated that POMC mRNA levels were significantly higher in the Heroin withdrawal + HC and Heroin withdrawal + Sal groups in comparison to Saline withdrawal + Sal and Saline withdrawal + HC groups.

4. POMC in AP

In Experiment 1 (**Figure 3C**), the ANOVA revealed a significant main effect of Treatment (F $_{(1,26)} = 5.50$, p < 0.05). Newman-Keuls *post-hoc* tests suggested that POMC mRNA levels in the Chronic heroin + HC and the Chronic heroin + Sal groups were significantly higher than those in Chronic saline + Sal and Chronic saline + HC groups. In Experiment II (**Figure 3D**), there were no significant differences.

5. Orexin in LH

In Experiment I (**Table 1**), the ANOVA revealed a significant effect of Treatment (F $_{(1,26)}$ = 4.84, p < 0.05). Newman-Keuls *post-hoc* tests further revealed that levels of orexin mRNA in the Chronic heroin + Sal group were significantly higher than those in the Chronic saline + Sal group (p < 0.05). No group differences were found in Experiment II.

6. MOP-r in LH

In Experiment I (**Table 2**), the ANOVA revealed a significant Treatment × Challenge interaction (F $_{(1,21)} = 4.59$, p < 0.05). Newman-Keuls *post-hoc* tests indicated that MOP-r mRNA levels in the Chronic heroin + Sal were significantly higher than those in the Chronic saline + Sal group (p < 0.05). In Experiment II, no group differences were found (**Table 2**).

7. CRF and AVP in MH or CRF-R1 and V1b in AP

In either Experiment I or II, there was no significant group difference in mRNA levels of CRF and AVP in MH (**Table 3A**), or CRF-R1 and V1b in AP (**Table 3B**).

8. Other measures

The mRNA levels of Dyn in LH, and Dyn, POMC or MOP-r in the amygdala were unaltered (**Tables 1** and **2**, **Figure 3E** and **F**).

Discussion

The present study confirmed that 12-hour acute heroin withdrawal, as stress, elevated HPA hormones. Under this stress condition, heroin inhibited the HPA activation. When the HPA hormonal levels returned to baseline following 10-day chronic withdrawal, the HPA activity was inhibited by heroin at a low dose, suggesting a long-lasting neuroadaptation during chronic heroin withdrawal. In acute withdrawal, a decreased POMC mRNA level in the medial hypothalamus was coupled with enhanced orexin and MOP-r gene expression in the lateral hypothalamus. After chronic withdrawal, conversely, an increased POMC mRNA level was found in the medial hypothalamus, with no lasting changes of orexin or MOP-r mRNA levels.

1. HPA axis

Consistent with earlier studies [8], we found that an acute heroin challenge (5 mg/kg) had a stimulatory effect on plasma ACTH and CORT release in heroin-naïve rats. After chronic (10-day) intermittent escalating-dose heroin administration, we have reported that the HPA axis did not respond to heroin, indicating tolerance of HPA activity, and in acute 12-h opiate withdrawal, activation of the HPA axis was observed [21]. The present experiment replicated the above results by showing that HPA hormonal levels were elevated after acute heroin withdrawal. Chronic intermittent exposure to opiates is a chronic stressor, and it may alter the responsivity of the HPA axis, as do many other stressors [23,24]. Under this particular stress condition (Experiment I), we examined the effect of heroin challenge on pituitary-adrenal function, and found that heroin challenge at 20 mg/kg decreased the elevation of plasma ACTH and CORT levels induced by acute opiate withdrawal stress.

In Experiment II, the present study examined basal levels of the stress hormones after chronic 10-day withdrawal as well as the alterations of HPA responses to heroin challenge, in order to compare with the HPA responsivity during acute withdrawal. We found that after 10-day heroin withdrawal, heroin challenge at a low dose, 5 mg/kg, did not stimulate the HPA axis as it did in heroin naïve rats. In contrast, it even had a slight, but significant, suppression on plasma ACTH levels. Our results indicate long-lasting neuroadaptation in the HPA system during chronic heroin withdrawal.

