

Nutritional Analysis of Cultivated Mushrooms in Bangladesh - *Pleurotus ostreatus*, *Pleurotus sajor-caju*, *Pleurotus florida* and *Calocybe indica*

Nuhu Alam^{1,2}, Ruhul Amin³, Asaduzzaman Khan⁴, Ismot Ara⁵, Mi Ja Shim⁶, Min Woong Lee⁷ and Tae Soo Lee^{1*}

¹Department of Biology, University of Incheon, Incheon-402-749, Korea

²Department of Botany, Jahangirnagar University, Savar, Dhaka-1342 Bangladesh

³National Mushroom Development and Extension Centre, Savar, Dhaka, Bangladesh

⁴Department of Biochemistry and Molecular Biology, Jahangirnagar University

⁵Department of Zoology, Jahangirnagar University

⁶Department of Life Science, University of Seoul, Seoul 130-743, Korea

⁷Department of Biology, Dongguk University, Seoul 100-715, Korea

(Received December 25, 2008. Accepted December 17, 2008)

Mushroom cultivation has been started recently in Bangladesh. Awareness of the nutritional and medicinal importance of mushrooms is not extensive. In this study, the nutritional values of dietary mushrooms- *Pleurotus ostreatus*, *Pleurotus sajor-caju*, *Pleurotus florida* and *Calocybe indica* that are very popular among the cultivated mushrooms in Bangladesh have been determined. These mushrooms were rich in proteins (20–25%) and fibers (13–24% in dry samples) and contained a lower amount of lipid (4 to 5%). The carbohydrate contents ranged from 37 to 48% (on the basis of dry weight). These were also rich in mineral contents (total ash content is 8–13%). The pileus and gills were protein and lipid rich and stripe was carbohydrate and fiber-rich. The moisture content of mushrooms ranged from 86 to 87.5%. Data of this study suggest that mushrooms are rich in nutritional value.

KEYWORDS : Carbohydrate, Lipid, Minerals, Oyster and milky mushroom, Protein

Mushroom have been a widely used as food and food supplements for millennia. It is an important food item concerning human health, nutrition and disease prevention (Chang, 1996). There is a common saying that “medicines and foods have a common origin” (Kaul, 2001). Dietary mushrooms provide a wide variety of medicinal properties and they are effective against certain life-threatening diseases. Major medicinal properties attributed to mushrooms include anticancer, antibiotic, antiviral activities, immunity and blood lipid lowering effects. *Pleurotus* spp. are also rich in medicinal values. *Pleurotus florida* has antioxidant and antitumor activities (Nayana and Janardhanan, 2000; Manpreet *et al.*, 2004), *Pleurotus sajor-caju* has hypertensive effects through its active ingredients which affect the renin-angiotensin system (Chang, 1996), *P. ostreatus* possesses antitumor activity (Yoshioka *et al.*, 1985) and hypoglycaemic effects in experimentally diabetic induced rats (Chorvathova *et al.*, 1993). Oyster mushrooms are very effective in reducing the total plasma cholesterol and triglyceride level (Nuhu Alam *et al.*, 2007) and thus reduce the chance of atherosclerosis and other cardiovascular and artery related disorders. These medicinal properties might be due to the presence of some important substance in dietary mushrooms. Mushrooms are rich in protein, minerals and vita-

mins and they contain an abundance of essential amino acids (Sadler, 2003). Nutritional analysis of several mushroom species of different origins had been carried out in many laboratories in the world. But nutritional values of locally cultivated mushrooms remain speculative. Moreover, nutritional composition is affected by many factors; these include differences among strains, the composition of growth substrate, the method of cultivation, stage of harvesting, specific portion of the fruiting bodies used for analysis (Benjamin, 1995).

Generally, people in Bangladesh are still not very aware of nutritional and medicinal importance of mushrooms. The history of mushroom cultivation is very recent in Bangladesh. Only some species of mushrooms are now cultivated in this country and among these *Pleurotus ostreatus*, *P. sajor-caju*, *P. florida* and *Calocybe indica* are popular and widely accepted (Ruhul Amin *et al.*, 2007). The aim of this investigation was to analyze the nutritional values of these mushrooms cultivated in Bangladesh, with a goal of increasing awareness of the beneficial effects of edible mushrooms among the consumers.

