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Organic Nitrate Metabolism and Action: Toward a Unifying Hypothesis and the Future --- A Dedication to Professor Leslie Z. Benet

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Abstract

This review summarizes the major advances that had been reported since the outstanding contributions that Professor Benet and his group had made in the 1980's and 1990's concerning the metabolism and pharmacologic action of organic nitrates (ORN). Several pivotal studies have now enhanced our understanding of the metabolism and the bioactivation of ORN, resulting in the identification of a host of cysteine-containing enzymes that can carry out this function. Three isoforms of aldehyde dehydrogenase, all of which with active catalytic cysteine sites, are now known to metabolize, somewhat selectively, various members of the ORN family. The existence of a long-proposed but unstable thionitrate intermediate from organic nitrate metabolism has now been experimentally observed. ORN-induced thiol oxidation in multiple proteins, called the "Thionitrate Oxidation Hypothesis", can be used not only to explain the phenomenon of nitrate tolerance, but also the various consequences of chronic nitrate therapy, viz., rebound vasoconstriction, and increased morbidity and mortality. Thus, a unifying biochemical hypothesis can account for the myriad of pharmacological events resulting from nitrate therapy. Optimization of future uses of ORN in cardiology and other diseases could benefit from further elaboration of this unifying hypothesis.

Keywords

Aldehyde Dehydrogenase; Bioactivation; Metabolism; Nitrate Tolerance; Organic nitrates; Redox Signaling; Thiol Oxidation; Thionitrate

Contributions of the Benet Group to the Understanding of Organic Nitrate Action

From the mid-1980's to the late 1990's, Professor Benet and his associates carried out a number of pivotal studies that greatly enhanced our understanding of the pharmacological properties of nitroglycerin (NTG) and its dinitrate metabolites, viz., 1,2-glyceryl dinitrate (1,2-GDN), and 1,3-glyceryl dinitrate (1,3-GDN). Expanding on the assay originally described by Miyazaki et al.¹, Benet's group utilized electron capture detection to determine the plasma concentrations of these organic nitrates (ORN),²⁻⁴ which allowed them to characterize the metabolism and pharmacokinetics of NTG and its dinitrate metabolites after different routes of administration,⁵ including the intravenous/vascular route,⁶⁻⁸ cutaneous/transdermal application,⁹⁻¹⁴ sublingual⁸ and oral dosing.¹⁵⁻¹⁷

Benet and his group then followed with several studies which delineated the pharmacokinetic/pharmacodynamic relationships of NTG and its dinitrate metabolites in dogs,^{18,19} healthy volunteers,^{20,21} and in elderly conscious patients.²² NTG metabolism was first examined in rabbit hepatic tissue,^{23,24} which identified the involvement of glutathione-S-transferases (GST). The relative roles of GST isozymes in mediating NTG-induced vascular relaxation were further examined in bovine coronary artery²⁵ and rabbit aorta,^{26–29} resulting in the finding that the cytosolic mu isozyme of GST may be substantially involved in these preparations. Additionally, cytochrome P450 3A (CYP 3A) activity was found to correlate with *in vivo* bioactivation in rats, but its contribution to the overall NTG bioactivation was limited.³⁰ Finally, the role of thiol content in mediating NTG action and tolerance was investigated.^{30–32}

Benet and his group therefore had provided the most comprehensive information on the pharmacokinetic/pharmacodynamic behavior of nitroglycerin and (uniquely) its dinitrate metabolites in humans after clinical doses. Their work had contributed to the understanding of the relationship between enzyme bioactivation and the vascular relaxant activity of NTG, and of the complexity in identifying the critical enzyme(s) involved in this process. These studies have led to subsequent efforts by other investigators to delineate the complex relationships among metabolism, action and tolerance of ORN.

Where are We Now Concerning ORN Metabolism and Bioactivation?

Besides GST and CYP 3A mentioned above, several enzymes have been implicated in the vascular metabolism of NTG and other ORN, including other cytochrome P450 isoforms,^{30,33} xanthine oxidoreductase (XOR),³⁴ and various isoforms of aldehyde dehydrogenase (ALDH)^{35–37} (Table 1). Complicating the investigations and interpretation of ORN metabolism and bioactivation are two factors. First, not all members of this drug class are metabolized by the same enzyme(s). ORN can be loosely classified into two groups: (a) the high-potency ORN containing three or four nitrate groups, as exemplified by NTG and pentaerythritol tetranitrate (PETN), (b) the low-potency ORN which contain only one or two nitro groups, as exemplified by isosorbide dinitrate (ISDN), isosorbide-5-mononitrate (IS-5-MN), nicorandil, and the novel nitro-conjugated drugs such as NO-aspirin.³⁸ As discussed below in relation to each enzyme, several investigative groups have found that different enzymes display selectivity toward one group of ORN vs. the other.^{36,39,40}