Although the mechanisms responsible for the interactions between heroin and withdrawal stress are not clear, the effects of opiates on HPA activity have been found to depend on the presence or absence of stress. In fact, an inhibitory effect of opiates on the HPA axis is found in heroin addicts [4]. It is consistently found that morphine suppresses cortisol release induced by surgical stress in humans [25,26]. In support of this concept, our animal study found that while either acute morphine or water restriction stress alone increased ACTH levels as an independent stimulus, morphine decreases plasma ACTH levels elevated by water restriction stress [27]. Both human and animal studies demonstrate that morphine or heroin effectively blunts the HPA activity caused by stress, indicating that opioids play a counter-regulatory role in modulating HPA stress responsivity under stress conditions. Although interesting, the HPA result of the current study should be interpreted with caution. No difference for the mRNA levels of CRF or AVP in the hypothalamus or of their receptors in the anterior pituitary was observed after acute or chronic heroin withdrawal with or without heroin challenge. In future experiments, information on the peptide levels of CRF and AVP in the hypothalamus, and the HPA axis responses at multiple time points or to other different stressors would be useful.

Although the mechanisms responsible for the interactions between heroin and withdrawal stress have not been elucidated, the inhibitory effects of opiates on HPA activity in rats have been reported to occur at the paraventricular nucleus [28] or median eminence [29], but not the anterior pituitary [30]. Moreover, although it has been reported that the stress responsive neuropeptides in the amygdala are involved in modulating HPA activity [3], we did not observe any change of POMC, MOP-r or Dyn in the amygdala during either acute (12 hours) or chronic (10 days) heroin withdrawal with or without heroin challenge.

2. POMC/MOP-r and orexin systems in the hypothalamus

In early studies using morphine pellets, it was found that POMC mRNA in the hypothalamus tended to decrease after chronic morphine in either normal rats [15,16], castrated male rats [31] or ovariectomized female guinea pigs [32]. In the current study, we examined POMC gene expression in two brain regions (medial hypothalamus and amygdala) after acute (12-h) withdrawal, and found a slight, but significant, decrease in the

POMC mRNA levels only in the medial hypothalamus. Our data suggests a decrease in opioidergic activity in the hypothalamus caused by reduced POMC gene expression in acute heroin withdrawal, although there was a parallel increase in POMC mRNA expression in the anterior pituitary, which could result in an increased opioid activity at peripheral level.

Given that POMC neurons in the arcuate nucleus project to the lateral hypothalamus, and that beta-endorphin inhibits orexin neurons [19], we also measured orexin mRNA levels in the lateral hypothalamus, with MOP-r in the same region. We found that both orexin and MOP-r mRNA levels were increased at 12-hour withdrawal from chronic escalating-dose heroin administration. Orexin A peptide has been found to decrease the excitability of brain reward systems in the lateral hypothalamus [33]. Also, alteration of the orexin activity is likely involved in the negative affective state during drug withdrawal, which may be important in the modulation of drug-seeking behaviors [34]. Our results suggest that activation of orexin gene expression in the lateral hypothalamic neurons may be associated with a new set point of brain reward function established by chronic heroin exposure and withdrawal [3,35].

In an early study, blockade of beta-endorphin activity in the brain by icv infusion of naloxone was found to reduce the reinforcing effect of lateral hypothalamic stimulation, suggesting that endogenous beta-endorphins are involved in self-stimulation reward [36]. Behaviorally, rats self-administer this opioid peptide into the brain and this supports a role for central beta-endorphin in reinforcement [37]. It has been hypothesized that heroin dependent individuals self-administer heroin to escape the negative affective and emotional states associated with drug withdrawal in humans [20,24]. There is also evidence in rats and primates that the motivational properties of opiates are altered by spontaneous and naloxone-precipitated withdrawal and that opiate withdrawal can motivate opiate-seeking behavior [38-40]. It is reasonable, therefore, to postulate that decreased POMC gene expression with resultant decreases in beta-endorphin may initiate behaviors leading to opiate intake and thus normalization of endogenous opioid tone. This opioid-deficiency hypothesis is also supported by evidence in humans that heroin addicts have decreased opioid activity in regulating HPA activity [5,41,42], which is normalized by methadone (a potent MOP-r agonist) maintenance treatment [4].

