Regulatory role of nitric oxide in lipopolysaccharides-triggered plant innate immunity

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Recent studies have suggested that lipopolysaccharides (LPS) induce nitric oxide (NO) production and defense gene expression in plants. Our current work investigated the signaling mechanism of NO and the role of NONEXPRESSOR OF PATHOGENESIS-RELATED GENES1 (NPR1) in LPS-induced innate immunity of Arabidopsis (Arabidopsis thaliana). We have provided evidence that LPS-elicited NO generation as well as increased antioxidant enzyme activities capable of maintaining the redox state could be important to protect plants against oxidative damage from pathogen attack. In addition, LPSactivated defense responses, including callose deposition and defense-related gene expression, are regulated through an NPR1-dependent signaling pathway. Our results contribute to elucidation of the signaling mechanism of NO and highlight an important role of NPR1 in modulating LPS-triggered innate immunity in plants. However, further research is necessary to clarify the cross-talk between mitochondria and NO on activating LPS-induced defense responses, and the regulatory mechanism of NO in LPS-induced innate immunity needs further improvement.

Emerging evidence suggests that pathogen-associated molecular patterns (PAMPs) are potent elicitors that could induce defense mechanisms against bacterial, fungal and viral pathogens.¹ Bacterial Lipopolysaccharides as the prototypical PAMP can induce various plant defenserelated responses, including the oxidative burst, nitric oxide (NO) generation, cell-wall alteration and pathogenesisrelated (PR) gene expression.²⁻⁴ NO is emerging as an important multifunctional signaling molecule in plants.^{5,6} During defense-related responses, NO production is an important hallmark of innate immunity elicited by LPS.7 Two key enzyme pathways are responsible for plant NO synthesis: oxidation of arginine to citrulline by a nitric oxide synthase (NOS)-like enzyme and reduction of nitrite to NO by a nitrate reductase (NR).8 Although no NOS gene has yet been identified in plants, NOS activity as well as inhibition of NO synthesis by animal NOS inhibitors has been reported in plants.9,10 NO mediates Tobacco mosaic virus (TMV)induced AOX induction that triggers systemic basal defense against viral pathogen, and as a redox regulator of the NPR1/ TGA1 system promotes the nuclear translocation of NPR1.11 NPR1 plays important roles in activating defense gene and mediating plant defense response.¹² These findings prompt us to further investigate the signaling mechanism of NO and the important role of NPR1 in LPS-induced innate immunity.

Accumulating experimental evidence support that LPS as a plant defense activator can lead to the production of NO and play a key role in plant disease resistance.^{7,13} Our current work has shown that LPS as a typical PAMP could directly induce defense-related responses including *PR1* gene induction and callose deposition.¹⁴ However, some biological agents and synthetic compounds cannot induce *PR1* expression and callose deposition per se until pathogen infection; these induced reactions are frequently associated with

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a primed state in which the plants are able to "recall" previous infection, root colonisation or chemical treatment.15-17 In consequence, primed plants respond more rapidly and/or effectively to subsequent attack. It has been suggested that priming comes at much lower costs than direct-defense induction.¹⁶ However, unlike direct defense induction, which could be determined directly by measuring various defense markers, priming is mostly analyzed by examining the speed and intensity of defense responses after disease exposure. To assess the mechanism and significant roles of priming will be a challenge for the future.