Materials and Methods

This study was carried out ‘Quality Control and Quality Assurance’ laboratory of National Mushroom Development and Extension Centre (NAMDEC), Savar, Dhaka,

*Corresponding author <E-mail : tslee@incheon.ac.kr>

Bangladesh. *Pleurotus ostreatus*, *Pleurotus sajor-caju*, *Pleurotus florida* and *Calocybe indica* mushrooms were cultivated and harvested in the laboratory of NAMDEC.

Moisture analysis. Twenty gram of fresh mushroom was weighed into a weighed moisture box (A&D company Ltd. N 92; P1011656; Japan) and dried in an oven at 100–105°C and cooled in a dessicator. The process of heating and cooling was repeated till a constant weight was achieved.

The moisture content was calculated as following equation:

$$\text{Moisture (\%)} = (\text{initial weight} - \text{final weight}) \times 100 / \text{weight of sample (Raghuramulu et al., 2003)}.$$

Determination of total protein. Five gram of grinded mushroom was taken with 50 ml of 0.1 N NaOH and boiled for 30 min. The solution was cooled in room temperature and centrifuged at 1000 × g by a DSC-200T tabletop centrifuge (Digisystem Laboratory Instruments, Taipei, Taiwan). The supernatant was collected and total protein content was measured according to the method of Lowry et al. (1951). For the determination of protein content from fresh mushroom, 5 g was taken with 50 ml phosphate buffer and homogenized with a tissue homogenizer (Polytron, Lucerne, Switzerland). Five milliliter of homogenized was taken with 50 ml of 0.1 N NaOH and protein content was determined as mentioned above.

Determination of total lipid. Total lipid was determined by slight modified method of Folch et al. (1957). Five gram of grinded mushroom was suspended in 50 ml of chloroform : methanol (2 : 1 v/v) mixture then mixed thoroughly and let stand for 3 days. The solution was filtered and centrifuged at 1000 g by a table centrifuge machine. The upper layer of methanol was removed by Pasteur pipette and chloroform was evaporated by heating. The remaining was the crude lipid. For the determination of total lipid from fresh mushroom, 5 g was taken with 50 ml phosphate buffer and homogenized with a tissue homogenizer. Five ml of homogenized was taken with 50 ml of chloroform : methanol (2 : 1 v/v) mixture and lipid content was determined as mentioned above.

Determination of crude fiber. Ten grams of moisture and fat-free sample was taken in a beaker and 200 ml of boiling 0.255 N H₂SO₄ was added. The mixture was boiled for 30 minutes keeping the volume constant by the addition of water at frequent intervals. The mixture was then filtered through a muslin cloth and the residue washed with hot water till free from acid. The material was then transferred to the same beaker, and 200 ml of boiling 0.313 N NaOH added. After boiling for 30 minutes

(keeping the volume constant as before) the mixture was filtered through a muslin cloth and the residue washed with hot water till free from alkali, followed by washing with some alcohol and ether. It was then transferred to a crucible, dried overnight at 80–100°C and weighed (We) in an electric balance (Keyi: JY-2003; China). The crucible was heated in a muffle furnace (Nebetherm: Mod-L9/11/c6; Germany) at 600°C for 5–6 hours, cooled and weighed again (Wa). The difference in the weights (We-Wa) represents the weight of crude fiber.

$$\text{Crude fiber (g/100 g sample)} = [100 - (\text{moisture} + \text{fat})] \times (\text{We-Wa}) / \text{Wt of sample (Raghuramulu et al., 2003)}.$$

Determination of total ash. One gram of the sample was weighed accurately into a crucible. The crucible was placed on a clay pipe triangle and heated first over a low flame till all the material was completely charred, followed by heating in a muffle furnace for about 5–6 hours at 600°C. It was then cooled in a dessicator and weighed. To ensure completion of ashing, the crucible was then heated in the muffle furnace for 1 h, cooled and weighed. This was repeated till two consecutive weights were the same and the ash was almost white or grayish white in color. Then total ash was calculated as:

$$\text{Ash content (g/100 g sample)} = \text{weight of ash} \times 100 / \text{weight of sample taken (Raghuramulu et al., 2003)}.$$