Of interest, beta-endorphin acting on the MOP-r exerts tonic inhibition of CRF release from the paraventricular nucleus, and then of the HPA axis in both humans and rodents [20]. In acute heroin withdrawal, we found that acute opiate withdrawal dramatically activated HPA activity, which is associated with decreased POMC mRNA levels in the hypothalamus. The results of the present study support the concept that there is an inverse association between hypothalamic POMC activity and the HPA axis. To date, there is no evidence that orexin acting at its receptors has any direct relationship with or regulation of the HPA axis, although there is extensive evidence that orexin has actions in reward-related nuclei, such as the hypothalamus, nucleus accumbens and ventral tegmental area [3].

In sharp contrast, after 10-day chronic heroin withdrawal, we observed a rebound increase in the POMC mRNA levels in the medial hypothalamus. Although the exact timing and maximal magnitude of POMC gene expression remain to be further resolved, our results suggest that after chronic withdrawal, there may be an increase in the POMC biosynthesis and/or opioidergic activity from the hypothalamus, although there was no alteration of POMC mRNA expression in the anterior pituitary observed. In line with these animal studies, human studies have demonstrated that supersensitivity to naloxone-induced HPA stimulation is evident after administration of a single low dose of naloxone to an opiate dependent individual, indicating that enhanced opioid activity is present during withdrawal [7].

3. Summary

The present study confirmed that acute heroin withdrawal stimulated HPA activity. When the rats were under this stress condition, heroin suppressed the HPA activation, indicating that opioids play a counter-regulatory role in the stress response by inhibiting the stress response cascade. Our data further showed that when basal HPA hormonal levels returned to baseline following chronic withdrawal, the HPA activity was inhibited by heroin challenge at a low dose, suggesting a long-lasting neuroadaptation during chronic heroin withdrawal. Also, in the hypothalamus, a relative deficit in basal opioid activity in response to acute withdrawal (as reflected by decreased POMC mRNA levels) may lead to enhanced orexin and MOP-r gene expression in the lateral hypothalamus. Conversely, relative excess in basal opioid activity was found after chronic withdrawal, as reflected by increased POMC mRNA levels in the hypothalamus.

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Zhou et al.

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Zhou et al.

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Chronic (10-day) intermittent escalating-dose (7.5 up to 60 mg/kg/day) heroin, acute (12-hour) withdrawal (WD) and chronic (10-day) WD with heroin challenge.

						Experiment I	Experiment II
	Days 1-2	Days 3-4	Days 5-6	Days 7-8	Days 9-10	Day 11	Day 21
Heroin (mg/kg/day)	3x2.5	3x5	3x10	3x15	3x20	12-h WD	10-day WD
Heroin challenge						20 mg/kg	5 mg/kg
	Days 1-2	Days 3-4	Days 5-6	Days 7-8	Days 9-10	Day 11	Day 21
Saline	Saline	Saline	Saline	Saline	Saline	12-h WD	10-day WD
Heroin challenge						5 mg/kg	5 mg/kg

Figure 1.

Timelines for heroin administration regimens with acute heroin challenge.



Exp I. 12-hour withdrawal with heroin challenge



Figure 2.

