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Ambient ultrafine particles reduce endothelial nitric oxide production via S-glutathionylation of eNOS

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Abstract

Exposure to airborne particulate pollutants is intimately linked to vascular oxidative stress and inflammatory responses with clinical relevance to atherosclerosis. Particulate matter (PM) has been reported to induce endothelial dysfunction and atherosclerosis. Here, we tested whether ambient ultrafine particles (UFP, diameter < 200 nm) modulate eNOS activity in terms of nitric oxide (NO) production via protein S-glutathionylation. Treatment of human aortic endothelial cells (HAEC) with UFP significantly reduced NO production. UFP-mediated reduction in NO production was restored in the presence of JNK inhibitor (SP600125), NADPH oxidase inhibitor (Apocynin), anti-oxidant (N-acetyl cysteine), and superoxide dismutase mimetics (Tempol and MnTMPyP). UFP exposure increased the GSSG/GSH ratio and eNOS S-glutathionylation, whereas over-expression of Glutaredoxin-1 (to inhibit S-glutathionylation) restored UFP-mediated reduction in NO production by nearly 80%. Thus, our findings suggest that eNOS S-glutathionylation is a potential mechanism underlying ambient UFP-induced reduction of NO production.

Keywords

Ultrafine Particles/UFP; Oxidative Stress; eNOS; S-Glutathionylation; Endothelial dysfunction; Air Pollution

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Conflict of interest

The authors report no conflict of interest.

Introduction

Exposure to ambient particulate matter (PM) is increasingly recognized as a modifiable risk factor to cardiovascular morbidity and mortality [1]. Mice exposed to PM displayed accelerated atherosclerosis and increased lesion size [2]. Both ambient and diesel exhausted particles have been shown to induce endothelial dysfunctions in ApoE-null mice [3,4]. However, the mechanisms underlying UFP-induced endothelial dysfunction remain fairly unexplored.

Endothelial nitric oxide synthase (eNOS) plays a pivotal role in maintaining vascular homeostasis. eNOS is a source of both nitric oxide (NO) and superoxide ($O_2^{\cdot -}$). In the absence of tetrahydrobiopterin (BH_4), eNOS is uncoupled to produce superoxide [5,6,7,8]. In the presence of BH_4 , NOS activity and NOS-dependent vasodilation were partially restored [5]. These observations led to the investigation of alternative mechanisms underlying the regulation of eNOS activity. Chen *et al* recently reported that S-glutathionylation uncouples eNOS activity via two mechanisms: 1) exchange of thiol-disulphide with oxidized glutathione, or 2) reaction of oxidant-induced protein thiol radicals with reduced glutathione [9]. In this context, we sought to assess whether exposure to ambient ultrafine particles (UFPs) modulates eNOS activity.

Ambient ultrafine particles (UFP, diameter < 200 nm), highly enriched in transition metals and redox cycling organic chemicals, harbor potent toxic properties and oxidizing potential [10,11]. UFPs induce vascular oxidative stress via JNK signaling in endothelial cells [12,13], promoting inflammatory responses and reducing anti-oxidant capacity of high-density lipoprotein (HDL) in mouse model of atherosclerosis [2,14]. Oxidative stress modulates cellular protein S-glutathionylation [15,16,17,18], and an increase in reactive oxygen species (ROS) is implicated in a decrease in NO production [19]. However, whether UFP-induced oxidative stress would reduce NO production remain unclear.

In this study, we assessed whether exposure to ambient ultrafine particulate pollutants modulated endothelial function via eNOS S-glutathionylation. We demonstrated that UFP significantly increased glutathione oxidation, protein S-glutathionylation, and eNOS S-glutathionylation, leading to a decrease in NO production, whereas over-expression of glutaredoxin-1 (Grx-1) to inhibit protein S-glutathionylation nearly rescued eNOS activity. Thus, S-glutathionylation of eNOS is implied as a potential mechanism underlying UFP-mediated reduction in NO production.