Second, there are two major chemical pathways of denitration, viz: mechanism-based vs. clearance-based.⁴¹ The mechanism-based pathway (i.e., bioactivation) is associated with the generation of vasoactive nitric oxide (NO), with 1,2-GDN being the predominant metabolite from NTG denitration. On the other hand, the clearance-based pathway generates the weakly vasoactive nitrite ion (NO₂⁻) and 1,3-GDN from NTG metabolism. While an enzyme may preferentially produce a specific dinitrate metabolite at low substrate concentrations, this selectivity may be lost at higher concentrations, as well as under different reaction conditions.^{42,43} Thus, while an enzyme can be shown to metabolize an ORN at a certain substrate concentration, it is possible that the reaction results primarily in metabolic clearance, with an insignificant role, if at all, in bioactivation.

More extensive information are available regarding ORN metabolism by several enzymes/enzyme families, viz; GST, XOR, CYP450 and ALDH, and these findings are discussed in more detail below.

ORN Metabolism by Glutathione-S-Transferase

GST represents a large superfamily of ubiquitously expressed enzymes containing at least 9 separate classes comprised of multiple isoforms. GST, found in various subcellular fractions, is responsible for the conjugation of reduced glutathione to an electrophilic center of various endogenous compounds and xenobiotics as a mechanism for detoxification.⁴⁴ Although cytosolic GST was found to generate the 1,2-GDN metabolite preferentially, thus suggesting their likely role as bioactivating enzymes,^{24,45} its metabolism of NTG was not associated with NO production in the microsomes of bovine coronary aorta smooth muscle cells.⁴⁶ It has also been demonstrated that microsomal associated GST (MGST1) possesses the ability to denitrate NTG^{24,47} but it is associated with a notable lack of cyclic guanylyl cyclase activation (a marker of NO generation), and the generation of 1,3-GDN as the predominant metabolite, suggesting that this enzyme may be involved in the clearance, rather than the bioactivation of NTG.⁴⁷ The role of GST in the metabolism of ORN other than NTG is very poorly understood. While cytosolic GST possesses the ability to denitrate ISDN,⁴⁵ it is not known whether NO is concomitantly produced.

The mechanism-based inactivation of GST by NTG was extensively studied by Lee and Fung⁴⁸ who found thiol redox reactions to be critically involved. Exogenous glutathione inhibited this inactivation, which was characterized by GST dimerization, and not by tyrosine nitration. Since the spontaneous NO donor S-nitroso N-acetylpenicillamine did not inactivate GST at equal molar concentration to NTG, the mechanism of inactivation was unlikely to be mediated via NO exposure as such.

ORN metabolism by Xanthine Oxidoreductase

XOR is a complex homodimeric flavoenzyme which contains one molybdenum center, two ferredoxin iron-sulfur clusters and one flavin adenine dinucleotide (FAD) cofactor within each dimer.⁴⁹ It is responsible for purine degradation, and oxidizes hypoxanthine to xanthine and then to uric acid using NAD⁺ and/or molecular oxygen as cofactors. When first discovered, XOR was thought to be two different enzymes, i.e., xanthine oxidase (XO) and xanthine dehydrogenase (XDH) based on different cofactor requirements and differing expression patterns in various animals. However, it is now clear that XO and XDH represent the same enzyme which however exists in differing post-translational cysteine modifications, resulting in the alterations of its conformational structure.⁴⁹ XO contains the intact disulfide and utilizes molecular oxygen as the preferred electron acceptor while XDH contains the reduced disulfide and prefers NAD⁺ as the terminal electron acceptor.⁴⁹

Direct measurement of NO showed that, under hypoxic conditions, purified XO was able to reduce inorganic nitrite, nitrate, and NTG to vasoactive NO in the presence of the cofactor NADH. This reduction was inhibited by oxygen and specific XOR inhibitors.⁵⁰ The ability of XOR to reduce NTG was later confirmed using a platelet aggregation inhibition bioassay which is highly sensitive for detecting NO generation.³⁴ It is currently unknown which dinitrate metabolite of NTG is predominantly formed by XOR. In addition to NTG, purified XOR (both XO and XDH forms) can also bioactivate the lower potency ORN, viz., ISDN, IS-5-MN, and isosorbide-2-mononitrate (IS-2-MN) *in vitro* using either xanthine or NADH as a cofactor.⁵¹ Despite these encouraging *in vitro* results, the role of XOR *in vivo* has not been established, although a couple of factors might argue for its useful involvement in maintaining vascular function, i.e., XOR is highly expressed in the vascular endothelium,⁵² thus its ability to liberate NO from ORN under hypoxic conditions would be highly beneficial to correct the existing ischemia.