Total carbohydrate estimation. The content of the available carbohydrate was determined by the following equation:

$$\text{Carbohydrate (g/100 g sample)} = 100 - [(\text{moisture} + \text{fat} + \text{protein} + \text{ash} + \text{crude fiber}) \text{ g/100 g}] \text{ (Raghuramulu et al., 2003)}.$$

Mineral analysis. Total ash was taken for the analysis of mineral contents. Two ml of conc. HNO₃ was added to the ash and heated for 2 minutes. One drop of hydrogen peroxide was added into the solution. The solution was then transferred into a volumetric flask and total volume was made 50 ml by adding deionized distilled water. This was then used to analyze the contents of calcium (Ca), iron (Fe), manganese (Mn), magnesium (mg), zinc (Zn), Selenium (Se) and arsenic (As) by flame and graphite method with atomic absorption spectrophotometer (Perkin Elmer: AS 80).

Results and Discussion

Several nutritional parameters were measured for both fresh mushrooms (Table 1) and dry mushrooms (Table 2). The moisture contents of *P. ostreatus*, *P. sajor-caju*, *P. florida* and *C. indica* were found about 86, 87, 87.5 and

Table 1. Nutrient contents of fresh mushrooms (g/100 g)

Mushroom species	Moisture (%)	Protein	Lipid	Fiber	Ash	Carbohydrate
<i>P. ostreatus</i>	86.0 ± 0.2	3.4 ± 0.4 ^a	0.68 ± 0.05 ^a	3.4 ± 0.2 ^a	1.18 ± 0.1 ^{a,b}	5.1 ± 0.25 ^a
<i>P. sajor-caju</i>	87.0 ± 0.4	3.26 ± 0.33 ^a	0.57 ± 0.05 ^b	2.97 ± 0.17 ^b	1.1 ± 0.1 ^a	5.09 ± 0.19 ^a
<i>P. florida</i>	87.5 ± 0.35	2.6 ± 0.13 ^b	0.54 ± 0.07 ^b	3.0 ± 0.12 ^b	1.13 ± 0.07 ^a	5.24 ± 0.4 ^a
<i>C. indica</i>	87.4 ± 0.55	2.75 ± 0.2 ^b	0.65 ± 0.06 ^a	1.63 ± 0.2 ^c	1.28 ± 0.06 ^b	6.8 ± 0.5 ^b

Results show mean ± SEM of 5 trials. Values in the same column that do not share a common superscript are significantly different at $P < 0.05$ (one way ANOVA then LSD post hoc comparison).

Table 2. Nutrient contents of dried mushrooms (g/100 g)

Mushroom species	Protein	Lipid	Fiber	Ash	Carbohydrate
<i>P. ostreatus</i>	23.91 ± 2.0 ^a	4.6 ± 0.26 ^{a,b}	24.34 ± 1.8 ^a	9.36 ± 0.5 ^a	37.8 ± 2.5 ^a
<i>P. sajor-caju</i>	24.63 ± 1.51 ^a	4.41 ± 0.2 ^a	22.87 ± 0.8 ^a	8.28 ± 0.29 ^b	39.82 ± 1.73 ^{a,b}
<i>P. florida</i>	20.56 ± 1.45 ^b	4.30 ± 0.29 ^a	23.29 ± 1.3 ^a	9.02 ± 0.48 ^{a,b}	42.83 ± 2.54 ^b
<i>C. indica</i>	21.4 ± 1.86 ^{a,b}	4.95 ± 0.4 ^b	12.9 ± 2.5 ^b	13.1 ± 0.45 ^c	48.5 ± 2.4 ^c

Results show mean ± SEM of 5 trials. Values in the same column that do not share a common superscript are significantly different at $P < 0.05$ (one way ANOVA then LSD post hoc comparison).