Materials and Methods

UFP collection, preparation, and characterization

UFP were collected at the University of Southern California (USC) campus near downtown Los Angeles on 20×25 cm Teflon coated filters (PALL Life Sciences, PTFE membrane, $2.0 \mu m$, R2PJO37) using a High-Volume Particle Sampler [20]. PM mass was determined gravimetrically by pre- and post-weighing the high volume sampler filters, as discussed in greater detail in a previous publication [21]. The Teflon-coated glass fiber filters were split into different portions: $\frac{1}{4}$ was used for chemical analysis, whereas $\frac{3}{4}$ was used for the preparation of the exposure suspensions, as discussed in subsequent paragraphs. The $\frac{1}{4}$ of the Teflon filter was split in four equal parts; one part was analyzed by Shimadzu TOC-5000A liquid analyzer [22] for water soluble organic carbon (WSOC) and another one by ion chromatography (IC) technique for their inorganic ion content (i.e. sulfate, nitrate and ammonium). A third portion of the high volume sampler filter was analyzed by gas chromatography-mass spectrometry (GC/MS) for organic compounds [23], whereas the remaining portion was analyzed for UFP bound water-soluble metals and elements by means

of Inductively Coupled Plasma – Mass Spectroscopy (ICP-MS) [24]. The chemical composition and size distribution of UFPs are reported in a previous publication by our groups [25](Chemical compositions are provided as a supplemental table). The remaining portion of the high volume samples was used to prepare for the UFP suspension for exposure tests. The filters were first soaked in 10 ml of ultra-pure milli-Q water (USP grade) for 30 minutes in endotoxin-free glass vial, followed by sonication for 30 minutes [26]. After the particle suspension was transferred to endotoxin-free tube, another 10 ml of ultra-pure water was used to repeat the aforementioned process. Our control (i.e. particle – free) suspension was prepared by extracting a blank filter, akin to the ones used for particulate matter (PM) collection in the USP grade water, using the aforementioned procedures. The UFP suspension was aliquoted and stored at -80°C to maintain chemical stability.

Measurement of NO production

eNOS activity was assessed by measuring NO production using Nitrate/Nitrite Colorimetric Assay Kit from Cayman Chemicals. Due to the presence of nitrate/nitrite in UFP that gave high background, we prepared cell lysate to measure NO production.

Human aortic endothelial cells (HAEC) (Cell Application) were cultured with endothelial cell growth media (Cell Application). The cells were used between passages 5 and 9. For UFP treatment, confluent HAEC were incubated in the presence or absence of UFPs in M199/0.1% FBS (Invitrogen) for 6 hours. After washing with PBS, the cells were lysed with PBS/0.2% Triton X-100 and scraped into 1.5ml tubes. After incubation at 4°C for 30 minutes, the supernatants were collected as lysate for NO production measurement following manufacturer's instruction and for protein assay using BioRad DCP protein assay kit. NO production was normalized to protein concentration relative to control.

To assess NO production in the presence of inhibitors, HAEC were pretreated with JNK inhibitor SP600125 ($2\mu\text{M}$), NADPH oxidase inhibitor Apocynin ($200\mu\text{M}$), an anti-oxidant N-acetyl cysteine (NAC, 1mM), or superoxide dismutase mimetics, Tempol ($200\mu\text{M}$), and MnTMPyP ($2\mu\text{g/mL}$) for 30 minutes, followed by treatment with or without $50\mu\text{g/mL}$ of UFP in the presence of inhibitors. Cell lysates were then prepared for the measurement of NO production. The inhibitor concentration used in the experiments was lower than that of standard usage and was optimized to minimize the interference with background NO production.

S-glutathionylation of eNOS

Quantification of eNOS S-glutathionylation was performed with a modified protocol from Cayman Chemical. Briefly, HAEC in 100mm dishes were grown to confluent. Cells were treated with or without $50\mu\text{g/mL}$ of UFP for 6 hours in M199/0.1% FBS. After washing with PBS, cells were collected by trypsinization and re-suspended in 1mL PBS. The cells were then fixed with PBS/3.7% Formaldehyde for 20 min at room temperature, followed by cell lysis. S-glutathionylation proteins were labeled with Biotin following the manufacturer's instructions. The protein lysate was used for ELISA to measure eNOS S-glutathionylation. Briefly, aforementioned protein lysate was added to the 96-well plates coated with NeutrAvidin (Pierce), and incubated for 30 minutes at room temperature. After rinsing with washing buffer (PBS/0.05% Tween-20), anti-eNOS antibody (1:1000, Cell signaling) in Blocking buffer (PBS/1%BSA/0.05% Tween-20) was added. After incubation for 1 hour at room temperature, the wells were rinsed and HRP-anti-Rabbit Ig secondary antibody (1:1000) was added. After 30 minutes, the wells were rinsed and TMB turbo substrate (Pierce) was added. After color development, 2M of sulfuric acid was added to cease the reaction. Optical density at 450nm (OD450) was measured as readout of eNOS S-glutathionylation

Measurement of Glutathione

Levels of glutathione (GSH) were quantified by using Glutathione Assay Kit (Cayman Chemical) according to the manufacturer's instruction. Oxidized glutathione (GSSG) was measured by using GSH derivatizing reagent 2-vinylpyridine using the alternative protocol from the manufacturer. The concentration of GSH and GSSG was normalized to control by protein concentration.