ORN metabolism by cytochrome P450

Servent et al.⁵³ first demonstrated that isolated rat hepatic microsomes were able to denitrate NTG and this reaction was prevented by CYP inhibition. Subsequent studies utilizing CYP enzyme inhibitors and inducing agents identified the principal isozymes involved, viz., CYP 3A and/or CYP 2B11.⁵⁴⁻⁵⁶ However, the bioactivating ability of CYP enzymes to release the vasoactive NO is less clear. Both positive³⁰ and negative^{57,58} findings regarding the *in vivo* role of the CYP enzymes in NTG bioactivation have been reported.

Only sparse data exist concerning CYP-mediated metabolism of ORN other than NTG. Using a microsomal preparation containing transfected human CYP, it was demonstrated that CYP 1A2, 2A6, 2C9, 2E1, 3A4 and 2J2 were able to bioactivate ISDN to liberate NO *in vitro*.^{59,60} In addition, it has been noted that CYP 1A2, 2E1, 3A4 and 2J2 are localized in human cardiac vascular tissue.^{59,60} The involvement of these CYP isozymes in the *in vivo* bioactivation of ORN is however still unexplored.

ORN metabolism by Aldehyde Dehydrogenase

ALDH is a super family of at least 17 ubiquitously expressed NAD/P⁺ dependent enzymes responsible for the oxidation of various endogenous and exogenous aldehydes to their corresponding carboxylic acids. They possess a range of functions including the detoxification of reactive aldehydes, biogenic amine synthesis, and maintenance of proper retinal (vitamin A) signaling.⁶¹⁻⁶³

Mitochondrial ALDH (ALDH2) was identified in 2002 as a major enzyme responsible for the bioactivation of NTG,³⁵ producing 1,2-GDN as the predominant metabolite, thus consistent with mechanism-based metabolism.⁴² This finding was confirmed via follow-up studies utilizing specific ALDH2 inhibitors, an ALDH2 null mouse model,⁶⁴⁻⁶⁶ and in clinical investigations.^{67,68} This enzyme is also critical to the bioactivation and *in vivo* activity of PETN. However although purified ALDH2 can bioactivate several lower potency ORN (e.g., ISDN and nicorandil) *in vitro*, it plays an insignificant role in the *pharmacological* activity of these agents.^{36,39,64}

Two cytosolic forms of ALDH, ALDH1a1 and ALDH3a1, have been demonstrated to bioactivate NTG *in vitro* producing the 1,2-GDN metabolite predominantly.^{43,69} In addition, these isoforms can metabolize several lower-potency ORN such as ISDN, IS-5-MN, and nicorandil, although the substrate selectivity varies somewhat between the isoforms.⁶⁹ While purified ALDH1a1 can bioactivate ORN *in vitro*, studies with ALDH1a1 null mice suggest that the isoform is not critical to the *in vivo* activity of ORN, most likely due to a lack of expression in the vascular tissue.⁷⁰ However, ALDH1a1 is heavily expressed in the liver where it may serve as a clearance enzyme for these drugs. The *in vivo* role of ALDH3a1 as well as the role of other ALDH isoforms in ORN metabolism is still unclear at present.

ORN Metabolism: A Tower of Babel? Or can all the findings be unified?

It is now clear that ORN metabolism and bioactivation are mediated by multiple enzymes, some perhaps even unidentified at this point. Why is there such a lack of enzyme specificity for this class of compounds? In 2004, we proposed an overall biochemical scheme, for all involved enzymes, that could account for ORN bioactivation and its subsequent mechanism-based inactivation. Because this mechanism proceeds through a unifying intermediate, viz., a thionitrate, we have termed it the “Thionitrate Oxidation Hypothesis”.⁷¹ While this unstable intermediate had long been proposed for the biochemical action of ORN, it avoided direct detection until we documented its existence recently.⁷²

We proposed that the locus of enzyme reaction with ORN, for most if not all metabolizing enzymes, involves one or more critical cysteine residue(s) to produce a thionitrate initially (Fig. 1). This highly facile intermediate can then interact with excess free thiol(s) to liberate inorganic nitrite ion while forming a disulfide between the two involved thiols, leading to clearance-based metabolism. On the other hand, the thionitrate may also liberate NO via a mechanism-based metabolic pathway, producing a sulfenyl intermediate [P_xS-OH] in the protein (P_x denotes the reactive protein, $x = 1, 2, \dots, n$). Subsequent oxidation of these unstable thiol intermediates would lead to formation of further oxidation modifications of the proteins (P_xS-SP_x , P_xSO_2H , P_xSO_3H and P_xS-SG , where G represents a glutathione residue).