Effects of chronic (10-day) intermittent escalating-dose heroin administration, 12-hour withdrawal, and 10-day withdrawal with heroin challenge on mean + SEM of plasma ACTH levels (pg/ml) (**A**, **B**) or corticosterone (CORT) levels (ng/ml) (**C**, **D**). **A** and **C**: In Experiment I (12-hour withdrawal with heroin challenge), rats received chronic heroin or saline for 10 days followed by heroin challenge (HC, 20 mg/kg) or saline (Sal) challenge on day 11, including four groups of rats: (1) <u>Chronic saline + Sal</u>, as saline control; (2) <u>Chronic saline + HC</u>, as acute heroin (heroin challenge at 5 mg/kg); (3) <u>Chronic heroin + Sal</u>: as 12-hour withdrawal; and (4) <u>Chronic heroin + HC</u>: as chronic heroin (heroin challenge at 20 mg/kg). **B** and **D**: In Experiment II (10-day withdrawal with heroin challenge), rats received chronic heroin or saline for 11 days, and then 10 days of withdrawal, followed by heroin challenge (HC, 5 mg/kg) or saline control; (2) <u>Saline withdrawal + HC</u>, as acute heroin (heroin withdrawal + Sal: 10-day heroin withdrawal + saline, as 10-day withdrawal; and (4) <u>Heroin withdrawal + HC</u>: as heroin challenge (5 mg/kg) after 10-day withdrawal. Significant differences are indicated: *, +, # p < 0.05 (n=6-9).



Figure 3.

Effects of chronic (10-day) intermittent escalating-dose heroin administration, 12-hour withdrawal, and 10-day withdrawal with heroin challenge on pro-opiomelanocortin (POMC) mRNA levels (attomole/ μ g total RNA) in the medial hypothalamus (**A**, **B**), anterior pituitary (**C**, **D**) and amygdala (**E**, **F**). **A**, **C** and **E**: In Experiment I (12-hour withdrawal with heroin challenge), rats received chronic heroin or saline for 10 days followed by heroin challenge (HC, 20 mg/kg) or saline (Sal) challenge on day 11. **B**, **D** and **F**: In Experiment II (10-day withdrawal with heroin challenge), rats received chronic heroin or saline for 11 days, and then 10 days of withdrawal, followed by heroin challenge (HC, 5 mg/kg) or saline on day

21. See details in Figure 2. There was a significant main effect for Chronic heroin, & p < 0.05. Newman-Keuls *post-hoc* tests showed significant effects: *, +, @ p < 0.05 (n=6-9).

Table 1

Effects of chronic (10-day) intermittent escalating-dose heroin administration, 12-hour withdrawal, and 10day withdrawal with heroin challenge on mRNA levels (attomole/µg total RNA) of orexin and preprodynorphin (Dyn) in the lateral hypothalamus (LHyp) and amygdala (Amy).

Experiment	Treatment	Orexin LHyp	Dyn LHyp	Dyn Amy
I. 12-hour withdrawal with heroin challenge	Chronic saline + Sal	4.6 ± 0.48	4.9 ± 0.52	2.2 ± 0.48
	Chronic saline + HC	4.8 ± 0.32	4.8 ± 0.49	1.9 ± 0.21
	Chronic heroin + Sal	6.9 ± 0.52 *	4.0 ±0.29	1.9 ± 0.21
	Chronic heroin + HC	5.9 ± 0.69	4.3 ± 0.25	2.0 ± 0.27
II. 10-day withdrawal with heroin challenge	Saline withdrawal + Sal	5.0 ± 0.44	4.7 ± 0.27	2.1 ± 0.35
	Saline withdrawal + HC	5.2 ± 0.58	4.6 ± 0.31	1.9 ± 0.16
	Heroin withdrawal + Sal	5.4 ± 0.48	4.7 ± 0.35	2.0 ± 0.21
	Heroin withdrawal + HC	5.1 ± 0.32	4.8 ± 0.33	2.2 ± 0.19

In Experiment I (12-hour withdrawal with heroin challenge), rats received chronic heroin or saline for 10 days followed by heroin challenge (HC, 20 mg/kg) or saline (Sal) on day 11, including four groups of rats: (1) <u>Chronic saline + Sal</u>, as saline control; (2) <u>Chronic saline + HC</u>, as acute heroin (heroin challenge at 5 mg/kg); (3) <u>Chronic heroin + Sal</u>: as 12-hour withdrawal; and (4) <u>Chronic heroin + HC</u>: as chronic heroin. In Experiment II (10-day withdrawal with heroin challenge), rats received chronic heroin or saline for 11 days, and then 10 days of withdrawal, followed by heroin challenge (HC, 5 mg/kg) or saline on day 21, including four groups of rats: (1) <u>Saline withdrawal + Sal</u>, as saline control; (2) <u>Saline withdrawal + HC</u>, as acute heroin (heroin challenge at 5 mg/kg); (3) <u>Heroin withdrawal + Sal</u>: as 10-day withdrawal; and (4) <u>Heroin withdrawal + HC</u>: as heroin challenge (5 mg/kg) after 10-day heroin withdrawal. Data shown in table are treatment group mean ± SEM. Significant differences are indicated:

p < 0.05 vs. Chronic saline + Sal (n=6-9).