Western Blots

HAEC were grown to confluence, and treated with or without UFP. Cell lysate preparation and western blots were performed as previously described [13]. To assess the role of oxidative stress on eNOS S-glutathionylation, we infected HAEC with glutaredoxin-1 (Grx-1) recombinant adenovirus (kindly provided by Dr. Mieyal at Case Western Reserve University) or with the control LacZ recombinant adenovirus overnight. The levels of Grx-1 expression were assessed by western blot with anti Grx-1 antibody (Abcam).

Immunohistochemistry

LDLR-null mice were exposed to filtered air (FA) or UFP as described [14]. After exposure for ten weeks, the heart tissues of mice were dissected and paraffin blocks were made. The heart sections were stained with anti-GSH antibody (1:100) (Virogen) for visualizing protein S-glutathionylation with standard immunohistochemistry procedure as described previously [27].

Statistical Analysis

All of the experiments were performed for three or more trials. Data were expressed as mean \pm standard deviation (SD). Student *t*-test was used for significance analysis. A *P* value of < 0.05 was considered statistically significant.

Results

UFP-induced oxidative stress reduced endothelial NO production

UFP induced JNK-dependent oxidative stress in vascular cells [13]. Here, we demonstrate that significantly reduced NO production in terms of nitrite/nitrate concentration in a dose dependent manner (control=9.5 \pm 3.0 μ M; UFP at 12.5 μ g/mL=6.4 \pm 2.2 μ M; UFP at 25 μ g/mL 4.0 \pm 1.0, *p* < 0.05 vs. control; UFP at 50 μ g/mL 2.7 \pm 1.4, *p* < 0.05 vs. Control; n = 3) (Fig. 1A). For this reason, UFP concentration at 50 μ g/mL was arbitrarily used for the subsequent studies.

In the presence of JNK inhibitor (SP600125) and NADPH oxidase inhibitor (Apocynin), UFP-mediated reduction in NO production was restored (Fig. 1B). Similarly, superoxide dismutase mimetics, both Tempol (SOD1) and MnTMPyP (SOD2), as well as anti-oxidant, N-acetyl cysteine (NAC), rescued UFP-mediated reduction in NO production (Fig. 1C). Thus, UFP-induced oxidative stress diminished endothelial NO production.

UFP exposure increased eNOS S-glutathionylation

Protein thiols can be glutathionylated by oxidized glutathione (GSSG) through disulfide exchange that modulates protein function [9]. To further elucidate the mechanism underlying UFP-induced reduction in NO production, we assessed the relative levels of oxidized (GSSG) and reduced glutathione (GSH) in response to UFP exposure. In HAEC exposed to UFP, GSH levels were decreased (control=17.1 \pm 1.8 μ M, UFP=12.0 \pm 2.4 μ M, n=4, *p* < 0.05), whereas GSSG levels were increased (control=0.62 \pm 0.26 μ M, UFP=1.60 \pm 0.20 μ M, n=4, *p* < 0.05) (Fig. 2A). The GSSG/GSH ratio was 0.134 in UFP-exposed cells

versus 0.036 in control cells. In corollary, UFP treatment increased protein S-glutathionylation as evidenced by the bands to anti-GSH antibody (Fig. 2B) in consistent with S-glutathionylated Actin in response to oxidative stress [18]. Increased protein S-glutathionylation was further recapitulated in mice exposed to UFP. The endothelium of endocardium of LDLR-null mice exposed to UFP revealed prominent staining to anti-GSH antibody (Fig. 2C). Thus, increased GSSG/GSH ratio was in parallel with the elevated protein S-glutathionylation in response to UFP exposure.