Our genetic and pharmacological analysis also provides evidence that arginine-dependent NOS-like enzyme is likely responsible for the biosynthetic mechanism involved in LPS-induced NO generation. Interestingly, the NR activity was found to be slightly inhibited by LPS treatment. The generated NO is not stable and would format the stable degradation of products, nitrate and nitrite. It has been proposed that post-translational regulation of NR takes place in response to various treatments, and nitrite accumulation is likely to be eliminated by post-translational inactivation of NR.18 We speculate that the inhibited NR activity might result from the feedback repression by nitrite, and it may be modulated by a post-translational modification as the transcript levels of NR genes, NIA1 and NIA2 do not change during LPS induction. Recently, generation of NO can be observed in mitochondria induced by bacterial protein harpin, which support the model of cross talk between NO and mitochondria in the activation of stress-related responses in plants.¹⁹ NO is required for TMV-induced mitochondrial AOX induction and promotes systemic basal defense against TMV.²⁰ AOX helps to maintain the electron flux and to reduce mitochondrial ROS levels,^{21,22} which is often induced by various treatment.^{23,24} Our data have determined the involvement of LPS-elicited NO in mediating the expression of the AOX gene and the upregulation of antioxidant enzyme activities, which might then modulate mitochondrial (or cellular) redox. Thus, it is important to clarify the function of mitochondria cross-talk with NO on LPS-induced defense activation, and requires further attention. In addition, the translocation of NPR1 into the nucleus is promoted by NO during LPS induction, which is consistent with previous study that the nuclear translocation of NPR1 is induced by GSNO/ NO.10 Although GSNO/NO-mediated S-nitrosylation of NPR1 facilitates its oligomerization, it is suggested that this S-nitrosylation-mediated oligomerization is not seen as an inhibitory effect of NPR1 signaling but rather as a step prior to monomer accumulation.¹⁰ We did not show evidence how the NPR1 activity was regulated by NO. Further studies to elucidate the more comprehensive mechanism are warranted.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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References

- Jones JD, Dangl JL. The plant immune system. Nature 2006; 444:323-9; PMID:17108957; http:// dx.doi.org/10.1038/nature05286.
- Braun SG, Meyer A, Holst O, Pühler A, Niehaus K. Characterization of the Xanthomonas campestris pv. campestris lipopolysaccharide substructures essential for elicitation of an oxidative burst in tobacco cells. Mol Plant Microbe Interact 2005; 18:674-81; PMID:16042013; http://dx.doi.org/10.1094/ MPMI-18-0674.
- Desaki Y, Miya A, Venkatesh B, Tsuyumu S, Yamane H, Kaku H, et al. Bacterial lipopolysaccharides induce defense responses associated with programmed cell death in rice cells. Plant Cell Physiol 2006; 47:1530-40; PMID:17018557; http://dx.doi. org/10.1093/pcp/pcl019.
- Newman MA, von Roepenack-Lahaye E, Parr A, Daniels MJ, Dow JM. Prior exposure to lipopolysaccharide potentiates expression of plant defenses in response to bacteria. Plant J 2002; 29:487-95; PMID:11846881; http://dx.doi.org/10.1046/ j.0960-7412.2001.00233.x.
- Delledonne M. NO news is good news for plants. Curr Opin Plant Biol 2005; 8:390-6; PMID:15922651; http://dx.doi.org/10.1016/j. pbi.2005.05.002.