The promiscuity of reaction between the nitrate moiety in ORN with protein cysteine residues is not surprising because it has long been known (see for example⁷³) that ORN react avidly even with non-protein thiols to produce NO. Site mutagenesis studies have also identified critical cysteine groups in ALDH2 (cysteine 319) and ALDH1a1 (cysteine 303) as essential for metabolizing ORN.³⁶ It was also recently demonstrated in an animal model that NTG-induced vasodilation was mediated by oxidation of critical cysteine residues in protein kinase G 1 alpha, the oxidation of which is associated with nitrate tolerance.⁷⁴ The sequential nature of the thiol oxidation scheme shown in Fig. 1 also provides an explanation why ORN-mediated enzyme inactivation can sometimes be reversible (e.g., with sulfenyl and S-glutathionylated proteins), while not so at other times because irreversible products are formed (e.g., sulfinic and sulfonic acids). This partially reversible nature of cysteine oxidation by NTG was demonstrated for GST⁴⁸ and ALDH2⁷⁵. All these reaction products with NTG have been observed recently by us using LCMS/MS.⁷²

How does the Thionitrate Oxidation Hypothesis explain pharmacological tolerance?

Another characteristic of repeated ORN therapy is the rapid development of pharmacological tolerance. This effect has been observed with all ORN via all routes of administration, and is accompanied by multiple consequences, including the inactivation of bioactivating enzymes,⁶⁸ depletion/oxidation of free thiol,⁷⁶ counter-regulatory responses such as rebound vasoconstriction,^{77,78} and increased oxidative stress in the vasculature leading to endothelial dysfunction.^{79,80} The multiplicity of these effects, some apparently unconnected, is not surprising when viewed in the context of the biochemical reaction scheme shown in Fig. 1. Because multiple proteins are affected by ORN-mediated oxidation, whether NO is produced or not as a result, their oxidation can lead to a wide array of pharmacological consequences, as determined by the specific proteins involved. In addition, abrupt discontinuation of short acting ORN leads to the phenomenon of withdrawal rebound.

Thus, as shown in Fig. 2, the oxidation of active site cysteine residues in bioactivating enzymes such as ALDH2 will result in reduced bioactivation of NO, leading to pharmacological tolerance observed in animal models⁶⁵ and in clinical studies⁶⁸. It has also been demonstrated that NTG can inactivate ALDH1a1,³⁶ ALDH3a1,⁶⁹ and several GST⁴⁸ and CYP isoforms.⁸¹ ISDN and IS-2-MN can inactivate ALDH1a1, ALDH2 and ALDH3a1 while IS-5-MN can inactivate ALDH1a1 *in vitro*.³⁶

Similarly, inactivation of clearance-based enzymes such as GST will lead to drug accumulation (concomitant with tolerance induction), as shown in patients for both NTG⁸² and ISDN.⁸³ The induction of counter-regulatory responses may potentially be mediated through the S-oxidation of signaling proteins, such as the small GTPase secondary messenger p21ras, and Kelch-like ECH-associated protein 1 (KEAP1), e.g., through the formation of sulfenyl and S-glutathionylated modifications. The actual roles of signaling

proteins or pathways in mediating nitrate tolerance remain undefined currently, as involvement of other thiol proteins is being discovered.⁷⁴ Although our hypothesis of thiol oxidation indeed produces a state of oxidative stress in the vasculature, it differs from the superoxide oxidative stress hypothesis that was quite prevalently accepted in the last decade.³⁸ We have shown recently that increased superoxide formation was not a cause of nitrate tolerance, but rather as a consequence of ORN-mediated S-oxidation.⁸⁴ Our hypothesis also differs from suggestion that endothelial nitric oxide synthase (eNOS) is critically involved in nitrate tolerance³⁸ since we have shown that eNOS knockout mice exhibits a similar degree of vascular tolerance toward NTG as the wild-type mice.⁸⁵

Viewing Tolerance Avoidance Findings through our Unifying Hypothesis

Tolerance avoidance strategies toward ORN can also be interpreted through Fig. 2. The use of nitrate-free period to regenerate nitrate sensitivity is consistent with the regeneration of the appropriate vascular redox state through, e.g., the thioredoxin or glutaredoxin enzyme system. Reversal or prevention of protein cysteine oxidation, and thereby nitrate tolerance, can be understandably accomplished by co-administration of antioxidants or reduced thiols. These agents include N-acetylcysteine,⁸⁶ vitamin C,^{87,88} vitamin E,⁸⁹ folic acid,^{90,91} lipoic acid,⁹² and possibly carvedilol,⁹³ hydralazine,⁹⁴ and statins.⁹⁵ Recently, ALDA-1, a small molecule inducer of ALDH2 expression and activity has been demonstrated to prevent tolerance formation through increased availability of this enzyme.⁹⁶