To examine if the reduction in NO production is associated with protein S-glutathionylation, we assessed eNOS S-glutathionylation in response to UFP exposure. By performing biotin-labeling of S-glutathionylation proteins and ELISA for eNOS, we demonstrated that UFP significantly increased eNOS S-glutathionylation in HAEC (control=0.008±0.001, UFP=0.042±0.006, BCNU (1,3-cis(2-chloroethyl)-1-nitrosourea, a positive control) = 0.032±0.004, n=3, $p < 0.01$) (Fig. 3).

S-glutathionylation mediated UFP-reduced NO Production

To assess whether eNOS S-glutathionylation mediated a reduction in NO production in response to UFP exposure, we over-expressed Glutaredoxin-1 (Grx-1), an inhibitor of protein S-glutathionylation. While NO production was lower in general in adenovirus-infected endothelial cells, over-expression of Grx-1 with recombinant adenoviruses (Fig. 4A) significantly attenuated UFP-mediated reduction in NO production by nearly 80% as compared to over-expression of control gene LacZ (control/LacZ=0.69±0.17; LacZ/UFP=0.34±0.11; Grx=0.78±0.25, Grx/UFP=0.63±0.13; LacZ/UFP vs. LacZ $p < 0.001$; Grx/UFP vs. Grx, $p = 0.24$; n=6) (Fig. 4B). Thus our findings corroborated UFP-mediated protein S-glutathionylation in modulating NO production.

Discussion

In this study, we elucidated a novel mechanism by which ambient ultrafine particulate pollutants reduced vascular endothelial NO production via eNOS S-glutathionylation, whereas over-expression of glutaredoxin-1 nearly rescued UFP-mediated reduction in NO production. Exposure to particulate matter (PM) has been shown to promote endothelial dysfunctions in ApoE^{-/-} mice [3,4]. Here, we demonstrate that eNOS S-glutathionylation is a novel mechanism underlying UFP-mediated reduction in NO production.

Oxidative stress regulates enzymatic activity of eNOS, and is intimately linked with production of reactive oxygen species (ROS) via eNOS uncoupling [28]. In this study, UFP-mediated reduction in NO production was restored in the presence of antioxidants; namely, N-acetyl cysteine (NAC), Tempol, and MnTMPyP, as well as NADPH oxidase inhibitor, Apocynin, and JNK inhibitor, SP600125. These findings support the previously published reports that UFP induced vascular oxidative stress via NADPH oxidase and JNK activation [13,29].

Oxidative stress was reported to increase protein S-glutathionylation [18], which has recently been reported to uncouple eNOS [9]. UFP increased the intensity of a dominant band by Western analysis, consistent with S-glutathionylated Actin [18]. We employed a modified S-glutathionylated protein Biotin labeling/ELISA methodology to establish the role of UFP in eNOS S-glutathionylation. Over-expression of glutaredoxin-1 (Grx-1) to inhibit protein S-glutathionylation corroborated the role of S-glutathionylation in the regulation of eNOS activity [30]. Oxidative stress was also implicated in the reduced bioavailability of BH₄, a cofactor of eNOS, resulting in eNOS uncoupling [31]. Thus, UFP-mediated reduction in NO production may be influenced by both S-glutathionylation and BH₄ reduction.

Various mechanisms are involved in protein S-glutathionylation. In the presence of ROS, cysteines form thiyl radicals which, in turn, react with glutathione (GSH). Alternatively, protein thiols can be glutathionylated by oxidized glutathione (GSSG) through disulfide exchange [9]. In this study, we revealed that UFPs increased intracellular GSSG levels as the potential underlying mechanism in the increase in eNOS S-glutathionylation. However, the precise mechanisms whereby UFP mediate eNOS S-glutathionylation and reduction in eNOS activity are in need of further investigation.

In summary, we demonstrate that ambient UFP collected in the urban environment of Los Angeles reduced vascular endothelial NO production via S-glutathionylation. In light of NO's role in vascular homeostasis, S-glutathionylation in eNOS may further influence UFP-mediated reduction in HDL anti-oxidant capacity [14] with clinical relevance to accelerated atherosclerosis [2].