Table 3. Mineral contents of dried mushrooms (mg/100 g)

Elements	<i>P. ostreatus</i>	<i>P. sajor-caju</i>	<i>P. florida</i>	<i>C. indica</i>
Calcium (Ca)	35.9 ± 3.8 ^a	22.15 ± 2.3 ^b	33.7 ± 1.9 ^a	20.65 ± 2.1 ^b
Iron (Fe)	55.45 ± 5.2 ^a	33.45 ± 3.8 ^b	43.2 ± 4.0 ^c	56.25 ± 4.1 ^a
Zinc (Zn)	26.565 ± 2.2 ^a	20.9 ± 1.5 ^b	16 ± 0.9 ^c	12.865 ± 1.9 ^d
Magnesium (Mg)	16.395 ± 2.1 ^a	20.22 ± 1.2 ^b	13.4 ± 3.1 ^c	12.825 ± 2.1 ^c
Manganese (Mn)	2.85 ± 0.9 ^a	2.87 ± 0.5 ^a	2.7 ± 0.3 ^a	1.64 ± 0.4 ^b
Selenium (Se)	0.011 ± 0.002 ^a	0.025 ± 0.004 ^b	0.0132 ± 0.003 ^a	0.0132 ± 0.001 ^a
Arsenic (As)	0.1 ± 0.002 ^a	0.095 ± 0.02 ^a	0.083 ± 0.009 ^a	0.54 ± 0.004 ^b

Results show mean ± SEM of 3 trials. Values in the same row that do not share a common superscript are significantly different at $P < 0.05$ (one way ANOVA then LSD post hoc comparison).

87.4% respectively. Hundred grams of fresh *P. ostreatus* contains 3~3.8 g of proteins, 0.63~0.73 g of lipids, 3.2~3.6 g of fiber and 5.0~5.4 g of carbohydrates. One hundred grams of fresh *P. sajor-caju* contained 3~3.6 g of proteins, 0.52~0.62 g of lipids, 2.8~3.1 g of fiber and 5.0~5.3 g of carbohydrates. In case of fresh *P. florida* these were as follows: 2.5~2.75 g of proteins, 0.5~0.6 g of lipids, 2.9~3.1 g of fiber and 5.0~5.6 g of carbohydrates and 100 g of *C. indica* contained 2.6~2.9 g of proteins, 0.6~0.7 g of lipids, 1.5~1.8 g of fiber and 6.3~7.3 g of carbohydrates (Table 1). The protein, lipid, fiber and carbohydrate contents in 100 g of dried *P. ostreatus* were found as 22~26 g, 4.4~4.8 g, 23~26 g and 35~40 g respectively. These were found as 23~26 g, 4.2~4.6 g, 22~23.6 g and 38~41.5 g respectively in *P. sajor-caju*. 100 g of dried *P. florida* contained 19~22 g of proteins, 4~4.6 g of lipids, 22~24.6 g of fiber and 40~45 g of carbohydrates and 100 g of dried *C. indica* contained 20~23 g of proteins, 4.6~5.3 g of lipids, 11~15 g of fiber and 46~51 g of carbohydrates (Table 2). The protein content of *P. ostreatus* and *P. sajor-caju* were found 15~20% greater than that of *P. florida* and *C. indica*. The total fat content was greater in *C. indica* which is significant to *P. sajor-caju* and *P.*

florida. *C. indica* was also significantly richer in carbohydrates than the three species of *Pleurotus*. On the other hand the fiber content in *C. indica* is significantly lower (about 40~50%) than that in *Pleurotus* spp.

Mushrooms are also rich in mineral contents. The total ash content found in *Pleurotus ostreatus*, *P. sajor-caju*, *P. florida* and *C. indica* were 1.1~1.3 g, 1~1.2 g, 1.1~1.2 g and 1.2~1.4 respectively. In case of dry mushrooms these were 9~10 g, 8~8.6 g, 8.6~9.5 g and 12.5~13.5 g respectively (Tables 1, 2).