Other tolerance avoidance agents may act to overcome the counter-regulatory responses produced by ORN. In the rat aorta, NTG exposure is associated with non-specific S-glutathionylation of proteins (a consequence of thiol oxidation), concomitant with decreased bioactivation and action of NTG.⁸⁴ Oxidation of proteins responsible for signal transduction may also be partly responsible for the observed changes in vascular gene expression.⁹⁷ In cultured endothelial cells, it was demonstrated that NTG exposure increased S-glutathionylation and activity of p21ras, which is integral to the signaling of the AKT and extracellular signal-regulated kinase 1/2 (ERK 1/2) pathways which play a part in the regulation of smooth muscle cell proliferation.⁹⁸ NTG incubation led to a significant increase in the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B p50/p65) activity in differentiated human monocytic leukemia cells,⁹⁹ possibly through nuclear factor kappa-B kinase subunit alpha (IKK- α) activation via its cysteine-179 residue.

To counter the vasoconstrictive effects of nitrate tolerance, diuretics¹⁰⁰ and anti-vasoconstrictive agent such as captopril and losartan have been used.¹⁰¹ Interestingly, PETN has been reported to induce less tolerance formation than the other ORN, likely through its unique ability to upregulate hemeoxygenase-1 (HO-1).¹⁰² In addition, this induction of HO-1 can prevent and reverse the oxidative stress-mediated endothelial dysfunction.¹⁰³ HO-1 is one of several antioxidant response elements (ARE) that are unregulated in response to increased oxidative stress as a protective mechanism. ARE expression is controlled via the NRF2 signaling pathway, and its upregulation is mediated by cysteine oxidation of NRF2's inhibitor KEAP1. Maintained as an inactive NRF2/KEAP1 dimer in the cytosol, oxidation of KEAP1 allows NRF2 dissociation and translocation into the nucleus to upregulate ARE.¹⁰⁴

Cyclic guanosine monophosphate, the secondary messenger responsible for NO mediated vasorelaxation, is degraded in the vasculature by various phosphodiesterase (PDE) isoforms and specific inhibition of PDE5 by 4-[[3,4-(Methylenedioxy)benzyl]amino]-6-chloroquinazoline (MBCQ) has been demonstrated to prevent tolerance formation presumably through increased persistence of this messenger.¹⁰⁵ This approach addresses the down-stream effects of ORN, and it does not prevent the initial tolerance-inducing

biochemical reactions of thionitrate formation and protein thiol oxidation. Thus, while the net potency of ORN is increased by PDE inhibitors, the root cause of nitrate tolerance is not abrogated by these agents.

Table 2 summarizes the various agents that have been used for the *in vivo* prevention of tolerance and the experimental systems employed, and their likely mechanisms of action. However, it is likely that some agents may exert multiple mechanism that have not yet been fully characterized.

Organic Nitrate Toxicity: Can it also be explained by thiol oxidation?

In the last decade, pharmacoepidemiological studies have revealed the existence of possible deleterious effects, in terms of increased mortality and morbidity, for patients who are on chronic nitrate therapy.^{106,107} Although prospective studies will need to be carried out to confirm these analyses, there are two potential mechanistic explanations for this phenomenon, based on the reaction scheme shown in Fig. 2.

In the heart, ALDH2 is responsible for the detoxification oxidation of reactive toxic aldehydes (e.g., 4-hydroxy-2-nonenal) that are generated via lipid peroxidation during periods of ischemia and reperfusion.^{108,109} As discussed earlier, NTG has the ability to reduce cardiac ALDH2 activity via mechanism-based inactivation of this enzyme, and this effect has been demonstrated, in an *ex vivo* heart model, to increase cardiac damage after ischemia/reperfusion injury.¹⁰⁸ Mechanisms to restore cardiac ALDH2 activity, such as administration of the small molecule inducer of ALDH2, ALDA-1, have been proposed as a therapy to minimize damage after cardiac infarction.¹⁰⁹

In addition, ORN have the ability to modulate proteins responsible for maintaining the fibrous cap of atherosclerotic plaques which is comprised of extracellular matrix components such as collagen. This cap can degrade and weaken, ultimately leading to rupture of the plaque, platelet adhesion and thrombus, and ultimately resulting in infarction. We have shown that NTG exposure, both *in vitro* and *in vivo*, can activate matrix metalloproteinases (MMP) which degrade components of the extracellular matrix, leading to weakening of the fibrous cap.^{99,110} The mechanism of activation of MMP by NTG involves the oxidation of the “cysteine switch”¹¹¹ which converts the inactive Pro-MMP to its active protein.

