

High-Quality Draft Genome Sequence of *Pseudomonas syringae* pv. Syringae Strain SM, Isolated from Wheat

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Pseudomonas syringae is one of the most widespread plant pathogens that can cause significant damage to crop plantations. Here, we announce a noncontiguous finished genome sequence of *Pseudomonas syringae* pv. syringae strain SM, isolated from hexaploid wheat. The genome sequence revealed the smallest described complement of type III effectors.

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Pseudomonas syringae strains have been isolated from >180 host species across the entire plant kingdom, including many agriculturally important crops (1). The observed wide host range is reflected in a relatively large genetic heterogeneity among different pathovars. This is most pronounced in the complement of virulence factors, which is also assumed to be the key factor defining the host specificity (2). *Pseudomonas syringae* pv. syringae strain SM was isolated from hexaploid wheat (*Triticum aestivum*) in the United States (3). The strain, which was also denoted D20, has been used in several studies addressing the issue of bacteriuminduced systemic resistance in plants (3–7) but never as an infection model for wheat.

A 3-kb paired-end library was generated and sequenced at the Functional Genomics Center Zurich on a Roche genome sequencer FLX+ platform. A total of 974,051 quality filtered reads with a total of 213,333,037 bases were obtained, resulting in 34.8fold average sequencing coverage. The obtained reads were further de novo assembled using Newbler 2.5.3 into 64 contigs combined into one 6.08-Mb-long superscaffold and 3 smaller scaffolds (46.5 kb, 9.09 kb, and 5.24 kb in size). The largest of the minor scaffolds turned out to be a pPT23A family plasmid, the 9-kb scaffold showed sequence similarity to nonribosomal peptide synthase (NRPS) modules, and the smallest scaffold constituted an rRNA operon. A portion of intrascaffold gaps was closed by sequencing of PCR products using Sanger technology, decreasing the total number of contigs to 26. However, it was not possible to precisely map the 9-kb scaffold, but due to its insignificance to the project, it was excluded from the assembly. Initial open reading frame (ORF) prediction and functional annotation were performed using the RAST server (8). The start codons of all the predicted ORFs were further manually verified using the position of potential ribosomal binding sites and BLASTp (9) alignments with homologous ORFs from other Pseudomonas strains as a reference. Functional annotations were also refined for every ORF using BLASTp searches against the nonredundant protein sequence database (nr) and the NCBI Conserved Domain search engine (10).

The estimated genome size of strain SM is 6,124,102 bp, with

an average G+C content of 58.73%. It contains 5,072 proteincoding sequences (excluding pseudogenes), five rRNA operons, and 64 tRNA genes for all of the amino acids. Notably, it contains a complete type III secretion system and seven known effector proteins: AvrE1, HopAA1, HopI1, HopM1, HopBA1, HopA2, and HopAZ1. In addition, there are two complete type VI secretion system gene clusters and 12 putative effector proteins belonging to the VgrG and Hcp1 families, as well as intact gene clusters for the biosynthesis of syringopeptin and mangotoxin. All of these genome components have previously been demonstrated to be involved in virulence and epiphytic fitness of *P. syringae*, as well as in competition of pseudomonads with other microbial species (11–16).

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. APWT00000000. The version described in this paper is the first version, APWT01000000.

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REFERENCES

- 1. Bradbury JF. 1986. Guide to plant pathogenic bacteria, p 175–177. CAB International Mycological Institute, Kew, England.
- Lindeberg M, Cunnac S, Collmer A. 2009. The evolution of *Pseudomonas syringae* host specificity and type III effector repertoires. Mol. Plant Pathol. 10:767–775.
- Smith JA, Métraux J-P. 1991. Pseudomonas syringae pv. syringae induces systemic resistance to Pyricularia oryzae in rice. Physiol. Mol. Plant Pathol. 39:451–461.
- Smith JA, Hammerschmidt R, Fulbright DW. 1991. Rapid induction of systemic resistance in cucumber by *Pseudomonas syringae* pv. syringae. Physiol. Mol. Plant Pathol. 38:223–235.
- Vincent JR, Fulbright DW. 1983. Transfer of pRD1 to *Pseudomonas* syringae and evidence for its integration into the chromosome. J. Bacteriol. 156:1349–1351.
- 6. Rasmussen JB, Hammerschmidt R, Zook MN. 1991. Systemic induction

of salicylic acid accumulation in cucumber after inoculation with *Pseudomonas syringae* pv syringae. Plant Physiol. **97**:1342–1347.

- Ramel C, Tobler M, Meyer M, Bigler L, Ebert MO, Schellenberg B, Dudler R. 2009. Biosynthesis of the proteasome inhibitor syringolin A: the ureido group joining two amino acids originates from bicarbonate. BMC Biochem. 10:26. doi:10.1186/1471-2091-10-26.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. doi:10.1186/1471-2164-9-75.
- 9. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J. Mol. Biol. 215:403–410.
- Marchler-Bauer A, Lu S, Anderson JB, Chitsaz F, Derbyshire MK, DeWeese-Scott C, Fong JH, Geer LY, Geer RC, Gonzales NR, Gwadz M, Hurwitz DI, Jackson JD, Ke Z, Lanczycki CJ, Lu F, Marchler GH, Mullokandov M, Omelchenko MV, Robertson CL, Song JS, Thanki N, Yamashita RA, Zhang D, Zhang N, Zheng C, Bryant SH. 2011. CDD: a conserved domain database for the functional annotation of proteins. Nucleic Acids Res. 39:D225–D229. doi:10.1093/nar/gkq1189.
- 11. Bender CL, Alarcón-Chaidez F, Gross DC. 1999. Pseudomonas syringae

phytotoxins: mode of action, regulation, and biosynthesis by peptide and polyketide synthetases. Microbiol. Mol. Biol. Rev. **63**:266–292.

- 12. Lindeberg M, Cunnac S, Collmer A. 2012. *Pseudomonas syringae* type III effector repertoires: last words in endless arguments. Trends Microbiol. 20:199–208.
- Cunnac S, Chakravarthy S, Kvitko BH, Russell AB, Martin GB, Collmer A. 2011. Genetic disassembly and combinatorial reassembly identify a minimal functional repertoire of type III effectors in *Pseudomonas syringae*. Proc. Natl. Acad. Sci. U. S. A. 108:2975–2980.
- Arrebola E, Cazorla FM, Codina JC, Gutiérrez-Barranquero JA, Pérez-García A, De Vicente A. 2009. Contribution of mangotoxin to the virulence and epiphytic fitness of *Pseudomonas syringae* pv. syringae. Int. Microbiol. 12:87–95.
- 15. Haapalainen M, Mosorin H, Dorati F, Wu RF, Roine E, Taira S, Nissinen R, Mattinen L, Jackson R, Pirhonen M, Lin NC. 2012. Hcp2, a secreted protein of the phytopathogen *Pseudomonas syringae* pv. tomato DC3000, is required for fitness for competition against bacteria and yeasts. J. Bacteriol. 194:4810–4822.
- Hood RD, Singh P, Hsu F, Güvener T, Carl MA, Trinidad RR, Silverman JM, Ohlson BB, Hicks KG, Plemel RL, Li M, Schwarz S, Wang WY, Merz AJ, Goodlett DR, Mougous JD. 2010. A type VI secretion system of *Pseudomonas aeruginosa* targets a toxin to bacteria. Cell Host Microbe 7:25–37.