According to Breene (1990) and Ço_kuner and Özdemir (2000), protein contents of mushrooms range from 19 to 39 g in 100 g dried matter. In our study we found the protein values (g/100 g dried matter) as 23.91 g in *P. ostreatus*, 24.63 g in *P. sajor-caju*, 20.56 g in *P. florida* and 21.4 g in *C. indica*. Lipid value was 4.3~4.9 g in 100 g in dry matter of four cultivated mushroom. These results are in conformity with Shin *et al.* (2007). 34.8% dietary fibre value was found in oyster mushrooms by Justo *et al.* (1999), which are higher than the values, we obtained in our study. Watanabe *et al.* (1994) found the carbohydrate value as 47.9 g in 100 g dry matter. Our carbohydrate values are almost similar the study made by Watanabe *et al.*

(1994).

Table 3 shows the contents of some important minerals. Hundred grams of dried *P. ostreatus* contained Ca (35.9 mg), Fe (55.5 mg), Mg (16.4 mg), Mn (2.9 mg), Zn (26.6 mg), Se (11 µg) and As (100 µg). 100 g of dried *P. sajor-caju* contained Ca (22.15 mg), Fe (33.45 mg), Mg (20.22 mg), Mn (2.87 mg), Zn (20.9 mg), Se (25 µg) and As (95 µg). In case of 100 g *P. florida* these were as follows: Ca (33.7 mg), Fe (43.2 mg), Mg (13.4 mg), Mn (2.7 mg), Zn (16 mg), Se (13.2 µg) and As (83 µg) and 100 g *C. indica* contained Ca (20.7 mg), Fe (56.2 mg), Mg (12.8 mg), Mn (1.65 mg), Zn (12.9 mg), Se (13.2 µg) and As (54 µg). The difference of total mineral (ash) content among the *C. indica* and *Pleurotus* spp is significant. The findings in this study are comparable to previous studies (Crisan and Sands, 1978; Chang, 1980; Bano and Rajarathnam, 1982; Hong *et al.*, 2007; Shin *et al.*, 2007; Dundar *et al.*, 2008).

Figures 1~4 show the variation of nutritional parameters among the different parts of mushrooms. The pileus and gills are richer in protein (about 40~60%), lipid (30~

60%) and ash content (5~10%) than stipe. On the other hand the stipe is richer in fiber (40~50%) and carbohydrate content (10~15%). The variation in protein, fat and fiber content is significant with the exception of *C. indica* in which protein variation is not significant. This result is agreeable with data collected from the previous study (Watanabe *et al.*, 1994; Justo *et al.*, 1999; Shin *et al.*, 2007; Dundar *et al.*, 2008).

In conclusion, the chemical composition of edible mushrooms determines their nutritional value and sensory properties as also mentioned by other authors (Shah *et al.*, 1997; Manzi *et al.*, 2001). They differ according to species but this difference also depends on the substratum, atmospheric conditions, age and part of the fructification. We found different nutritional values in the different part of cultivated mushrooms. These data suggest that dietary mushrooms cultivated in Bangladesh are good source of nutrients specially protein and fiber. Mushrooms are rich in protein, edible fiber and minerals but lipid content is low. These results also indicate that the studied mushrooms have good nutritive value for human. Protein is an

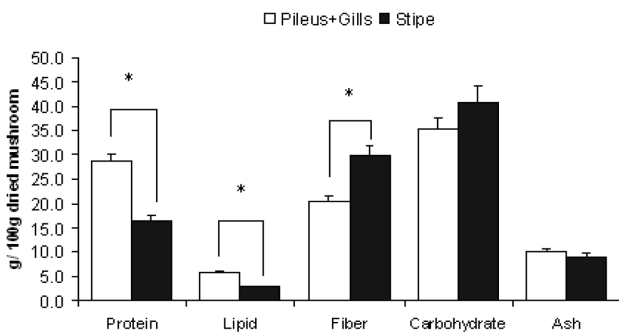


Fig. 1. Nutritional variation in different parts of *Pleurotus ostreatus*. Bars show mean + SEM of 5 trials. Data was analyzed by one way ANOVA and then post hoc LSD test. * indicates that difference is significant at $P \leq 0.05$ level.