Thus, based on the analysis of the available findings, a potential unifying hypothesis (Fig. 2) that focuses on the oxidation of protein cysteine residues in multiple proteins can be used to rationalize on the multi-faceted actions of organic nitrates, including its metabolism, bioactivation, enzyme inactivation, vascular tolerance, and long-term toxicity.

Looking Forward: Clinical Uses of Organic Nitrates beyond Cardiology

The involvement of NO in many physiological and pathological conditions is now well documented. Because ORN can serve as clinical NO donors, their pharmacological use in humans have been extended beyond cardiology. Below we summarize some of the more mature areas of studies that suggest the potential extension of ORN use in other diseases.

Treatment of osteopenia/osteoporosis

Bone turnover is a tightly controlled process that relies on a variety of humoral and micro-environmental signals that ultimately alter the function of the catabolic osteoclasts and the anabolic osteoblasts. One of these signal molecules is NO which is endogenously generated by both osteoblasts and osteoclasts, with a net inhibitory effect on osteoclast activity preventing bone resorption. In a variety of animal models, it has been demonstrated that

modulating NO exposure via stimulation of endogenous NOS activity^{112,113} or administration of topical ORN^{114,115} can attenuate the reduction of bone mineral density due to osteoporosis. A prospective case-controlled study in Denmark had examined the administration of ORN and the incidence of fracture in over 100,000 subjects. Use of ORN (IS-5-MN, ISDN, or NTG) was associated with an approximately 11% reduction in any fracture in both men and women and a 15% reduction in hip fractures in women. The risk reduction was dose- and duration-dependent.¹¹⁶ While these studies have demonstrated promising results, randomized controlled studies have yielded mixed conclusions regarding the therapeutic benefits of ORN. A small study randomized 186 female post-menopausal subjects to receive topical NTG or placebo. At the end of their 3-year study, topical ORN administration was associated with no change in bone mineral density compared to baseline although the results were confounded by compliance problems due to adverse events such as headaches.¹¹⁷ A second randomized study examined 205 post-menopausal women who received topical NTG or placebo for 2 years. At the end of the study period, there was a small but significant improvement in bone density as well as markers in bone turnover in the subjects receiving NTG.¹¹⁸ Neither study was powered sufficiently to examine fractures. Because of these conflicting results, more studies are required to define the role of ORN in the treatment of osteoporosis.

Protection of the Gastric Mucosa

NO exerts a protective effect on the gastric mucosa via several mechanisms, including promotion of angiogenesis around gastric ulcers, promotion of adequate blood flow, modulation of the local inflammatory response and the promotion of mucus secretion.^{119,120} Inhibition of endogenous NO production via inhibition of nitric oxide synthase (NOS) slows ulcer healing in animal models of gastric ulceration^{121,122} while supplementation of exogenous NO prevented ischemia-induced gastrointestinal damage.¹²³

The ability of NO to prevent gastric ulcers has garnered the most interest in the prevention of ulcers induced by non-steroidal anti-inflammatory drugs (NSAID) including aspirin. These agents act through cyclooxygenase (COX) 1 and 2 inhibition, leading to altered prostaglandin signaling in the gastrointestinal tract, direct irritation of the mucosal lining and, for aspirin, irreversible inhibition of platelet aggregation.¹²⁴ This has led to a new class of drug, the NO-releasing drug hybrids called NO-NSAIDs including NO-aspirin. These drugs contain a nitro group attached to the parent drug via a labile linker (generally an ester linkage) which is hydrolyzed *in vivo* to release the parent drug and the linker containing the nitro moiety which is further metabolized to liberate NO.¹²⁰ Through these hybrid drugs, the severity and incidence of the gastric adverse effects of NSAIDs may be reduced without compromising their beneficial effects. Thus, NO-aspirin has been demonstrated to prevent and reverse animal models of gastric damage.¹²⁵ Two small proof-of-concept studies using healthy volunteers showed that after 7 days and 14 days of administration, NO-aspirin was associated with less gastric damage (determined via endoscopic study) compared to aspirin.^{126,127} Both NO-aspirin and NO-naproxen are currently in clinical trial.^{128,129}

Use of NTG to normalize ALDH expression in overexpressing malignancies

The over-expression of ALDH1a1 and subsequent alteration of retinal (vitamin A) signaling is a potential factor of malignancy aggressiveness that has been noted in a variety of cancers, including lung,¹³⁰ prostate,¹³¹ and breast cancer.¹³² It has been demonstrated that downregulation of over expressed ALDH1a1 *in vitro* via siRNA has the potential to inhibit tumor growth.¹³⁰ In addition, the increased expression of ALDH1a1 serves as a chemotherapeutic resistance mechanism because this enzyme has the ability to detoxify the active metabolites of cyclophosphamide-based alkylating agents.