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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Highlights

1. Ultrafine particles (UFP) reduced nitric oxide (NO) production
2. UFP-induced oxidative stress mediated the reduction in NO production
3. UFP induced eNOS S-glutathionylation
4. Inhibition of S-glutathionylation attenuated UFP-induced reduction in NO production

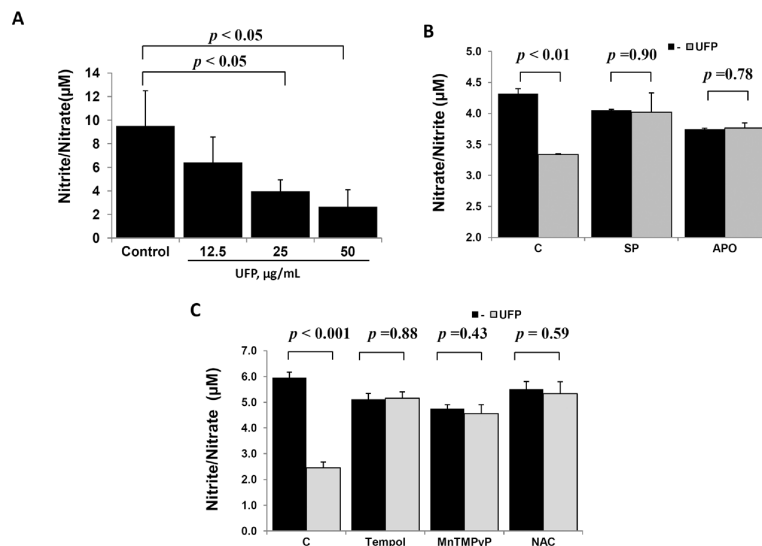


Fig. 1. UFP-induced oxidative stress reduced NO production

(A) HAEC were treated with an incremental concentration of UFP for 6 hours. NO production was assessed in terms of Nitrite /Nitrate concentration in cell lysate. (B) and (C) HAEC were pre-treated with $2\mu\text{M}$ of SP600125 (SP) or $200\mu\text{M}$ of Apocynin (APO), and with SOD1 mimetics, Tempol, at $200\mu\text{M}$, SOD2 mimetic, MnTMPyP, at $2\mu\text{g/mL}$, or an antioxidant N-acetyl Cysteine (NAC, 1mM) for 30 minutes, followed by co-treatment with or without $50\mu\text{g/mL}$ of UFP for 6 hours. Inhibition of JNK or NADPH oxidase in (B) as well as mitigation of ROS by antioxidants in (C) restored UFP-induced inhibition on NO production ($n=3$).

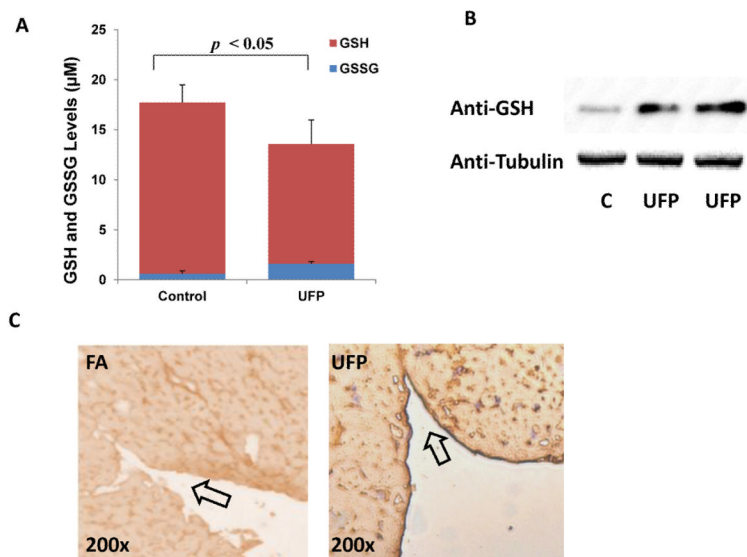


Fig. 2. UFP increased protein S-glutathionylation in HAEC

(A) HAEC were treated with or without 50µg/mL of UFP for 6 hours, cellular levels of GSH and GSSG were measured as described in Methods. UFP decreased GSH level, but increased GSSG level. (n=4) (B) HAEC were treated with or without 50µg/mL of UFP for 6 hours and protein lysates were collected. Western blots with antibody against GSH revealed an increase in a dominant band to S-Glutathionylated Actin. The western blot with anti-Tubulin was performed as loading reference. (C) Sections of endocardium from LDLR-null mice exposed to filtered air (FA) or UFP for 10 weeks were stained with anti-GSH antibody for visualization of protein S-glutathionylation. UFP exposure led to a prominent staining in the endothelium of endocardium (Arrow).