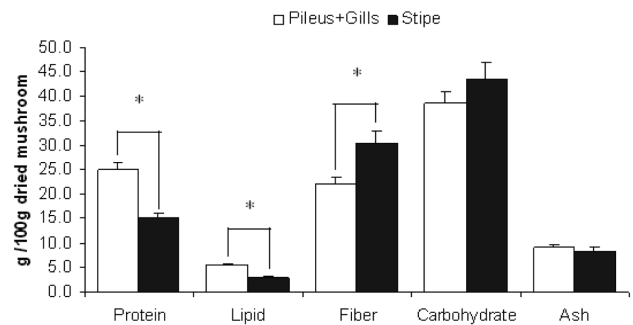


Fig. 3. Nutritional variation in different parts of *Pleurotus florida*. Bars show mean + SEM of 5 trials. Data was analyzed by one way ANOVA and then post hoc LSD test. * indicates that difference is significant at $P \leq 0.05$ level.

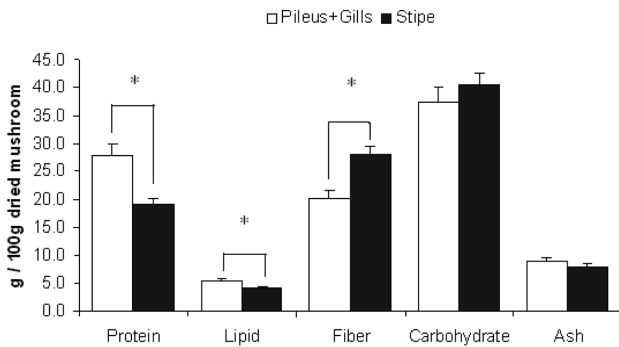


Fig. 2. Nutritional variation in different parts of *Pleurotus sajor-caju*. Bars show mean + SEM of 5 trials. Data was analyzed by one way ANOVA and then post hoc LSD test. * indicates that difference is significant at $P \leq 0.05$ level.

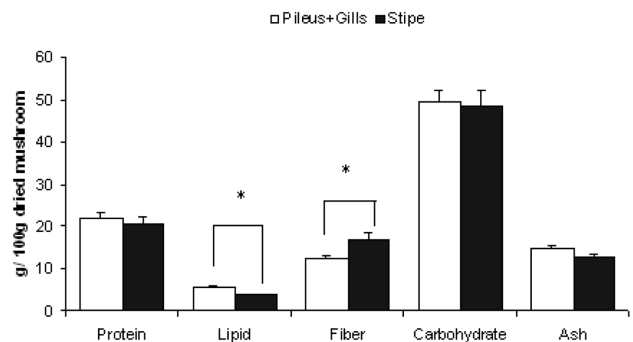


Fig. 4. Nutritional variation in different parts of *Calocybe indica*. Bars show mean + SEM of 5 trials. Data was analyzed by one way ANOVA and then post hoc LSD test. * indicates that difference is significant at $P \leq 0.05$ level.

important nutritional component and protein deficiency is the world's most serious human nutritional problem, especially in third world countries like Bangladesh. So mushroom is a promising food that may overcome protein-energy malnutrition problem in the third world.