The ability of NTG to inactivate various ALDH isoforms, via irreversible oxidation of the enzyme's catalytic cysteine residues,^{36,40} could potentially be exploited to reverse this over-expression of this enzyme's activity, with the potential to reduce tumor virulence while sensitizing the tumor toward alkylating agents. In contrast to the more complex (and therapeutically difficult) approach of employing siRNA, NTG has a proven track record of clinical safety with minimal and easily managed adverse effects (headaches and hypotension). We believe, therefore, that ORN use in modulating cancer growth deserves further investigation.

Finally, several non-cardiovascular uses of ORN rely in its vasodilatory action to restore normalized blood flow in pathological tissue/organs. These applications include the treatment of erectile dysfunction,^{133,134} anal fissures,^{135,136} and the management of esophageal varices due to portal hypertension.¹³⁷

Conclusion

From the early groundwork by Benet and his associates who provided critical information on the pharmacokinetics and pharmacodynamics of NTG and its dinitrate metabolites, significant progress has been made to gain understanding of this well-established class of drugs. We believe that ORN are metabolized by a variety of enzymes because of the promiscuous reaction between ORN and the many cysteine residues available in proteins. However, not all of these reactions produce the necessary vasoactive NO. The promiscuous cysteine-nitrate reaction also explains the multiplicity of pharmacologic effects that have been observed after chronic nitrate use, including pharmacological tolerance, reduced ORN systemic clearance, induction of counter-regulatory vasoconstriction and increased morbidity/mortality. Most of the findings relating to tolerance avoidance strategies can also be explained by this mechanism. Thus, a unifying hypothesis linking the metabolism and action of organic nitrates is available, and it may find utility as a biochemical basis to further optimize and extend the use of this group of useful pharmacological agents.

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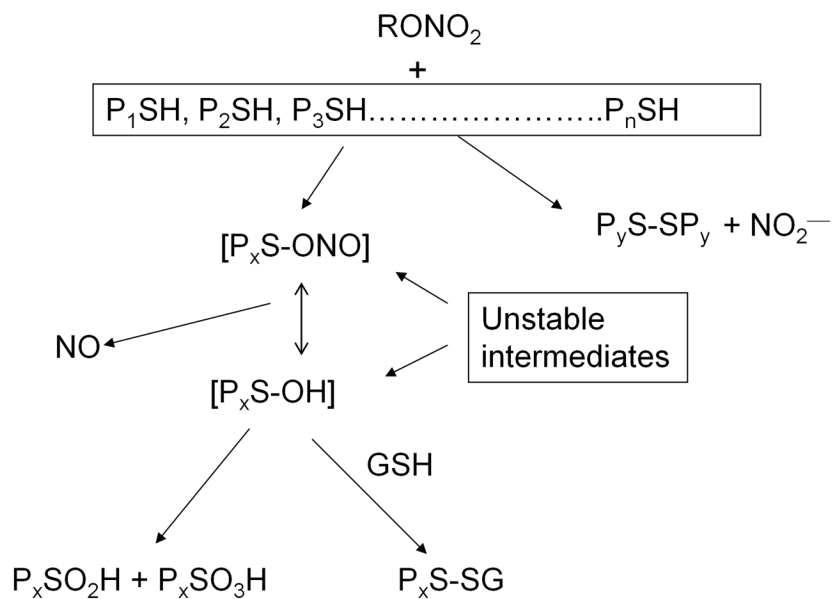


Fig. 1. The Thionitrate Oxidation Hypothesis of organic nitrate metabolism, bioactivation, and enzyme inactivation. PS = cysteine-containing proteins; SG = glutathione residue, $-\text{SO}_2\text{H}$ = sulfinic acid protein modification and $-\text{SO}_3\text{H}$ = sulfonic acid modification. Adapted from Fung⁷¹