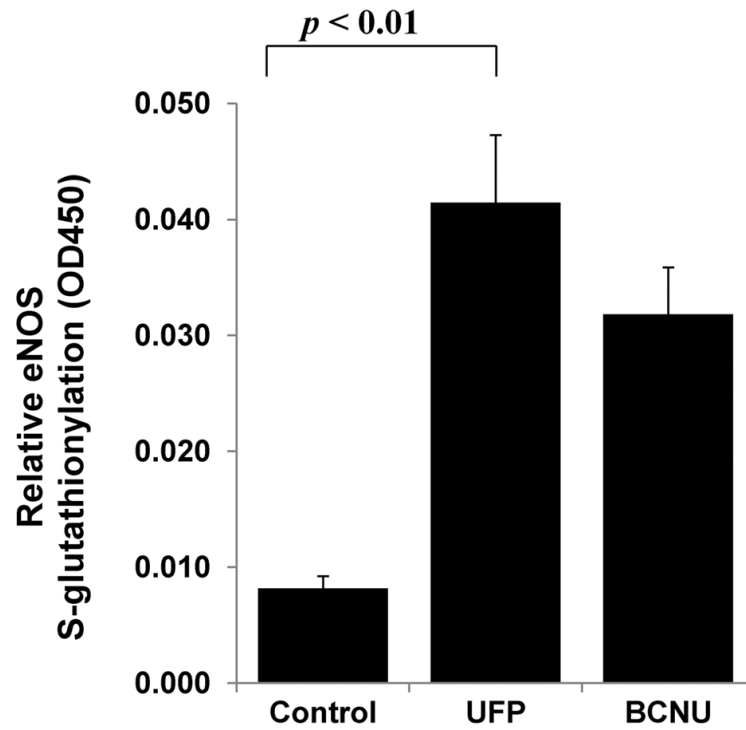


Fig. 3. UFP stimulated eNOS S-glutathionylation
HAEC were treated with 50 μ g/mL of UFP or 25 μ M BCNU (a positive control) for 6 hours. eNOS S-glutathionylation was measured as described in Methods. UFP significantly increased eNOS S-glutathionylation (n=3, $p < 0.01$).

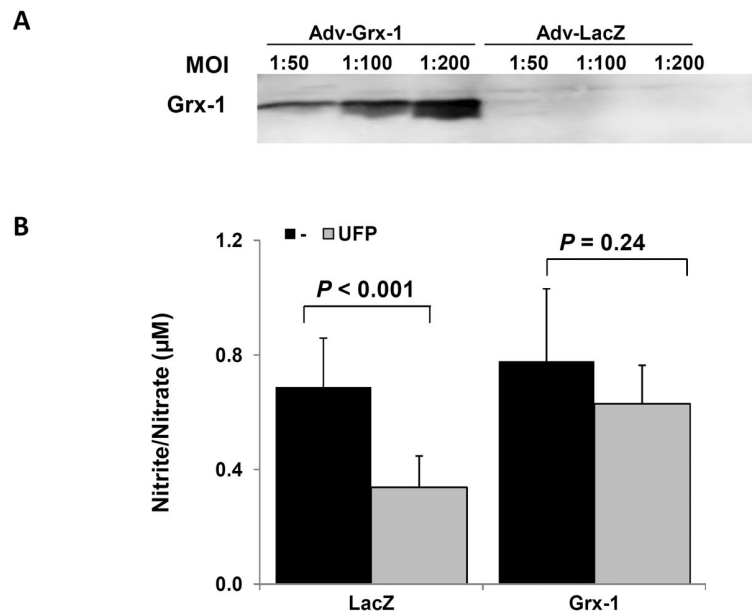


Fig. 4. S-glutathionylation mediated UFP-reduced NO Production

(A) HAEC were infected with control (Adv-LacZ) or Glutaredoxin-1(Adv-Grx-1) adenoviruses at different multiple of infection (MOI) overnight. Grx-1 expression was assessed by western blot. (B) HAEC were infected with Adv-LacZ or Adv-Grx-1 at MOI of 1:100 overnight. The cells were then treated with or without 50µg/mL of UFP for 6 hours. NO production was measured. Over-expression of Grx-1 attenuated UFP-mediated inhibition in NO production (n=6).