References

- Bano, Z. and Rajarathnam, S. 1982. Pleurotus mushroom as a nutritious food. *Tropical Mushrooms-Biological Nature and Cultivated Methods*, pp. 362-363. The Chinese University Press, Hong Kong.
- Benjamin, D. R. 1995. Mushroom, Poisons and Panaceas, pp. 151-165. W. H. Freeman & Company, New York, USA.
- Breene, W. M. 1990. Nutritional and medicinal value of specialty mushrooms. *J. Food Protect.* 53:883-894.
- Chang, R. 1996. Functional properties of mushrooms. *Nutr. Rev.* 54:91-93.
- Chang, S. T. 1980. Mushrooms as human food. *Bioscience.* 30: 399-400.
- Chorvathoba, V., Bobek, P., Ginter, E. and Klavanova, J. 1993. Effect of the oyster fungus on glycemia and cholesterolemia in rats with insulin depended diabetes. *Physiol. Res.* 42:175-179.
- Ço_kuner, Y. and Özdemiir, Y. 2000. Acid and EDTA blanching effects on the essential element content of mushrooms (*Agaricus bisporus*). *J. Sci. Food Agric.* 80:2074-2076.
- Crisan, E. V. and Sands, A. 1978. Nutritional value. In: S. T. Chang and W. A. Hayes. *The biology and cultivation of edible mushrooms*, pp. 137-165. New York Academic Press, U.S.A.
- Dundar, A., Hilal Acay and Yildiz, A. 2008. Yield performance and nutritional contents of three oyster mushroom species cultivated on wheat stalk. *Afr. J. Biotechy* 7:3497-3501.
- Folch, J., Lees, M. and Sloane-Stanely, G. H. 1957. A simple method for the isolation and purification of total lipids from animal tissues. *J. Biol. Chem.* 226:497-509.
- Hong, I. P., Nam, S. H., Sung, G. B., Chung, I. M., Hur, H., Lee, M. W., Kim, M. K. and Guo, S. X. 2007. Chemical components of *Paecilomyces tenuipes* (Peck) Samson. *Mycobiology* 35:215-218.
- Justo, M. B., Guzman, G. A., Mejia, E. G., Diaz, C. L. G., Martinez, G. and Corona, E. B. 1999. Calidad proteínica de tres cepas mexicanas de setas (*Pleurotus ostreatus*). *Archivos Latinoamericanos de Nutricion.* 49:81-85.
- Kaul, T. N. 2001. *Biology and Conservation of Mushrooms*, pp. 117-145. Oxford & IBH Publishing Co. Pvt. Ltd. New Delhi, India.
- Lowry, O. H., Rosebrough, N. J., Farr, A. L. and Randall, R. J. 1951. Protein measurement with the folin phenol reagent. *J. Biol. Chem.* 193:265-275.
- Manpreet, K., Giridhar, S. and Khanna, P. K. 2004. *In vitro* and *in vivo* antioxidant potentials of *Pleurotus florida* in experimental animals. *Mushroom Res.* 13:21-26.
- Manzi, P., Aguzzi, A. and Pizzoferrato, L. 2001. Nutritional value of mushrooms widely consumed in Italy. *Food Chem.* 73:321-325.
- Nayana, J. and Janardhanan, K. K. 2000. Antioxidant and antitumor activity of *Pleurotus florida*. *Curr. Sci.* 79:941-943.
- Nuhu Alam, Shahdat Hossain, Abul Khair, Ruhul Amin and Asaduzzaman, K. 2007. Comparative effects of oyster mushrooms on plasma lipid profile of hypercholesterolaemic rats. *Bangladesh J. Mushroom.* 1:15-22.
- Raghuramulu, N., Madhavan, N. K. and Kalyanasundaram, S. 2003. *A Manual of Laboratory Techniques*, pp. 56-58. National Institute of Nutrition. Indian Council of Medical Research, Hyderabad, India.
- Ruhul Amin, Nirod, C. S., Mahbuba, M., Jebunnahar, K. and Mahfuzur Rahman. 2007. Officer's Training Manual, pp. 13-17. National Mushroom Development and Extension Centre, Savar, Dhaka, Bangladesh.
- Sadler, M. 2003. Nutritional properties of edible fungi. *Br. Nutr. Found. Nutr. Bull.* 28:305-308.
- Shah, H., Iqtidar, A. K. and Shagufta, J. 1997. Nutritional composition and protein quality of *Pleurotus* mushroom. *Sarhad J. Agric.* 13:621-626.
- Shin, C. K., Yee, C. F., Lee, J. S. and Atong, M. 2007. Nutritional properties of some edible wild mushrooms in Sabah. *J. Appl. Sci.* 7:2216-2221.
- Watanabe, T., Tsuchihashi, N., Takai, Y., Tanaka, K. and Suzuki, A. 1994. Effects of ozone exposure during cultivation of oyster mushroom (*Pleurotus ostreatus*) on chemical components of the fruit bodies. *J. Jpn. Soc. Food Sci. Technol.* 41:705-708.
- Yoshioka, Y., Tabeta, R., Saito, H., Uehara, N. and Fukoaka, F. 1985. Antitumor polysaccharides from *P. ostreatus* (Fr.) Quel. isolation and structure of a β -glucan. *Carbohydrate Res.* 140: 93-100.