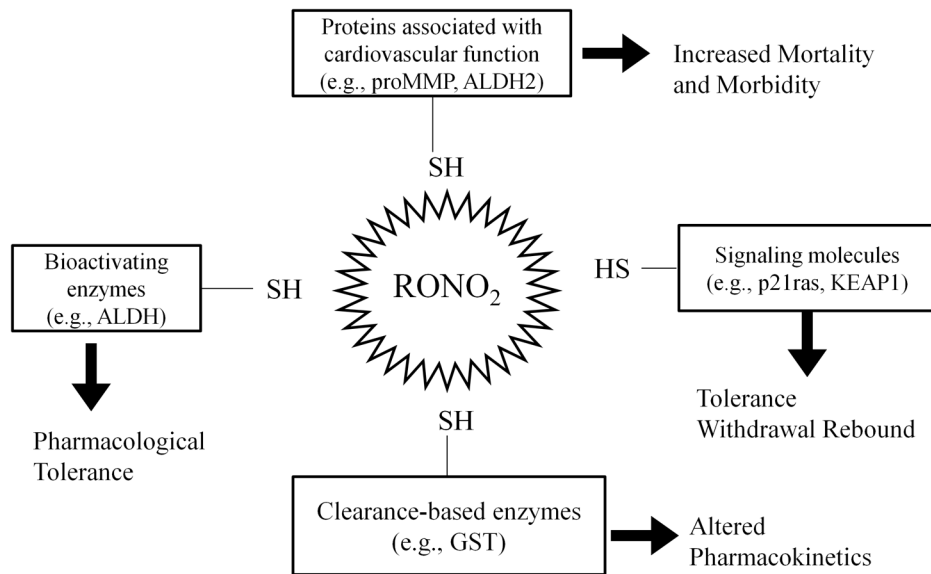


Fig. 2. Schematic showing how thiol oxidation in different proteins by organic nitrates (RONO_2) can lead to a wide array of pharmacological consequences. Adapted from Page and Fung¹⁴³

Table 1

Summary of enzymes known to metabolize organic nitrates

Class	Isoforms	ORN Metabolized	ORN NOT Metabolized	Predominant NTG metabolite	References
GST	GSTmu	NTG, ISDN	nicorandil	1,2-GDN	24,45
	MGST1	NTG		1,3-GDN	24,47
Xanthine Oxidase		NTG, ISDN, IS-5-MN, IS-2-MN		1,2-GDN	34,50,51
	3A4	NTG, ISDN, NO-aspirin		N.D.	54-56,59,60
Cytochrome P450	2B11	NTG		N.D.	54-56
	2J2, 1A2, 2A6, 2C9, 2E1	ISDN, NO- aspirin		N.D.	59,60
Aldehyde Dehydrogenase	ALDH1a1	NTG, ISDN, IS-5-MN, IS-2-MN, nicorandil		1,2-GDN	42,43
	ALDH2	PETN, NTG, ISDN, nicorandil	IS-2-MN, IS-5-MN	1,2-GDN	35,64,62,138
	ALDH3a1	NTG, ISDN, IS-2-MN, nicorandil	IS-5-MN	1,2-GDN	69

(N.D. = no available data)

Table 2Agents demonstrated to prevent or avoid organic nitrate tolerance *in vivo*

Agent	ORN Examined	Likely Mechanism	Experimental Model	References
ALDA-1	NTG	ALDH2 upregulation	Normal Rats	96
Atorvastatin	NTG	Anti-oxidant	Health Volunteers	95
Captopril	ISDN	RAS pathway modulation and anti-oxidant	CAD Patients	101
Carvedilol	NTG	Anti-oxidant	HTN Patients	93
Folic Acid	NTG	Anti-oxidant	Health Volunteers	90,91
Hydralazine	NTG	Anti-oxidant and vasodilation	CHF-induced Rats	94
			Normal Rabbit	139
Hydrochlorothiazide with amiloride	ISDN	Diuresis	CAD Patients	100
L-arginine	NTG	eNOS mediated	CAD patients	140
Lipoic acid	NTG	Anti-oxidant	Normal Rats	92
Losartan	ISDN	RAS pathway modulation	CAD Patients	101
MBCQ	NTG	PDE inhibition	Normal Rats	105
N-acetylcysteine	NTG	Anti-oxidant and thiol supplementation	CHF Patients	86
Pravastatin/atorvastatin	NTG	Anti-oxidant	Normal Rats	141
Vitamin C	IS-5-MN	Anti-oxidant	Health Volunteers	80
Vitamin C	NTG	Anti-oxidant	CHF Patients	87
Vitamin E	NTG	Anti-oxidant	Obese Zucker Rat	142
			Health Volunteers and CAD patients	89

CHF=congestive heart failure, HTN=hypertension, CAD=coronary artery disease, MBCQ=4-[[3,4-(Methylenedioxy)benzyl]amino]-6-chloroquinazoline