Table 2

Effects of chronic (10-day) intermittent escalating-dose heroin administration, 12-hour withdrawal, and 10day withdrawal with heroin challenge on mu-opioid receptor (MOP-r) mRNA levels (attomole/µg total RNA) in the lateral hypothalamus (LHyp) and amygdala (Amy).

Experiment	Treatment	MOP-r LHyp	MOP-r Amy
I. 12-hour withdrawal with heroin challenge	Chronic saline + Sal	0.38 ± 0.05	0.17 ± 0.02
	Chronic saline + HC	0.37 ± 0.11	0.14 ± 0.01
	Chronic heroin + Sal	0.62 ± 0.07 *	0.17 ± 0.01
	Chronic heroin + HC	0.53 ± 0.11	0.15 ± 0.01
II. 10-day withdrawal with heroin challenge	Saline withdrawal + Sal	0.44 ± 0.03	0.19 ± 0.01
	Saline withdrawal + HC	0.43 ± 0.04	0.18 ± 0.02
	Heroin withdrawal + Sal	0.46 ± 0.03	0.17 ± 0.01
	Heroin withdrawal + HC	0.39 ± 0.04	0.19 ± 0.01

See details in Table 1. Data shown in table are treatment group mean \pm SEM. Significant differences are indicated:

p < 0.05 vs. Chronic saline + Sal (n=6-9).

Table 3

Effects of chronic (10-day) intermittent escalating-dose heroin administration, 12-hour withdrawal, and 10day withdrawal with heroin challenge on mRNA levels (attomole/µg total RNA) of arginine vasopressin (AVP) and corticotropin-releasing factor (CRF) in the medial hypothalamus (A), and AVP V1b type receptor (V1b) and CRF type I receptor (CRF-R1) in the anterior pituitary (B).

A.					
Experiment	Treatment	AVP	CRF		
I. 12-hour withdrawal with heroin challenge	Chronic saline + Sal	103 ± 11	1.13 ± 0.09		
	Chronic saline + HC	104 ± 9	1.10 ± 0.15		
	Chronic heroin + Sal	112 ± 9	1.30 ± 0.10		
	Chronic heroin + HC	77 ± 8	0.94 ± 0.11		
II. 10-day withdrawal with heroin challenge	Saline withdrawal + Sal	93 ± 9	1.06 ± 0.18		
	Saline withdrawal + HC	90 ± 7	1.10 ± 0.09		
	Heroin withdrawal + Sal	94 ± 15	1.13 ± 0.19		
	Heroin withdrawal + HC	99 ± 9	1.11 ± 0.12		

Experiment	Treatment	V1b	CRF-R1		
I. 12-hour withdrawal with heroin challenge	Chronic saline + Sal	1.7 ± 0.19	0.92 ± 0.10		
	Chronic saline + HC	1.8 ± 0.11	0.95 ± 0.18		
	Chronic heroin + Sal	1.9 ± 0.34	1.20 ± 0.25		
	Chronic heroin + HC	2.2 ± 0.41	1.11 ± 0.30		
II. 10-day withdrawal with heroin challenge	Saline withdrawal + Sal	1.9 ± 0.19	0.89 ± 0.16		
	Saline withdrawal + HC	2.0 ± 0.29	0.88 ± 0.27		
	Heroin withdrawal + Sal	1.9 ± 0.10	0.98 ± 0.33		
	Heroin withdrawal + HC	1.9 ± 0.26	0.92 ± 0.31		

See details in Table 1. Data shown in table are treatment group mean \pm SEM (n=6-9).