- Ye Y, Li Z, Xing D. Nitric oxide promotes MPK6mediated caspase-3-like activation in cadmiuminduced Arabidopsis thaliana programmed cell death. Plant Cell Environ 2012; In press; PMID:22621159; http://dx.doi.org/10.1111/j.1365-3040.2012.02543.x.
- Melotto M, Underwood W, Koczan J, Nomura K, He SY. Plant stomata function in innate immunity against bacterial invasion. Cell 2006; 126:969-80; PMID:16959575; http://dx.doi.org/10.1016/j. cell.2006.06.054.
- Neill S, Bright J, Desikan R, Hancock J, Harrison J, Wilson I. Nitric oxide evolution and perception. J Exp Bot 2008; 59:25-35; PMID:17975211; http:// dx.doi.org/10.1093/jxb/erm218.
- D. Durner J, Wendehenne D, Klessig DF. Defense gene induction in tobacco by nitric oxide, cyclic GMP, and cyclic ADP-ribose. Proc Natl Acad Sci USA 1998; 95:10328-33; PMID:9707647; http://dx.doi. org/10.1073/pnas.95.17.10328.
- Corpas FJ, Palma JM, del Río LA, Barroso JB. Evidence supporting the existence of L-arginine-dependent nitric oxide synthase activity in plants. New Phytol 2009; 184:9-14; PMID:19659743; http://dx.doi.org/10.1111/j.1469-8137.2009.02989.x.
- Lindermayr C, Sell S, Müller B, Leister D, Durner J. Redox regulation of the NPR1-TGA1 system of *Arabidopsis thaliana* by nitric oxide. Plant Cell 2010; 22:2894-907; PMID:20716698; http://dx.doi. org/10.1105/tpc.109.066464.
- Zhang Y, Fan W, Kinkema M, Li X, Dong X. Interaction of NPR1 with basic leucine zipper protein transcription factors that bind sequences required for salicylic acid induction of the *PR-1* gene. Proc Natl Acad Sci USA 1999; 96:6523-8; PMID:10339621; http://dx.doi.org/10.1073/ pnas.96.11.6523.
- Zeidler D, Zähringer U, Gerber I, Dubery I, Hartung T, Bors W, et al. Innate immunity in Arabidopsis thaliana: lipopolysaccharides activate nitric oxide synthase (NOS) and induce defense genes. Proc Natl Acad Sci USA 2004; 101:15811-6; PMID:15498873; http://dx.doi.org/10.1073/ pnas.0404536101.
- Sun A, Nie S, Xing D. Nitric oxide-mediated maintenance of redox homeostasis contributes to NPR1dependent plant innate immunity triggered by lipopolysaccharides. Plant Physiol 2012; 160:1081-96; PMID:22926319; http://dx.doi.org/10.1104/ pp.112.201798.
- Ton J, Mauch-Mani B. β-amino-butyric acidinduced resistance against necrotrophic pathogens is based on ABA-dependent priming for callose. Plant J 2004; 38:119-30; PMID:15053765; http:// dx.doi.org/10.1111/j.1365-313X.2004.02028.x.
- van Hulten M, Pelser M, van Loon LC, Pieterse CMJ, Ton J. Costs and benefits of priming for defense in *Arabidopsis*. Proc Natl Acad Sci USA 2006; 103:5602-7; PMID:16565218; http://dx.doi. org/10.1073/pnas.0510213103.
- Ahn IP, Kim S, Lee YH, Suh SC. Vitamin B1-induced priming is dependent on hydrogen peroxide and the NPR1 gene in Arabidopsis. Plant Physiol 2007; 143:838-48; PMID:17158583; http://dx.doi.org/10.1104/pp.106.092627.
- Lillo C, Meyer C, Lea US, Provan F, Oltedal S. Mechanism and importance of post-translational regulation of nitrate reductase. J Exp Bot 2004; 55:1275-82; PMID:15107452; http://dx.doi. org/10.1093/jxb/erh132.
- Palmieri MC, Lindermayr C, Bauwe H, Steinhauser C, Durner J. Regulation of plant glycine decarboxylase by s-nitrosylation and glutathionylation. Plant Physiol 2010; 152:1514-28; PMID:20089767; http://dx.doi.org/10.1104/pp.109.152579.

- Fu LJ, Shi K, Gu M, Zhou YH, Dong DK, Liang WS, et al. Systemic induction and role of mitochondrial alternative oxidase and nitric oxide in a compatible tomato-*Tobacco mosaic virus* interaction. Mol Plant Microbe Interact 2010; 23:39-48; PMID:19958137; http://dx.doi.org/10.1094/ MPMI-23-1-0039.
- Li Z, Xing D. Mechanistic study of mitochondria-dependent programmed cell death induced by aluminium phytotoxicity using fluorescence techniques. J Exp Bot 2011; 62:331-43; PMID:20937730; http://dx.doi.org/10.1093/jxb/ erq279.
- Umbach AL, Fiorani F, Siedow JN. Characterization of transformed Arabidopsis with altered alternative oxidase levels and analysis of effects on reactive oxygen species in tissue. Plant Physiol 2005; 139:1806-20; PMID:16299171; http://dx.doi.org/10.1104/ pp.105.070763.
- Zhang LR, Xing D. Methyl jasmonate induces production of reactive oxygen species and alterations in mitochondrial dynamics that precede photosynthetic dysfunction and subsequent cell death. Plant Cell Physiol 2008; 49:1092-111; PMID:18535010; http://dx.doi.org/10.1093/pcp/pcn086.
- 24. Li Z, Yue H, Xing D. MAP Kinase 6-mediated activation of vacuolar processing enzyme modulates heat shock-induced programmed cell death in Arabidopsis. New Phytol 2012; 195:85-96; PMID:22497243; http://dx.doi.org/10.1111/ j.1469-8137.2012.04131.x.